Occurrence and Genetic Characteristics of Third-Generation Cephalosporin-Resistant Escherichia coli in Swiss Retail Meat

Debora Vogt, Gudrun Overesch, Andrea Endimiani, Alexandra Collaud, Andreas Thomann, and Vincent Perreten

Prevalence and genetic relatedness were determined for third-generation cephalosporin-resistant Escherichia coli (3GC-R-Ec) detected in Swiss beef, veal, pork, and poultry retail meat. Samples from meat-packing plants (MPPs) processing 70% of the slaughtered animals in Switzerland were purchased at different intervals between April and June 2013 and analyzed. Sixty-nine 3GC-R-Ec isolates were obtained and characterized by microarray, PCR/DNA sequencing, Multi Locus Sequence Typing (MLST), and plasmid replicon typing. Plasmids of selected strains were transformed by electroporation into E. coli TOP10 cells and analyzed by plasmid MLST. The prevalence of 3GC-R-Ec was 73.3% in chicken and 2% in beef meat. No 3GC-R-Ec were found in pork and veal. Overall, the blaCTX-M-1 (79.4%), blaCMY-2 (17.6%), blaCMY-4 (1.5%), and blashv-12 (1.5%) β-lactamase genes were detected, as well as other genes conferring resistance to chloramphenicol (cmrlA1-like), sulfonamides (sul), tetracycline (tet), and trimethoprim (dfrA). The 3GC-R-Ec from chicken meat often harbored virulence genes associated with avian pathogens. Plasmid incompatibility (Inc) groups IncI1, IncFIB, IncFII, and IncB/O were the most frequent. A high rate of clonality (e.g., ST1304, ST38, and ST93) among isolates from the same MPPs suggests that strains persist at the plant and spread to meat at the carcass-processing stage. Additionally, the presence of the blaCTX-M-1 gene on an IncI1 plasmid sequence type 3 (IncI1/pST3) in genetically diverse strains indicates interstrain spread of an epidemic plasmid. The blaCMY-2 and blaCMY-4 genes were located on IncB/O plasmids. This study represents the first comprehensive assessment of 3GC-R-Ec in meat in Switzerland. It demonstrates the need for monitoring contaminants and for the adaptation of the Hazard Analysis and Critical Control Point concept to avoid the spread of multidrug-resistant bacteria through the food chain.

Introduction

Since the introduction of third-generation cephalosporins, a large number of extended-spectrum β-lactamase (ESBL), chromosomally mediated, and plasmid-mediated AmpC β-lactamase (cAmpC and pAmpC) producers have emerged in Gram-negative bacteria, particularly in Enterobacteriaceae such as Escherichia coli. ESBL enzymes belong to Ambler class A and are mainly represented by the SHV-, TEM-, and CTX-M-type families. The latter is the most frequent, with ~150 different types categorized into five groups. ESBLs hydrolyze penicillins, first- to fourth-generation cephalosporins and monobactams, but they are inhibited by the standard β-lactamase inhibitors (e.g., clavulanate and tazobactam). Plasmid-mediated AmpC β-lactamases belong to Ambler class C, hydrolyze penicillins and all cephalosporins (with the exception of the fourth-generation, e.g., cefepime), and are not inhibited by β-lactamase inhibitors. They are divided into six clusters, the most common being the CMY family.

E. coli is a normal inhabitant of the gut of humans and animals; however, certain E. coli have acquired different virulence factors causing host-specific infections. Extraintestinal pathogenic E. coli (ExPEC) are responsible for colibacillosis in various animal species, including poultry (avian pathogenic E. coli [APEC]), urinary tract infections (uropathogenic E. coli), and newborn meningitis (newborn meningitis-causing E. coli). Enterotoxigenic E. coli can cause diarrhea in calves, pigs, and humans. Enterohemorrhagic E. coli (EHEC) can be commensal or pathogenic in cattle and have been of great interest as meat and vegetable contaminants, as they cause severe illness in humans. E. coli are also responsible for wound infections, pneumonia, and septicemia in people.

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The emergence of third-generation cephalosporin-resistant *E. coli* (3GC-R-Ec) isolates is associated with prolonged hospitalization, higher health costs, and mortality.\textsuperscript{41} In addition, 3GC-R-Ec can carry other antibiotic resistances, such as those against aminoglycosides, chloramphenicol, trimethoprim, tetracycline, sulfonamides, and quinolones. These genes are frequently located on the same genetic elements (e.g., plasmids, integrons) as the *bla*\textsubscript{ESBL} and *bla*\textsubscript{pAmpC} genes. It has been shown that these genetic elements can be transferred from commensals to pathogens and vice versa.\textsuperscript{41,49} This phenomenon was well demonstrated by Leverstein-van Hall et al., who found that in 19% of isolates from human patients, the plasmids containing the *bla*\textsubscript{ESBL} were indistinguishable from those isolated from poultry meat.\textsuperscript{39,49}

Therefore, treating 3GC-R-Ec infections is becoming a challenge for clinicians and a growing concern for human health. Furthermore, the use of carbapenems to treat 3GC-R bacteria favors the selection of carbapenemase producers, which are able to hydrolyze carbapenems, leaving only colistin, fosfomycin, and tigecycline as last therapeutic options.\textsuperscript{44,49}

Since the beginning of the 21st century, ESBL- and pAmpC-producing isolates have been increasingly reported in livestock such as broilers, cattle, and pigs, but data about 3GC-R-Ec in meat are still scarce.\textsuperscript{5,22,49} This situation raised public health and food safety concerns about whether food-producing animals act as reservoirs for 3GC-R-Ec, which may contaminate meat and thus reach consumers, where 3GC-R-Ec could transfer genetic elements to human pathogenic *E. coli*.\textsuperscript{16,37,40,49}

In Switzerland, preliminary studies indicated a high prevalence of 3GC-R-*Enterobacteriaceae* in both indigenous and imported poultry meat.\textsuperscript{1,50} Another study recently showed a high rate of contamination of chicken meat received at a Swiss hospital kitchen.\textsuperscript{52} The aim of this study was to determine a more general and representative prevalence of 3GC-R-Ec in different lots of Swiss beef, pork, poultry, and veal retail meat and to determine their molecular characteristics. This study represents the first national surveillance of 3GC-R-Ec in meat, using a sampling plan based on representative production data from Swiss meat processing plants, and could therefore also serve as a basis for national monitoring of antibiotic resistance in fresh meat products at retail in Switzerland.

**Materials and Methods**

**Sampling**

Meat from meat-packing plants (MPPs) processing at least 70% of the slaughtered animals in Switzerland was analyzed in a time frame spanning from March 2013 to June 2013. This set encompasses a total of seven different MPPs, including two MPPs (MPPA and MPPB) processing solely poultry meat, four MPPs (MPPC\textsubscript{1}, MPPD, MPPE\textsubscript{1}, and MPPF\textsubscript{1}) processing beef, three MPPs (MPPC\textsubscript{2}, MPPE\textsubscript{2}, and MPPF\textsubscript{2}) processing pork, and three MPPs (MPPC\textsubscript{3}, MPPE\textsubscript{3}, and MPPG) processing veal. One type of meat among chicken breast, pork, beef, and veal escalope was analyzed every 4 weeks to avoid sampling the same batch twice. Twenty-five chicken meat samples selected in different stores selling products from MPPA (*n* = 37) and MPPB (*n* = 38) were purchased three times at 4-week intervals for a total of 75 chicken breast samples. Twenty-five pork, beef, and veal escalopes were purchased twice at 4-week intervals from different stores selling meat from MPPC (*n* = 44), MPPD (*n* = 12), MPPE (*n* = 54), MPPF (*n* = 28), and MPPG (*n* = 12), for a total of 50 samples each.

**Isolation and identification of 3GC-R-Ec**

Twenty-five grams of meat was homogenized in 25 ml of LB broth in a stomacher (Stomacher 400 Circulator, Seward) for 1 min and incubated overnight at 37°C with agitation. A loop full (10 µl) of this overnight culture was streaked onto MacConkey agar (Oxoid), Chrom ID\textsuperscript{®} ESBL agar (bioMérieux), and Brilliance\textsuperscript{TM} CRE agar (Oxoid) selective plates and incubated overnight at 37°C.

Lactose-positive colonies on MacConkey agar, reddish or white colonies on Chrom ID ESBL agar, and every colony growing on the Brilliance CRE agar plate were transferred onto Tryptone soy agar plates containing 5% sheep blood ( TSA-SB; Becton Dickinson) and incubated overnight at 37°C. The colonies were identified using a matrix-assisted laser desorption/ionization time-of-flight mass spectrometer (MALDI-TOF MS, microflex LT; Bruker Daltonics). Isolates growing on Brilliance CRE agar plates were further tested by ertapenem (Becton Dickinson) disk diffusion tests according to the Clinical and Laboratory Standards Institute (CLSI) guidelines 2013.\textsuperscript{11}

**Antimicrobial susceptibility testing**

Minimum inhibitory concentrations (MIC) were obtained by broth microdilution in cation adjusted Mueller-Hinton using ESB1F and EUVMV2 Sensititre\textsuperscript{®} plates (TREK Diagnostics Systems). Interpretations of MIC were performed according to the CLSI guidelines.\textsuperscript{11}

**Detection of antibiotic resistance and virulence factor genes**

DNA was obtained by incubating half a loop full of bacteria in 400 µl of lysis buffer (0.1 M Tris-HCl pH 8.5, 0.05% Tween 20 and 0.24 mg/ml proteinase K) for 45 min at 60°C followed by 15 min at 95°C.

Antibiotic resistance and virulence factor genes were detected using AMR08 ArrayStrip\textsuperscript{™} microarrays (an upgrade of Card \textit{et al.} 2013\textsuperscript{5}) and the HybridisationPlus (+) Kit (Alere Technologies GmbH). A signal intensity of 0.4 or higher was considered positive. The presence of β-lactam resistance genes *bla*\textsubscript{SHV} and *bla*\textsubscript{TEM}, and virulence factor genes was confirmed by PCR and sequencing with already published primers.\textsuperscript{26,31,38,43,51} New primers were designed with the NCBI primer-Blast tool (www.ncbi.nlm.nih.gov/tools/primer-blast): For the amplification and sequencing of the entire CTX-M Group 1 genes: CTX-M-Gr1 (CTX-MGr1_Fout2, 5'-GACTATTCATGTTGTTGTTAWTTCG; CTX-MGr1_RinSeq, 5'-TTCCCCATTCCGTTTCCG [PCR and sequencing] and CTX-MGr1-FinSeq, 5'-CGCGTGATACCACTTCAC; CTX-MGr1-Rout, 5'-CCGACTGCCGCTCTAAAT [sequencing], [designed from consensus sequences obtained from the GenBank accession no. X92506, GQ274934, AY458016, JQ434850]). For *cba*-F (5'-AAGGCACTGGAGTTCCTC TCC; *cba*-R, 5'-AAGGCACTGGAGTTCCTC TCC; cba-R, 5'-CCGACTGCCGCTCTAAAT [sequencing], [designed from consensus sequences obtained from the GenBank accession no. NC_014382.1]), *cif*-F (cif-F, 5'-CCATCGCA ACACACTTCAGCC; cif-F, 5'-GCTGTCGAGCGTATAG...
TGTC [GenBank accession no. AY128544.1], ireA (ireA-F, 5'-CACTGGGTFAGCAGCATGGAA; ireA-R, 5'-TGAAGGAGCCCGAAATGCTT [GenBank accession no. NC_008563.1]), and nleA (nleA-F, 5'-ACCGAGCCAAAAGACCCAAT; nleA-R, 5'-CTTGCCCAACCATGGCACC [GenBank accession no. NC_013008]). blaCMY genes were amplified using previously described primers and sequenced with both the recommended primers, and using an additional internal sequencing primer blaCMY-2_IntF1 5'-ACAACTTGACGCCGAAGC (GenBank accession no. X91840).

Amino acid substitutions in the quinolone resistance-determining region of GyrA and ParC were detected as described previously. PCR was performed using 5× HOT FIREPol® Master Mix Ready to Load (Solis BioDyne) under the following conditions: initialization for 15 min at 94°C followed by 35 cycles of amplification for 30 s at 94°C, 30 s at 48–60°C, and 1 min at 72°C and a final elongation step at 72°C for 5 min. Selected DNA amplicons were sequenced using Big Dye by the Sanger method/chain termination method on an ABI Prism 3100 Genetic Analyzer (Applied Biosystems).

Genotyping and determination of phylogenetic group

Genotyping was performed by Multi Locus Sequence Typing (MLST) using the method described by Wirth et al. 200656 and the sequence types (ST) were obtained with the database provided by the University College Cork, Ireland (http://mlst.ucc.ie/mlst/dbs/Ecoli). Plasmid incompatibility (Inc) groups were determined using a multiplex PCR-based replicon typing kit (PBRT kit MBK0038; Diatheva). Plasmids were extracted using the alkaline lysis method, including the phenol/chloroform purification step, and transformed into E. coli TOP10 cells by electroporation (Invitrogen). The transformants were selected on Mueller Hinton II agar containing either 70 mg/L of ampicillin or 2 mg/L cefotaxime and confirmed for the presence of blaCTX-M and blaCMY genes by colony PCR. Inc groups of the transformed plasmids were confirmed by PCR using specific primers described previously. Plasmids of the IncI1 group were further typed by Plasmid MLST28 and the STs were obtained using the PubMLST database (http://pubmlst.org/plasmid).

The phylogenetic groups of the isolates were obtained using the tripex PCR method previously described. Genotyping and determination of phylogenetic group

Statistical analysis


Results

Prevalence

The prevalences of 3GC-R-Ec were 73.3% (confidence interval [CI] 95%: 62–83%, n = 55/75) in chicken, 2% (CI 95%: 0–10%, n = 1/50) in beef, and 0% (CI 95%: 0–7.1%, n = 0/50) in pork and veal retail meat. No carbapenem-resistant E. coli (Fig. 1) were detected, even for those colonies growing on Brilliance CRE agar. However, colonies containing CMY-2 (n = 8/9) and CTX-M-1 (n = 1/9) were found growing on this agar, as already reported. 3GC-R-Ec isolates

The only 3GC-R-Ec isolated from beef retail meat belonged to ST453, phylogenetic group B1, and it contained a blaCTX-M-1 gene as well as genes conferring resistance to sulfamethoxazole (sul2), tetracycline [tet(B)], and trimethoprim (dfrA17) and a class 1 integrase as well as an IncI1 plasmid (Table 1).

FIG. 1. Percentage of resistance and resistance determinants among isolates from chicken meat. 3GC, third-generation cephalosporins; CHL, chloramphenicol; CIP, ciprofloxacin; COL, colistin; GEN, gentamicin; IMI, imipenem; KAN, kanamycin; MER, meropenem; NAL, nalidixic acid; SMX, sulfamethoxazole; TET, tetracycline; TMP, trimethoprim; CMY, pAmpC β-lactamase; CTX-M-1, ESBL; SHV-12, ESBL; DfrA, dihydrofolate reductase for trimethoprim resistance; Sul, dihydropteroate synthetase for sulfonamide resistance; Tet, tetracycline efflux.
Table 1. Genetic Characteristics, Origin, and Association of 3GC-R Isolates with Sequence Types, Phylogenetic and Plasmid Incompatibility Groups, Antibiotic Resistance, and Virulence Profiles

<table>
<thead>
<tr>
<th>ST (n)</th>
<th>PhG (n)</th>
<th>MPP (n)</th>
<th>Phenotype (n)</th>
<th>3GC-R genes (n)</th>
<th>Other antibiotic resistance genes and integrons (n)</th>
<th>Virulence factor genes (n)</th>
<th>Plasmid Inc groups and pMLST (n)</th>
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</table>

<sup>a</sup>The only isolate from beef retail meat.

(n): Number of strains; the absence of brackets means that only one strain is associated with the corresponding information. Bold and underlined plasmid incompatibility groups represent the plasmid on which the CTX-M-1, CMY-2, or CMY-4 gene has been located after transformation into *Escherichia coli* TOP10 cells. MPPA, MPPB, MPPE, meat-packing plant (MPP) A, B, and E; PhG, phylogenetic group; ST, sequence type; 3GC, third-generation cephalosporin; CHL, chloramphenicol; CIP, ciprofloxacin; KAN, kanamycin; NAL, nalidixic acid; SMX, sulfamethoxazole; TET, tetracycline; TMP, trimethoprim; astA, enteroaggregative toxin gene; biaC, colicin B activity protein gene; cba, colicin B activity protein gene; cmlA1-like, chloramphenicol efflux gene;bla<sub>CMY-2</sub>, pAmpC β-lactamase gene; bla<sub>CTX-M-1</sub>, extended-spectrum β-lactamase gene; bla<sub>SHV-12</sub>, extended-spectrum β-lactamase gene; dfrA, dihydrofolate reductase gene for trimethoprim resistance; ereB, erythromycin esterase gene; IntI, class I integrase gene; strAB, streptomycin phosphotransferase gene; sul, dihydropteroate synthetase genes for sulfonamide resistance; hylE, avian *E. coli* hemolysin gene; ireA, siderophore receptor gene; iroN, enterobactin siderophore receptor protein gene; iss, increased serum survival protein gene; lpfA, long polar fimbriae gene; nleA, non-LEE-encoded receptor A gene; tir, translocated intimin receptor gene; tsh, temperature-sensitive hemagglutinin gene; Inc, incompatibility; MLST, Multi Locus Sequence Typing; pST3, plasmid sequence type 3.
The 68 isolates from chicken retail meat (nine samples contained two and two samples contained three different STs) carried *bla*$_{CTX-M-1}$ (79.4%), *bla*$_{CMY-2}$ (17.6%), *bla*$_{CMY-4}$ (1.5%), or *bla*$_{SHV-12}$ (1.5%) and were frequently associated with additional resistance genes, such as sul sulfonamide resistance genes, *dfra* trimethoprim resistance genes, and *tet* tetracycline resistance genes (Fig. 1). Multidrug resistance was present in 61.8% of the isolates, which were resistant to three to six different antibiotics. The most frequent resistances were to third-generation cephalosporins, sulfamethoxazole, nalidixic acid, trimethoprim, and tetracycline (Fig. 1 and Table 1). Multidrug resistance was present in 61.8% of the isolates, which were resistant to three to six different antibiotics. The most frequent resistances were to third-generation cephalosporins, sulfamethoxazole, nalidixic acid, trimethoprim, and tetracycline (Fig. 1 and Table 1). One strain exhibited resistance to ciprofloxacin, which was associated to amino acid substitutions Ser83Leu, Asp87Asn in GyrA, and Ser80Arg in ParC.

The isolates from poultry belonged to 18 different STs, with the most frequent being ST1304 ($n=15$), ST38 ($n=12$), ST93 ($n=9$), ST770 ($n=6$), ST1158 ($n=6$), ST371 ($n=5$), and ST155 ($n=4$) (Table 1). Isolates belonging to the same ST also belonged to the same phylogenetic group except for ST93 isolates that formed two different groups (Table 1). The *bla*$_{SHV-12}$ and *bla*$_{CMY-4}$ genes were each only present in one ST, and *bla*$_{CMY-2}$ genes were present in two STs, whereas *bla*$_{CTX-M-1}$ was widespread in 15 different STs (Fig. 2). Isolates belonging to the same ST also contained the same resistance profiles and the same antibiotic resistance genes. Most of them also contained the same plasmid replicon types. The predominant Inc groups were IncI1 (91.2%), IncFIB (83.8%), IncFII (45.6%), IncK (36.8%), IncY (11.8%), and IncI2 (8.8%), whereas IncFIA, IncHI1, IncP, and IncX1 were only detected once (1.5%) (Table 1). Plasmids belonging to IncI1 (98.1%), IncFIB (85.2%), and IncFII (81.5%) were the most frequent in isolates harboring *bla*$_{CTX-M-1}$, and the Inc groups IncB/O (100%), IncFIB (100%), IncFII (100%), IncK (100%), and IncI1 (83.3%) were most common in *bla*$_{CMY-2}$ carriers. Analysis of the *bla*$_{CTX-M-1}$-positive plasmids transformed from six genetically diverse strains (ST93, ST1155, ST1304, ST2248, ST2307, and ST4007) as well as four genetically related strains (all of ST1304) into *E. coli* Top10 cells revealed that they all belonged to the IncI1 group and were of plasmid sequence type 3 (pST3). Similarly, the plasmids of three strains containing *bla*$_{CMY-2}$ (two strains belonging to ST38 and one to ST1364) as well as of one strain containing *bla*$_{CMY-4}$ (ST38) all belonged to the IncB/O group as determined by PCR (Table 1 and Fig. 2).

**Virulence factors**

A total of 12 different virulence factor genes could be identified from the isolates (Table 1). The APEC-associated *iroN* and *iss* genes were found in 59.4% and 76.8% of the isolates, respectively; in 59.4% of the isolates, they were detected together. Other APEC-associated virulence genes, including *tsh*, *astA*, and *lylE*, were found less frequently (<4.3%). The *lpfA* (long polar fimbriae) gene was only detected in one predominant clonal lineage ST1304 (100%). One strain belonging to ST752 contained enteropathogenic *E. coli* (EPEC)/EHEC-associated virulence factor genes *cif*, *espB*, *nleA*, and *tir* additionally to *iss*, *iroN*, and *tsh*. The isolates belonged to different phylogenetic groups: A: 18.8%, B1: 33.3%, B2: 2.9%, D: 27.5%, E: 15.9%, and F: 1.4% (Table 1).

**Discussion**

A very high prevalence (i.e., 73.3%) of 3GC-R-*Ec* was found at retail in poultry meat from indigenous animal
production in Switzerland, whereas beef (2%), pork (0%),
and veal (0%) meats were less or not contaminated. The
same observation has already been made for meat destined
for hospital meals in Switzerland as well as in other coun-
tries.\(^4,5,17,22\) However, the analysis of meat packed in
two different MPPs and purchased at different time points indi-
cates that contamination is not a sporadic event, but likely
occurs during slaughter and meat processing (Table 2). In-
deed, the prevalence of 3GC-R-Ec in Swiss poultry meat is
higher as that observed in Swiss broiler herds (38%).\(^6\)
The high rate of clonality for samples from the same slaughter-
house found here underlines the possible spread and persist-
ence of bacteria in meat packing plants, whereas 3GC-R-Ec
containing CTX-M-1 from broilers are very diverse.\(^21\)

Swiss meat contained CTX-M-1-, CMY-2-, and SHV-12-
possessing E. coli, which are also the most frequent 3GC-R-
Ec detected in animals in Switzerland and Europe.\(^49\) The
E. coli also belong to the ST commonly detected among
poultry isolates (ST10, ST48, ST57, ST69, ST93, ST155,
ST770, and ST1304) around Europe and in Switzerland\(^49,50\)
(personal communication). Similarly to other studies, iso-
lates from poultry meat harboring \(\text{bla}_{\text{CTX-M-1}}\) were mainly
associated with the presence of IncI1 plasmids, and those
containing \(\text{bla}_{\text{SHV-12}}\) with IncFIB, IncFI1, and IncI1 plas-
mids.\(^2,18,20,28,49,54\) The \(\text{bla}_{\text{CMY-2}}\) and \(\text{bla}_{\text{CMY-4}}\) genes were
associated with IncB/O plasmids, which is uncommon.
In fact, the association of \(\text{bla}_{\text{CMY-2}}\) with IncB/O plasmids has
only been recently reported in E. coli isolates from broilers
in Japan,\(^50\) while \(\text{bla}_{\text{CMY-2}}\) has mainly been associated with
IncK and IncFl1 plasmids in other European countries.\(^2,20,54\)
This suggests that a particular CMY-2 plasmid has been
selected and is circulating among E. coli from the Swiss poultry
meat production. Of note, all the CTX-M-1-expressing plasmids analyzed belonged to IncI1/pST3, an epidemic
plasmid propagating in Enterobacteriaceae from different
human and animal sources, including poultry in several
European countries (plasmid MLST isolate database [http://
pubmlst.org/databases]).\(^2,15,28,29\) In our study, the \(\text{bla}_{\text{CTX-M-1}}\)-
containing IncI1/pST3 plasmid was detected in both geneti-
cally related and genetically diverse E. coli strains, further
emphasizing the interstrain exchange of this promiscuous
plasmid in the poultry meat production.

The detection of chicken-associated toxins in the 3GC-R-
Ec emphasizes that the isolates originated from poultry.
Indeed, the predominant virulence factors \(\text{iroN}\) and \(\text{iss}\) as
well as \(\text{tsh}\) have been associated with plasmids in APEC,
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Disclosure Statement

None to declare.

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