Integration of two herbivore-induced plant volatiles results in synergistic effects on plant defence and resistance

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Abstract
Plants can use induced volatiles to detect herbivore- and pathogen-attacked neighbors and prime their defenses. Several individual volatile priming cues have been identified, but whether plants are able to integrate multiple cues from stress-related volatile blends remains poorly understood. Here, we investigated how maize plants respond to two herbivore-induced volatile priming cues with complementary information content, the green leaf volatile (Z)-3-hexenyl acetate (HAC) and the aromatic volatile indole. In the absence of herbivory, HAC directly induced defence gene expression, whereas indole had no effect. Upon induction by simulated herbivory, both volatiles increased jasmonate signalling, defence gene expression, and defensive secondary metabolite production and increased plant resistance. Plant resistance to caterpillars was more strongly induced in dual volatile-exposed plants than plants exposed to single volatiles. Induced defence levels in dual volatile-exposed plants were significantly higher than predicted from the added effects of the individual volatiles, with the exception of induced plant volatile production, which showed no increase upon dual-exposure relative to single exposure. Thus, plants can integrate different volatile cues into strong and specific responses that promote herbivore defence induction and resistance. Integrating multiple volatiles may be beneficial, as volatile blends are more reliable indicators of future stress than single cues.

KEYWORDS
(Z)-3-hexenyl acetate, benzoazinoids, indole, induced resistance, insects, jasmonic acid, maize, plant defence, plant herbivore interactions, volatile communication

1 | INTRODUCTION

The capacity to perceive and respond to fluctuating environments is essential to all life on earth. As primary producers in terrestrial ecosystems, plants are constantly dealing with limiting resources, adverse abiotic conditions, competitors, pests, and pathogens (Cramer, Urano, Delrot, Pezzotti, & Shinozaki, 2011; Van Dam, 2009). By consequence, they have evolved systems to detect these stressors and respond to them appropriately (Cui, Tsuda, & Parker, 2015; Felton & Tumlinson, 2008; Hirayama & Shinozaki, 2010). Plants can, for example, perceive pathogens and herbivores directly via associated molecular patterns or indirectly via volatile cues from attacked neighbors (Bonaventure, 2012; Heil, 2014; Zipfel, 2014). The induction and priming of defence responses by herbivore- and pathogen-induced volatiles in particular is increasingly recognized as an important aspect of plant immunity and resistance (Baldwin, Halitschke, Paschold, von Dahl, & Preston, 2006;
Based on these considerations, we investigated how simultaneous exposure of maize plants to HAC and indole affects maize defenses. We first quantified the impact of HAC and indole individually on phytohormone production, defence gene expression, and defence metabolite accumulation in plants that were induced by simulated herbivory and measured the influence of these volatiles on plant resistance to herbivores. We then compared the effects of individual volatile exposure with the effects of simultaneous exposure to HAC and indole. We tested for synergistic effects of HAC and indole exposure by comparing the effects elicited by simultaneous exposure with the calculated additive effects of the individual exposures (Machado, Arce, McClure, Baldwin, & Erb, 2018). Our experiments reveal that maize plants integrate two different herbivore-induced volatiles into strong and specific defence signatures.

2 MATERIALS AND METHODS

2.1 Plants and herbivores

The maize (Z. mays) genotype B73 was used in this study. Maize seedlings were grown as previously described (Erb et al., 2011). Fourteen-day-old plants were used for all experiments. Spodoptera littoralis eggs were provided by the University of Neuchâtel and reared on artificial diet as previously described (Maag et al., 2014). Herbivore oral secretions were collected from third instar S. littoralis larvae, which had been feeding on maize leaves for 48 hr. Briefly, the S. littoralis larvae were held with a pair of lightweight forceps, and regurgitation was induced by gently pinching their heads with another pair of forceps. Oral secretions were collected using a micropipette and collected in Eppendorf tubes on ice. Oral secretions were stored at −80°C and diluted 1:1 in autoclaved Milli-Q water prior to use.

2.2 Volatile dispensers

Volatile dispensers were manufactured as previously described (Erb et al., 2011; von Merey et al., 2011). Dispensers consisted of 2-ml amber glass vials (11.6 × 32 mm−2; Sigma, St. Louis, USA) containing 20 mg of synthetic indole (>98%, GC, Sigma, St. Louis, USA) or 0.2-ml (Z)-3-hexenyl acetate (HAC, >98%, Sigma, St. Louis, USA). The vials were closed with open screw caps that contained a PTFE/rubber septum, which was pierced with a 2-µl micropette (Drummond, Millan SA, Switzerland). The vials were sealed with parafilm and wrapped in aluminium foil for heat protection and to avoid photodegradation. The dispensers release approximately 150 ng h−1 of indole and 70 ng h−1 of HAC, which corresponds to amounts typically emitted by herbivore-attacked maize plants (Erb et al., 2015, von Merey et al., 2011). Control dispensers were prepared the same way using empty glass vials. Dispensers were prepared 24 hr before the start of experiments.

2.3 Plant volatile exposure

To expose maize plants to synthetic indole and/or HAC, different sets of dispensers were individually introduced into 2-L glass vessels containing maize seedlings. The glass vessels were connected to a multiple air-delivery system via PTFE tubing. Purified air entered the glass...
vessels at a flow rate 0.3 L min⁻¹ and was released through additional openings. This set-up ensured sufficient ventilation to avoid the buildup of unnatural volatile concentrations while effectively isolating the headspaces of the different plants. The volatile exposure system was placed into a greenhouse cabin (26 ± 2°C; 14:10 hr, light [8 a.m.–10 p.m.]; dark: 55% relative humidity). Dispensers were added into the glass vessels in the evening (8 p.m.) before herbivore induction. The following treatment combinations were used in all experiments: Control (empty dispenser), HAC (HAC dispenser), indole (indole dispenser); HAC + indole (HAC dispenser and indole dispenser). Although HAC is released 1 hr earlier than indole upon simulated herbivory (Erb et al., 2015), both volatiles are released continuously and simultaneously from maize leaves that are attacked by real caterpillars (Erb et al., 2011). We therefore exposed maize plants to HAC and indole using the same timing. After 16 hr of exposure (at 10 a.m.), the plants were carefully removed from the glass vessels, placed on a table in the same greenhouse cabin, and induced as described in the next section.

2.4 | Plant induction by simulated herbivory

To test how indole and HAC influence herbivore-induced plant responses, the pre-exposed maize plants were induced by wounding two leaves over an area of ~0.5 cm² on both sides of the central vein with a razor blade, followed by the application of 10 μl of S. littoralis oral secretions. This treatment results in plant defence responses similar to real S. littoralis attack (Erb et al., 2009) and is referred to as "simulated herbivory" or "induction" throughout the rest of the manuscript. In three different experiments, leaves were either harvested at 0 min (no herbivore induction), 45 min, or 5 hr after simulated herbivory and then flash frozen and used to quantify phytohormones, expression of defence-related genes, benzoxazinoids, and volatiles. Whole maize leaves, excluding the damaged area, were harvested. All analyses within time points were performed on the same leaf samples.

2.5 | Gene expression analysis

The influence of volatile exposure on the herbivore-induced expression of signalling and defence genes was determined by quantitative real-time PCR (QRT-PCR, n = 5). On the basis of earlier studies, we measured the induction of hormone biosynthesis genes and hormonal signalling markers 45 min upon simulated herbivory (n = 5) and the induction of defence-related genes 5 hr upon simulated herbivory (n = 5; Seidl-Adams et al., 2015). In addition, we measured the effect of HAC and indole on all marker genes at the 0-min time point to evaluate direct induction. Maize leaves were ground to a fine powder under liquid nitrogen. Total RNA of 80-mg maize leaf powder was isolated using the GeneJET Plant RNA Purification Kit (Thermo Scientific, Waltham, MA, USA). Three hundred nanograms of total RNA of each sample were then reverse transcribed with the SuperScript® II Reverse Transcriptase (Invitrogen, Carlsbad, CA, USA). The QRT-PCR assay was performed on the LightCycler® 96 Instrument (Roche, Switzerland) using the KAPA SYBR FAST qPCR Master Mix (Kapa Biosystems, Wilmington, MA, USA). The maize actin gene ZmActin was used as an internal standard to normalize cDNA concentrations (Erb et al., 2009). The relative gene expression levels of the target genes were calculated using the 2^ΔΔCt method (Wong & Medrano, 2005). The primers of all tested genes are provided in Table S1.

2.6 | Phytohormone analysis

The influence of volatile exposure on herbivore-induced phytohormone levels were measured 45 min after induction by simulated herbivory (n = 5). This time point was selected on the basis of established hormone accumulation kinetics and volatile priming effects, both of which peak at 35–45 min after herbivore induction in maize (Engelberth et al., 2004; Erb et al., 2015). Jasmonic acid (JA), 12-oxophytodienoic acid (OPDA), JA-isoleucine (JA-Ile), abscisic acid (ABA), and salicylic acid (SA) were extracted from 80-mg frozen maize leaf powder in ethyl acetate spiked with isotopically labelled standards (1 ng for d₅-JA, d₅-ABA, d₅-SA, and d₁₃C₆-JA-Ile) and analysed by UHPLC–MS–MS as previously described (Glauser, Vallat, & Balmer, 2014).

2.7 | Benzoxazinoid analysis

To evaluate the influence of volatile exposure on benzoxazinoid defence metabolites, maize leaves were measured 5 hr after simulated herbivory (n = 5). Seventy milligrams of frozen maize leaf powder was extracted in 700 μl of acidified H₂O/MeOH (50:50 v/v; 0.1% formic acid) and then analysed with an Acquity UHPLC–MS system equipped with an electrospray source (Waters i-Class UHPLC-QDA, USA) using a previously established method (Robert et al., 2017). Compounds were separated on an Acquity BEH C18 column (2.1 × 100 mm i.d., 1.7-μm particle size). Water (0.1% formic acid) and acetonitrile (0.1% formic acid) were employed as mobile phases A and B. The elution profile was 0–9.65 min, 97–83.6% A in B; 9.65–13 min, 100% B; 13.1–15 min 97% A in B. The mobile phase flow rate was 0.4 ml/min. The column temperature was maintained at 40°C, and the injection volume was 5 μl; 2-(2-hydroxy-4,7-dimethoxy-1,4-benzoxazin-3-one)-β-d-glucopyranose (DIMBOA-Glc), 2-(2,4-dihydroxy-7-methoxy-1,4-benzoxazin-3-one)-β-d-glucopyranose (DIMBOA-Glc), and 2,4-dihydroxy-7-methoxy-1,4-benzoxazin-3-one (DIMBOA) were quantified in positive mode using single ion monitoring (SIM) at m/z 194 with cone voltage of 20 V; 2-(2,4-dihydroxy-6,7-dimethoxy-1,4-benzoxazin-3-one)-β-d-glucopyranose (DIM₇BOA-Glc), 2-(2,4-dihydroxy-1,4-benzoxazin-3-one)-β-d-glucopyranose (DIBOA-Glc), 2-(2-hydroxy-4,7,8-trimethoxy-1,4-benzoxazin-3-one)-β-d-glucopyranose (HD₈MOA-Glc), and 6-methoxy-benzoxazolin-2-one (MBOA) were acquired in negative scan mode (m/z 150–650) using a cone voltage of 10 V. The ESI capillary voltage was set to 0.8 kV. Theprobe temperature was maintained at 600°C. The detector gain was set to 1 and the sampling frequency was 5 Hz. Absolute quantities were determined using standard curves obtained from purified or synthetic DIMBOA, DIMBOA-Glc, HD₈MOA-Glc, and MBOA as described (Maag et al., 2015).

2.8 | Volatile analyses

To assess the impact of volatile exposure to herbivore-induced volatile production, maize leaves were analysed 5 hr upon simulated herbivory. At this time point, volatile priming significantly increases terpene release in maize (Engelberth et al., 2004; Erb et al., 2015). Frozen leaf powder was analysed with solid-phase microextraction-gas chromatography–mass spectrometry (SPME-GC–MS; n = 5). This approach
allows for the measurement of leaf volatile contents, which are highly correlated with volatile release rates in maize during daytime (Seidl-Adams et al., 2015). Fifty milligrams of leaf powder were placed in a 10-ml glass vial. An SPME fibre (100-μm polydimethylsiloxane coating; Supelco, USA) was then inserted into the vial and incubated at 60°C for 35 min. The incubated fibre was immediately analysed by GC-MS (Agilent 7820A GC interfaced with an Agilent 5977E MSD, USA) following previously established protocols (Huang et al., 2016). Major volatile compounds were identified by comparing mass spectra with the NIST Mass Spectral Library (USA) as well as authentic standards, and the abundance of each compound was determined by integrating individual peak areas.

2.9 | Herbivore resistance assays

To quantify the impact of volatile exposure on herbivore growth and plant resistance, individual preweighed second instar *S. littoralis* larvae were introduced into cylindrical mesh cages (1-cm height and 5-cm diameter) and then clipped onto the leaves of individual maize plants that were previously exposed to different volatile combinations (*n* = 10). The position of the cages was moved every day to provide sufficient food supply for the larvae. Larval weight was recorded 4 days after the start of the experiment. For damage quantification, the remaining leaves were scanned, and the removed leaf area was quantified with Digimizer 4.6.1 (Digimizer).

2.10 | Statistical analyses

Gene expression, phytohormone, benzoxazinoid, volatile, larval growth, and leaf damage data were analysed by analysis of variance (ANOVA) followed by pairwise or multiple comparisons of least squares means (LSMeans), which were corrected using the false
discovery rate (FDR) method (Benjamini & Hochberg, 1995). Normality was verified by inspecting residuals, and homogeneity of variance was tested through Shapiro–Wilks’s tests using the “plotresid” function of the R package "RVAideMemoire" (Herve, 2015). Datasets that did not fit assumptions were loge-transformed to meet the requirements of equal variance and normality. Potential synergism was evaluated using a previously described approach (Machado et al., 2018). Briefly, we calculated additive effects by randomly pairing replicates of individual volatile treatments (an indole-treated plant [I] and a HAC-treated plant [H]) for each random pair, we calculated theoretical additive values (A) for the different defence parameters using the following formula: A = I + H - C, where C corresponds to the average level of nonexposed control plants. The calculated additive values were then compared with the measured treatment values of the double volatile treatment using Student’s t tests. Cases in which the measured level of the double volatile treatment was significantly greater than the calculated additive level were classified as synergistic. Principal component analysis (PCA) was furthermore employed to compare the response profiles at 0 min (defence and signalling gene expression), 45 min (signalling gene expression, phytohormones), and 5 hr (defence gene expression, benzoxazinoids, volatiles) in an integrated manner (Chapman, Schenk, Kazan, & Manners, 2002). Raw data were scaled with the “scale” function in R, and PCAs were then performed using the “MVA” function of the “RVAideMemoire” package and the “rda” function of the “vegan” package (Herve, 2015; Oksanen et al., 2013). Permutational ANOVAs were then conducted using the “adonis” function of the “vegan” package with 999 permutations. All statistical analyses were conducted with R 3.2.2 (R Foundation for Statistical Computing, Vienna, Austria) using the packages “car,” “lsmeans,” “vegan,” and “RVAideMemoire” (Bates, Machler, Bolker, & Walker, 2015; Herve, 2015; Lenth, 2016; Oksanen et al., 2013).

2.11 Accession numbers and data availability

The sequence data of maize genes can be found in the GenBank/EMBL database under the following accession numbers: ZmActin (MZEACT1G), ZmLOX10 (DQ335768), ZmAOS (AY488135), ZmPR1 (U82200), ZmPR5 (U82201), ZmMPI (X78988), ZmSerPIN (BM382058), ZmCyst (CK371502), ZmRIP2 (L26305), ZmCYP92C5 (ACG28049), ZmTPS2 (AY928081), ZmTPS3 (AY928082), ZmTPS10 (AY928078), ZmIGL (AF271383), ZmBx10 (GRMZM2G311036), ZmBx11 (GRMZM2G336824), and ZmBx14 (GRMZM2G127418). All relevant data supporting the findings of this study can be downloaded from the Dryad repository (doi:10.5061/dryad.f21g54g).

3 RESULTS

3.1 Pre-exposure to HAC and indole synergistically increases JA and ABA biosynthesis in induced plants

As reported before (Engelberth et al., 2004; Erb et al., 2015), exposure to HAC and indole individually increased the production of jasmonates, including OPDA, JA, and JA-Ile as well as ABA 45 min after induction by simulated herbivory. Simultaneous exposure to HAC and indole increased jasmonate and ABA levels beyond their calculated additive levels (Figure 1a–c). SA levels were not changed by volatile exposure (Figure 1e). Similar to jasmonates themselves, transcript levels of the JA related genes ZmAOS10 and ZmAOS (Christensen et al., 2013; Engelberth, Seidl-Adams, Schultz, & Tumlinson, 2007) were enhanced by exposure to HAC and indole individually and synergistically increased by simultaneous HAC and indole exposure (Figure 1f). The expression levels of ZmPR1 and ZmPR5 were not changed by volatile exposure (Figure 1f; Morris et al., 1998). Thus,
HAC and indole enhance ABA and JA biosynthesis in induced plants in a synergistic manner.

3.2 Pre-exposure to HAC and indole specifically and synergistically increases the expression of defence genes in induced plants

To further explore the interactions of HAC and indole in regulating plant defence responses, we measured the expression levels of four defensive marker genes in volatile pre-exposed plants 5 h after induction by simulated herbivory: the putative proteinase inhibitors ZmMPI (Farag et al., 2005; Tamayo, Rufat, Bravo, & San Segundo, 2000), ZmSerPIN and ZmCyst (Erb et al., 2011; Ton et al., 2007), and the insecticidal ribosome-inactivating protein ZmRIP2 (Chuang et al., 2014). Exposure to HAC and indole individually increased the expression of ZmMPI, ZmSerPIN, and ZmRIP2 (Figure 2a-c). ZmCyst expression was increased by HAC, but not by indole (Figure 2d). Simultaneous exposure to HAC and indole increased the expression of ZmMPI, ZmSerPIN, and ZmRIP2 in a synergistic manner (Figure 2a-c). By contrast, ZmCyst expression was not further increased by the double volatile treatment in comparison with individual HAC exposure (Figure 2d). Thus, HAC and indole differentially regulate the expression of defence marker genes in induced plants, with combined effects ranging from neutral to synergistic.

3.3 Pre-exposure to HAC and indole synergistically regulates BX biosynthesis in induced plants

Benzoxazinoids (BXs) are important secondary metabolites, which strongly respond to herbivore attack (Glauser et al., 2011) and protect cereals against herbivores (Wouters, Blanchette, Gershenzon, & Vassao, 2016). Five hours after induction by simulated herbivory, pre-exposure to HAC and indole individually did not significantly change the production of BXs. By contrast, simultaneous exposure to HAC and indole increased the production of HDMBOA-Glc, DIBOA-Glc, and HDMBOA-Glc compared with nonexposed plants (Figure 3a). HDMBOA-Glc and HDMBOA-Glc were regulated synergistically by the two volatiles, whereas the effect on DIBOA-Glc was not significantly different from the calculated additive effect. The expression levels of the O-methyltransferases that produce
HDMOA-Glc (ZmBx10/11, (Mehlis et al., 2013)) and HDM2BOA-Glc (ZmBx14; Handrick et al., 2016) followed the same pattern: The expression of both genes was not further increased by individual HAC or indole exposure in induced plants but strongly responded to simultaneous HAC and indole exposure (Figure 3b,c). Therefore, HAC and indole synergistically regulate the production of BXs in induced plants.

### 3.4 Pre-exposure to HAC and indole does not synergistically regulate volatile production in induced plants

Exposure of plants to both HAC and indole individually can prime herbivore-induced terpene emissions (Engelberth et al., 2004; Erb et al., 2015). As terpene biosynthesis in maize are regulated by jasmonates (Schmelz, Alborn, Banchio, & Tumlinson, 2003; Schmelz, Alborn, & Tumlinson, 2003), we expected additive or synergistic effects of simultaneous HAC and indole exposure on volatile production similar to the defence marker genes and BXs. Exposure of maize plants to HAC and indole individually followed by simulated herbivory increased the production of linalool, (3E)-4,8-dimethyl-1,3,7-nonatriene (DMNT), (E)-α-bergamotene, (E)-α-farnesene and indole 5 hr after induction (Figure 4a-e). Simultaneous exposure to HAC and indole did not further increase volatile production. For indole, we even detected significantly lower amounts in plants exposed to both volatiles than would be expected in an additive scenario. Transcript levels of genes involved in terpene synthesis, including ZmCYP92C5, ZmTPS2, ZmTPS3, ZmTPS10, and ZmIGL (Frey et al., 2000; Richter et al., 2016; Schnee et al., 2006), showed a similar pattern (Figure 4f).

**FIGURE 4** Simultaneous pre-exposure to (Z)-3-hexenyl acetate (HAC) and indole does not synergistically regulate volatile production in induced maize plants. (a)-(e) Average relative amounts (peak areas) of linalool (a), (3E)-4,8-dimethyl-1,3,7-nonatriene (DMNT, b), (E)-α-bergamotene (c), (E)-α-farnesene (d), and indole (e) in plants that were pre-exposed to HAC, indole, or both volatiles simultaneously (HAC + Indole) and induced by simulated herbivory (+SE, n = 5). (f) Average transcript levels of ZmCYP92C5, ZmTPS2, ZmTPS3, ZmTPS10, and ZmIGL (+SE, n = 5). FW, fresh weight. n.s., not significant. Treat., treatment. Gene expression is shown relative to the expression level of the control treatment. P values of one-way analyses of variance (ANOVA) are shown (*P < 0.05, **P < 0.01, ***P < 0.001). Dashed lines indicate calculated additive effects of single volatile exposures. Letters indicate significant differences between different volatile exposure treatments (P < 0.05, one-way ANOVA followed by multiple comparisons through FDR-corrected LSMeans). Stars indicate a significant difference between the double exposure treatment and the calculated additive effect of both single treatments (*P < 0.05, Student’s t tests)
3.5 | Pre-exposure to HAC and indole increases herbivore resistance of maize in an additive manner

To investigate how HAC and indole pre-exposure influences herbivore performance and plant resistance, we measured *S. littoralis* growth and damage on volatile-exposed plants. Pre-exposure to HAC or indole individually reduced *S. littoralis* growth and plant damage (Figure 5). Simultaneous pre-exposure to HAC and indole further increased this effect, with reductions of larval growth and damage attaining 40% (Figure 5). Thus, HAC and indole enhance plant resistance against herbivores in an additive manner.

3.6 | Pre-exposure of HAC, but not indole, directly induces defence gene expression

To investigate whether the observed synergistic effects on plant defenses are due to priming or direct induction by volatile exposure, we measured the expression of the different defense marker genes upon HAC and indole exposure without further induction. HAC pre-exposure significantly increased the expression of the tested jasmonate, volatile, and benzoxazinoid biosynthesis genes as well as other defence genes (Figure 6). By contrast, indole pre-exposure did not directly induce any defense marker genes (Figure 6). Expression of the SA-responsive genes *ZmPR1* and *ZmPR5* was not changed by HAC or indole exposure (Figure 6). Simultaneous exposure to HAC and indole resulted in similar gene expression patterns as HAC alone, with the exception of the DMNT biosynthesis gene *ZmCYP92C5*, whose expression was synergistically enhanced by double exposure (Figure 6). Thus, HAC, but not indole, directly induces a broad spectrum of defence genes. Furthermore, most of the synergistic effects observed upon double exposure after induction by simulated herbivory (Figures 1–4) are likely due to priming rather than direct induction by HAC and indole.

3.7 | Individual and simultaneous exposure to HAC and indole results in specific defence signatures

To evaluate whether HAC and indole double exposure results in specific defence signatures, we performed PCAs for the individual time points. Permutational multivariate analysis revealed clear treatment effects at all time points (Figure 7). Without induction by simulated herbivory, HAC pre-exposure resulted in a defence signature that was clearly separated from control and indole pre-exposure (Figure 7a). Double-exposure clustered together with HAC pre-exposure (Figure 7a), reflecting the fact that indole pretreatment does not affect HAC-induced signature changes. By contrast, 45 min and 5 hr after induction by simulated herbivory, a clear separation between controls, individual volatile exposures, and double volatile exposure was observed (Figure 7b,c). At 45 min, the treatments were predominantly separated along PC axis 1 (Figure 7b). The major vectors contributing to treatment separation were related to jasmonate and abscisic acid biosynthesis. No clear separation was observed between individual HAC and indole exposure, but double exposure was clearly separated from single exposure. The profiles at 5 hr showed a similar structure, with both PC axes 1 and 2 contributing to the separation of individual volatile exposures and double exposure (Figure 7c). In this case, the vectors contributing most to the separation of double and single exposure were benzoxazinoids and a subset of defence marker genes. Thus, double exposure to HAC and indole leads to distinct defence signatures.

4 | DISCUSSION

Plants can perceive various environmental cues, but whether they can integrate multiple cues to regulate defence responses is poorly understood. The present study shows that simultaneous exposure of maize plants to two different herbivore-induced volatile cues results in specific defence signatures, with most defence markers responding in an additive or synergistic fashion to double exposure. Below, we discuss the underlying mechanisms and ecological context of this phenomenon.

Maize plants that are induced by simulated herbivory responded to simultaneous HAC and indole exposure by markedly increasing their defence responses compared with nonexposed and single volatile-exposed plants. In particular, HAC and indole synergistically enhanced the deployment of jasmonates, the expression of defence...
marker genes, and the production of defensive secondary metabolites in plants. Dual exposure also markedly suppressed herbivore growth and plant damage. These patterns are unlikely due to direct induction, as HAC, but not indole, directly increased defence gene expression. Instead, HAC and indole primed maize plants together to respond more strongly upon induction. A likely mechanism to explain this pattern is convergence of HAC and indole in early defence signalling. Both HAC and indole act upstream of the jasmonate signalling pathway, possibly by priming the activity of MAP kinases (Ye, Glauer, Lou, Erb, & Hu, 2018) and/or WRKY transcription factors (Engelberth, Contreras, Dalvi, Li, & Engelberth, 2013; Mirabella et al., 2015). As predicted from single exposure responses and cannot be explained by signal convergence and amplification alone. In Arabidopsis thaliana, the GLV (E)-2-hexenal regulates GABA signalling and the redox status of mitochondria (Ameye et al., 2017; Mirabella et al., 2015; Scala et al., 2017). Indole on the other hand has been shown to inhibit auxin signalling in A. thaliana roots at high doses (Bailly et al., 2014). Thus, it is well possible that HAC and indole fine-tune defence expression through signalling crosstalk (Machado et al., 2016; Pieterse, Leon Reyes, Van der Ent, & Van Wees, 2009), leading to specific patterns of defence priming. Further experiments aiming at understanding the
early signalling events that are directly elicited by indole and HAC and how they affect hormonal signalling networks from a more holistic perspective may help to test this hypothesis.

From an ecological point of view, the integration of HAC and indole into stronger defence priming may allow plants to adjust their defence investment according to the reliability of the perceived cues. As GLVs can be emitted in response to many stresses, including for instance mechanical injury in the absence of herbivory (Ebel et al., 1995; Scala et al., 2013), they cannot be used as reliable cues by plants to anticipate herbivory. The same is true for indole alone, which can emanate from various environmental sources (Bailly et al., 2014; Stamm et al., 2005) but is emitted from leaves in much greater quantities upon contact with herbivore-elicitors than wounding alone (Frey et al., 2000; Zhuang et al., 2012). The simultaneous presence of indole and GLVs on the other hand may be a relatively robust predictor of herbivore attack due to the complementary nature of their information contents. Given that priming can be costly (van Hulten, Pelser, van Loon, Pieterse, & Ton, 2006), adjusting the magnitude of priming according to the reliability of the perceived cues may be beneficial. Especially when the reliability of individual volatile cues is low, the ability to integrate multiple volatile cues may confer important advantages to plants. However, it is important to point out that the integration of multiple volatile cues is not always necessary to obtain reliable information from the environment. Insect pheromones, for instance, can be fairly specific and may be sufficient to reliably indicate the presence of a herbivore. In line with this argument, Solidago altissima plants respond similarly to the exposure to a single pheromone component of the goldenrod gall fly as to the full volatile blend of the herbivore (Helms et al., 2017).

Double exposure to HAC and indole enhanced direct defenses but had no clear effect on the emission of induced volatiles, which are often viewed as indirect defenses that attract natural enemies (Turlings & Erb, 2018). Recent work in tomato furthermore demonstrates that changes in light quality leading to phytochrome B inactivation shifts tomato defenses from direct to volatile-mediated indirect defenses (Cortés, Weldegergis, Boccalandro, Dicke, & Ballaré, 2016). Thus, plants seem to be able to integrate various environmental cues to regulate their relative investment into direct and indirect defenses. Regarding the results of the present study however, we would like to remain cautious with our interpretation, as the effects of the observed patterns on indirect defenses have not been quantified, and the ecological interpretation of defence responses of a domesticated plant warrants caution due to possible pleiotropic effects of domestication. Nevertheless, exploring if and how the composition of volatile cues influences the relative investment of plants into direct and indirect defenses is an exciting prospect of this work.

5 | CONCLUSIONS

Plants perceive a variety of volatiles from the environment. Our work lends support to the concept that plants are also able to integrate multiple volatile cues into specific, and possibly adaptive, defence responses. Understanding the mechanisms and ecological factors that shape the evolution of signal integration will be important to improve our understanding of plant responses to complex volatile blends.
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**AUTHOR CONTRIBUTIONS**

L. H. conceived, designed, performed, and analysed experiments and wrote the first draft of the manuscript. M. Y. designed, performed, and analysed experiments. M. E. acquired funding, conceived the project, designed, supervised, and analysed experiments, and wrote the first draft of the manuscript. All authors contributed to the final version of the paper.

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**SUPPORTING INFORMATION**

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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