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Monocyte chemoattractant protein-1 (MCP-1) in the kidney: does it more than simply attract monocytes?

Christiane Viedt¹ and Stephan R. Orth²¹Department of Internal Medicine, Division of Cardiology, Ruperto Carola University, Heidelberg, Germany and²Division of Nephrology and Hypertension, University Hospital of Berne (Inselspital), Switzerland

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MCP-1: a player in the pathogenesis of progression of renal diseases

There is an increasing body of evidence that the CC chemokine monocyte chemoattractant protein-1 (MCP-1) plays a major role in the pathogenesis of progression of renal failure. This is based on observations both in various animal models of renal damage and in different types of human renal disease (for review, see [1]). Locally produced MCP-1 seems to be

particularly involved in the initiation and progression of tubulointerstitial damage. The latter has been documented in experiments using transgenic mice with nephrotic serum-induced nephritis: compared with wild-type mice, MCP-1-deficient mice exhibit less tubulointerstitial lesions, but they exhibit no differences in glomerular lesions [2]. There is, however, evidence that MCP-1 also plays a role in the progression of glomerular lesions, since glomerular expression of MCP-1 correlates with the degree of renal damage in inflammatory [3] and non-inflammatory [4] models of glomerular injury. Furthermore, in humans with crescentic glomerulonephritis, MCP-1 is not only expressed in tubular epithelial cells and leukocytes infiltrating the tubulointerstitium, but also in crescents and parietal epithelium [5]. In experimental crescentic glomerulonephritis, administration of antibodies to MCP-1 decreases the extent of proteinuria, reduces glomerulosclerosis and improves renal dysfunction [6].

Of importance, a consistent increase of urinary MCP-1 concentration is found in patients or animals

Correspondence and offprint requests to: Stephan R. Orth, Department of Nephrology and Hypertension, University Hospital of Berne (Inselspital), CH-3010 Berne, Switzerland.
Email: stephan.orth@insel.ch

with a diseased kidney, and this correlates with the degree of urinary albumin/protein excretion and renal damage [7–10]. In addition to mesangial cells, endothelial cells and infiltrating mononuclear cells [11,12], tubular epithelial cells [12,13] seem to be the major source of MCP-1 in urine. The increased production of MCP-1 by tubular epithelial cells is due to: (i) stimulation by cytokines [14,15]; and (ii) exposure to urinary proteins [16]. Both are mediators of progressive renal damage [17].

Interestingly, therapy with an angiotensin-converting enzyme (ACE) inhibitor or an angiotensin II receptor type 1 (AT₁) antagonist is able to abrogate the renal expression of MCP-1 [18,19]. This may not only be due to reduction of proteinuria. Rat mesangial cells *in vitro* produce increasing amounts of MCP-1 in response to increasing external pressure, suggesting that MCP-1 may be a mediator of the adverse effects of intraglomerular hypertension [20]. The latter is an important factor in the genesis of progressive glomerular damage, which is attenuated by therapy with an ACE inhibitor or an AT₁ receptor antagonist. Taken together, MCP-1 seems to be of pivotal importance in the pathogenesis of progression of renal damage and failure.

To date, the classic perception of MCP-1 was that its main property is the attraction of monocytes/macrophages. These inflammatory cells in turn secrete cytokines and chemokines, fostering inflammation and fibrosis. Increased synthesis of cytokines and chemokines by infiltrating inflammatory cells and resident renal cells has been documented in various forms of

glomerulonephritis and tubulointerstitial nephritis. It has been suggested that they are important mediators of progressive renal failure *in vivo* [1]. Recent data [21,22] indicate that the role of MCP-1 goes beyond that of a simple chemoattractant protein.

MCP-1 induces an inflammatory activation of human tubular epithelial cells

Recent data from our group provide evidence that increased concentrations of MCP-1 in the diseased kidney may be an important factor contributing *per se* to increased production of cytokines and adhesion molecules. We demonstrated that MCP-1 specifically activates tubular epithelial cells *in vitro* [21], leading to a time- and dose-dependent increase in secretion of the proinflammatory cytokine interleukin-6 (IL-6) and expression of the intracellular adhesion molecule-1 (ICAM-1) via G_i-protein-, protein kinase C (PKC)- and intracellular Ca²⁺-dependent mechanisms (Figure 1). MCP-1 activated: (i) the transcription factor nuclear factor- κ B (NF- κ B), a transcription factor commonly involved in inflammatory and immune responses, and (ii) activating protein-1 (AP-1), a transcription factor involved in inflammatory and growth responses. Both NF- κ B and AP-1 were involved in the MCP-1-mediated induction of IL-6. In contrast to IL-6 release, MCP-1-induced ICAM-1 expression was predominantly dependent on NF- κ B activation (Figure 2). IL-6 has been suggested to contribute to

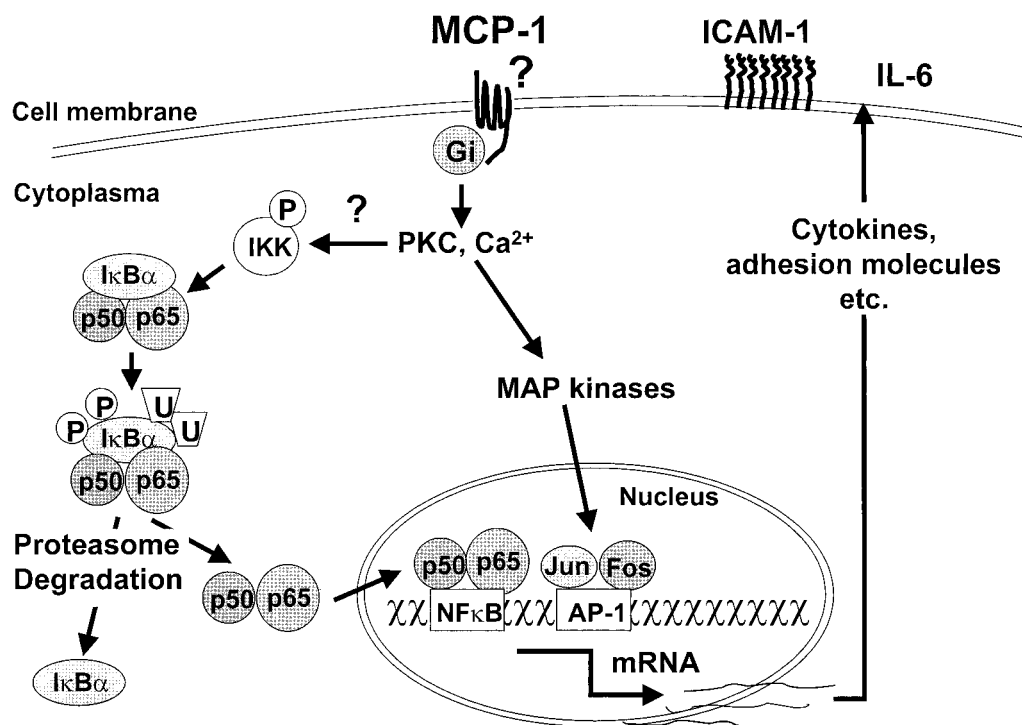


Fig. 1. Scheme of the intracellular signalling pathways of MCP-1 in a human tubular epithelial cell.

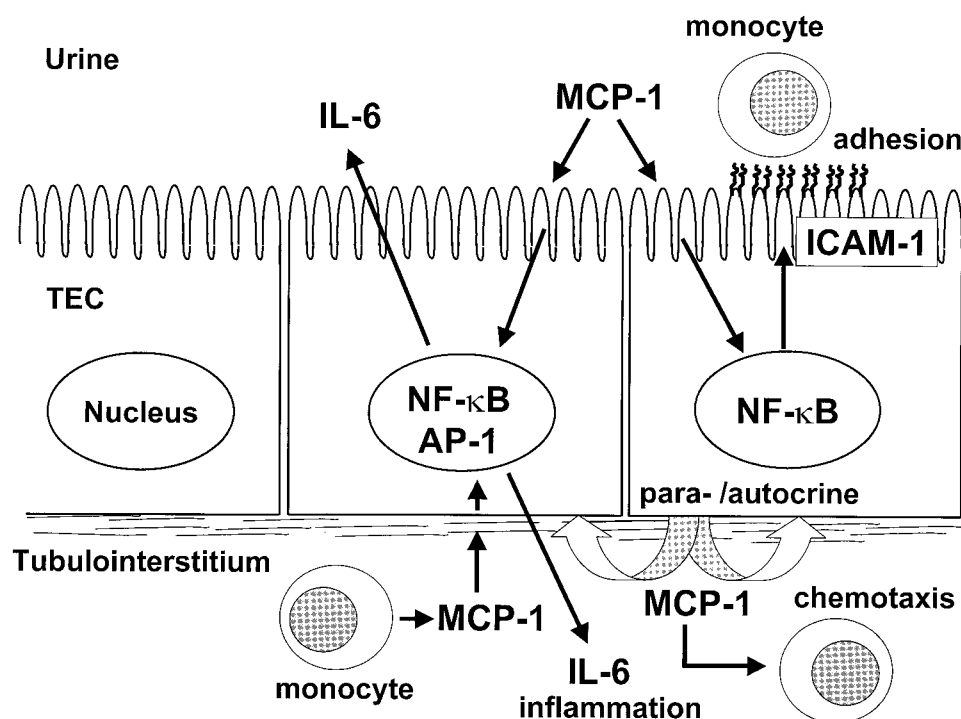


Fig. 2. Scheme of the direct and indirect effects of MCP-1 on human tubular epithelial cells (TECs).

progression of renal diseases [23,24] because its renal expression correlates with the degree of mesangial hyperproliferation, tubular atrophy, and the intensity of interstitial infiltrates [25,26]. De novo expression of the adhesion molecule ICAM-1 by tubular epithelial cells and increased expression by interstitial and glomerular cells has been observed in different forms of glomerulonephritis, tubulointerstitial inflammation and renal allograft rejection [27]. Thus, the observed effects of MCP-1 on tubular epithelial cells *in vitro* may be critical steps during progression *in vivo*.

MCP-1 does not only exhibit direct effects on tubular epithelial cells, but also on another cell type that plays a role in progressive renal damage, i.e. vascular smooth muscle cells [22]. Stimulation of vascular smooth muscle cells with MCP-1 induced proliferation and resulted in a concentration- and time-dependent release of IL-6. Similar to tubular epithelial cells, this effect was mediated via G_i-protein, PKC and NF-κB. MCP-1 also induced extracellular signal-regulated kinase (ERK), which, along with IL-6 release, was G_i-protein-dependent. MCP-1-induced, proliferation of vascular smooth muscle cells, which was ERK-dependent. MCP-1 stimulated the binding activity of NF-κB and of AP-1. NF-κB was involved in IL-6 release by MCP-1, whereas proliferation was dependent on AP-1, demonstrating that, as in tubular epithelial cells, MCP-1 induces differential activation of NF-κB and AP-1 in vascular smooth muscle cells. Thus, these latter data do not only propose a new mechanism for the proatherogenic effect of MCP-1 in the progression of cardiovascular disease, but also implicate MCP-1 as a factor leading to vascular damage in the diseased kidney.

Which receptor(s) is responsible for the mediation of effects of MCP-1 on human tubular epithelial cells?

Chemokines exert their effects through binding to G-protein-coupled receptors on the surface of leukocytes targeted for activation and migration. These receptors, once activated, trigger a set of cellular reactions that result in inositol triphosphate formation, intracellular calcium release, and PKC activation [28]. The classic MCP-1 receptors belong to the family of heptahelical, pertussis-sensitive, G-protein-coupled receptors [29]. The MCP-1 receptor on tubular epithelial cells appears to be coupled to G_i-protein activation [21]. These findings are in accordance with our observations in vascular smooth muscle cells [22], as well as data of others [29,30].

As in leukocytes, MCP-1-induced up-regulation of IL-6 synthesis and ICAM-1 expression by human tubular epithelial cells are dependent on PKC and intracellular Ca²⁺ (Ca_i²⁺) [21]. Similarly, Schecter *et al.* [30] demonstrated that the induction of tissue factor by MCP-1 in human vascular smooth muscle cells required Ca_i²⁺ mobilization and that it was PKC-dependent. Therefore, although currently unproven, it is likely that MCP-1-signalling is mediated by the classical PKC subgroups α, β or γ.

To date, in renal tissue of humans and experimental animals, expression of CCR1-5 mRNA transcripts were detected only in infiltrating mononuclear cells. Two MCP-1 receptors, generated by alternative splicing and designated as CCR2A and -B, have been cloned in human monocytes [31]. On the basis of our polymerase chain reaction and flow cytometry

analysis studies, the MCP-1 receptor on human tubular epithelial cells is distinct from these two receptors [21]. Furthermore, the MCP-1 receptor on human tubular epithelial cells is also distinct from the CCR1, CCR3, CCR4 and CCR5 receptors. Antibodies against CCR1–5 failed to inhibit the specific effects of MCP-1 on tubular epithelial cells. However, binding studies for MCP-1 revealed that cultured human tubular epithelial cells express a MCP-1 binding protein on the cell surface. The possibility that the effects of MCP-1 on tubular epithelial cells are mediated via an endosome-lysosomal pathway was ruled out [21]. Taken together, the above results implicate that it is likely that the MCP-1 receptor on human tubular epithelial cells is different from previously cloned CC chemokine receptors. The nature of the MCP-1 receptor(s) on human tubular epithelial cells, which seems to be G_i-protein coupled, remains to be determined (Figure 1).

Conclusion

MCP-1 can activate tubular epithelial cells directly *in vitro*. This action of MCP-1 on a resident renal cell had so far remained unknown, but had been described in cultured vascular smooth muscle cells [22,30]. The MCP-1-mediated activation of tubular epithelial cells is consistent with the notion that MCP-1 contributes to tubulointerstitial inflammation, which is a hallmark of progressive renal disease [17]. Importantly, tubulointerstitial rather than glomerular damage correlates best with the loss of renal function and the risk of progression to end-stage renal failure. Actually, there is a strict correlation between tubular atrophy, interstitial fibrosis, the extent of interstitial infiltrates and renal dysfunction [32]. The direct effects of MCP-1 on vascular smooth muscle cells [22] may not only be of importance in the progression of cardiovascular damage in general, but also in the progression of vascular lesions in the kidney, which could contribute further to progression of renal failure.

Previous studies focused on the role of MCP-1 in renal inflammation and its induction of inflammatory signals [1]. Our recent data suggest that MCP-1 is more than just a chemoattractant. Rather, MCP-1 can directly elicit an inflammatory response by inducing cytokine and adhesion molecule expression in the kidney. This is an important new mechanism in the pathogenesis of tubulointerstitial inflammation.

Note added in proof

Krüger *et al.* [33] recently reported that recipients of a renal allograft who are carriers of the MCP-1-2518 (G/G) polymorphism, i.e. a genotype characterized by increased production of MCP-1 by mononuclear cells, have a significantly reduced mean graft survival compared with the heterozygous (A/G) or wild-type (A/A) allele carriers. In contrast, carriers of the 64I

mutation of the CCR2 receptor, a genotype which presumably alters CCR2 expression or function, had no increase of mean renal allograft survival. These *in vivo* data support our *in vitro* findings [21], i.e. specific inflammatory activation of human tubular epithelial cells by MCP-1 despite the lack of CCR2 receptors on these cells.

References

1. Segerer S, Nelson PJ, Schlöndorff D. Chemokines, chemokine receptors, and renal disease: from basic science to pathophysiologic and therapeutic studies. *J Am Soc Nephrol* 2000; 11: 152–176
2. Tesch GH, Schwarting A, Kinoshita K *et al.* Monocyte chemoattractant protein-1 promotes macrophage-mediated tubular injury, but not glomerular injury, in nephrotoxic serum nephritis. *J Clin Invest* 1999; 103: 73–80
3. Panzer U, Thaïss F, Zahner G *et al.* Monocyte chemoattractant protein-1 and osteopontin differentially regulate monocytes recruitment in experimental glomerulonephritis. *Kidney Int* 2001; 59: 1762–1769
4. Taal MW, Zandi-Nejad K, Weening B *et al.* Proinflammatory gene expression and macrophage recruitment in the rat remnant kidney. *Kidney Int* 2000; 58: 1664–1676
5. Segerer S, Cui Y, Hudkins KL *et al.* Expression of the chemokine monocyte chemoattractant protein-1 and its receptor chemokine receptor 2 in human crescentic glomerulonephritis. *J Am Soc Nephrol* 2000; 11: 2231–2242
6. Wada T, Yokoyama H, Furuichi K *et al.* Intervention of crescentic glomerulonephritis by antibodies to monocyte chemoattractant and activating factor (MCAF/MCP-1). *FASEB J* 1996; 10: 1418–1425
7. Rovin BH, Doe N, Tan LC. Monocyte chemoattractant protein-1 levels in patients with glomerular disease. *Am J Kidney Dis* 1996; 27: 640–646
8. Wada T, Yokoyama H, Su SB *et al.* Monitoring urinary levels of monocyte chemotactic and activating factor reflects disease activity of lupus nephritis. *Kidney Int* 1996; 49: 761–767
9. Banba N, Nakamura T, Matsumura M *et al.* Possible relationship of monocyte chemoattractant protein-1 with diabetic nephropathy. *Kidney Int* 2000; 58: 684–690
10. Stephan M, Conrad S, Eggert T *et al.* Urinary concentration and tissue messenger RNA expression of monocyte chemoattractant protein-1 as an indicator of the degree of hydro-nephrotic atrophy in partial ureteral obstruction. *J Urol* 2002; 167: 1497–1502
11. Baggiolini M, Dewald B, Moser B. Interleukin-8 and related chemotactic cytokines: CXC and CC chemokines. *Adv Immunol* 1994; 55: 97–179
12. Wada T, Furuichi K, Sakai N *et al.* Up-regulation of monocyte chemoattractant protein-1 in tubulointerstitial lesions of human diabetic nephropathy. *Kidney Int* 2000; 58: 1492–1499
13. Mezzano SA, Barria M, Droguett MA *et al.* Tubular NF- κ B and AP-1 activation in human proteinuric renal disease. *Kidney Int* 2001; 60: 1366–1377
14. Prodjosudjadi W, Gerritsma JS, Klar-Mohamad N *et al.* Production and cytokine-mediated regulation of monocyte chemoattractant protein-1 by human proximal tubular epithelial cells. *Kidney Int* 1995; 48: 1477–1486
15. Gerritsma JS, van Kooten C, Gerritsen AF *et al.* Transforming growth factor-beta 1 regulates chemokine and complement production by human proximal tubular epithelial cells. *Kidney Int* 1998; 53: 609–616
16. Wang Y, Rangan GK, Tay YC *et al.* Induction of monocyte chemoattractant protein-1 by albumin is mediated by nuclear factor κ B in proximal tubule cells. *J Am Soc Nephrol* 1999; 10: 1204–1213
17. Remuzzi G, Bertani T. Pathophysiology of progressive nephropathies. *N Engl J Med* 1998; 339: 1448–1456

18. Donadelli R, Abbate M, Zanchi C *et al.* Protein traffic activates NF- κ B gene signaling and promotes MCP-1-dependent interstitial inflammation. *Am J Kidney Dis* 2000; 36: 1226–1241
19. Hilgers KF, Hartner A, Porst M *et al.* Monocyte chemoattractant protein-1 and macrophage infiltration in hypertensive kidney injury. *Kidney Int* 2000; 58: 2408–2419
20. Suda T, Osajima A, Tamura M *et al.* Pressure-induced expression of monocyte chemoattractant protein-1 through activation of MAP kinase. *Kidney Int* 2001; 60: 1705–1715
21. Viedt C, Dechend R, Fei J *et al.* MCP-1 induces inflammatory activation of human tubular epithelial cells: involvement of the transcription factors, nuclear factor- κ B and activating protein-1. *J Am Soc Nephrol* 2002; 13: 1534–1547
22. Viedt C, Vogel J, Athanasiou T *et al.* Monocyte chemoattractant protein-1 induces proliferation and interleukin-6 production in human smooth muscle cells by differential activation of nuclear factor- κ B and activator protein-1. *Arterioscler Thromb Vasc Biol* 2002; 22: 914–920
23. Fukatsu A, Matsuo S, Tamai H *et al.* Distribution of interleukin-6 in normal and diseased human kidney. *Lab Invest* 1991; 65: 61–66
24. Leonard M, Ryan MP, Watson AJ *et al.* Role of MAP kinase pathways in mediating IL-6 production in human primary mesangial and proximal tubular cells. *Kidney Int* 1999; 56: 1366–1377
25. Ranieri E, Gesualdo L, Petrarulo F *et al.* Urinary IL-6/EGF ratio: a useful prognostic marker for the progression of renal damage in IgA nephropathy. *Kidney Int* 1996; 50: 1990–2001
26. Ryffel B, Car BD, Gunn H *et al.* Interleukin-6 exacerbates glomerulonephritis in (NZB \times NZW) F1 mice. *Am J Pathol* 1994; 144: 927–937
27. Brady HR. Leukocyte adhesion molecules and kidney diseases. *Kidney Int* 1994; 45: 1285–1300
28. Lodi PJ, Garrett DS, Kuszewski J *et al.* High-resolution solution structure of the beta chemokine hMIP-1 beta by multidimensional NMR. *Science* 1994; 263: 1762–1767
29. Myers SJ, Wong LM, Charo IF. Signal transduction and ligand specificity of the human monocyte chemoattractant protein-1 receptor in transfected embryonic kidney cells. *J Biol Chem* 1995; 270: 5786–5792
30. Schechter AD, Rollins BJ, Zhang YJ *et al.* Tissue factor is induced by monocyte chemoattractant protein-1 in human aortic smooth muscle and THP-1 cells. *J Biol Chem* 1997; 272: 28568–28573
31. Charo IF, Myers SJ, Herman A *et al.* Molecular cloning and functional expression of two monocyte chemoattractant protein 1 receptors reveals alternative splicing of the carboxyl-terminal tails. *Proc Natl Acad Sci USA* 1994; 91: 2752–2756
32. Bohle A, Müller GA, Wehrmann M *et al.* Pathogenesis of chronic renal failure in the primary glomerulopathies, renal vasculopathies, and chronic interstitial nephritides. *Kidney Int Suppl* 1996; 54: S2–S9
33. Krüger B, Schröppel B, Ashkan R *et al.* A monocyte chemoattractant protein-1 (MCP-1) polymorphism and outcome after renal transplantation. *J Am Soc Nephrol* 2002; 13: 2585–2589