Short communication: Differential immunoglobulin transfer during mastitis challenge by pathogen-specific components

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ABSTRACT

Mastitis induced by Escherichia coli is often characterized by severe clinical signs, indicating a more powerful combat of the immune system against the pathogen compared with Staphylococcus aureus infections, which are often represented by chronic and subclinical diseases. The aim of this study was to test the major pathogenic component lipopolysaccharide (LPS) from E. coli and lipoteichoic acid (LTA) from Staph. aureus for their effects on blood-milk barrier integrity and the related transfer of immunoglobulins and lactate from blood into milk. A similar somatic cell count (SCC) increase was achieved by intramammary challenge of 1 quarter of 5 cows with 20 μg of LTA, and 8 cows with 0.2 μg of LPS (maximum log SCC/mL: 7). Milk IgG1 concentrations increased in LPS- but not in LTA-challenged quarters. Milk IgG2 concentrations were increased in treated quarters at 3 h after LPS, and 6 h after LTA challenge. Higher maximum levels of IgG2 were reached in milk of LPS-treated quarters (173 ± 58 μg/mL) than of LTA-challenged quarters (62 ± 13 μg/mL). Immunoglobulin G1 and IgG2 levels did not change in control quarters. l-Lactate concentrations in milk increased 4 h after LPS and 5 h after LTA challenge and reached higher maximum levels in LPS- (221 ± 48 mg/L) than in LTA-treated quarters (77 ± 18 mg/L). In conclusion, a mammary inflammation on a quantitatively similar level based on SCC increase achieves a more efficient transfer of blood components such as IgG2 via the blood-milk barrier if induced by LPS from E. coli than by LTA from Staph. aureus. This pathogen-specific difference may play an important role in the development of different mastitis severities.

Key words: mastitis, blood-milk barrier, lipoteichoic acid, lipopolysaccharide
L-Lactate (hereafter referred to as lactate) is another blood component that increases in milk during mastitis and is proposed to be used as an early indicator to detect mastitis (Davis et al., 2004). Leukocytes as a source of lactate in milk during an immune response have been considered (Davis et al., 2004). Recently, we described the blood as a major source of milk lactate that leaks into milk as a result of the impairment of the blood-milk barrier during the immune response (Lehm-ann et al., 2013). The aim of the current study was to investigate the transfer of immunoglobulins and lactate from blood into milk due to a change in the blood-milk barrier integrity after an intramammary challenge with LPS from *E. coli* and LTA from *Staph. aureus* with a comparable SCC increase in milk.

In 13 dairy cows, a similar SCC increase (maximum log SCC/mL: 7) was achieved by intramammary challenge of 1 quarter with 20 μg of LTA (n = 5) from a *Staph. aureus* strain that induced a chronic bovine mastitis, or with 0.2 μg of LPS (n = 8) from *E. coli* that induced acute bovine mastitis, as previously described (Figure 1; Wellnitz et al., 2011). In plasma (jugular vein) and milk samples (~10 mL) taken hourly from challenged and control quarters, IgG1 and IgG2 concentrations were analyzed using ELISA (bovine IgG1/IgG2 ELISA Quantitation Set; Bethyl Laboratories Inc., LuBioScience GmbH, Lucerne, Switzerland). The procedure was performed according to the manufacturer’s protocol. A blocking reagent consisting of fish gelatin [1 mL of fish skin gelatin (G7765; Sigma-Aldrich, Steinheim, Germany) in 20 mL of bidistilled water] was used to avoid matrix effects. Coefficients of variation, calculated using a control sample on each plate, were 10 and 20% within and between assays, respectively. Lactate concentrations were measured using the test kit Lactate PAP (bioMérieux, Marcy l’Étoile, France) with an automated analyzer (Cobas Mira; Roche Diagnostics International AG, Rotkreuz, Switzerland) according to the manufacturer’s instructions.

Data are presented as means ± standard error of the mean. Lactate concentrations are presented and statistically evaluated on a logarithmic scale (log10) to ensure normal distribution. Differences within treatment group to time point 0 and between-LPS and -LTA treatments within each time point (hourly) were tested for significance (*P* < 0.05) by ANOVA using PROC MIXED SAS (1999–2001, release 8.02; SAS Institute Inc., Cary, NC). The model included time, treatment, and their interaction as fixed effects, and quarter within cow as repeated subject. A Tukey-Kramer adjustment was used to compensate for multiple comparisons. The significant (*P* < 0.001) Pearson correlation coefficient (SigmaPlot v11; Systat Software Inc., Chicago, IL) between SCC and IgG1 and IgG2 in LTA-challenged quarters was 0.42 and 0.33, respectively, and 0.45 and 0.68 between SCC and IgG1 and IgG2 in LTA-challenged quarters, respectively.

In blood IgG1 and IgG2, concentrations were 16.5 ± 1.1 mg/mL and 35.4 ± 6.8 mg/mL, respectively, and did not change throughout the experiment. Although IgG concentrations in serum are known to be variable due to different factors such as age and lactational stage (Mallard et al., 1983), these are relatively high values compared with those in other studies where concentrations around 10 mg/mL were found for both immunoglobulins (Butler, 1983; Caffin and Poutrel, 1988). Reasons for that remain unclear. The test kits were validated according the manufacturer recommendations. Milk IgG1 concentrations (Figure 2A) were 68 ± 6, 63 ± 5, and 83 ± 12 μg/mL in control, LPS-, and LTA-challenged quarters before (0 h) challenge, respectively. In LPS-challenged quarters, IgG1 concentrations increased at 4 and 5 h and from 7 h after challenge until the end of the experiment. The maximum of 105 ± 13 μg/mL was reached 5 h after challenge. In control and LTA-challenged quarters, milk IgG1 concentrations did not significantly increase.

Milk IgG2 concentrations (Figure 2B) were 30 ± 6, 32 ± 8, and 23 ± 8 μg/mL, in control, LPS-, and LTA-challenged quarters before (0 h) challenge, respectively. Milk IgG2 concentrations increased at 3 h in LPS-challenged quarters, reached the maximum of 173 ± 58 μg/mL at 6 h after challenge, and stayed elevated until the end of the experiment. In LTA-challenged quarters, IgG2 was increased at 6 h, reached a maximum of 67 ± 9 μg/mL at 8 h, and stayed elevated until 11 h after challenge.

![Figure 1](https://example.com/figure1.png)  
*Figure 1. Milk SCC in LPS-challenged quarters (□; n = 8), in lipoteichoic acid (LTA)-challenged quarters (●; n = 5), and in control quarters (x; n = 13). Means without common letters (a–c) are significantly different between groups within a time point (*P* < 0.05). Data are presented as means ± SEM. Reproduced with permission from Wellnitz et al. (2011).*
Concentrations of IgG2 in milk raised to a greater extent (5 fold after LPS challenge) than IgG1 (3 fold after LPS challenge) although these molecules have comparable molecular weights of approximately 160 kDa (Butler, 1983) and should be able to pass the blood-milk barrier in a comparable way. The IgG1:IgG2 ratio in milk at 5 h after LPS challenge was 1:1.5, whereas the ratio in blood was 1:2.1. This was mainly due to the specific transport of IgG1 (Mayer et al., 2005) that was also responsible for a 2.5:1 ratio in milk before LPS challenge. In addition, specific transport of IgG2 from the blood is assumed (Guidry et al., 1980).

As IgG2 plays a particular role in the mammary immune defense, the availability of antibodies in the milk can be crucial for mastitis defense, specifically if antibodies against the invading mastitis pathogens are available in the blood. Although IgG2 also enters the mammary gland, passive transfer of IgG2 from the blood is assumed (Guidry et al., 1980).

Lactate is another blood component whose appearance in milk was tested with the intramammary LPS or LTA challenge. The blood lactate concentration was 54.9 ± 1.2 mg/L before LPS challenge and did not change throughout the experiment. Lactate concentrations in milk (Figure 3) were increased at 4 h after challenge in LPS-treated quarters and 5 h after challenge in LTA-treated quarters and both stayed elevated until the end of the experiment. At 4 h after challenge to 11 h after challenge, the lactate concentrations in milk were higher in LPS- than in LTA-treated quarters. This greater increase in lactate concentration in milk of quarters challenged with LPS compared with quarters challenged with LTA also shows a different characteristic in the change of blood-milk barrier permeability in response to E. coli compared with Staph. aureus endotoxin. The small molecular size (90 Da) of lactate facilitates transfer from blood into milk. However, milk concentration of lactate increased up to 4.0-fold higher levels of those in blood (220.7 ± 48.1 mg/L) at 8 h after LPS challenge and to 1.4 fold (76.8 ± 18.3 mg/L) 10 h after LTA challenge. This effect was seen before (Lehmann et al., 2013). This effect was suggested to be due to additional lactate production and release during anaerobic metabolism by milk and epithelial cells in the gland (Silanikove et al., 2011) or to be an effect of an
additional transporter system for lactate, which could be via aquaporins, as aquaporins transport lactate (Conde et al., 2010) and aquaporins are present in the bovine mammary gland (Mobasheri et al., 2011).

Mammary challenge equalized for SCC increases produces a greater and differential permeability of the blood-milk barrier for immunoglobulins when induced by LPS from E. coli compared with LTA from Staph. aureus. Thus, more efficient transfer of blood components such as IgG2 is achieved with Staph. aureus. Thus, more efficient transfer of blood components such as IgG2 is achieved with Staph. aureus compared with LTA from E. coli compared with LTA from E. coli.

Figure 3. Milk lactate concentrations in LPS-challenged quarters (□; n = 8), in lipoteichoic acid (LTA)-challenged quarters (●; n = 5), and in control quarters (x; n = 13). * indicates the first and subsequent time point with a significant (P < 0.05) difference compared with time 0; # indicates significant (P < 0.05) differences between LPS- and LTA-challenged quarters within a time point. Data are presented as means ± SEM.

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REFERENCES


