1 An Isotopic Dilution Approach for Quantifying Mercury Lability in

Soils

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42 Abstract

The accurate estimation of soil mercury lability is crucial for risk assessment. In 43 comparison to chemical fractionation and speciation, isotopic dilution (ID) offers precise 44 definition of labile mercury fractions while maintaining natural equilibrium. We developed 45 and applied an ID protocol with ¹⁹⁹Hg to estimate the soil mercury (Hg) isotopically 46 exchangeable (labile) pool or HgE using a range of industrially contaminated soils in 47 48 Switzerland. The measured HgE values were consistent for the same soil against different spike levels (50, 100 and 200% of native ¹⁹⁹Hg) indicating that the spiked and soil isotopes 49 50 achieved required dynamic equilibrium at the soil-water interface. Total soil Hg (THg; mg kg⁻¹) was the best predictor of HgE (mg kg⁻¹) and %HgE and accounted for 96 and 63 % 51 52 of the variance, respectively. Nonetheless, despite the wide range of THg (0.37 - 310 mg kg^{-1}) in the studied soils, Hg lability spanned across a narrow range (~ 12 – 25 % of THg), 53 54 highlighting the large capacity of soils to sequester Hg in a very stable form. The 'exchangeable pool' of Hg extracted by CH_3COONH_4 and $MgCl_2$ (<0.25% and <0.32% of 55 56 THg, respectively) largely underestimated Hg lability in comparison to ID suggesting the potential usefulness of the ID approach. 57

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59 Keywords:

60 Soil pollution, Air pollution, Heavy metals, Stable isotopes, ICP-MS

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68 **1. Introduction**

Mercury (Hg) is a global pollutant that has drawn public concern due to its toxicity and 69 70 substantial bioaccumulation ¹⁻³. The UNEP Minamata Convention on Mercury aims to reduce Hg use and to curb global anthropogenic emissions of Hg ^{3, 4}. However, legacy Hg 71 72 in soils and sediments will continue to pose significant risks through re-emission to the atmosphere and the surrounding environments. For example, the land-atmosphere flux 73 may actually exceed the primary anthropogenic emissions of Hq⁰ and thus prolongs its 74 atmospheric residence ⁵⁻⁷. Moreover, a considerable fraction of soil mercury can 75 potentially accumulate in crops ⁸ or migrate to ground and surface waters ⁹. 76

77 Soil Hg mobility and bioavailability are largely linked to its labile pool; therefore, accurate 78 estimation of Hg lability is pivotal. To date, chemical extraction methods are the most 79 popular way to liberate the labile fraction of soil Hg. Nonetheless, they suffer numerous limitations e.g. inter-species conversion, re-adsorption and redistribution of Hg between 80 soil phases ^{10, 11}. In addition, there is no universal protocol available such as the sequential 81 82 extraction procedure developed by Tessier, et al. ¹² for classic hard metals ¹⁰ and selecting the type and sequence of extractants has always been an empirical decision based on the 83 nature of soil or the targeted Hg species ^{10, 11}. Moreover, when compared to Hg uptake by 84 flora and fauna, good correlations were found between bioavailable Hg and Hg from all soil 85 86 fractions indicating that labile Hg is not exclusively bound to specific soil phases ^{13, 14}. 87 Isotopic dilution (ID) assesses the labile metal pool in soils by defining the fraction of metal 88 that is isotopically-exchangeable or its 'E-Value' ¹⁵. Currently, ID is the most promising method to estimate the potentially labile metal fraction regardless of its speciation or soil 89 phase ¹⁶. Isotopic dilution has been successfully applied to determine the E-value of several 90 metals ¹⁵⁻²⁹. 91

92 To our knowledge, ID protocols have never been applied to try and measure the lability of 93 soil Hg. Working with Hg isotopes is a challenging task due to (i) ICP-MS low sensitivity 94 towards Hg caused by its naturally occurring seven isotopes and its high first ionization

95 energy ³⁰, and (ii) the potential reduction of soluble Hg⁺² to Hg⁰ and subsequent loss via 96 evaporation or adsorption to glassware and ICP-MS tubing system ³¹. In view of the need 97 to accurately estimate the availability of Hg in contaminated soils and to assess the 98 associated environmental risks, this study aims to develop and apply a working ID protocol 99 to quantify the pool size of labile soil Hg, and to provide preliminary insights into the 100 factors that likely control its mobility.

101 **2. Materials and Methods**

102 **2.1. Soil sampling and characterisation**

Three topsoil cores (0-10 cm) were taken in 9 residential areas in South-Switzerland where past industrial contamination with Hg is suspected (more in SI; S.1.1). Soils were air dried and sieved to <2 mm and soil properties including pH, organic carbon (Org-C), and total soil Hg (THg) were determined as described in SI (S.1.1).

107 **2.2. Preparation of Hg stable isotope standards**

Enriched mercury standard (HgCl₂, Trace Sciences Inc., Texas, USA, certified isotopic abundances of 30% for ¹⁹⁶Hg and 36.8% for ¹⁹⁹Hg) was dissolved in 2 % HNO₃ and 1% HCl to prevent Hg volatilization and improve its washout during analysis ^{30, 31}.

111 **2.3.** Determination of isotopically exchangeable (labile) Hg (Hg_E) in soil

112 Our ID protocol was adapted from several protocols developed for other heavy metals ^{17,} ^{18, 20, 25}. For all soils, two sets of soil suspensions (2 g dry soils in 30 mL 0.01 M Ca (NO₃)₂), 113 each with 4 replicates, were prepared and shaken for 72 h. Two of the four replicates were 114 then spiked with enriched ¹⁹⁶Hg or ¹⁹⁹Hg before all tubes were re-shaken for another 72 h. 115 Spiking solutions were prepared to deliver 50%, 100% and 200% of native ¹⁹⁶Hg or ¹⁹⁹Hg 116 to the soil suspension in three major groups (SI, S.1.2). The volume of the acidic spike 117 solution was minimized to avoid altering natural soil pH. Suspensions were then 118 centrifuged (3500 rpm, 25 min) and filtered with 0.45 µm syringe filters. The isotopic 119 ratios ¹⁹⁶Hg/²⁰⁰Hg, ¹⁹⁶Hg/²⁰¹Hg, ¹⁹⁶Hg/²⁰²Hg, ¹⁹⁹Hg/²⁰⁰Hg, ¹⁹⁹Hg/²⁰¹Hg and ¹⁹⁹Hg/²⁰²Hg, in the 120

supernatants, were measured by an Agilent 7700x ICP-MS and the isotopic abundances of spike and reference isotopes in soil (unspiked soil suspensions) were then inferred from their natural abundances ^{20, 24, 32} (details in SI; S.1.3.). The isotopically exchangeable Hg pool or HgE (mg kg⁻¹) of each soil was calculated from Equation 1 (adapted from Garforth, et al. ²⁰).

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$$HgE = \frac{M_{sl} C_{sp} V_{sp} ({}^{spIA}_{sp} - {}^{rfIA}_{sp}R)}{M_{sp} W_{sl} ({}^{rfIA}_{sl}R - {}^{spIA}_{sl})}$$
(1)

where M_{sl} and M_{sp} are the average atomic masses of Hg in soil and spike, respectively. C_{sp} is the gravimetric concentration (mg L⁻¹) of ¹⁹⁶Hg or ¹⁹⁹Hg in the spike solution, V_{sp} is the volume of the spike (L) and W_{sl} is the soil weight (kg). ^{sp}IA_{sp}, ^{rf}IA_{sp} are the spike and reference isotopes abundances in the spike solution, respectively, and ^{sp}IA_{sl}, ^{rf}IA_{sl} are their abundances in soils (un-spiked solutions). R is the equilibrium ratio of the spike to the reference isotope as measured by ICP-MS.

While HgE gives the absolute amount of isotopically exchangeable Hg (mg HgE kg⁻¹ soil),
Hg lability (%HgE) is the percentage of HgE to total soil Hg (THg).

For comparison with chemical extraction, 'exchangeable' soil Hg was extracted by
ammonium acetate and magnesium chloride ^{10, 33, 34} (more details in SI; S.1.4.).

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138 **3. Results and Discussion**

139 **3.1. Soil parameters**

Key soil parameters are displayed in Table S1. All soils were slightly acidic or alkaline (pH
6.2 - 8.2) with average Org-C content (2.6 to 4.5 %). Total soil Hg (THg) ranged in average
from 0.37 to 310 mg kg⁻¹ (more in SI, S.2.1.).

143 **3.2. Selection of spike and reference isotopes**

Mercury is a challenging element to analyse by ICP-MS because, in addition to its seven stable isotopes and high first ionization energy, soluble mercuric ions (Hg⁺²) can be readily 146 reduced to elemental mercury (Hg⁰) and lost from solution by sorption to containers' walls and/or sample introduction system walls ³⁰. To eliminate cross-contamination and memory 147 effect, all working solutions were prepared in a 0.5% ultra-pure HCl matrix to keep Hg in 148 its oxidised form and thus improve its washout from the system ³¹; in addition, a triple-149 step washing protocol (SI, S.1.3.) was adopted. Nevertheless, results still showed 150 substantial cross contamination in transition from ¹⁹⁶Hg spiked samples to the periodic Hg 151 standard. This was likely due to the very low natural abundance of ¹⁹⁶Hg in the periodic 152 standard. As a result, the natural isotopic ratio of the periodic Hg standard was significantly 153 distorted and fluctuated the external mass discrimination correction factors beyond 154 acceptable levels (0.59 - 0.93) (Table S2). In comparison, no significant cross-155 contamination was observed for ¹⁹⁹Hg and the mass-bias correction factors were close to 156 the unity ($\pm < 0.02$; Table S2) rendering it more favourable as a spike isotope than ¹⁹⁶Hg. 157

Usually, unless significant isobaric interference is predicted, the 'main' isotope is selected 158 159 as a reference isotope for E-value calculations (Eq. 1); this is normally either the most abundant isotope or the closest to the average mass e.g. ²⁰⁸Pb, ⁶⁵Cu and ¹¹⁴Cd ^{16, 17, 20}. 160 The values of HgE calculated using ²⁰¹Hg (average mass), ²⁰⁰Hg and ²⁰²Hg (most 161 162 abundant), were significantly different (paired t-test; p < 0.05, Table S3). Nevertheless, 163 equivalence analysis (Minitab 17) showed that the differences between HgE calculated by any two reference Hg isotopes were within a range of \pm 5% of their means (95%) 164 165 confidence interval, Figure 1.); moreover, systematic shifts of the confidence intervals towards the more abundant isotopes were evident (Figure 1.). In other words, the 166 calculated HgE was directly proportional to the natural abundance of the selected reference 167 isotope which is most likely due to the higher sensitivity of ICP-MS for heavier and more 168 abundant isotopes ³⁵. Accordingly, the HgE values calculated using ¹⁹⁹Hg as a spike isotope 169 170 and ²⁰²Hg as a reference isotope were regarded the most precise of all.

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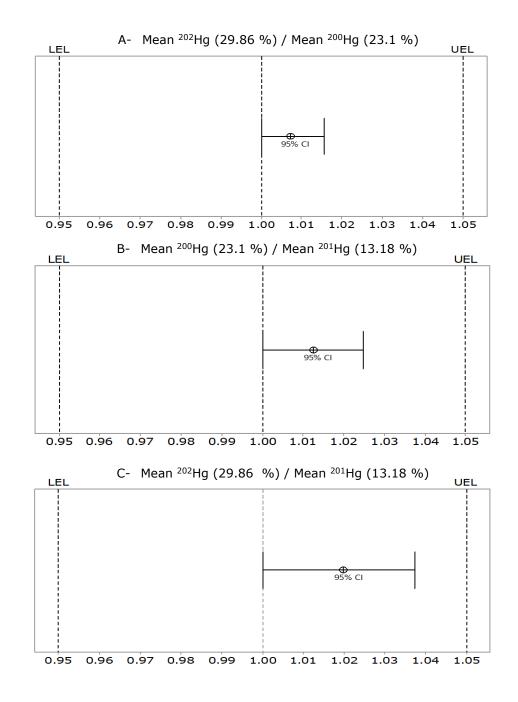


Figure 1. Comparison of equivalence tests (Minitab 17 package) showing the differences between the mean HgE values, at all sampling locations, measured by: A- 202 Hg vs 200 Hg, B- 200 Hg vs 201 Hg and C- 202 Hg vs 201 Hg, as reference isotopes. Dashed lines represent Lower Equivalence Limit (LEL) and Upper Equivalence Limit (UEL) set at ± 5% difference. Horizontal lines show confidence interval (95%) for the difference in HgE measured by 202 Hg, 201 Hg or 200 Hg. Three soil samples were taken from each location and two spiked and two un-spiked replicates were prepared for each sample.

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184 **3.3. Validation of Hg E-value**

In principle, E-value estimation is based on the assumption that the sorption reaction of 185 the spiked isotopes is reversible and that the isotopes in soil solution and solid phases are 186 in dynamic equilibrium ^{15, 16, 28, 29}. Hamon, et al. ³⁶ and Marzouk, et al. ²⁵ demonstrated 187 188 that the spiked isotopes may disturb the natural isotopic equilibrium leading to a systematic variation in the measured E-values at higher spike levels. Moreover, in 189 190 elements with several oxidation states e.g. As, Se, interspecies conversion may produce 191 an error in the measured E-values rendering it no longer represents the 'isotopically exchangeable' pools ^{37, 38}. Since Hg is known to interact significantly with organic matter 192 ^{3, 4, 39} and may undergo interspecies conversion e.g. evasion ^{40, 41} and methylation ^{1, 42, 43}, 193 validation of the measured HgE was essential. 194

We examined the validity of the proposed protocol by comparing HgE in all soils at different spike levels. Results showed that the apparent %HgE was consistent against different ¹⁹⁹Hg spike concentrations (Figure 2 & Table S4). This indicates that the dominant process is the reversible adsorption of Hg and that the potential chemical transformations of the spiked Hg did not preclude the accurate determination of HgE. This is also true for the expected disturbance in natural isotopic equilibrium at greater spike levels which seems to have minimal effect on the measured HgE.

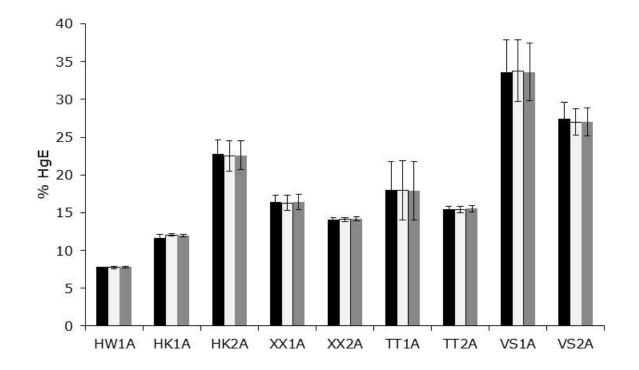


Figure 2. Hg lability (% HgE) in nine different soil from all sampling locations. Black, light grey and dark grey columns represent %HgE estimated at 50%, 100% and 200% ratios of spike to native ¹⁹⁹Hg, respectively. Error bars are standard error between 2 spiked (for each spike ratio) and 2 unspiked replicates. More data in Table S4.

218 **3.4. Mercury lability and soil properties**

Across the whole range of data, both HgE (mg kg⁻¹) and %HgE showed good correlations 219 220 (r= 0.96, 0.63, respectively) with THg while they showed no significant correlation with 221 any other soil parameter (Table S5). Stepwise regression, performed by Minitab 17, 222 confirmed that THg is solely the best predictor of both HgE and %HgE accounting for 96 223 and 63 % of the variance, respectively (Table S6). Figures S1 and S2 show that although 224 a very good 'power' relationship (R²=0.97) between HgE and THg is evident, there is only 225 a very broad 'logarithmic' relationship ($R^2=0.18$) between THg and %HgE. For HgE (mg kg⁻¹), the strong correlation with THg is expected since THg spanned across four orders of 226 227 magnitude while falls within a relatively narrow range of pH (6.2 - 8.2) and Org-C (2.63 -228 4.48 %) (Table S1). The broad correlation of THg with %HgE may reflect that 229 anthropogenic Hg remains more mobile than geogenic Hg or indicates a greater Hg 230 reactivity due to weaker adsorption at higher THg. However, this relationship could 231 possibly be an artefact especially that THg is used to calculate %HgE. To investigate this, 232 we used the 'lognorm.inv' function in Microsoft Excel to produce random distributions of THg and %HgE at 10000 values. In addition, %HgE was estimated against ln(THg) 233 234 assuming that HgE (mg kg⁻¹) values are constant around their means \pm standard deviation 235 ⁴⁴. Figure 3 clearly shows that, opposite to the experimental data, the calculated random relationships between THg and %HgE displayed negative trends. 236

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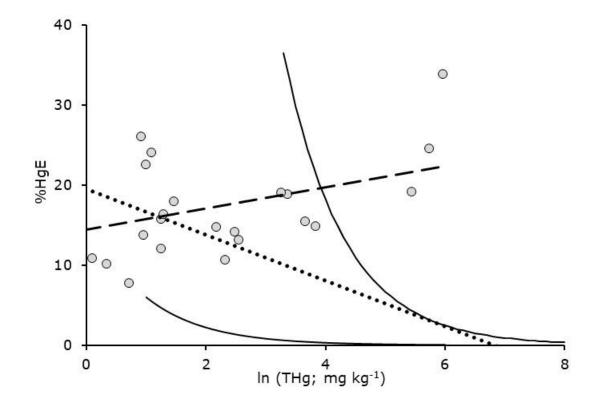
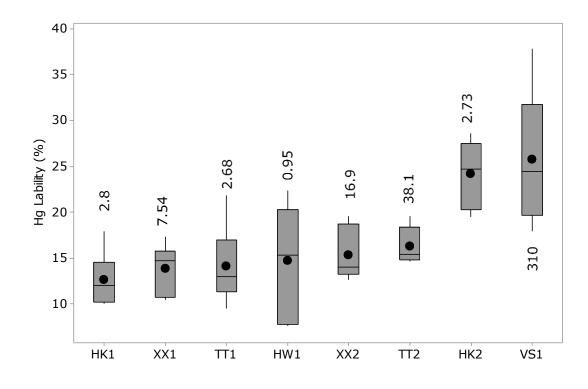




Figure 3. Relationship between In (THg; mg kg⁻¹) and experimentally measured %HgE values is displayed as grey circles and the dashed line is the a linear regression of the data. The dotted line represents linear regression of 10000 random pairs of %HgE and In(THg) allocated using the 'lognorm.inv(rand)' function of Microsoft Excel. The two solid curves assumes that HgE values are fixed at the means of In(HgE)±SD, thus resulting in a variation of %HgE with In(THg).

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Overall, the observed range of %HgE (12.2 – 24.7; Figure 4) was very narrow as opposed to the very wide range of THg (0.37 - 310 mg kg⁻¹; Table S1) and was considerably lower than other heavy metals' labilities e.g. Cd (up to 80%), Pb (up to 60%), and Zn (up to 40%) ^{17, 18, 20, 23, 24}. The exceptional affinity of Hg toward soil organic matter among heavy metals is well documented ^{3, 45}. This highlights the extraordinarily large capacity of top soils and sediments with low/average organic content to scavenge high concentrations of Hg and mitigate its environmental mobility ^{7, 46-48}.



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Figure 4. Values of % Hg lability (% HgE of total Hg) in all sampling locations. The 'box and whisker' plot shows median (horizontal lines), mean (black circle symbols). The box demarcates the boundaries of the second and third quartile; the whiskers extend to the furthest data point within 1.5 box heights (of the box). Locations are arranged according to their mean %HgE values in an ascending order. Each sampling location is represented by three soil samples and 4 replicates (two spiked and two un-spiked). Numbers above or below boxes are average total Hg (THg) in each location (mg kg⁻¹).

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264 **3.5. Comparison with extraction methods**

265 Ammonium acetate and magnesium chloride extractable Hg (exchangeable Hg) in our soils was only detectable in VS1 soils with pools of 0.01 to 0.25 % and 0.01 to 0.32 %, 266 respectively, which is substantially lower than the labile Hg estimated by ID for the same 267 268 location (19 - 33.7 %). This very low extractability was in line with previous findings (usually 0-5%) ^{10, 34, 48} but was substantially lower than those of Pb, Zn and Cd (up to 30 269 % at similar pH range) ^{17, 23-25}. This reveals the clear distinction between Hg as a classic 270 example of soft metal that has high affinity toward soft ligands e.g. S-bearing groups and 271 organic matter as opposed to hard metals which tend to react with hard ligands such as 272 O-bearing groups and Fe, Mn and Al hydrous oxides ^{3, 47, 49, 50}. Therefore, in case of hard 273

acids (hard metals and protons), competition with Mg²⁺ and NH₄⁺ on negatively charged oxygen sites may displace large amounts of sorbed metals especially in acidic soils. On the other hand, as a soft metal attached to soft ligands, Hg will face no such competition and the amounts released to soil solution will be minimal over the whole natural soil pH range ⁴⁷.

279 **3.6. Highlights, limitations and outlooks**

280 The soils used in this study were sampled at sites relatively close to each other that had been contaminated by one industrial Hg point source. They were thus relatively 281 282 homogenous in their Hg speciation and this may have contributed to the apparent 283 consistency of HgE at different spike levels. Highly acidic/alkaline soils or those Org-C 284 enriched that are simultaneously highly mineralized with Hg are scarce and the investigation of HgE in pristine soils will require an analytical system that offers greater 285 286 detection limits e.g. by using a cold vapour introduction system and/or higher resolution ICP-MS. This may also help in overcoming the ¹⁹⁶Hg carryover issues we encountered 287 288 enabling its usage as a spike isotope at much lower levels than ¹⁹⁹Hg.

Nevertheless, our set of soils covered the pH (slightly acidic to slightly basic) and Org-C (c. 2-5%) ranges mostly found in nature and the results demonstrated clearly that, under common circumstance, THg was the most decisive factor for both HgE and %HgE and highlighted the great capacity of surface soils and sediments to immobilize large quantities of Hg. However, future work on Hg contaminated soils with wider range of parameters, and studies that link HgE to bioavailable and reactive Hg (uptake by plants and animals, migration and evasion) will be of great importance.

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306 Reference

- 1. Mahaffey, K. R., Methylmercury: A new look at the risks. *Public Health Reports* **1999**, *114*, (5), 397-+.
- 2. Eto, K.; Marumoto, M.; Takeya, M., The pathology of methylmercury poisoning (Minamata disease). *Neuropathology* **2010**, *30*, (5), 471-479.
- 311 3. Beckers, F.; Rinklebe, J., Cycling of Mercury in the Environment: Sources, Fate, 312 and Human Health Implications-A Review. *Critical Reviews in Environmental Science and* 313 *Technology* **2017**, (just-accepted), 00-00.
- Driscoll, C. T.; Mason, R. P.; Chan, H. M.; Jacob, D. J.; Pirrone, N., Mercury as a
 Global Pollutant: Sources, Pathways, and Effects. *Environmental Science & Technology* **2013**, *47*, (10), 4967-4983.
- 5. Grigal, D., Inputs and outputs of mercury from terrestrial watersheds: a review. *Environmental Reviews* **2002**, *10*, (1), 1-39.
- 6. Reis, A. T.; Rodrigues, S. M.; Davidson, C. M.; Pereira, E.; Duarte, A. C., Extractability and mobility of mercury from agricultural soils surrounding industrial and mining contaminated areas. *Chemosphere* **2010**, *81*, (11), 1369-1377.
- Tipping, E.; Wadsworth, R. A.; Norris, D. A.; Hall, J. R.; Ilyin, I., Long-term mercury
 dynamics in UK soils. *Environmental Pollution* **2011**, *159*, (12), 3474-3483.
- 8. Zhang, H.; Feng, X.; Larssen, T.; Qiu, G.; Vogt, R. D., In inland China, rice, rather than fish, is the major pathway for methylmercury exposure. *Environmental Health Perspectives* **2010**, *118*, (9), 1183.
- 327 9. Xue, T.; Wang, R. Q.; Zhang, M. M.; Dai, J. L., Adsorption and Desorption of
 328 Mercury(II) in Three Forest Soils in Shandong Province, China. *Pedosphere* 2013, *23*, (2),
 329 265-272.
- Issaro, N.; Abi-Ghanem, C.; Bermond, A., Fractionation studies of mercury in soils
 and sediments: A review of the chemical reagents used for mercury extraction. *Analytica Chimica Acta* 2009, 631, (1), 1-12.
- Reis, A. T.; Lopes, C. B.; Davidson, C. M.; Duarte, A. C.; Pereira, E., Extraction of
 available and labile fractions of mercury from contaminated soils: The role of operational
 parameters. *Geoderma* **2015**, *259*, 213-223.

12. Tessier, A.; Campbell, P. G.; Bisson, M., Sequential extraction procedure for the speciation of particulate trace metals. *Analytical chemistry* **1979**, *51*, (7), 844-851.

Han, F. X.; Su, Y.; Shi, Z.; Xia, Y.; Tian, W.; Philips, V.; Monts, D. L.; Gu, M.;
Liang, Y., Mercury distribution and speciation in floodplain soils and uptake into native
earthworms (Diplocardia spp.). *Geoderma* **2012**, *170*, 261-268.

341 14. Jing, Y.; He, Z.; Yang, X.; Sun, C., Evaluation of Soil Tests for Plant-available
342 Mercury in a Soil–Crop Rotation System. *Communications in soil science and plant analysis*343 **2008**, *39*, (19-20), 3032-3046.

- 15. Young, S. D.; Tye, A.; Carstensen, A.; Resende, L.; Crout, N., Methods for determining labile cadmium and zinc in soil. *European Journal of Soil Science* **2000**, *51*, (1), 129-136.
- 16. Gabler, H. E.; Bahr, A.; Heidkamp, A.; Utermann, J., Enriched stable isotopes for determining the isotopically exchangeable element content in soils. *European Journal of Soil Science* **2007**, *58*, (3), 746-757.
- Atkinson, N. R.; Bailey, E. H.; Tye, A. M.; Breward, N.; Young, S. D., Fractionation
 of lead in soil by isotopic dilution and sequential extraction. *Environmental Chemistry* **2011**, *8*, (5), 493-500.
- 18. Chenery, S. R.; Izquierdo, M.; Marzouk, E.; Klinck, B.; Palumbo-Roe, B.; Tye, A. M., Soil-plant interactions and the uptake of Pb at abandoned mining sites in the Rookhope catchment of the N. Pennines, UK - A Pb isotope study. *Science of the Total Environment* **2012**, *433*, 547-560.
- 19. Gäbler, H.-E.; Bahr, A.; Mieke, B., Determination of the interchangeable heavymetal fraction in soils by isotope dilution mass spectrometry. *Fresenius' journal of analytical chemistry* **1999**, *365*, (5), 409-414.
- 360 20. Garforth, J. M.; Bailey, E. H.; Tye, A. M.; Young, S. D.; Lofts, S., Using isotopic
 361 dilution to assess chemical extraction of labile Ni, Cu, Zn, Cd and Pb in soils. *Chemosphere*362 2016, 155, 534-541.
- Izquierdo, M.; Tye, A.; Chenery, S., Sources, lability and solubility of Pb in alluvial
 soils of the River Trent catchment, UK. *Science of the Total Environment* **2012**, *433*, 110122.
- 22. Mao, L.; Young, S.; Tye, A.; Bailey, E., Predicting trace metal solubility and fractionation in Urban soils from isotopic exchangeability. *Environmental Pollution* **2017**.

Mao, L. C.; Bailey, E. H.; Chester, J.; Dean, J.; Ander, E. L.; Chenery, S. R.; Young,
S. D., Lability of Pb in soil: effects of soil properties and contaminant source. *Environmental Chemistry* **2014**, *11*, (6), 690-701.

- Marzouk, E. R.; Chenery, S. R.; Young, S. D., Predicting the solubility and lability
 of Zn, Cd, and Pb in soils from a minespoil-contaminated catchment by stable isotopic
 exchange. *Geochimica Et Cosmochimica Acta* **2013**, *123*, 1-16.
- Marzouk, E. R.; Chenery, S. R.; Young, S. D., Measuring reactive metal in soil: a
 comparison of multi-element isotopic dilution and chemical extraction. *European Journal of Soil Science* **2013**, *64*, (4), 526-536.

26. Nolan, A. L.; Ma, Y. B.; Lombi, E.; McLaughlin, M. J., Measurement of labile Cu in soil using stable isotope dilution and isotope ratio analysis by ICP-MS. *Analytical and Bioanalytical Chemistry* **2004**, *380*, (5-6), 789-797.

380 27. Sivry, Y.; Riotte, J.; Munoz, M.; Sappin-Didier, V.; Dupre, B., Study of labile Cd 381 pool in contaminated soil using stable isotope analysis, radioactive isotope dilution and 382 sequential extraction. *Geochimica Et Cosmochimica Acta* **2006**, *70*, (18), A594-A594.

28. Tongtavee, N.; Shiowatana, J.; McLaren, R. G.; Gray, C. W., Assessment of lead availability in contaminated soil using isotope dilution techniques. *Science of the Total Environment* **2005**, *348*, (1-3), 244-256.

Young, S.; Crout, N.; Hutchinson, J.; Tye, A.; Tandy, S.; Nakhone, L., Techniques
for measuring attenuation: isotopic dilution methods. *Natural attenuation of trace element availability in soils* 2007, 19-37.

389 30. McCurdy, E., Successful Low Level Mercury Analysis using the Agilent 7700 Series 390 ICP-MS. *Agilent ICP-MS J* **2011**, *45*, (1).

391 31. Nelms, S., Determination of Mercury in Urine. *AMERICAN BIOTECHNOLOGY* 392 *LABORATORY* **2004**, *22*, (2), 50-55.

393 32. Hieftje, G. M.; Barnes, J. H.; Grøn, O. A.; Leach, A. M.; McClenathan, D. M.; Ray, 394 S. J.; Solyom, D. A.; Wetzel, W. C.; Denton, M. B.; Koppenaal, D. W., Evolution and 395 revolution in instrumentation for plasma-source mass spectrometry. *Pure and applied* 396 *chemistry* **2001**, *73*, (10), 1579-1588.

397 33. Han, F. X.; Su, Y.; Monts, D. L.; Waggoner, C. A.; Plodinec, M. J., Binding,
398 distribution, and plant uptake of mercury in a soil from Oak Ridge, Tennessee, USA.
399 Science of the Total Environment **2006**, *368*, (2-3), 753-768.

400 34. Panyametheekul, S., An operationally defined method to determine the speciation 401 of mercury. *Environmental Geochemistry and Health* **2004**, *26*, (1), 51-57.

- 402 35. Nelms, S. M.; Beauchemin, D., *Inductively coupled plasma mass spectrometry* 403 *handbook*. Oxford, Boca Raton, FL: 2005.
- 404 36. Hamon, R. E.; Parker, D. R.; Lombi, E., Advances in isotopic dilution techniques in 405 trace element research: a review of methodologies, benefits, and limitations. *Advances in* 406 *agronomy* **2008**, *99*, 289-343.
- 407 37. Hamon, R. E.; Lombi, E.; Fortunati, P.; Nolan, A. L.; McLaughlin, M. J., Coupling 408 speciation and isotope dilution techniques to study arsenic mobilization in the 409 environment. *Environmental Science & Technology* **2004**, *38*, (6), 1794-1798.

410 38. Collins, R. N.; Tran, N. D.; Bakkaus, E.; Avoscan, L.; Gouget, B., Assessment of 411 isotope exchange methodology to determine the sorption coefficient and isotopically 412 exchangeable concentration of selenium in soils and sediments. *Environmental Science* & 413 *Technology* **2006**, *40*, (24), 7778-7783.

414 39. Fitzgerald, W.; Lamborg, C., Geochemistry of mercury in the environment. 415 *Environmental Geochemistry* **2005**, *9*, 107.

40. Fritsche, J.; Osterwader, S.; Nilsson, M. B.; Sagerfors, J.; Akerblom, S.; Bishop,
K.; Alewell, C., Evasion of Elemental Mercury from a Boreal Peat land Suppressed by LongTerm Sulfate Addition. *Environmental Science & Technology Letters* **2014**, *1*, (10), 421425.

- 420 41. Schluter, K., Review: evaporation of mercury from soils. An integration and 421 synthesis of current knowledge. *Environmental Geology* **2000**, *39*, (3-4), 249-271.
- 422 42. Morel, F. M. M.; Kraepiel, A. M. L.; Amyot, M., The chemical cycle and 423 bioaccumulation of mercury. *Annual Review of Ecology and Systematics* **1998**, *29*, 543-424 566.
- 425 43. Mousavi, A.; Chavez, R. D.; Ali, A. M. S.; Cabaniss, S. E., Mercury in Natural 426 Waters: A Mini-Review. *Environmental Forensics* **2011**, *12*, (1), 14-18.
- 427 44. Shetaya, W. H.; Marzouk, E. R.; Mohamed, E. F.; Elkassas, M.; Bailey, E. H.; Young, 428 S. D., Lead in Egyptian soils: Origin, reactivity and bioavailability measured by stable 429 isotope dilution. *Science of The Total Environment* **2018**, *618*, 460-468.
- 430 45. Tipping, E., Modelling the interactions of Hg(II) and methylmercury with humic 431 substances using WHAM/Model VI. *Applied Geochemistry* **2007**, *22*, (8), 1624-1635.
- 432 46. Soares, L. C.; Egreja, F. B.; Linhares, L. A.; Windmoller, C. C.; Yoshida, M. I., 433 Accumulation and oxidation of elemental mercury in tropical soils. *Chemosphere* **2015**, 434 *134*, 181-191.
- 435 47. Wallschlager, D.; Hintelmann, H.; Evans, R. D.; Wilken, R. D., VOLATILIZATION
 436 OF DIMETHYLMERCURY AND ELEMENTAL MERCURY FROM RIVER ELBE FLOODPLAIN
 437 SOILS. *Water Air and Soil Pollution* **1995**, *80*, (1-4), 1325-1329.
- 438 48. Renneberg, A. J.; Dudas, M. J., Transformations of elemental mercury to inorganic
 439 and organic forms in mercury and hydrocarbon co-contaminated soils. *Chemosphere*440 **2001**, *45*, (6-7), 1103-1109.
- 441 49. Schuster, E., THE BEHAVIOR OF MERCURY IN THE SOIL WITH SPECIAL EMPHASIS
 442 ON COMPLEXATION AND ADSORPTION PROCESSES A REVIEW OF THE LITERATURE.
 443 Water Air and Soil Pollution 1991, 56, 667-680.
- 50. Skyllberg, U.; Bloom, P. R.; Qian, J.; Lin, C.-M.; Bleam, W. F., Complexation of mercury (II) in soil organic matter: EXAFS evidence for linear two-coordination with reduced sulfur groups. *Environmental science & technology* **2006**, *40*, (13), 4174-4180.
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