Role of wavefront curvature in propagation of cardiac impulse

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Abstract

It is traditionally assumed that impulse propagation in cardiac muscle is determined by the combination of two factors: (1) the active properties of cardiac cell membranes and (2) the passive electrical characteristics of the network formed by cardiac cells. However, advances made recently in the theory of generic excitable media suggest that an additional factor—the geometry of excitation wavefronts—may play an important role. In particular, impulse propagation strongly depends on the wavefront curvature on a small spatial scale. In the heart, excitation wavefronts have pronounced curvatures in several situations including waves initiated by small electrodes, waves emerging from narrow tissue structures, and waves propagating around the sharp edges of anatomical obstacles or around a zone of functional conduction block during spiral wave rotation. In this short review we consider the theoretical background relating impulse propagation to wavefront curvature and we estimate the role of wavefront curvature in electrical stimulation, formation of conduction block, and the dynamic behavior of spiral waves.

Keywords: Myocardium; Wave front; Impulse propagation; Conduction; Reentry; Spiral wave

1. Introduction

The mechanisms of impulse propagation in the heart have been extensively studied for several decades both theoretically and experimentally [1–4]. It has been firmly established that impulse propagation is dependent on two tissue properties: (1) the passive electrical properties of cardiac muscle defined by the tissue micro-architecture, the cell shapes, the passive membrane characteristics, and the distribution of gap junctions, and (2) the excitable membrane properties defined by the distribution, conductances, and kinetic characteristics of ionic channels, transporters, and pumps. From the biophysical point of view, cardiac tissue belongs to the class of so-called ‘reaction–diffusion systems’ in which a local reaction, such as an action potential, propagates through a medium due to the release of stored biochemical energy. It shares important aspects of its biophysical behavior with a variety of other excitable systems not only of biological, but also of physical and chemical origin [5]. Investigation of these systems, first by means of mathematical modeling and subsequently in experiments, has shown that in addition to the passive and active properties of the medium, a third factor—the geometry of the excitation wavefront—contributes to wave propagation [6,7]. Particularly, the curvature of a wavefront may cause slowing of propagation and conduction block. Also, wavefront curvature is important for defining the properties of spiral waves which are responsible for some types of cardiac tachyarrhythmias. The concept of wavefront curvature has recently been applied to studies of impulse conduction in cardiac muscle, provoking new experimental and modeling work. The purpose of this short article is to discuss the new data obtained in these studies together with the theoretical background and to evaluate the role of wavefront curvature in normal and abnormal impulse propagation and in the mechanisms of re-entrant excitation. The role of wavefront curvature in the behavior of spiral waves will be discussed in detail.
2. Effects of wavefront curvature on wave propagation

2.1. Wavefront curvature and conduction velocity

The basic mechanism relating wavefront curvature to velocity of propagation (\( \theta \)) in an excitable medium is illustrated in Fig. 1. For simplicity, the medium is assumed to be 2-dimensional and isotropic; the effects of wavefront curvature in 3-dimensional and anisotropic media will be considered later. In the case of a flat wavefront, conduction velocity is equal to the steady-state velocity in a one-dimensional strand. The steady-state velocity (\( \theta_0 \)) is solely determined by the passive and active properties of excitable tissue (e.g., Ref. [1]). When the excitation front is curved outwards (convex), the conduction velocity is lower than \( \theta_0 \). This is because the local excitatory current supplied by the cells at the front of a convex wave distributes over a larger membrane area downstream. An opposite process takes place when the excitation front is curving inwards (concave). In this case, excitatory current converges in front of the propagating wave, producing a more rapid membrane depolarization. As a result, the conduction velocity of a concave wavefront is larger than \( \theta_0 \). The degree of wavefront bending is characterized by the local curvature (\( \rho \)) which is defined as the negative reciprocal of the local radius of curvature (\( r \)):

\[
\rho = -\frac{1}{r}
\]

It follows from this definition that the convex front has a negative \( \rho \), the concave front has a positive \( \rho \), and \( \rho = 0 \) for a flat front. A quantitative description of the dependence of conduction velocity on curvature in a continuous isotropic 2-dimensional excitable medium can be obtained analytically for small values of \( \rho \). Zykov and Morozova [8] have shown that under such conditions the velocity is given by the following equation:

\[
\theta = \theta_0 + D\rho \tag{1}
\]

The coefficient \( D \) is determined by the passive properties of the medium. For a continuous isotropic model of the electrical structure of cardiac muscle, \( D \) is equal to \( 1/C_m S_m R_1 \), where \( C_m \) is the specific membrane capacitance, \( S_m \) is the cell surface-to-volume ratio, and \( R_1 \) is the intracellular resistivity.

![Diagram](image)

Fig. 1. Effect of wavefront geometry on propagation velocity \( \theta \). \( \theta_0 \) denotes the steady-state velocity in a one-dimensional medium.

2.2. Critical curvature and conduction block

Since conduction velocity decreases as the wavefront curvature becomes more negative, it follows that the velocity will become zero at some critical level of \( \rho \) (\( \rho_c \)), and the excitation wave will be blocked. Moreover, in the case of a circular nucleus of excitation, propagation will not take place if the radius of the nucleus is less than a critical radius, \( r_c \). A rough theoretical estimate of \( \rho_c \) and \( r_c \) can be obtained from Eq. (1). Assuming that the linear relationship between the wavefront velocity and the curvature is preserved at high \( \rho \) values, the velocity will become zero when \( \rho \) and \( r \) satisfy the following equations:

\[
\rho_c = -\frac{\theta_0}{D} \quad \text{or} \quad r_c = \frac{D}{\theta_0} \tag{2}
\]

The effects of curvature on wave propagation have been extensively studied in mathematical models [6,7,9–12]. Experimentally, they have been demonstrated in several excitable systems including the chemical Belousov-Zhabotinsky reaction [13,14] and a biological system producing calcium waves in *Xenopus laevis* oocytes [15].

The fact that these effects have been considered only rarely in heart tissue until recently is explained by the small radii at which wavefront curvature significantly affects conduction. In order to estimate quantitatively the effect of \( \rho \) on the conduction velocity in ventricular myocardium, we assume \( R_1 \) to be 400 \( \Omega \) cm (longitudinal direction [16]), \( C_m \) to be 1 \( \mu \)F/cm\(^2\), and \( S_m \) to be 0.33 \( \mu \)m\(^{-1}\) [17]. Under such conditions the coefficient \( D \) is approximately 0.76 cm/s. The propagation velocity (\( \theta_0 \)) of the flat excitation front is approximately 50 cm/s in the longitudinal direction [16,18]. Introducing \( D \) and \( \theta_0 \) into Eq. (2) yields a critical curvature \( \rho_c \) = 66 cm\(^{-1}\) which corresponds to a critical radius of curvature \( r_c \) = 152 \( \mu \)m. Detection of a circular nucleus of excitation of such a small radius requires a spatial resolution of \( < 100 \mu \)m, which is difficult to achieve by using conventional mapping techniques with arrays of extracellular electrodes. It should be noted that estimates of \( \rho_c \) and \( r_c \) based on Eq. (2) may include two types of errors: (1) the dependence of \( \theta \) on \( \rho \) in cardiac muscle may be non-linear at high values of \( \rho \) and (2) the dependence of \( \theta \) on \( \rho \) may show a discontinuity near \( \rho_c \) and, correspondingly, conduction velocity may decrease to zero abruptly. The precise shape of the function \( \theta = f(\rho) \) in cardiac tissue is not yet known. Examples of the dependence of \( \theta \) on \( \rho \) for several excitable models (Fig. 2) suggest that the true value for the critical curvature may be somewhat smaller and the critical radius larger than predicted by Eq. (2).

An important factor affecting wave propagation in ventricular myocardium is its 3-dimensional (3D) structure. The theory linking conduction velocity to wavefront curvature according to Eq. (1) is valid for the 3D case as well. However, the relation between the curvature and the local radius in 3D tissue can be more complex than in 2-dimen-
sional (2D) tissue. For a cylindrical wavefront which is a 3D analogue of a circular wavefront, the relationship between the curvature and the local radius is the same as in the 2D medium \( \rho = \frac{-1}{r} \). For a sphere with radius \( r \) the local 3D curvature \( \rho_{3D} \) is:

\[
\rho_{3D} = -\frac{2}{r}
\]

(3)

It follows that the critical radius \( r_{c} \) in a 3D medium is 2 times larger than \( r_{c} \) in a 2D medium. Thus, the estimate for \( r_{c} \) of 152 \( \mu \text{m} \) in isotropic 2D cardiac tissue corresponds to 300 \( \mu \text{m} \) in 3D tissue. This difference reflects the fact that, for a given radius, the dissipation of excitatory current from a spherical 3D wavefront is larger than from the circular 2D wavefront.

2.3. Dependence of critical curvature on active and passive tissue properties

Eq. (2) allows one to predict how modulation of either passive or active tissue properties will affect the critical curvature. It follows from this equation that the effect of a change in membrane excitability on \( \rho_e \) is solely determined by the change of \( \theta_0 \) and can, therefore, be estimated from experimental measurements of \( \theta_0 \). For example, it is well known that conduction velocity decreases with high excitation rates. In atrial myocardium, velocity decreases approximately 2-fold in response to a very early premature stimulus or during pacing at the shortest possible interval [19]. According to Eq. (2), a 2-fold increase of the critical radius \( r_{c} \) is expected in these conditions. This frequency-dependent increase of \( r_{c} \) may be particularly important in tachycardias including flutter, or fibrillation.

According to Eq. (2), cell-to-cell coupling affects \( \rho_e \) and \( r_{c} \) via both the conduction velocity \( \theta_0 \) and the coefficient \( D \). In a continuous uniform medium, the conduction velocity is inversely proportional to the square-root of intracellular resistivity \( R_i \) while the coefficient \( D \) is inversely proportional to \( R_i \). It follows that \( \rho_e \) is inversely proportional to \( R_i^{1/2} \). This dependence also allows one to evaluate the effect of anisotropy on critical curvature and critical radius. Thus, a wavefront propagating in the longitudinal direction has a critical radius of curvature which is \( (R_T/R_I)^{1/2} \) times smaller than the \( r_{c} \) for a wavefront propagating in the transverse direction, where \( R_T \) and \( R_I \) are axial resistivities in longitudinal and transverse directions, respectively. Also, a circle of critical radius in an isotropic medium corresponds to an ellipse in an anisotropic medium with the transverse axis of this ellipse shortened by the factor \( (R_T/R_I)^{1/2} \).

3. Occurrence of curved wavefronts in cardiac muscle

The estimates of critical curvature given above indicate that curvature effects are expected to become apparent in cardiac muscle when excitation fronts have bending radii < 150–300 \( \mu \text{m} \). In cardiac tissue curved wavefronts with small radii are encountered in a variety of situations which include: (1) centrifugal propagation from a small stimulating electrode or from a small group of pacemaker cells; (2) abrupt changes in tissue geometry with propagation emerging from small narrow strands or isthmuses into a large mass of excitable tissue. Examples of such structures are Purkinje–muscle junctions, insertions of accessory pathways into ventricular muscle in the WPW syndrome, and probably surviving cell strands connecting regions of intact tissue in infarcted muscle; (3) propagation of a wave around a fixed anatomical or functional conduction block; (4) spiral wave rotation.

3.1. Wave propagation following point stimulation

Direct experimental evidence that wavefront curvature affects propagation of waves initiated from small stimulating electrodes was recently obtained by Knisley and Hill [20]. They investigated impulse conduction in 2-dimensional rims of epicardial tissue stimulated either by a single electrode or by a linear array of electrodes as shown in Fig. 3. Optical mapping of activation spread with a laser scanning technique demonstrated that stimulation with a single electrode resulted in an elliptic spread of excitation while stimulation with a linear array produced a nearly flat activation front. As a result of increased wavefront curvature, the velocity of the elliptical propagation was on the average 13% smaller than that of the flat wavefront. In these measurements, the spatially averaged velocity values were compared which underestimate the effect of steep wavefront curvature on local conduction velocity immediately near the stimulating electrode. Nevertheless, these data clearly demonstrate the importance of wavefront curvature. Measurements of conduction velocity as a function of intracellular resistivity \( R_i \) while the coefficient \( D \) is inversely proportional to \( R_i \). It follows that \( \rho_e \) is inversely proportional to \( R_i^{1/2} \). This dependence also allows one to evaluate the effect of anisotropy on critical curvature and critical radius. Thus, a wavefront propagating in the longi-
of curvature at small values of $r$ were taken to determine the coefficient $D$ in Eq. (1). The mean longitudinal $D$ value was 0.47 cm$^2$/s. According to Eq. (2) this corresponds to a critical radius of 92 $\mu$m.

3.1.1. Wavefront curvature and ‘liminal area’

The inability of excitable tissue to support propagation of waves with curvatures higher than $r_c$ suggests that a critical number of cells encompassed within a nucleus of excitation with radius $r_c$ must be excited to achieve a propagated response. A similar requirement has long been recognized for 1D excitable strands and formulated in the concept of ‘liminal length’ [21,22]. The ‘liminal length’ was defined as the length of an excited strand segment necessary to produce the local current required for a propagated response. Accordingly, in a 2D tissue this critical number of cells is characterized by a ‘liminal area’ [23]. Although the ‘liminal area’ of impulse initiation and a curved wavefront with a ‘critical radius’ may differ with respect to membrane potential distribution within the excited portion of the tissue, the common features between these processes can be used to estimate the ‘critical radius’ from the dimension of the ‘liminal area’, independently of the parameters $D$ and $\theta_0$ in Eq. (2).

The liminal area has been calculated in a 2-dimensional computer model by Ramza et al. [24]. They studied im-
pulse initiation produced by a point current injection in a continuous, isotropic model described by Beeler-Reuter ionic kinetics [25]. The liminal area necessary to generate sufficient inward current during stimulation was determined as a function of the maximal sodium conductance. At a level of excitability estimated to correspond to that of adult ventricular myocardium, the radius of the liminal area was 200–250 \( \mu \text{m} \). This compares well with the estimates of the critical radius calculated from Eq. (2). Experimentally, the liminal area was estimated from measurements of stimulation threshold as a function of electrode size by Lindemans and co-workers [23,26]. Stimulation current was applied to canine epicardium via disk electrodes with radii varying between 0.01 and 9 mm. As illustrated in Fig. 4A, it was found that the current threshold was independent of the electrode size when the disk radius was less than 0.2 mm. However, the current threshold was proportional to the electrode radius to the power 1.5 when the radius was larger than 0.4 mm. This behavior was explained by noting that, with small electrodes, all current passed through the liminal area and, therefore, current density at the edge of the liminal area was independent of the electrode size. With large electrodes, more current had to be provided to maintain a constant stimulatory current density as the electrode surface area increased. Thus, the estimated radius of the liminal area was approximately 0.3 mm. If we assume that the disk electrode excited a semi-spherical area of cardiac tissue under the epicardial surface, then this situation is equivalent to excitation of a spherical region in a tissue filling the whole 3D space. Therefore, the measured radius of the liminal area can be taken as an estimate of the critical radius of curvature. Scaled down from 3 to 2 dimensions, the estimated critical radius of 300 \( \mu \text{m} \) in the 3D tissue corresponds to 150 \( \mu \text{m} \) in the 2D tissue, which is close to the other estimates of the critical radius. Lindemans and Zimmerman [26] and Winfree [27] also pointed out a practical consequence of the ‘liminal area’ effect: an electrode with a radius matching the radius of liminal area is the most efficient (i.e., it requires the lowest stimulation energy, as illustrated in Fig. 4B).

3.2. Curved wavefronts at geometrical expansions

Abrupt geometrical expansions such as transitions from narrow strands to a large volume of myocardium or narrow isthmuses of tissue bridging two large areas of atrial or ventricular myocardium have been implicated in the formation of unidirectional conduction block and re-entry. Examples of such structures include the Purkinje–muscle junction [28], the junction between accessory pathways and myocardium in the WPW syndrome [29,30], and the junction between thin cell strands surviving within an infarcted myocardium and intact tissue [31,32]. The failure of impulse transmission at such structures was attributed to the ‘impedance mismatch’ between the strands and the wide tissue regions in analogy to conduction block in branching axons or cell strands [33–35]. More recently, however, a different approach was proposed which is based on consideration of the curvature of the wave front emerging from an expansion instead of the impedance mismatch at the transitional region. Cabo et al. [36] investigated impulse propagation across narrow isthmuses in isolated sheets of ventricular epicardial muscle using a video-imaging technique. As shown in Fig. 5, cuts were made in the preparations so that only a narrow isthmus of tissue (0.9–2.2 mm in width) bridging two large areas was left intact. The excitation wave emerging from such an isthmus had an elliptical shape with a pronounced curvature (panels B and C): the smaller the isthmus width, the higher the curvature of the elliptical wavefront beyond the isthmus. In accordance with the expected effect of increased curvature, conduction velocity in the region immediately beyond the isthmus decreased as the isthmus width became smaller (panel D). At the smallest isthmus shown in panel D the conduction velocity is 19 cm/s, 42% smaller than the velocity of the flat wave (33 cm/s). The critical radius, \( r_c \), was estimated from the minimal isthmus width allowing passage of excitation waves between two large areas at various stimulation frequencies. At long cycle lengths (200 to 500 ms) the critical width was estimated to be \( <1 \text{ mm} \) in the longitudinal direction, corresponding to \( r_c < 0.5 \text{ mm} \). At a short excitation interval (150 ms) excitability was reduced and the critical isthmus width increased to 1.3–2 mm (critical radius 0.65–1 mm). At the maximal frequency of stimulation (117 ms interval) isthmuses with width \( <2.5 \text{ mm} \) (\( r_c < 1.2 \text{ mm} \)) resulted in conduction block.

![Fig. 5. Wave propagation across a narrow tissue isthmus in an isolated ventricular preparation of sheep heart. (A) Map of activation spread before an isthmus was produced. (B) Activation spread in the same preparation with isthmus 2.26 mm wide. The isthmus was produced by two tissue cuts (gray zones). (C) Activation spread after the isthmus was reduced to 0.88 mm. (D) Local conduction velocity measured across the isthmus as a function of isthmus width. Reproduced with permission [36].](image)
A similar expanding structure, consisting of a thin long strand emerging into a large tissue area, has been recently investigated both experimentally [37,38] and theoretically [39]. In computer simulations impulse propagation from a strand into a large area failed when the strand width was reduced below 200 μm in a 2D model or 350 μm in a 3D model. Fig. 6 illustrates the block of impulse propagation in the 2D model. The precise localization of the site of conduction block was obtained from the recordings of the excitatory inward sodium current (panel C). These recordings demonstrate that conduction block occurred beyond the site of geometrical expansion. A small circular nucleus of excitation emerged into the large area. Pronounced curvature of this nucleus imposed a large current drain on the wavefront and excitation did not propagate further. Experimentally, wave propagation from a narrow strand into a large cell area has also been investigated in cell cultures as a function of the strand width using high-resolution optical mapping of transmembrane potential [38]. In these studies, narrow myocyte strands emerging into a cell monolayer were produced using a patterned-growth technique [40]. As in simulations, wave propagation was either slowed or blocked when the strand width was reduced below a critical level. The critical strand width was approximately 30 μm. This is significantly smaller than the estimates of critical radius given above. This difference was explained by several geometrical and electrophysiological features of cell cultures: a shorter electrotonic space constant than in the adult myocardium, electrophysiological heterogeneity between the strands and the large areas, and a gradual increase of the strand width before the transition to the large area. When these properties were included into the computer model, a close fit between experimental and simulated results was obtained [39].

We would like to point out that the two approaches—the ‘wavefront curvature’ or ‘impedance mismatch’—are related to the same biophysical mechanism: namely, the balance of electrical currents during wave propagation or the ‘source–sink’ relationship. The approach based on consideration of the wavefront curvature has an advantage of being able to relate, in a simple way, conduction block to electrophysiological parameters which can be directly measured in experiments (i.e., steady-state conduction velocity and passive tissue properties). Also, it helps to understand the occurrence of conduction block at an isthmus connecting two equal (i.e., symmetrical) areas where no impedance mismatch is present.

3.3. Wavefront curvature and propagation around sharp obstacles

Sharp, longitudinally oriented obstacles, which may be structural or functional in nature, are thought to play an important role in so-called ‘anisotropic re-entry’ [41]. They have been described in the border zone of experimental infarcts and in the aging heart [41,42]. Computer simulations and recent experiments demonstrate that wavefront curvature has a profound effect on wave propagation near the sharp edges of such obstacles. Fig. 7 demonstrates wave propagation in a computer model of an excitatory medium with a thin resistive obstacle. Ionic currents in this model are described by the Luo-Rudy kinetics [43] with maximal sodium conductance reduced to approximately 30% of its nominal value. As in Fig. 6, local activation is defined by the flow of sodium inward current. The refractory state is defined by inactivation of the sodium current. The isochronal activation map (panel A) demonstrates that
as the propagation wave approaches the pivoting point, it detaches from the obstacle and describes a trajectory around a circular area which remains in the resting state. The detachment takes place since the critical curvature of the wavefront prohibits an abrupt turn around the sharp edge of the obstacle. Several electrophysiological features of this effect are of special importance not only for characterizing the process of propagation around unexcitable obstacles but also for describing the behavior of spiral waves in general (see below). As illustrated in Fig. 7B, there is a point on the wavefront (detached from the obstacle) where 3 states—the excited state (black), the refractory state (gray), and the resting state (white)—meet. We define this singular point as the tip of the wavefront. Accordingly, it is used to define the tip of a spiral wave and to describe spiral wave rotation (see below). The radius of the pivoting trajectory \( r_p \) is determined by the critical wavefront curvature or the critical radius, \( r_c \). At present, the exact value of \( r_p \) in cardiac muscle is not known. The lack of detachment of a wave front from unexcitable obstacles in cardiac muscle under normal physiological conditions indicates that \( r_p \) is rather small (i.e., smaller than the spatial resolution of the mapping techniques which amounted to several hundred micrometers in these experiments [44,45]). However, reduction of tissue excitability (e.g., by application of tetrodotoxin or by stimulation at high rates) increases \( r_c \) and \( r_p \) and causes detachment of wavefronts from a sharp obstacle [44]. Under conditions of very low excitability, the radius of the pivoting trajectory can become larger than the wavelength of excitation, the product of conduction velocity and refractory period. In this case, the wave may perform a complete turn without reconnecting to the obstacle and form a spiral wave. The detachment of excitation waves from sharp obstacles and subsequent formation of spiral waves was first described in a computer model at high excitation rates [46]; experimentally this has been observed in the chemical Belousov-Zhabotinsky reaction [47] and more recently in heart muscle [44].

4. Wavefront curvature and spiral waves

One of the most remarkable examples of wave propagation in which curvature plays an important role is a spiral wave of excitation. Spiral waves occur universally in excitable media including chemical reactions [48,49], neural tissue (depression waves in the retina [50] and cerebral cortex [51]), intracellular calcium signaling systems (\( Xenopus laevis \) oocytes [15], cardiac myocytes [52]), and amoebae colonies [53]. One of the most extensively studied examples is the Belousov-Zhabotinsky (BZ) reaction. In this reaction, malonic acid is reversibly oxidized by bromate in the presence of ferroin. In this process, ferroin changes in color from red to blue and then back to red, which allows the visual observation of the reaction. Fig. 8A shows a rotating spiral wave in a thin 2-dimensional layer of the BZ reaction. In the center of the rotating wave (core) the tip of the wave moves along a complex trajectory and radiates waves into the surrounding medium. Since the velocity of a convex wavefront cannot exceed the speed of the flat wave \( (\theta_0) \) the rotating wave always acquires the shape of an Archimedean spiral at the periphery, independently of the behavior of its core. Because of this shape, rotating waves were given the name ‘spiral waves’. Other names used in the literature include ‘vortices’ and ‘reverberators’. In some cases, the term ‘rotor’ was used to refer to the core of a spiral wave. Important properties of spiral waves including the rotation period, size, and dynamic behavior are determined by
Fig. 8. Rotating waves in various excitable media. (A) Spiral wave in the Belousov-Zhabotinskii reaction. Reproduced with permission [49]. (B) Isochronal activation map of ‘leading circle’ re-entry in an isolated preparation of rabbit atrial muscle. Reproduced with permission [64]. (C) Spiral wave in an isolated preparation of canine epicardial muscle imaged using voltage-sensitive dye. Reproduced with permission [71].

the wave propagation in the core where the wavefront has a pronounced curvature. The theory of excitable waves suggests that two factors—the wavefront curvature and the refractory period—contribute to the period of spiral wave rotation [6,10]. Mutual dependence and interaction between these parameters result in a complex behavior of spiral waves. Because of this complexity, the understanding of spiral wave dynamics relies heavily on computer modeling. A convenient way to relate the behavior of spiral waves with model parameters is to use cellular automata models of cardiac excitation. In such models, excitatory elements are described by 4 states (excited, absolute refractory, relative refractory, and resting) and the transition from one state to another is governed by formal rules [54]. One of the advantages of these models is that integrative properties of excitable media such as excitability and the durations of the absolute and relative refractory periods are set as model parameters and can be modified independently. Importantly, these models can be constructed to reproduce the effects of wavefront curvature on conduction velocity. Although cellular automata models oversimplify active and passive properties of cardiac tissue, they provide an easy insight into spiral wave dynamics.

4.1. Curvature and spiral wave dynamics

Fig. 9 demonstrates 3 different types of spiral wave rotation in a cellular automata model of excitable medium [55]. As in Fig. 7, the wavefront tip is defined as the point where 3 states—the excited, the refractory, and the resting—are in contact with each other. Fig. 9A shows the simplest type of spiral wave rotation when the excitability in the model is very low. In this case, the critical radius (\( r_c \)) and therefore the pivoting radius of the wave tip (\( r_p \)) are large. By contrast, the wavelength of excitation (\( \lambda \)) given by the product of the velocity of propagation (\( \theta \)) and the refractory period [\( R, (\lambda = \theta \cdot R) \)] is small as a result of the small \( \theta \). Under these conditions the length of the pivoting trajectory (\( 2\pi r_p \)) is larger than \( \lambda \), and therefore propagation of the excitation wavefront is not affected by the refractory tail. The only constraint on the wavefront propagation is imposed by the curvature at its tip. Because the wavefront curvature cannot exceed the critical curvature (\( r_c \)) the wave tip does not extend towards the center of the rotation but follows a circular trajectory (panel A, bottom). The area circumscribed by the circle is never excited and remains at rest. In other words, the spiral wave contains a fully excitatory gap.

At a higher level of excitability, when \( r_p \) decreases and the length of the pivoting trajectory becomes comparable with the wavelength of excitation (\( 2\pi r_p \approx \lambda \)), the front of the excitation wave meets its own refractory tail, as shown in Fig. 9B. As a result, the wavefront velocity decreases and the front retreats from the refractory tail. It now propagates in a fully recovered medium and the velocity increases again. The alternation of wavefront acceleration and retardation repeats itself in a complex dynamic manner. Such a front–tail interaction results in a ‘meandering’ of the spiral wave tip as shown at the bottom of panel B. The movement of the wave tip in this case follows a cycloidal or ‘flower’ trajectory. Such a meandering movement of spiral waves was observed in the BZ reaction.

Fig. 9. Effect of wavefront curvature on spiral wave rotation in a cellular automata model of an excitable medium. Upper panels show snapshots of activation. Excited, absolute refractory, relative refractory, and resting states are shown in black, dark gray, light gray, and white, respectively. Lower panels show enlarged trajectories of the spiral wave tip. (A) Circular type of rotation in a model with large critical curvature (\( 2\pi r_p \)) and \( \lambda \). (B) Cycloidal type of rotation in a model with intermediate level of critical curvature (\( \lambda \approx 2\pi r_p \)). (C) ‘Z’ type of rotation in a model with small critical curvature (\( \lambda > 2\pi r_p \)). Reproduced with permission [55].
and Z types C and D of rotation in the FitzHugh-Nagumo model. Bars at the bottom show relative scaling. Reproduced with permission.

The types of spiral wave rotation described by the cellular automata model in Fig. 9 are also observed in computer models based on partial differential equations (FitzHugh-Nagumo model [60] and Beeler-Reuter model [61]). Fig. 10 shows trajectories of the spiral wave tip in the FitzHugh-Nagumo model upon increasing the wavelength of excitation (\(\lambda\)), while the pivoting radius (\(r_p\)) remains constant [60]. As in the cellular automata model (Fig. 9), spiral wave rotation changes from circular to cycloidal, and then to Z type as the ratio between \(\lambda\) and \(r_p\) increases. Interestingly, the rotation of spiral waves in the computer simulations shown in Figs. 9 and 10 is never stable, except for the extreme case of circular rotation when the length of the pivoting trajectory is larger than \(\lambda\). The strongest meandering is observed at intermediate levels of the ratio \(\lambda/r_p\). Such meandering of spiral wave rotation has been implicated in the generation of polymorphic ECG’s related to cardiac arrhythmias such as torsades de pointes [62,65]. With high values of \(\lambda/r_p\) the linear zone of functional conduction block also moves, although more slowly.

The results presented above were obtained in isotropic models. They can be directly extended to anisotropic tissue with continuous electrical properties. In such a case, the effect of changing anisotropic tissue ratio is equivalent to geometrical scaling in the transverse direction by a factor which equals the square root of the ratio between resistivities in transverse and longitudinal directions. Introducing anisotropy in such a way does not change the period of spiral wave rotation or the duration of excitable gap [66].

### 4.2. Occurrence of spiral waves in cardiac muscle

In the heart, spiral waves have long been implicated in the generation of cardiac arrhythmias [48,67–69]. However, the first experimental observation of rotating waves in atrial muscle [70] and later mapping studies in a variety of preparations revealed a pattern of activation spread that differed in part from the spiral shape (Fig. 8B). Thus, the isochrone lines were only slightly bent and deviated from the spiral shape shown in Fig. 8A. There are two possible explanations for this discrepancy: (1) Only the central portion of the wave near its tip was observed in a tissue approximately 2 cm in diameter and, therefore, the spiral shape was less prominent. (2) The spiral shape might have been further masked by electrophysiological heterogeneities present in atrial muscle. However, provided that the size of the preparation is large and/or the excitation wavelength is small, the wavefront will inevitably acquire a spiral shape. Indeed, in preparations with short excitation wavelength (low excitability and/or low degree of cellular coupling), rotating waves show a distinct spiral shape (Fig. 8C) [71].

As described above (Figs. 9 and 10), the type of rotation (i.e., the trajectory followed by the spiral wave tip) is determined by the relation between the excitation wavelength (\(\lambda\)) and the pivoting radius (\(r_p\)). The wavelength of excitation during sustained rotation is well known and amounts to approximately 3 cm (a velocity of 30 cm/s times an absolute refractory period of 0.1 s). The value of the pivoting radius in cardiac muscle has not been measured yet. As discussed above, it is very small at normal excitability, perhaps in the range of several hundred micrometers. Thus, \(\lambda\) is significantly larger than the estimated value of the pivoting radius in the normal case. Therefore, for cardiac tissue in a normal state of cell-to-cell coupling and with normal excitability, the trajectory of the tip is expected to assume the linear or Z-type of shape shown in Fig. 9C and 10D. This theoretical consideration
is supported by experimental observation from mapping experiments showing that the zone of functional conduction block often has a linear shape [19,72,73]. The linear trajectory of the rotating wave tip is also seen in the work of Allessie et al. (Fig. 3 of Ref. [64]). Under conditions of reduced excitability spiral waves with cycloidal or even circular types of rotation (Fig. 9A and 10A) can be anticipated.

4.3. Wavefront curvature and drift of spiral waves

In addition to meandering in the center of a spiral wave, spiral wave instability may involve drift due to the electrophysiological heterogeneity of cardiac tissue. The first experimental observation of spiral wave drift in cardiac muscle was reported by Fast and Pertsov [74]. In this work, spiral waves were initiated in isolated 2-dimensional preparations of rabbit ventricular epicardium. Preparations were placed in a chamber with two compartments divided by a thin rubber barrier. A functional heterogeneity in refractoriness and conduction velocity was created by separate superfusion of two compartments with normal versus quinidine-containing solutions. As shown in Fig. 11, premature stimulation from the area with the low refractoriness induced a conduction block at the border between the two compartments (panel A) and the formation of a spiral wave rotating in a counterclockwise direction. On subsequent cycles (panel B) the spiral wave moved along the border of heterogeneity (perpendicular to the gradient of refractoriness) and continued to drift until it died at the border of the preparation (panel C). The drift velocity amounted to about one-fifth of the propagation velocity, and its direction was determined by the direction of wave rotation and the gradient of refractoriness. When a spiral wave with a clockwise rotation was initiated, it drifted in the opposite direction.

Due to the drift of a spiral wave, the frequency of excitation at a given measurement site depends on the location of this site relative to the moving spiral. This effect is known in the theory of electromagnetic and acoustic waves as the Doppler effect. Fig. 11D shows excitation intervals measured during spiral wave drift at different tissue locations. The measuring sites were distributed along the boundary of heterogeneity which determined the direction of the drift. Because of the drift, the sites located in front of the drifting spiral wave were excited significantly faster than the sites located behind it. The difference in excitation intervals measured ahead of and behind the spiral wave amounted to 30%. Drift of spiral waves and the Doppler effect were also observed in isolated preparations of sheep ventricular muscle imaged with an optical mapping technique [66,71]. In this case, no artificial heterogeneity was created and the spiral wave drift was likely to be a result of intrinsic spatial gradients of electrophysiological properties. The Doppler effect and the coexistence of different excitation frequencies within the same preparation have been used to explain a possible mechanism of the ECG pattern observed during arrhythmias (torsades de pointes [66,75]).

Theoretical analysis of the drift mechanism in mathematical models has demonstrated that, similarly to spiral wave meandering, the drift is strongly dependent on the relationship between the wavelength of excitation and the critical radius of curvature. Depending on the $\lambda/r_p$ ratio, two different mechanisms of spiral wave drift can be distinguished. In one extreme situation when $\lambda$ is much larger than $r_p$, spiral wave rotation is mainly determined by the tissue refractory period. In this case, the drift of spiral waves is governed by spatial gradients in refractoriness [54,76]. Accordingly, in a medium with a stepwise heterogeneity, spiral waves drift along the borders separating regions with different refractoriness as shown in Fig. 11. With an increase in $r_p$ and/or a decrease in $\lambda$ the drift is influenced by a new component: in addition to drifting along the boundary, the spiral wave shifts into the region with the larger rotation periods. In the extreme case of $2\pi r_p \gg \lambda$, the spiral wave rotation is no longer affected by refractoriness and the drift is governed by local gradi-
ents in $r_p$ alone. In the case of a stepwise gradient in $r_p$, the drift is directed predominantly along the border separating regions with different $r_p$ [77,78], similar to the case of purely refractoriness-dependent drift.

4.4. Stationary spiral waves, anchoring effect

In many of the experimentally-induced tachycardias, the initial transition from a normal propagation pattern during a basic beat to a rotating pattern is caused by a premature wave propagating through tissue with heterogeneous refractoriness. According to the mechanism of spiral wave instability described above, such waves should be unstable due to refractoriness-dependent drift. However, mapping experiments have shown that such waves are often stable, rotating rigidly around a fixed core [41,73]. This discrepancy can be explained by the stabilizing effects of small localized discontinuities in tissue structure corresponding to unexcitable obstacles [71]. Fig. 12 shows an example of the anchoring of an initially drifting spiral wave [79]. Electrical activity in this case is represented in the form of a time–space plot (panel B) where signals from all measuring points were compressed into a single line which is displayed as a function of time (see Ref. [71] for details). In such diagrams, a propagating wave is represented by a narrow band and the location of the spiral wave core is equivalent to the point of band branching. Initially the spiral wave drifted in a downward direction as shown by the straight line. After 8 cycles of rotation, the spiral wave was anchored and became stationary. In most cases of stable rotation a band of connective tissue or a small branch of the coronary artery was identified as the site of anchoring.

![Image of anchoring of a drifting spiral wave](image)

Fig. 12. Anchoring of a drifting spiral wave. (A) Electrocardiographic recordings showing that premature stimulation ($S_2$) produced polymorphic arrhythmic activity followed by a transition to sustained monomorphic tachycardia. (B) Time–space plot of activation spread obtained from video-imaging of the fluorescence of a voltage-sensitive dye. In these plots activity from the whole image is projected onto a single direction (vertical axis) and displayed as a function of time. White bands show planar wave propagation while the branching of bands indicates the presence of a spiral wave induced by $S_2$ stimulation. The spiral wave drifted during the first 7 cycles and became stationary thereafter. Reproduced with permission [79].
The anchoring of spiral waves by unexcitable obstacles depends on the relation between the critical radius of curvature ($r_c$) and the size and shape of the obstacle. The smaller the $r_c$, the stronger the interaction between the spiral wave tip and the obstacle. As a result, anchoring of spiral waves is more likely to occur in conditions of relatively high excitability. In a medium with large $r_c$, excitation waves tend to detach from sharp obstacles (Fig. 7), and therefore obstacles are less likely to anchor spiral waves.

4.5. Spiral waves in 3 dimensions

Representation of myocardium as a 2-dimensional excitable medium is valid in a case of relatively thin myocardial tissue such as atrial muscle or a thin layer of epicardium surviving after myocardial infarction or the cryo-ablation procedure. However, in other cases such as a left ventricular wall, the 3-dimensional structure of cardiac muscle must be taken into account. Analogues of spiral waves in 3 dimensions are called ‘scroll waves’. A spiral wave core in a 2-dimensional medium corresponds to a filament in a 3-dimensional medium around which rotation of a scroll wave occurs. The simplest case of a scroll wave is an extension of a 2-dimensional spiral wave into the third dimension. Provided that cardiac tissue is homogeneous and the scroll filament is a straight line, the behavior of such a wave is equivalent to the behavior of a 2-dimensional spiral wave. The behavior of a scroll wave may become different when muscle properties vary with depth or when the scroll wave filament is bent or twisted. The specific 3-dimensional effects related to scroll wave rotation were quite extensively investigated in computer models. A brief review of these data has been published recently. One of the interesting effects specific to the 3-dimensional media is the unstable rotation of scroll waves with filaments which are bent or closed into rings. The mechanism for such instability relates to the 3-dimensional wavefront curvature, which may occur even if the rotation of a corresponding 2-dimensional spiral wave is stable and which may result in termination of the scroll wave. Another intriguing theoretical possibility is a persistent but disordered rotation of a scroll wave in homogenous 3-dimensional media resulting in a fibrillation-like electrical activity when projected to a surface of the medium. The experimental verification of these effects in cardiac muscle is still lacking due to the inability to map 3-dimensional activation spread with sufficiently high spatial resolution.

4.6. Concluding remarks

In summary, the experimental and theoretical data available at present clearly demonstrate the importance of the wavefront geometry in cardiac impulse propagation and in the behavior of spiral waves. Critical wavefront curvature contributes to a number of physiologically relevant effects: (1) conduction block at structures with abrupt tissue isthmuses or expansions; (2) dependence of stimulation strength on electrode size; (3) formation of an excitable gap during spiral wave rotation; (4) meandering, drift and anchoring of spiral waves. The application of theory based on continuous model to cardiac muscle in a quantitative way may have limitations, especially taking into account the fact that the estimated value of the critical radius is comparable to the dimensions of an individual cardiac cell. However, the fact that the value of critical curvature predicted from the theory and from the computer simulations in continuous models is close to the experimental estimates suggests that the continuous model is a good approximation, at least for healthy tissue. That may be different in aging tissue and in pathological conditions such as infarction and hypertrophy where larger discontinuities in axial resistivity may be present. Another limitation of the present approach is that it does not take into account the extracellular space. It is well established that impulse initiation and propagation in a tissue with a restricted extracellular space are strongly affected by differences in the anisotropic resistivity ratio between intra- and extracellular spaces as described by bi-domain models. The extent to which these factors affect critical curvature remains unknown.

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