REVIEW



Orphan crops: their importance and the urgency of improvement

Zerihun Tadele¹

Received: 8 February 2019 / Accepted: 5 June 2019 / Published online: 12 June 2019 © Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

Main conclusion Due to significant contributions of orphan crops in the economy of the developing world, scientific studies need to be promoted on these little researched but vital crops of smallholder farmers and consumers.

Abstract Food security is the main challenge in the developing world, particularly in the least developed countries. Orphan crops play a vital role in the food security and livelihood of resource-poor farmers and consumers in these countries. Like major crops, there are members of all food types-cereals, legumes, vegetables and root and tuber crops, that are considered to be orphan crops. Despite their huge importance for present and future agriculture, orphan crops have generally received little attention by the global scientific community. Due to this, they produce inferior yields in terms of both quantity and quality. The major bottlenecks affecting the productivity of these crops are little or no selection of improved genetic traits, extreme environmental conditions and unfavorable policy. However, some orphan crops have recently received the attention of the global and national scientific community where advanced research and development initiatives have been launched. These initiatives which implement a variety of genetic and genomic tools targeted major constraints affecting productivity and/or nutritional quality of orphan crops. In this paper, some of these initiatives are briefly described. Here, I provide key suggestions to relevant stakeholders regarding improvement of orphan crops. Concerted efforts are urgently needed to advance the research and development of both the major and orphan crops so that food security will be achieved and ultimately the livelihood of the population will be improved.

Keywords Crops for the future · Disadvantaged crops · Food security · Minor crops · Orphan crops · Understudied crops · Underutilized crops

CLAT

International Contar for Transcol

Development

	CIAT	International Center for Tropical
frican Orphan Crops Consortium		Agriculture
orld Vegetable Center	CIMMYT	International Maize and Improvement
iotechnology and Biological Sciences		Center
esearch Council	CIP	International Potato Center
eijing Genomics Institute	CRISPR/Cas9	Clustered regularly interspaced short
ill and Melinda Gates Foundation		palindromic repeats/CRISPR-associated 9
ollaborative Crop Research Program,	ECA	Economic Commission for Africa
cKnight Foundation	FAO	Food and Agriculture Organization
luster of Excellence in Plant Science	GBS	Genotyping by sequencing
rops for Future	GWAS	Genome wide association study
onsultative Group for International	ICARDA	International Center for Agricultural
gricultural Research		Research in the Dry Areas
	ICRAF	World Agroforestry Centre
	ICRISAT	International Crops Research Institute for
		the Semi-Arid Tropics
a unika ak	IFAD	International Fund for Agricultural
	orld Vegetable Center otechnology and Biological Sciences search Council bijing Genomics Institute II and Melinda Gates Foundation ollaborative Crop Research Program, cKnight Foundation uster of Excellence in Plant Science ops for Future onsultative Group for International gricultural Research	Trican Orphan Crops ConsortiumCIMMYTorld Vegetable CenterCIMMYTotechnology and Biological SciencesCIPessearch CouncilCIPetijing Genomics InstituteCRISPR/Cas9II and Melinda Gates FoundationECAoblaborative Crop Research Program, cKnight FoundationECAouster of Excellence in Plant ScienceGBSops for FutureGWASonsultative Group for International gricultural ResearchICRAFICRAFICRAFIFADIFAD

Zerihun Tadele zerihun.tadele@ips.unibe.ch

1 Institute of Plant Sciences, University of Bern, Bern, Switzerland

International Institute of Tropical Agriculture
Least Developed Countries
New Partnership for Africa's
Development
National Research Council of the USA
β- <i>N</i> -Oxalyl-L-α, β-diaminopropanoic acid
Public-private partnership
Sustainable Agriculture for Sub-Saharan Africa
Syngenta Foundation for Sustainable Agriculture
Targeting induced local lesions in genomes

Introduction

Food security is the main challenge to the developing world, especially in the Least Developed Countries (LDCs). Sixtynine percent of the 45 total LDCs are in Africa. Among 52 countries with food deficits, 35 are in Africa (67%) while 11 are in Asia (FAOSTAT 2018). Due to the inability of these nations to grow enough food for their people, nations with large food deficits are obliged to import large quantities of agricultural products. For instance, in 2016, African nations imported agricultural products worth 71 billion USD out of which 29% were spent on importing cereal crops. This indicates the severity of food insecurity in Africa compared to other parts of the world.

Food security is based on four pillars: (1) food availability which refers to the availability of sufficient quantities; (2) food access which refers to having sufficient resources for acquiring appropriate food for a nutritious diet; (3) stability which refers to availability and accessibility of quality food at all times; and (4) utilization which refers to the appropriate use of food based knowledge of basic nutrition and care, as well as adequate water and sanitation (FAO 2006). Under all the four pillars which focus on the availability, access, stability and utilization of food, Africa is in the worst situation followed by Asia. While food availability is defined by the adequacy of dietary energy supply, food access is a measure of the prevalence of undernourishment. A parameter which represents food stability is the variability in per capita food production. Similarly, a single parameter used to represent food utilization is the proportion of children under 5 years of age who are stunted. As expected, there were large variabilities within a continent for these food security parameters. Although the average undernourishment for Africa in 2016 was 20.4%, the values ranged from a high of 31.4% in East Africa to a low of 8.4% in Northern and Southern Africa (FAOSTAT 2018).

Food security could be improved by focusing on both the major and minor (also known as orphan) crops of the world (Fahey 1998). Orphan crops play an important role in the economy of the developing world since they provide nutrition to a large number of people and provide income for small-scale farmers. The need to focus on orphan crops to meetthe demand for food has been emphasized (Raheem 2011; Ejeta 2010).

The objective of this review is to provide highlights on the importance and constraints related to orphan crops as well as highlights on initiatives associated with the improvement of these disadvantaged crops. Suggestions are made for future research and development of orphan crops.

Types and significance of orphan crops

Orphan crops are also known as underutilized crops (Dawson and Jaenicke 2006), lost crops (NRC 1996, 2006, 2008), neglected crops (Bermejo and León 1994) or crops for the future (CFF 2019) (Table 1). According to Crops for the Future (CFF 2019), the diverse names given to these crops reflect the following characteristics: 'neglected' (by science and development), 'orphan' (without champions or crop experts), 'minor' (relative to global crops), 'promising' (for emerging markets, or because of previously unrecognized value traits), 'niche' (of marginal importance in production systems and economies), and 'traditional' (used for centuries or even millennia).

Orphan crops belong to the major groups of crops, which include cereals, legumes, and fruit as well as root crops (Table 2). Selected orphan crops are briefly described below.

Cereals

Cereals are rich sources of nutrients for both humans and animals. Among these, millets are a rich source of vitamin, calcium, iron, potassium, magnesium and zinc (Léder 2004). Millets represent different types of millets which include Barnyard millet (Echinochloa crus-galli), finger millet (Eleusine coracana), foxtail millet (Setaria italica), kodo millet (Paspalum scrobiculatum), little millet (Panicum sumatrense), pearl millet (Pennisetum glaucum) and prosomillet (Panicum miliaceum) as well as tef (Eragrostis tef) and fonio (Digitaria sp.). They are annually cultivated on a total of 32.2 million hectares of land mainly in India, Niger, Sudan, Nigeria and Chad (FAOSTAT 2018). Among major types of millets, pearl millet is dominantly cultivated as a food crop due to its extreme tolerance to moisture deficit (Kholová et al. 2010). Similarly, finger millet is extensively cultivated in the semi-arid regions of Asia and Africa due to its adaptation to unfavorable climatic and soil conditions especially drought (Williams and Haq 2000; Dawson and

Table 1 Different names given to the orphan crops, which refer to the current status or future potential of these crops

Name	Name refer to	Reference
Abandoned crops	Neglect from research and development	Padulosi (2017)
Alternative crops	Options under extreme environment	Padulosi (2017)
Crops for the future	High potential in future agriculture	CFF (2019)
Disadvantaged crops	Unfavored by research and development	Massawe et al. (2015)
Forgotten crops	Little focus on their research	FAO (2017); Pearce (2013)
Future smart food	High contribution to future food security	Li and Siddique (2018)
Indigenous crops	Native crops	Kamadi (2014)
Life-style crops	Health-related benefits	Cannarozzi et al. (2018a)
Local crops	With domestic importance	Padulosi (2017)
Lost crops	Genetic erosion of the germplasm	NRC (2006, 2008, 1996)
Minor crops	Relative to global (major crops) crops	Padulosi (2017)
Neglected crops	Little focus to science and development	Bermejo and León (1994)
Niche crops	Marginal importance in production systems	Padulosi (2017)
Orphan crops	Without champions or crop experts	AOCC (2018); Tadele (2009a, b)
Promising crops	For emerging markets	FAO (2017)
Superfood	Due to nutritional- and health-related benefits	Provost and Jobson (2014)
Traditional crops	Used for centuries or even millennia	Padulosi (2017)
Underdeveloped crops	With limited investment	Padulosi (2017)
Understudied crops	Due to little scientific research	Tadele and Assefa (2012)
Underused crops	Due to little scientific advancement	EcoBusiness (2015)
Underutilized crops	Due to little research	Dawson and Jaenicke (2006); Massawe et al. (2015)
Wonder plants	Superiority in nutrition- and health-related benefits	EcoBusiness (2015)

Jaenicke 2006). Finger millet is also a popular food among diabetic patients because of its low glycemic index and slow release of glucose to blood due to slow digestion associated with high fiber content (Chandrashekar 2010). Another small cereal, which is extensively cultivated in the Horn of Africa due to its tolerance to abiotic stresses especially to poorly drained soil, is tef. Tef has recently received attention and is considered as a life-style crop due to the lack of gluten, the cause for celiac disease (Spaenij-Dekking et al. 2005). The panicles of selected orphan cereal crops are shown in Fig. 1.

Pseudocereals

Pseudocereals refer to the group of crops, which do not belong to the Grass Family due to their two cotyledons unlike grasses with a single cotyledon. However, pseudocereals show their close relationship to the 'true cereals' by their nutritional content especially with regard to the carbohydrate composition. Amaranths (*Amaranthus spp.*), buckwheat (*Fagopyrum esculentum*) and quinoa (*Chenopodium quinoa*) are the major representatives of this category. While buckwheat is mainly cultivated in several Asian and European countries, quinoa is extensively cultivated in three South American countries, namely Bolivia, Peru and Ecuador. In addition to being free of gluten, pseudocereals have a number of health-related benefits, which include reduction in oxidative stress, anti-cancer, anti-diabetic, anti-inflammatory, anti-hypertensive and prevention of cardiovascular diseases (Mir et al. 2018).

Legumes or pulses

Legumes are the major protein source for consumers. Due to their ability to fix atmospheric nitrogen and convert it to ammonium, legumes contribute towards soil improvement. While Bambara groundnut (Vigna subterranean) and cowpea (Vigna unguiculata) are extensively cultivated in Africa, horse gram (Macrotyloma uniflorum) is mainly cultivated in Asia (FAOSTAT 2018). The seeds of Bambara groundnut are known as a complete food since they contain adequate quantities of protein (19%), carbohydrate (63%), and fat (6.5%) (NRC 2006). Cowpea is annually grown on 12.3 million hectares mainly in three western African countries, namely Niger, Nigeria and Burkina Faso. The crop is tolerant to drought and heat, and it also performs better than many other crops on poor soil conditions (Sanginga et al. 2000). Similarly, grass pea (Lathyrus sativus) which is cultivated in Asia, Africa and Europe is extremely tolerant to drought. Due to this property, the crop is considered to

Beneficial traits R Erra Resistance to biotic and abiotic stresses L Fast maturing M Ki- Abiotic stress tolerance; nutritious and healthy food F Abiotic stress tolerance, free of gluten K Abiotic stress tolerance K Abiotic stress tolerance F Abiotic stress tolerance F Abiotic stresses, fast growing F Abiotic stresses, fast growing F Abiotic stresses, fast growing F		מווז ווזהו אינט אינען גערים צ	voğtapıncai III	ומטר ב שטרטנינים טו אוומון נטטט נוטאס איונו נווטון ציטטגומטוונימו וווויזטו ומוועט מוום שטוומטוט וומווא		
tice Dryza glaberrina Nigeria, Burkina Faso, Liberia, Sierra Resistance to biotic and abiotic stresses L ha Diyerse types 0.91 Guinea, Nigeria, Mali, Nigeria, Chad, Abiotic stress tolerance; nutritious and E N biverse types 3.10 Ethopia, Eritera Abiotic stress tolerance; nutritious and E E east <i>Eragrostis (ef</i> 3.10 Ethopia, Eritera Abiotic stress tolerance; nutritious and brown and mutritious E east <i>Amaratilus syp</i> Abiotic stress tolerance; nutritious and brown and mutritious E E east <i>Amaratilus syp</i> Abiotic stress tolerance; nutritious E E east <i>Amaratilus syp</i> Nutritious E E E east <i>Chemopodium quinou</i> 0.19 Bolivia, Pera, Erandor Nutritious E E east <i>Chemopodium quinou</i> 0.19 Bolivia, Pera, Erandor Nutritious E E east <i>Chemopodium quinou</i> 0.19 Bolivia, Pera, Mann Nutritious E E a <i>Chemopodium quinou</i> <	Common Name	Scientific name	Global area (million ha)	Major cultivating countries	Beneficial traits	Reference
fice Oryca glabertina Nigeria, Bukina Faso, Liberia, Sierra Resistance to botic and abiotic stresses L tha Diverse types 3.10 Guina, Nigeria, Mali, Nigeria, Chad, Abiotic stress tolerance; nuritious and Engroutis tef Note Note </td <td>Cereals</td> <td></td> <td></td> <td></td> <td></td> <td></td>	Cereals					
flat Digitaria calis 0.91 Guinea, Nigeria, Mali Fast maturing N Diverse types 3.2.2 India, Niger, Sudan, Mali, Nigeria, Chad, Abiotic stress tolerance: nutritious and E E eals <i>Engrostis tef</i> 3.10 Ethiopia Eritrea Abiotic stress tolerance: nutritious and E E eals <i>Ameranthus sp.</i> Abiotic stress tolerance, rutritious and the static stress tolerance, free of gluen K eals <i>Ameranthus sp.</i> Abiotic stress tolerance, free of gluen K eals <i>Ameranthus sp.</i> Abiotic stress tolerance, free of gluen K eal <i>Ameranthus sp.</i> Abiotic stress tolerance, free of gluen K eal <i>Ameranthus sp.</i> Nutritious and healthy E eal <i>Amerantan</i> 12.8 Niger, Jan, Man Nutritious E a <i>Cleer arietinum</i> 12.65 India, Bastal, Faso, Mozan Nutritious E E a <i>Cleer arietinum</i> 12.65 India, Bastal, Bastal, Paso, Mozan Nutritious E E a <i>Cleer arietinum</i>	African rice	Oryza glaberrima		Nigeria, Burkina Faso, Liberia, Sierra Leone	Resistance to biotic and abiotic stresses	Linares (2002); NRC (1996)
Diverse types 3.2.2 India, Nigeri, Sudan, Mali, Nigeria, Chad, Abiotic stress tolerance: nurritions and starting feas, strengal, China, Paki, Ready food Free of gluten K east Engrostic tef 3.10 Ethiopia, Eritrea Abiotic stress tolerance, free of gluten K east Amarenthus spp. Ethiopia, Eritrea Abiotic stress tolerance, free of gluten K east Fagoryirum ecculentum 4.18 Poland Nutritious Ethiopia east Chempodium quinoa 0.19 Bolivia, Peru, Ecuador Nutritious and healthy Ethiopia east Chempodium quinoa 0.19 Bolivia, Peru, Ecuador Nutritious and healthy Ethiopia east Ciner arietinum 12.65 India, Pakistan, Australia, Iran, Man- Nutritious and healthy Ethiopia a Cicer arietinum 12.65 India, Pakistan, Australia, Iran, Man- Nutritious Ethiopia a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Man- Nutritious Ethiopia a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Man- Nutritious Ethiopia a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Man- Nutritious Ethiopia a <i>Ladiyrus sativus</i> <	Fonio/acha	Digitaria exilis	0.91	Guinea, Nigeria, Mali	Fast maturing	Williams and Haq (2000); FAOSTAT (2018); NRC (1996)
Engrostis tef 3.10 Ethiopia, Eritrea Abiotic stress tolerance, free of gluten exals Amaranthus spp. Fast growing and nutritious exit Fagopyrum exculentum 4.18 Nutritious exit Fagopyrum exculentum 4.18 Nutritious exit Fagopyrum exculentum 1.8 Nutritious exit Fagopyrum exculentum 1.8 Nutritious citerropodium quinoa 0.19 Bolivia, Peru, Ecuador Nutritious a Citer arietinum 1.265 Niget, Cameroon, Burkina Faso, Mosan Nutritious a Citer arietinum 1.205 Niget, Sudan Nutritious am Ladhyrus sativus 1.50 India, Russia, Intrés, Ethiopia Nutritious am Ladhyrus sativus 1.50 India, Russia, Intrés, Ethiopia Nutritious am Ladhyrus sativus 1.50 India, Russia, Intrés, Australia, Intrés, Intrés, Australia, Intrés, Intrés, Australia, Nutritious Nutritious am Ladhyrus sativus 1.50 India, Russia, Intrés, Stri Lanka, Nutritious am	Millets ^a	Diverse types	32.2	India, Niger, Sudan, Mali, Nigeria, Chad, Burkina Faso, Senegal, China, Paki- stan, Ethiopia	Abiotic stress tolerance; nutritious and healthy food	FAOSTAT (2018); Dawson and Jaenicke (2006); Williams and Haq (2000); Gupta et al. (2017)
eals in <i>Amaranthus spr.</i> eat <i>Eagopyram ecolentum</i> 4.18 China, Russia, Ukraine, Kazakhstan, Nutritious <i>Tagopyram ecolentum</i> 4.18 China, Russia, Ukraine, Kazakhstan, Nutritious <i>Chempodium quinoa</i> 0.19 Bolivia, Feru, Ecuador Nutritious and healthy <i>Chempodium quinoa</i> 0.19 Bolivia, Feru, Ecuador Nutritious and healthy <i>Chempodium quinoa</i> 0.19 Niger, Cameroon, Burkina Faso, Mali, Nutritious and drought tolerance <i>Cicer arietinum</i> 1.2.65 India, Pakistan, Australia, Faso, Mali, Nutritious and drought tolerance <i>Vigra unguiculata</i> 1.2.65 Niger, Nuessi, Turkey, Etihopia <i>Cicer arietinum</i> 1.2.65 Niger, Nigeria, Burkina Faso, Mozam- <i>Vigra unguiculata</i> 1.2.50 Niger, Nigeria, Burkina Faso, Mozam- <i>Lathyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Ethiopia and <i>Macrooftoma uniforum</i> 1.640 Chanda, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.46 Chanda, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.49 India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.49 India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.49 India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.40 Chanda, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.40 Chanda, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 5.40 Chanda, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butuan</i> <i>Leus culinaris</i> 2.41 Nufer, Australia, Nutritious <i>Moringa oleffera</i> 1.41 Nufer, Chue d'Ivoire, Tolerance to buict stresses, fast growing and <i>Abelnoschus esculeruns</i> 1.44 India, Niger, Chue d'Ivoire, Healthy	Tef	Eragrostis tef	3.10	Ethiopia, Eritrea	Abiotic stress tolerance, free of gluten	Ketema (1997); Spaenij-Dekking et al. (2005)
Ih Amacranthus spp. Fast growing and nurritious eat <i>Fagopyrum exculentum</i> 4.18 China, Rusia, Ukraine, Kazakhstan, Nurritious Nurritious Nurritious eat <i>Chenopodium quinoa</i> 0.19 Bolivia, Petu, Ecuador Nurritious and healthy a groundnut <i>Vigua unperculent</i> 1.265 Niget, Cameroon, Burkina Faso, Mali, Nurritious and healthy a <i>Cicer arietinum</i> 12.65 Niget, Cameroon, Burkina Faso, Mali, Nurritious and healthy a <i>Cicer arietinum</i> 12.65 Niget, Nuessia, Turkey, Ethopia Nurritious a <i>Ladhyrus sativus</i> 1.50 Niget, Nigeria, Burkina Faso, Mozam- Drought tolerance a <i>Ladhyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nurritious am <i>Macroyloma uniforum</i> 1.265 Niget, Nigeria, Burkina Faso, Mozam- Drought tolerance and nurritious am <i>Ladhyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nurritious am <i>Ladhyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nurritious am <i>Ladhyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Nurritious am <i>Ladhyrus caifurus</i> <td>Pseudocereals</td> <td></td> <td></td> <td></td> <td></td> <td></td>	Pseudocereals					
eat <i>Fagopyrum esculentum</i> 4.18 China, Russia, Ukraine, Kazakhstan, Nutritious and healthy <i>Chenopodium quinoa</i> 0.19 Bolivia, Peru, Ecuador Nutritious and healthy agroundhut <i>Vigna subterranean</i> 0.26 Niger, Cameroon, Burkina Faso, Mali, Nutritious and drought tolerance <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Myan- <i>Vigna unguiculata</i> 12.25 Niger, Niger, Burkina Faso, Mozam- <i>Vigna unguiculata</i> 12.25 Niger, Niger, Burkina Faso, Mozam- <i>Uany vus sativus</i> 1.50 India, Burkina Faso, Mozam- <i>Drought tolerance and nutritious</i> <i>Macroyloma uniforum</i> 1.5.60 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious <i>Lens cultuaris</i> 5.46 Canada, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butan</i> <i>Lens cultuaris</i> 5.46 Canada, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butan</i> <i>Macroyloma uniforum</i> 5.49 India, USA, Turkey, Australia, Nutritious and healthy food <i>Butan</i> <i>Lens cultuaris</i> 5.46 Canada, India, USA, Turkey, Australia, Nutritious and healthy food <i>Butan</i> <i>Macroyloma digitate</i> 5.49 India, USA, Turkey, Australia, Nutritious <i>Adamsonia digitate</i> 1.41 India, Narania, Malawi, Alawi, Alawi, Alawi, Alawi, Alawi, Adamsonia digitate <i>Moringa oleifera</i> 2.41 Dira, Matania, Malawi, Alawi, Alawi, Alawi, Abernoschus excultutus 2.41 India, Mozane, Sudan, Nutritious <i>Moringa oleifera</i> 1.41 India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing and <i>Ricinus connunis</i> 1.44 India, Mozanbique, Brazil, China Healthy	Amaranth	Amaranthus spp.			Fast growing and nutritious	NRC (2006)
<i>Chenopodium quinoa</i> 0.19 Bolivia, Peru, Ecuador Nutritious and healthy a crownia <i>Vigna subierranean</i> 0.26 Niger, Cameroon, Burkina Faso, Mali, Nutritious and healthy a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Myan- Nutritious and drought tolerance a <i>Cicer arietinum</i> 12.25 Niger, Nigeria, Burkina Faso, Mozam- Drought tolerance and nutritious a <i>Luthyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious a <i>Luthyrus sativus</i> 1.50 India, Bangladesh, Myanmar, Sri Lanka, Nutritious and healthy food am <i>Macroyloma uniflorum</i> India, Bangladesh, Myanmar, Sri Lanka, Nutritious Nutritious am <i>Lathyrus sativus</i> 5.49 India, USA, Turkey, Australia, Nutritious am <i>Lathyrus sativus</i> 5.49 India, USA, Turkey, Australia, Nutritious am <i>Lathyrus sativus</i> 5.49 India, USA, Turkey, Australia, Nutritious am <i>Lathyrus sativus</i> 5.49 India, USA, Turkey, Australia, Nutritious am <i>Lathyrus sativus</i> 5.49 India, Nigar, Git Nutritious sea <i>Cajarus cajan</i> 5.49 India, Nigar, Git Nutritiou	Buckwheat	Fagopyrum esculentum	4.18	China, Russia, Ukraine, Kazakhstan, Poland	Nutritious	FAOSTAT (2018)
strondnut Vigna subterranean 0.26 Niger, Cameroon, Burkina Faso, Mali, Nutritious and drought tolerance a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Myan- Nutritious a <i>Cicer arietinum</i> 12.65 India, Pakistan, Australia, Iran, Myan- Nutritious a <i>Uigna unguiculata</i> 12.25 Niger, Nigeria, Burkina Faso, Mozam- Drought tolerance and nutritious a <i>Lathyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Entiopia am <i>Macroyloma uniforum</i> 1.50 India, Bangladesh, Nepal, Pakistan, Entiopia am <i>Lathyrus sativus</i> 1.50 India, Bangladesh, Nepal, Pakistan, Entiopia am <i>Lathyrus sativus</i> 1.50 India, USA, Turkey, Australia, Nutritious and healthy food am <i>Macroyloma uniforum</i> Butan Butan Nutritious and healthy food am <i>Macroyloma uniforum</i> Butan Nutritious and healthy food am <i>Lens culinaris</i> 5.46 Canada, India, USA, Turkey, Australia, <i>Nutritious acularus</i> 5.46 Canada, India, USA, Turkey, Australia, <i>Nutritious acularus culinaris</i> 5.46 Canada, India, USA, Turkey, Australia, <i>Nutritious acularus culinaris</i> 5.49 India, Myanmar	Quinoa	Chenopodium quinoa	0.19	Bolivia, Peru, Ecuador	Nutritious and healthy	FAOSTAT (2018)
a groundnut Vigra subterranean 0.26 Niger, Cameroon, Burkina Faso, Mali, Nutritious and drought tolerance a Cicer arietinum 12.65 India, Pakistan, Australia, Iran, Myan- Nutritious a Cicer arietinum 12.65 Niger, Ukey, Ethiopia Nutritious a Uigna unguiculata 12.25 Niger, Nigeria, Burkina Faso, Mozan- Drought tolerance and nutritious a Ladhyrus sativus 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Macropyloma uniflorum 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Ladhyrus sativus 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Ladhyrus sativus 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Macropyloma uniflorum Bangladesh, Namar, Sti Lanka, Nutritious Nutritious am Macropyloma uniflorum Bangladesh, Iran, Syria, Ethio- Nutritious Nutritious am Cajanus cajan 5.49 India, USA, Turkey, Australia, Nutritious ea Cajanus cajan 5.49 India, Maawi, Macros s Admsonia digiate Madagascar, Ind	Legumes					
a Cicer arietirum 12.65 India, Pakistan, Australia, Iran, Myan- Nutritious Vigra unguiculata 12.25 Niger, Nigeria, Burkina Faso, Mozam- Drought tolerance and nutritious a Lathyrus sativus 1.50 India, Pakistan, Kepal, Pakistan, Extreme drought tolerance and nutritious am Macronytoma unifforum 1.50 India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Macronytoma unifforum India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Macronytoma unifforum India, Bangladesh, Nepal, Pakistan, Extreme drought tolerance and nutritious am Macronytoma unifforum India, Bangladesh, Iran, Syria, Ethio- Nutritious and healthy food Butan Lens culinaris 5.49 India, Waanmar, Siri Lanka, Nutritious ea Cajanus cajan 5.49 India, Manmar, Tanzania, Malawi, Butan Lens calinaria S.49 India, Myanmar, Tanzania, Malawi, sea Cajanus cajan S.49 India, Malawi, adansonia digitare Mariana Nutritious sea Mansonia digitare Madagscar, India, Niger, Côte d'Ivoire, diansonia digitare Mariana Nutritious dennoschus exculentus 2.41 <t< td=""><td>Bambara groundnut</td><td>Vigna subterranean</td><td>0.26</td><td>Niger, Cameroon, Burkina Faso, Mali, Togo, DR Congo</td><td>Nutritious and drought tolerance</td><td>FAOSTAT (2018); NRC (2006)</td></t<>	Bambara groundnut	Vigna subterranean	0.26	Niger, Cameroon, Burkina Faso, Mali, Togo, DR Congo	Nutritious and drought tolerance	FAOSTAT (2018); NRC (2006)
Vigna unguiculata12.25Niger, Nigeria, Burkina Faso, Mozam- bique, SudanDrought tolerance and nutritious bique, SudananLathyrus sativus1.50India, Bangladesh, Nepal, Pakistan, EthiopiaExtreme drought tolerance and nutritious EthiopiaamMacrotyloma uniflorum1.50India, Bangladesh, Nepal, Pakistan, EthiopiaExtreme drought tolerance and nutritiousamMacrotyloma uniflorumIndia, Bangladesh, Myanmar, Sri Lanka, BhutanNutritious and healthy food BhutanamMacrotyloma uniflorum5.46Canada, India, USA, Turkey, Australia, Nepal, Bangladesh, Iran, Syria, Ethio- piaNutritious and healthy food BhutaneaCajanus cajan5.49India, USA, Turkey, Australia, NatritiousNutritioussCajanus cajan5.49India, USA, Turkey, Australia, NatritiousNutritiouseaCajanus cajan5.49India, Myanmar, Tanzania, Malawi, Haiti, KenyaNutritioussAdansonia digitareMadagascar, IndiaNutritiousnMoringa olefferaIndia, Niger, Côte d'Ivoire, Cameroon, Sudan, MaliNutritiouseanRicinus communis1.44India, Mozambique, Brazil, ChinaHealthy	Chickpea	Cicer arietinum	12.65	India, Pakistan, Australia, Iran, Myan- mar, Russia, Turkey, Ethiopia	Nutritious	FAOSTAT (2018)
an Lathyrus sativus 1.50 India, Bangladesh, Nepal, Pakistan, Ethiopia Extreme drought tolerance and nutritious anm Macrotyloma uniflorum Ethiopia India, Bangladesh, Myanmar, Sri Lanka, Nutritious and healthy food anm Macrotyloma uniflorum India, Bangladesh, Iran, Syria, Ethio- Bhutan Nutritious Nutritious caimus caim 5.46 Canada, India, USA, Turkey, Australia, Nutritious ken Caimus caim 5.49 India, Myanmar, Tanzania, Malawi, Haiti, Kenya Nutritious s Adansonia digitate Madagascar, India Nutritious h Moringa oleifera Madagascar, India Nutritious abelmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, and nutritious Polerance to biotic stresses, fast growing	Cowpea	Vigna unguiculata	12.25	Niger, Nigeria, Burkina Faso, Mozam- bique, Sudan	Drought tolerance and nutritious	FAOSTAT (2018); NRC (2006)
amMacrotyloma uniflorumIndia, Bangladesh, Myanmar, Sri Lanka, BhutanNutritious and healthy food BhutanLens culinaris5.46Canada, India, USA, Turkey, Australia, Nepal, Bangladesh, Iran, Syria, Ethio- piaNutritiousLens culinaris5.49India, USA, Turkey, Australia, Nepal, Bangladesh, Iran, Syria, Ethio- piaNutritiouseaCajanus cajan5.49India, Myanmar, Tanzania, Malawi, Haiti, KenyaNutritioussAdansonia digitateMadagascar, IndiaNutritioushMoringa oleiferaIndia, Niger, Côte d'Ivoire, Cameroon, Sudan, MalNutritiouseanRicinus commuis2.41Niger, Côte d'Ivoire, and nutritiousTolerance to biotic stresses, fast growingeanRicinus commuis1.44India, Mozambique, Brazil, ChinaHealthy	Grass pea	Lathyrus sativus	1.50	India, Bangladesh, Nepal, Pakistan, Ethiopia	Extreme drought tolerance and nutritious	Campell (1997); Kumar et al. (2013)
Lens culinaris5.46Canada, India, USA, Turkey, Australia, Nepal, Bangladesh, Iran, Syria, Ethio- piaNutritiouseaCajanus cajan5.49India, Myanmar, Tanzania, Malawi, Haiti, KenyaNutritioussCajanus cajan5.49India, Myanmar, Tanzania, Malawi, Haiti, KenyaNutritioussAdansonia digitateMadagascar, IndiaNutritioushMoringa oleiferaIndia, Niger, Côte d'Ivoire, Cameron, Sudan, MaliNutritiouseanRicinus communis1.44India, Mozambique, Brazil, ChinaHealthy	Horsegram	Macrotyloma uniflorum		India, Bangladesh, Myanmar, Sri Lanka, Bhutan	Nutritious and healthy food	Chahota et al. (2013)
cea Cajanus cajan 5.49 India, Myanmar, Tanzania, Malawi, Haiti, Kenya s Haiti, Kenya s Adansonia digitate Madagascar, India n Moringa oleifera India, helmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, and nutritious ean Ricinus communis 1.44 India, Mozambique, Brazil, China	Lentil	Lens culinaris	5.46	Canada, India, USA, Turkey, Australia, Nepal, Bangladesh, Iran, Syria, Ethio- pia	Nutritious	FAOSTAT (2018)
s Adansonia digitate Madagascar, India Nutritious n Moringa oleifera India, Nutritious Nutritious India, Nigeria, India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing Abelmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing can Ricinus communis 1.44 India, Mozambique, Brazil, China Healthy	Pigeon pea	Cajanus cajan	5.49	India, Myanmar, Tanzania, Malawi, Haiti, Kenya		FAOSTAT (2018); Pal et al. (2011)
Adansonia digitate Madagascar, India Nutritious India, Moringa oleifera India, Moringa oleifera India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing Abelmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing and nutritious Cameroon, Sudan, Mali and nutritious ean Ricinus communis 1.44 India, Mozambique, Brazil, China Healthy	Vegetables					
Moringa oleifera India, Nutritious Abelmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing Cameroon, Sudan, Mali and nutritious and nutritious ean Ricinus communis 1.44 India, Mozambique, Brazil, China Healthy	Baobab	Adansonia digitate		Madagascar, India	Nutritious	NRC (2006)
Abelmoschus esculentus 2.41 Nigeria, India, Niger, Côte d'Ivoire, Tolerance to biotic stresses, fast growing Cameroon, Sudan, Mali and nutritious ean Ricinus communis 1.44 India, Mozambique, Brazil, China Healthy	Moringa	Moringa oleifera		India,	Nutritious	NRC (2006)
ean <i>Ricinus communis</i> 1.44 India, Mozambique, Brazil, China Healthy	Okra	Abelmoschus esculentus	2.41	Nigeria, India, Niger, Côte d'Ivoire, Cameroon, Sudan, Mali	Tolerance to biotic stresses, fast growing and nutritious	FAOSTAT (2018)
Ricinus communis 1.44 India, Mozambique, Brazil, China Healthy	Oil seeds					
	Castor bean	Ricinus communis	1.44	India, Mozambique, Brazil, China	Healthy	FAOSTAT (2018)

 Table 2
 Selected orphan food crops with their geographical importance and desirable traits

Table 2 (continued)					
Common Name	Scientific name	Global area (million ha)	Major cultivating countries	Beneficial traits	Reference
Linseed	Linum usitatissimum	3.02	Russia, Kazakhstan, Canada, India, China, USA, Ethiopia, Ukraine	Nutritious and healthy	FAOSTAT (2018)
Noug	Guizotia abyssinica		Ethiopia, India	Quality oil, abiotic stress tolerance	Getinet and Sharma (1996)
Sesame	Sesamum indicum	10.60	Sudan, India, Myanmar, Tanzania, South Sudan, Nigeria, Ethiopia	Oxidatively stable oil	Dawson and Jaenicke (2006); FAOSTAT (2018)
Root crops					
Cassava	Manihot esculentum	26.1	Nigeria, DR Congo, Brazil, Thailand, Mozambique, Tanzania, Uganda, Ghana, Angola, Indonesia, Côte d'Ivoire	Drought tolerance	Ceballos et al. (2004); FAOSTAT (2018)
Enset	Ensete ventricosum	0.3	Ethiopia	Drought tolerance	Brandt (1997); Olango et al. (2014)
Sweet potato	Ipomoea batatas	12.25	China, Nigeria, Tanzania, Uganda, Ethio- pia, Angola, Madagascar	Rich in riboflavin and calcium	Dawson and Jaenicke (2006); FAOSTAT (2018)
Taro cocoyam	Colocasia esculenta	1.83	Nigeria, Cameroon, Ghana, China, Côte d'Ivoire, Rwanda, Madagascar	Nutritious and healthy	FAOSTAT (2018); Okpala (2015)
Yam	Dioscorea spp	8.38	Nigeria, Côte d'Ivoire, Ghana, Benin, Togo, Sudan	Drought tolerance	Williams and Haq (2000); FAOSTAT (2018)
Fruits					
Banana	Musa spp.	5.81	India, Brazil, Tanzania, China, Philip- pines, Rwanda, Burundi, Ecuador, Uganda	Healthy and nutritious	Heslop-Harrison and Schwarzacher (2007); FAOSTAT (2018)
Plantain	Musa spp.	5.43	DR Congo, Uganda, Nigeria, Côte d'Ivoire, Colombia, Ghana Cameroon, Tanzania, Philippines	Healthy and nutritious	Heslop-Harrison and Schwarzacher (2007); FAOSTAT (2018)
The list of countries ^a Millets refer to all ty <i>tum</i>), little millet (<i>Pa</i>	The list of countries is presented according to the descending or ^a Millets refer to all types of millets which include Barnyard mill <i>tum</i>), little millet (<i>Panicum sumatrense</i>), pearl millet (<i>Pennisetun</i>	he descending c ide Barnyard m nillet (Penniset,	The list of countries is presented according to the descending order of the area the crop is cultivated in 2016 ^a Millets refer to all types of millets which include Barnyard millet (<i>Echinochloa crus-galli</i>), finger millet (<i>Eleusin</i> tum), little millet (<i>Panicum sumatrense</i>), pearl millet (<i>Pennisetum glaucum</i>) and proso-millet (<i>Panicum miliaceum</i>)	16 (Eleusine coracana), foxtail millet (Setar liaceum)	ler of the area the crop is cultivated in 2016 let (<i>Echinochloa crus-galli</i>), finger millet (<i>Eleusine coracana</i>), foxtail millet (<i>Setaria italica</i>), kodo millet (<i>Paspalum scrobicula- n glaucum</i>) and proso-millet (<i>Panicum miliaceum</i>)

681



Fig. 1 Panicles of selected orphan cereal crops. Left to right: proso-millet (*Panicum miliaceum*), foxtail millet (*Setaria italica*), finger millet (*Eleusine coracana*) and two tef (*Eragrostis tef*) panicles with red and green color. Scale bar: 10 cm. Photo: Annett Weichert, University of Bern

be an insurance crop as it produces reliable yields when all other crops fail due to extreme moisture scarcity.

Vegetables

There are many indigenous or locally important vegetables in the developing world. Several of them have benefits in some agronomic and/or nutritional traits: for example, okra (*Abelmoschus esculentus*) is fast growing and nutritious (Gemede et al. 2014) or baobab (*Adansonia digitate*), which is a multi-purpose tree with leaves rich in iron and fruits rich in vitamin C (Sanchez 2018).

Oilseeds

Among oilseeds, sesame (*Sesamum indicum* L) is annually cultivated on over 10 million hecatres of land mainly in Sudan, India, Myanmar and Tanzania (FAOSTAT 2018). Other oilseeds with some importance in developing countries are linseed (*Linum usitatissimum*), castor bean (*Ricinus communis*) and noug (*Guizotia abyssinica*).

Root and tuber crops

Among root crops, cassava (*Manihot esculenta*), sweet potato (*Ipomoea batatas*) and yam (*Dioscorea sp.*) are the source of food for a large number of people in the developing

world. Cassava or manioc is annually cultivated on 26.1 million ha of land globally where the major producing countries are Nigeria, DR Congo, Brazil, Thailand and Mozambique (FAOSTAT 2018). Cassava is tolerant to drought, and performs better than other crops on soils with poor nutrients. Other extensively cultivated orphan root and tuber crops include sweet potato on 12.3 million ha and yam on 8.4 million ha (FAOSTAT 2018). While sweet potato is cultivated in different parts of the world, yam is exclusively cultivated in Africa. It is worthwhile to mention the plant called enset (Ensete ventricosum), which is also known as 'false banana' because it resembles the domesticated banana plant. Enset is a staple food for over 20 million people in the densely populated regions of Ethiopia. Unlike banana where the fruit is consumed, in enset the pseudo-stem and the underground corm are the edible parts. Enset is an extremely drought-tolerant crop that adapts to different soil types (Brandt 1997).

Fruits

Banana and plantain (*Musa spp.*) are important fruit crops in the developing world although genetic resources and research are very limited. The global cultivation area of these fruits was 5.8 million ha for banana and 5.4 million for plantain in 2016 (FAOSTAT 2018). Banana is extensively cultivated in Asia, Africa and South America. Plantain is the staple food in Africa especially in DR Congo, Uganda and Nigeria. It is often considered to be a vegetable rather than a fruit since the plantain is cooked like a vegetable. According to Fungo (2009), banana especially the orange pulped type with high carotenoid and iron content could reduce iron deficiency anemia by over 50% and also vitamin A deficiency in East Africa, where a large part of the population lack iron and vitamin A. Banana and plantain are considered to be healthy foods as they are also rich in essential nutrients for humans.

Need for improving orphan crops

There are many reasons to invest in research on orphan crops. The main reasons are described below.

Exclusion of orphan crops from advanced research

Orphan crops play a vital role in the livelihood of the developing world (Tadele 2014, 2017, 2018; Tadele and Assefa 2012). However, compared to major crops of the world, research and development of orphan crops are lagging behind that of major crops at both the global and at the national levels. These two scenarios are briefly discussed below.

Neglect at the global level: exclusion from the Green Revolution

The famous Green Revolution contributed to a significant boost in total production and productivity of wheat and rice in Asia. However, the Green Revolution did not occur either in Africa or on orphan crops (Ejeta 2010; Godfray et al. 2010). In terms of area of cultivation, the top three crops in Africa are maize (23%), sorghum (16%) and cassava (12%), while those in Asia are rice (35%), wheat (24%) and maize (16%) [Fig. 2, (FAOSTAT 2018)]. Similarly, in terms of total production, the top three crops in Africa are cassava (34%), maize (16%) and yam (14%), while in Asia these are rice (28%), maize (15%) and wheat (13%). In Africa, cereals produce inferior yield while root crops such as cassava and yam, which are associated with a high risk of post-harvest losses due to a short shelf-life, contribute to the bulk of production. On the contrary, the major cereal crops in Asia can be stored for long periods, at least until just the harvest of the next cropping season, which corresponds to the critical time of food shortage. This indicates that the dominant African crops in terms of both the area under cultivation and the total production were not given a policy-related focus which is also referred to as an enabling environment (Tadele 2019).

Neglect at the national level: a tale of two cereals from Ethiopia

Here, comparison is made between wheat and tef, the two extensively cultivated cereals in Ethiopia. While tef is an orphan crop with local importance, wheat is a major crop with both local and global importance. In Ethiopia, tef is annually cultivated by 6.7 million farmers while wheat by 4.8 million farmers (Fig. 3a). Similarly, the area of tef and wheat cultivations is 2.9 and 1.7 million hectares, respectively (Fig. 3b). This 40% more of tef farmers over wheat farmers and about 70% more area allocated to tef compared to wheat shows the importance of tef in the Ethiopian agriculture. However, the total production of 4.5 million tons of tef was only 7% higher than that of wheat (4.2 million tons) (Fig. 3c). This small gap in production between the two crops but large gap in the area under cultivation is mainly due to the more advanced research and development implemented in wheat. This can be witnessed from the number of improved varieties. The total number of improved varieties released to the farming community in Ethiopia until the year 2015 was only 35 for tef but 167 for wheat (Fig. 3d). It is also important to note that the total number of varieties released during the 3-year period between 2012 and 2015 was only 3 for tef while it was 15 for wheat. Due to the release and wide dissemination of improved wheat varieties, the national average yield of wheat is currently 2.5 t ha⁻¹ compared to only 1.6 t ha⁻¹ for tef. The improvement of wheat substantially exceeded that of tef as witnessed by the number of improved varieties available to farmers. This is mainly due to significant contributions to wheat improvement by international organizations such as CIMMYT (International Maize and Improvement Center) from where most wheat breeding lines for improved varieties originated. On the contrary, the bulk of research on tef has been done with little support by domestic and international organizations. Not only the orphan crops but also researchers of orphan crops have a disadvantage due to very limited access to advanced research and technical know-how as most researchers do not have training opportunities in advanced laboratories.

It is not the intention of this author to either call for the complete replacement of wheat fields by tef or to focus only on the orphan crop tef. Wheat is also equally important for farmers and consumers since it is extensively cultivated in the country. However, the intention is to pay attention to the improvement of tef, which is annually cultivated on about 30% of the total cereal area and a staple food for over 60 million people in Ethiopia alone.

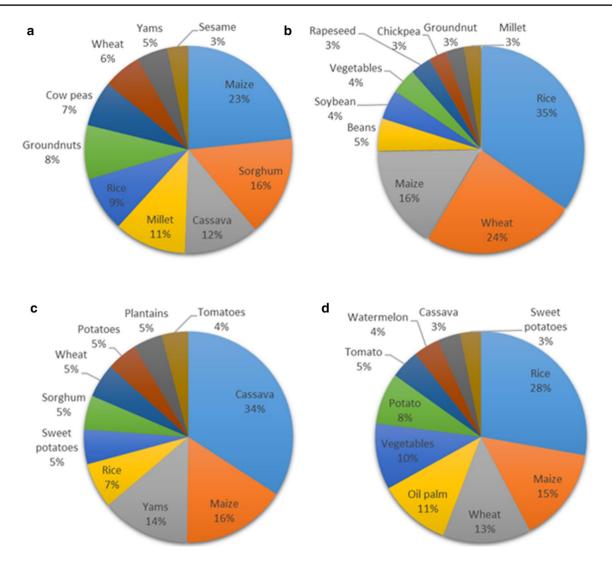


Fig. 2 The proportion of the top ten food crops in 2017 in Africa (\mathbf{a} , \mathbf{c}) and Asia (\mathbf{b} , \mathbf{d}) in terms of total area (\mathbf{a} , \mathbf{b}) and total production (\mathbf{c} , \mathbf{d}) Adapted from (FAOSTAT 2018)

Constraints related to inherent properties of orphan crops

Orphan crops fit to the socio-economic and diet of the population in the developing world. However, orphan crops are not without constraints. Some orphan crops are associated with undesirable traits, which need to be altered to improve their productivity and/or nutritional quality. Constraints which are directly related to some orphan crops are the following.

Inferior in productivity

Due to lack of genetic improvement, orphan crops produce inferior yield in terms of both quality and quantity. For instance, the seed yields of African rice (*Oryza glaberrima*), tef and millets are extremely low. The main reasons for poor productivity in these crops are lodging, seed shattering or other crop specific constraints (Assefa et al. 2011; Linares 2002).

Poor or unbalanced nutrition

Root and tuber crops such as cassava and enset produce high yields, but they are largely composed of starches that are deficient in other essential nutrients particularly in protein. Studies showed that large populations in low-income countries who depend on these food products as a sole food source suffer from malnutrition (Stephenson et al. 2010; Gegios et al. 2010). Hence, supplementation with other essential nutrients is necessary. Alternatively, biofortification of staple food crops with essential nutrients can be made to enhance the nutrient content and/or remove undesirable food quality traits.

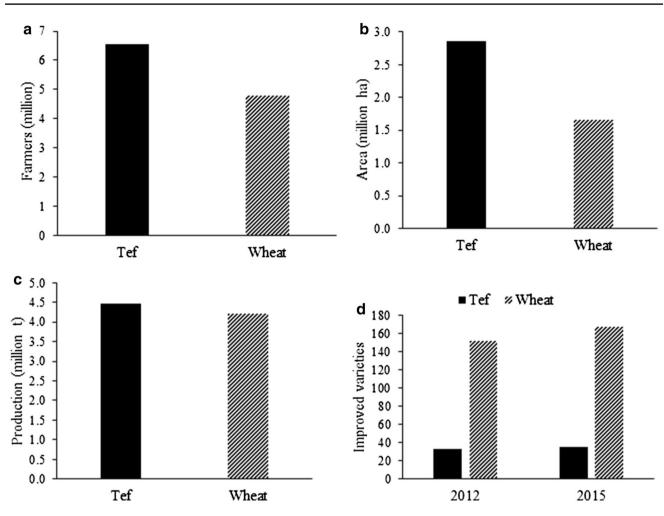


Fig. 3 Comparison between an orphan crop tef and a major crop wheat in Ethiopia. **a** Number of farmers cultivating in 2016; **b** total area under cultivation in 2016; **c** total grain production in 2016; and **d**

total number of varieties released by 2012 and by 2015 Adapted from (Mabaya et al. 2017; CSA 2015; WheatAtlas 2019)

Toxic products

Widely cultivated crops such as cassava and grass pea produce toxic compounds that affect the health of human and/or livestock. The edible parts of cassava produce a poisonous substance called cyanogenic glycosides which release cyanide (Ceballos et al. 2004). Hence, the consumption of unprocessed cassava results in a paralytic disease called *konzo*. Similarly, the grains of grass pea contain a neurotoxic substance called ODAP (β -*N*-oxalyl-L- α , β -diaminopropanoic acid) which causes irreversible paralysis of the lower body when consumed over long periods (Yan et al. 2006).

Extreme environmental conditions

Since most fertile lands are used to grow crops other than indigenous crops, native crops are mostly cultivated on less fertile and moisture-deficit soils. In addition, crop productivity is affected by a variety of abiotic and biotic stresses (Tadele 2019). Abiotic stresses which are prevalent in developing countries are drought, flooding, heat, soil salinity and soil acidity. At the same time, biotic stresses which include a variety of diseases, insects and weeds also tremendously reduce crop productivity. The adverse effects of biotic factors on crop productivity are more obvious in the tropical regions due to their presence in high density and diversity. Hence, the extent and type of environmental challenges are more intense for orphan crops.

Opportunities related to orphan crops

Orphan or indigenous crops are associated with some desirable properties which can be useful in the promotion of these crops to a higher level of production. The compatibility of these crops to the agro-ecology and socio-economic conditions as well as their potential to adapt to changing climate are a few examples of opportunities for using orphan crops.

Compatibility with agro-ecology and socio-economic conditions

Most orphan or indigenous crops perform better under adverse climatic and soil conditions than the exotic or introduced crops. In addition, orphan crops are compatible with the agro-ecology and socio-economic conditions. However, when other crops that are new to the locality replace indigenous crops, some problems might occur. For example, the incidence of malaria has been increased when the indigenous crop tef was replaced by maize in the northwestern part of Ethiopia since mosquitos prefer to feed on maize pollen rather than on indigenous cereals (Ye-Ebiyo et al. 2000).

Orphan crops play a vital role in changing climate conditions

Climate change poses a significant threat to the present crop production systems, infrastructures, and markets (Muller et al. 2011). It is expected that the total amount and distribution of rainfall will be substantially affected in the near future. Based on in situ station data and satellite observations, up to 15% decrease in rainfall is expected in the Eastern and Southern Africa (Funk et al. 2008). However, according to other studies, a wetter climate with more intense wet seasons and less severe droughts is predicted in East Africa (Shongwe et al. 2011) while a substantial decrease in rainfall and an increase in temperature in West Africa (Sarr 2012). According to other investigators, the long-term variability and changes of rainfall in the southern Africa will lead to more intense and widespread drought (Fauchereau et al. 2003). Orphan crops are expected to perform better than major crops under the changing climate due to their resilience to diverse environmental constraints (Chivenge et al. 2015).

In general, the positive traits of orphan crops should initiate research approaches to abolish the undesirable traits associated with orphan crops towards developing high yielding, nutritionally superior and resilient crops.

Initiatives on the awareness and improvement of orphan crops

Recently, some institutions in both, the developed and developing counties, are actively participating in the promotion of orphan crops. A number of promotions have been made to introduce the potential role of orphan crops in agriculture. Selected examples to increase the awareness of orphan crops are described. *Media*: famous newspapers heralded the nutrition- and health-related benefits of orphan crops. For instance, the story of the benefits of tef as a life-style crop has been recently presented in the New York Times (O'Connor 2016), the Washington Post (Gordon 2014), the Guardian (Provost and Jobson 2014), BBC (Jeffrey 2015) and many others.

The Lugar Center (http://www.thelugarcenter.org/ourwo rk-Orphan-Crops.html), under the leadership of the former United States Senator Richard G. Lugar, provides a platform for debates on global issues and also includes key information on orphan crops which are useful for researchers and other stakeholders including policymakers (Lugar 2019).

International Year of Quinoa

The United Nations General Assembly has declared 2013 as the International Year of Quinoa, in recognition of ancestral practices of the Andean people, who have managed to preserve quinoa in its natural state as food for present and future generations (FAO 2013). This was followed by publishing a comprehensive book on the global importance of this particular crop (Bazile et al. 2015).

Conferences and podium discussions

The first African Orphan Crops Conference was held in September 2007 at the University of Bern in Switzerland with the participation of researchers from all over the world (Tadele 2009a). This was followed by a conference held in September 2018 by the Cluster of Excellence in Plant Science (CEPLAS) in Cologne Germany (CEPLAS 2018).

Publications

In addition to original research articles in scientific journals and research reports, some journals dedicated a special issue to promote the research and knowledge of orphan crops; among them, a recently published report which describes orphan crops as 'Future Smart Food', in eight countries in South and Southeast Asia. The countries included in the report are Bangladesh, Bhutan, Cambodia, Lao PDR, Myanmar, Nepal, Viet Nam and West Bengal in India. The potential crops were selected based on four criteria such as nutrition, production, ecology and socio-economy. The crops include ten cereals, six root and tuber crops, nine pulses, nine fruit and vegetable crops, and five nuts and spices. The special issue in the African Technology Development Forum Journal (Tadele 2009b) and this issue of *Planta* should also be mentioned.

Conservation and maintenance of germplasm

National institutions

Worldwide, more than 1700 gene banks hold collections of food crops for safekeeping (CropTrust 2019). However,

many of these are not only vulnerable to natural catastrophes and war, but also suffer from poor funding. Hence, the conditions are not optimal for the conservation of valuable genetic resources.

International institutions

The consultative group for international agricultural research (CGIAR) plays a vital role among international institutions involved in germplasm collection and conservation. From the 15 CGIAR centers, those with the mandate on orphan crops are: Africa Rice on rice; International Center for Agricultural Research in the dry areas (ICARDA) on lentil and chickpea; International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) on chickpea, pigeon pea, and pearl millet; International Institute of Tropical Agriculture (IITA) on banana, plantain, cassava, cowpea and yam; International Center for Tropical Agriculture (CIAT) on beans and cassava; International Potato Center (CIP) on sweet potato and Bioversity International on banana (CGIAR 2019b). The total number of crop accessions including major and orphan crops deposited at CGIAR centers are estimated to be around 760,000 (CGIAR 2019a). These centers have been providing substantial technical and financial support to the national institutions in the developing world. However, highly nutritious and healthy crops such as tef are not included in the mandate of any CGIAR center.

Crop diversity trust

The trust supports crop conservation through developing global strategies for the ex situ conservation of 21 crops which include orphan crops such as cassava, chickpea, cowpea, finger millet, grass pea and sweet potato (CropTrust 2019).

Svalbard Global Seed Vault

This seed vault with the capacity to store 4.5 million varieties of crops is located at Svalbard in Norway which is only 1000 km from the North Pole. The vault is an ideal location for long-term seed storage due to the following reasons: (1) Svalbard is a remote location, so there are few humanrelated activities around it, (2) the area is geologically stable and humidity levels are low, (3) the vault is well above sea level, protected from ocean flooding according to worst case scenario sea level rises, and (4) the permafrost offers the vault room with a natural freezing, providing a costeffective and safe method to conserve seeds (CropTrust 2019). Hence, the Svalbard Global Seed Vault provides insurance against catastrophic loss of crop diversity. Currently, 983,524 varieties from all over the world have been conserved at the storage facility of the vault. It is interesting to note that some researchers have already started requesting the seeds they earlier deposited at the vault. The typical example is ICARDA, an International Agricultural Research Center, which sent thousands of its germplasm samples of different crops to the vault before the conflict in Syria destroyed its gene bank in Aleppo. Later, the center received over 38,000 seed samples from the seed vault to grow the plants at the newly relocated centers in Lebanon and Morocco, enabling continuation of the research (ICARDA 2015).

Advanced research on orphan crops

Improvement of orphan crops mostly uses conventional techniques particularly selection of lines from landraces. Some breeding programs of orphan crops also implement introgressions involving interspecific or intraspecific crosses. Recently also advanced breeding tools have been implemented for selected orphan crops through partnerships between institutions in developed and developing countries. Examples of advanced research on selected orphan crops are shown in Table 3. These advanced techniques include marker-based studies including GBS (genotyping by sequencing) and GWAS (genome-wide association study). High-throughput techniques which identify alleles such as TILLING (Targeting Induced Local Lesions in Genomes) from induced mutagenized populations and EcoTILLING from natural populations have been applied to selected orphan crops. Similarly, omic tools have been implemented on a few orphan crops. The African Orphan Crops Consortium (AOCC) was formed with primary goal of sequencing the genomes of 101 African food plants and improving nutrition in the continent through genome-assisted breeding resources and training (AOCC 2018). The consortium is formed by the New Partnership for Africa's Development (NEPAD), World Agroforestry Centre (ICRAF), Beijing Genomics Institute (BGI), University of California at Davis and Mars Incorporated. Other omic tools which include transcriptomics and proteomics have been used to describe gene expression profiles of some orphan crops. Genome editing techniques especially the CRISPR/Cas9 (clustered regularly interspaced short palindromic repeats/CRISPRassociated protein9) approach has been implemented on cassava and recently on ground cherry (Physalis pruinosa) (Lemmon et al. 2018; Bull et al. 2018; Gomez et al. 2018; Odipio et al. 2017; Hummel et al. 2018). The CRISPR/Cas9 technology has a high potential to modify undesired traits in orphan crops in a short time and should be implemented to orphan crops such as cassava, millet or tef (Mollins 2017).

Technique	Crop targeted	Reference
Speed breeding	Cassava, chick pea, finger millet	Chiurugwi et al. (2018)
Marker assisted		
GBS	Cassava, chickpea, cowpea, finger millet, pearl millet, tef	Kumar et al. (2016); Girma et al. (2018); Rabbi et al. (2014, 2015); Moumouni et al. (2015); Deokar et al. (2014); Xiong et al. (2016); Fatokun et al. (2018)
GWAS	Cassava, chickpea, finger millet, foxtail millet	Zhang et al. (2018); Jia et al. (2013); Li et al. (2018); Sharma et al. (2018)
Genome editing		
CRISPR/Cas 9	Cassava, ground cherry (Physalis pruinosa)	Lemmon et al. (2018); Bull et al. (2018); Gomez et al. (2018); Odipio et al. (2017); Hummel et al. (2018)
High throughput		
TILLING	Tef	Esfeld et al. (2013a)
EcoTILLING	Chickpea	Bajaj et al. (2016)
Omics		
Genome sequence	Chickpea, cowpea, enset, pearl millet, pigeon pea, tef, 101 African crops	Cannarozzi et al. (2014); Hittalmani et al. (2017); Hatakeyama et al. (2017); AOCC (2018); Varshney et al. (2017b); Mahato et al. (2018); Varshney et al. (2017a); Yematawa et al. (2018); Parween et al. (2015); Munoz-Amatriain et al. (2017)
Transcriptome sequence	Chickpea, tef	Cannarozzi et al. (2014); Verma et al. (2015)
RNASeq	Chickpea, finger millet, tef	Cannarozzi et al. (2018b); Hittalmani et al. (2017); Kudapa et al. (2018)
Proteome sequence	Chickpea, pearl millet, tef	Ghatak et al. (2016); Kamies et al. 2017; Bhushan et al. (2007)

Table 3 Examples of advanced breeding and genomic techniques implemented on selected orphan crops

GBS genotype by sequencing, GWAS genome-wide association study, CRISPR/Cas 9 clustered regularly interspaced short palindromic repeats/ CRISPR-associated protein9, TILLING Targeting Induced Local Lesions in Genomes

Institutions dedicated to research and improvement of orphan crops

National research institutions

Research and higher learning institutions in developing countries are involved in the improvement of significant crops of their respective countries. However, the expected level of advancement has not been made on many locally important crops due to limited investment and lack of skilled personnel.

International institutions

In addition to CGIAR centers indicated above, some higher learning institutions in developed countries are also interested in improving a single or multiple orphan crops. These institutions have been providing substantial technical and financial support. Few of these institutions are briefly presented below.

Crops for the Future (CFF)

The center, based at Semenyih in Malaysia, is dedicated to the development of underutilized crops, increasing income for producers and enhancing nutrition (CFF 2019). The center is affiliated with Nottingham University (UK) where advanced research on Bambara groundnut is performed.

African Orphan Crops Consortium (AOCC)

The main partners of consortium are new partnership for Africa's Development (NEPAD), Mars incorporated, World Agroforestry Centre (ICRAF), BGI, University of California, and Biosciences Eastern and Central Africa (BecA). The consortium is committed to sequence the genome of 101 traditional African food crops with high nutritional value which include cereals, vegetables, root and tubers and fruit trees (AOCC 2018).

World Vegetable Center

The center is based at Shanua in Taiwan and has six regional centers in Asia and Africa. The mandate of the World Vegetable Center is to conduct research and promote vegetables including orphan vegetables. To date, over 60,000 accessions of 440 species from 151 countries are conserved at the center out of which 10,000 accessions are traditional vegetables (AVRDC 2019).

Tef Improvement Project

The project aims at improving the orphan cereal crop tef and it is hosted at the Institute of Plant Sciences at the University of Bern in Switzerland. The project employs key genetic and genomic tools (Esfeld et al. 2013b; Girma et al. 2018) and collaborates with the National Tef Improvement Program in Ethiopia to develop tef varieties with desirable traits (Cannarozzi et al. 2018a; Kebede et al. 2018). The group led an international group of scientists which resulted in the genome sequence of the draft genome of tef (Cannarozzi et al. 2014).

Funding institutions

A list of institutions involved in orphan crop research, development and funding was earlier presented (Tadele 2009c). Here, only few institutions which are currently providing financial support to orphan crops research are presented.

Biotechnology and Biological Sciences Research Council (BBSRC): through Sustainable Agriculture for Sub-Saharan Africa (SASSA) program, it provides financial grant to address research challenges relating to the sustainable intensification of agricultural systems in Africa (BBSRC 2019).

Bill and Melinda Gates Foundation (BMGF): In addition to supporting the germplam conservation at Svalbard Seed Vault and sequencing the 101 African crops, the foundation supports the genetic improvement of cassava (BMGF 2019).

McKnight Foundation: throughits Collaborative Crop Research Program (CCRP), the foundation provides financial support for research and development of vital crops in Andes and Africa. Among orphan crops, Bambara groundnut, cowpea and finger millet are currently receiving grants from the foundation (CCRP 2019).

Syngenta Foundation for Sustainable Agriculture (SFSA). It provides both financial and technical support to projects in Africa, Asia and South America. Among these, the foundation supports the scientific work on an orphan crop tef (SFSA 2019).

International organizations: this includes organizations such as the World Bank, Food and Agriculture Organization (FAO) and International Fund for Agricultural Development (IFAD) who provide support to the work on orphan crop through their projects in developing countries.

Suggestions for improving orphan crops

Diverse types of orphan crops are cultivated at diverse agro-ecological and socio-economic conditions of developing world. Due to these extensive variabilities, the same types of suggestions cannot be forwarded for the research and development of these neglected crops. However, the following general suggestions can be provided to promote orphan crops.

Policy-related suggestions

A favorable policy or enabling environment is required to promote the research and development of orphan crops (Tadele 2017). These include the following:

Implementation of the right type of strategy

Locally appropriate technologies need to be developed in addition to human and institutional capacity building as well as forming conducive policy. Governments also need to implement land, marketing, and credit policies conducive to support agricultural development.

Investment on innovative agriculture

Stakeholders involved in agricultural research and development need to invest in agricultural innovation as it contributes towards improving the production, marketing or distribution system. An innovative Push–Pull Technology earlier developed in Kenya to control insect pests and a parasitic weed from maize fields has received a high acceptance in East Africa (Murage et al. 2015).

Provision of inputs and credit

In addition to policy-related and environmental constraints, poor crop productivity of orphan crops is due to the use of inefficient agricultural practices which begin from land preparation, weeding, harvesting and finally to threshing and storage. In addition, sub-optimal use of input such as fertilizers, herbicides and pesticides contributes to the low productivity of crops (Denning et al. 2009). The fertility of the land and the land tenure system has a huge impact on crop productivity. The access of farmers to the inputs and credit is important as they affect the productivity of crops.

Creation of robust extension system

Success in agricultural development is not achieved without the adoption of improved technologies by a large number of farmers. This calls for the establishment of strong extension system, which links the research community to the farming community. The transfer of new technologies to end users and their acceptance by farmers are facilitated when farmers are involved in decision-making starting at an early stage of technology development.

Germplasm collection and utilization

The germplasm of many under-studied crops have not been properly collected and utilized by researchers. To avoid loss of genetic materials due to human and natural disasters, it is advisable to save the duplicate of all germplasm of orphan crops at Svalbard Seed Vault. To harness the genetic diversity among the landraces, the Germplasm needs to be available to researchers from both developed and developing countries.

Developing crops that adapt to changing climate

Since abiotic stresses such as drought, salinity and heat as well as climate change substantially affect the productivity of crops and food security, future research should also focus on developing resistance or tolerance against these environmental calamities. The ability of indigenous crops to resist one or more stresses increases the chance to be selected as a potential crop to be used as an alternative crop in the changing climate. However, some undesirable traits associated with these crops (e.g., low productivity) need to be improved.

Partnership with relevant stakeholders

Establishing efficient partnerships with the national, regional and international institutions are important for improving orphan crops. Nowadays, public–private partnership (PPP) is considered as an effective mechanism to bring together the public and the private sectors towards enhancing agricultural sustainability in the developing world.

Conclusions

Orphan crops are very important for food security especially in the developing world and for the diversity of genetic resources. Despite their huge importance, these crops have generally received little attention by the global scientific community. It is important to promote research on both the major and orphan crops. Hence, major changes in agriculture are required to increase food production for underresearched crops to feed the ever-increasing population of the developing world. The next Green Revolution needs to include these locally adapted crops that are mostly known as orphan or under-studied crops. Crop improvement by itself does not lead us to the final goal unless supported by policies that include extension, land tenure and credits. Strategies and policies conducive for boosting productivity need to be devised in addition to establishing partnerships with relevant stakeholders. These partnerships among relevant stakeholders are necessary to tackle the challenges especially in the face of climate change.

Author contribution statement ZT conceived the idea, wrote the paper and approved for publication.

Acknowledgements The Research in my lab is financially supported by the Syngenta Foundation for Sustainable Agriculture and University of Bern.

Compliance with ethical standards

Conflict of interest I declare that there is no financial interest or conflict of interest.

References

- AOCC (2018) African orphan crops consortium: healthy Africa through nutritious, diverse and local food crops. http://africanorp hancrops.org/. Accessed 8 Jan 2019
- Assefa K, Yu JK, Zeid M, Belay G, Tefera H, Sorrells ME (2011) Breeding tef [*Eragrostis tef* (Zucc.) trotter]: conventional and molecular approaches. Plant Breed 130(1):1–9. https://doi.org/1 0.1111/j.1439-0523.2010.01782.x
- AVRDC (2019) World vegetable center. https://avrdc.org. Accessed 29 Jan 2019
- Bajaj D, Srivastava R, Nath M, Tripathi S, Bharadwaj C, Upadhyaya HD, Tyagi AK, Parida SK (2016) EcoTILLING-based association mapping efficiently delineates functionally relevant natural allelic variants of candidate genes governing agronomic traits in chickpea. Front Plant Sci 7:450. https://doi.org/10.3389/ fpls.2016.00450
- Bazile D, Bertero D, Nieto C (2015) State of the art report of Quinoa in the world in 2013. Food and Agriculture Organization of the United Nations (FAO), Santiago
- BBSRC (2019) Sustainable agriculture for Sub-Saharan Africa (SASSA). https://bbsrc.ukri.org/news/food-security/2018/18082 8-pr-sustainable-agriculture-for-sub-saharan-africa/. Accessed 3 Feb 2019
- Bermejo JEH, León J (eds) (1994) Neglected crops: 1492 from a different perspective. Plant Production and Protection Series no. 26. FAO, Rome. (ISBN: 9251032173)
- Bhushan D, Pandey A, Choudhary MK, Datta A, Chakraborty S, Chakraborty N (2007) Comparative proteomics analysis of

differentially expressed proteins in chickpea extracellular matrix during dehydration stress. Mol Cell Proteom 6(11):1868–1884. https://doi.org/10.1074/mcp.M700015-MCP200

- BMGF (2019) Nutritious rice and cassava aim to help millions fight malnutrition. https://www.gatesfoundation.org/Media-Center/ Press-Releases/2011/04/Nutritious-Rice-and-Cassava-Aim-to-Help-Millions-Fight-Malnutrition. Accessed 3 Feb 2019
- Brandt SA (1997) The "tree against hunger": enset-based agricultural system in Ethiopia. American Association for the Advancement of Science, Washington, D.C.
- Bull SE, Seung D, Chanez C, Mehta D, Kuon JE, Truernit E, Hochmuth A, Zurkirchen I, Zeeman SC, Gruissem W, Vanderschuren H (2018) Accelerated ex situ breeding of GBSS- and PTST1-edited cassava for modified starch. Sci Adv. https://doi. org/10.1126/sciadv.aat6086
- Campell CG (1997) Grass pea (*Lathyrus sativus* L.). Promoting the conservation and use of underutilized and neglected crops 18. IPK, Gartersleben/IPGRI, Rome
- Cannarozzi G, Plaza-Wuthrich S, Esfeld K, Larti S, Wilson YS, Girma D, de Castro E, Chanyalew S, Blosch R, Farinelli L, Lyons E, Schneider M, Falquet L, Kuhlemeier C, Assefa K, Tadele Z (2014) Genome and transcriptome sequencing identifies breeding targets in the orphan crop tef (Eragrostis tef). BMC Genomics 15:581. https://doi.org/10.1186/1471-2164-15-581
- Cannarozzi G, Chanyalew S, Assefa K, Bekele A, Blosch R, Weichert A, Klauser D, Plaza-Wuthrich S, Esfeld K, Jost M, Rindisbacher A, Jifar H, Johnson-Chadwick V, Abate E, Wang WY, Kamies R, Husein N, Kebede W, Tolosa K, Genet Y, Gebremeskel K, Ferede B, Mekbib F, Martinelli F, Pedersen HC, Rafudeen S, Hussein S, Tamiru M, Nakayama N, Robinson M, Barker I, Zeeman S, Tadele Z (2018a) Technology generation to dissemination: lessons learned from the tef improvement project. Euphytica 214(2):31. https://doi.org/10.1007/s10681-018-2115-5
- Cannarozzi G, Weichert A, Schnell M, Ruiz C, Bossard S, Blösch R, Plaza-Wüthrich S, Chanyalew S, Assefa K, Tadele Z (2018b) Waterlogging affects plant morphology and the expression of key genes in tef (Eragrostis tef). Plant Direct 2018:1–22
- CCRP (2019) Collaborative crop research program (CCRP), McKnight foundation. http://www.ccrp.org/. Accessed 3 Feb 2019
- Ceballos H, Iglesias CA, Perez JC, Dixon AGO (2004) Cassava breeding: opportunities and challenges. Plant Mol Biol 56(4):503–516
- CEPLAS (2018) Conference report: first cologne conference on food for future. CEPLAS (Cluster of Excellence in Plant Science). https://www.ceplas.eu/en/discover/competence-area-food-secur ity/conference-on-food-for-future/. Accessed 10 Jan 2019
- CFF (2019) Crops for the future. http://www.cffresearch.org/. Accessed 7 Jan 2019
- CGIAR (2019a) Genebank platform. https://www.genebanks.org/. Accessed 30 Jan 2019
- CGIAR (2019b) Research centers. https://www.cgiar.org/research/resea rch-centers/. Accessed 10 Jan 2019
- Chahota RK, Sharma TR, Sharma SK, Kumar N, Rana JC (2013) Horsegram. In: Singh M, Upadhyaya HD, Bisht IS (eds) Genetic and genomic resources of grain legume improvement. Elsevier, London, Waltham
- Chandrashekar A (2010) Finger millet *Eleusine coracana*. Adv Food Nutr Res 59:215–262. https://doi.org/10.1016/S1043 -4526(10)59006-5
- Chiurugwi T, Kemp S, Powell W, Hickey LT (2018) Speed breeding orphan crops. Theor Appl Genet. https://doi.org/10.1007/s0012 2-018-3202-7
- Chivenge P, Mabhaudhi T, Modi AT, Mafongoya P (2015) The potential role of neglected and underutilised crop species as future crops under water scarce conditions in Sub-Saharan Africa. Int J Env Res Pub He 12(6):5685–5711. https://doi.org/10.3390/ijerp h120605685

- CropTrust (2019) Svalbard global seed vault. https://www.cropt rust.org/our-work/svalbard-global-seed-vault/. Accessed 11 Jan 2019
- CSA (2015) Agricultural sample survey 2014/2015, vol I. Report on area and production of major crops. Statistical Bulletin 578. Central Statistical Agency (CSA), Addis Ababa
- Dawson I, Jaenicke H (2006) Underutilised plant species: the role of biotechnology. International Centre for Underutilized Crops, Position Paper No. 1, Colombo
- Denning G, Kabambe P, Sanchez P, Malik A, Flor R, Harawa R, Nkhoma P, Zamba C, Banda C, Magombo C, Keating M, Wangila J, Sachs J (2009) Input subsidies to improve smallholder maize productivity in Malawi: toward an African Green Revolution. PLoS Biol 7(1):2–10. https://doi.org/10.1371/journ al.pbio.1000023
- Deokar AA, Ramsay L, Sharpe AG, Diapari M, Sindhu A, Bett K, Warkentin TD, Tar'an B (2014) Genome wide SNP identification in chickpea for use in development of a high density genetic map and improvement of chickpea reference genome assembly. BMC Genomics 15(1):708. https://doi.org/10.1186/1471-2164-15-708
- EcoBusiness (2015) What can nearly 12,000 'orphan crops' do to address the nutrition gap? October 14, 2015. https://www.ecobusiness.com/news/what-can-nearly-12000-orphan-crops-do-toaddress-the-nutrition-gap/. Accessed 7 Jan 2019
- Ejeta G (2010) African green revolution needn't be a Mirage. Science 327:831–832
- Esfeld K, Plaza-Wüthrich S, Tadele Z (2013a) TILLING as a highthroughput technique of tef improvement. In: Assefa K, Chanyalew S, Tadele Z (eds) Achievements and prospects of tef improvement. EIAR-University of Bern, Bern, pp 53–65
- Esfeld K, Uauy C, Tadele Z (2013b) Application of TILLING for Orphan Crop Improvement. In: Jain SM, Gupta SD (eds) Biotechnology of neglected and underutilized crops. Springer, Berlin, pp 83–113
- Fahey JW (1998) Underexploited African grain crops: a nutritional resource. Nutr Rev 56(9):282–285
- FAO (2006) Food security. Policy Brief. Food and Agriculture Organization (FAO), Rome
- FAO (2013) The international year of Quinoa. Food and Agriculture Organization, Rome. http://www.fao.org/quinoa-2013/en/. Accessed 10 Jan 2019
- FAO (2017) Promoting neglected and underutilized crop species. Food and Agriculture Organization (FAO), Rome. http://www.fao.org/ news/story/en/item/1032516/icode/. Accessed 7 Apr 2019
- FAOSTAT (2018) FAO (Food and Agricultural Organization) statistical data. FAO statistics division http://www.fao.org/faostat/en/#data. Accessed 8 Jan 2019
- Fatokun C, Girma G, Abberton M, Gedil M, Unachukwu N, Oyatomi O, Yusuf M, Rabbi I, Boukar O (2018) Genetic diversity and population structure of a mini-core subset from the world cowpea (*Vigna unguiculata* (L.) Walp.) germplasm collection. Sci Rep 8(1):16035. https://doi.org/10.1038/s41598-018-34555-9
- Fauchereau N, Trzaska S, Rouault M, Richard Y (2003) Rainfall variability and changes in Southern Africa during the 20th century in the global warming context. Nat Hazards 29(2):139–154
- Fungo R (2009) Potential of bananas in alleviating micronutrient efficiencies in the great lakes region of East Africa. African Crop Sci Conf Proc 9:8
- Funk C, Dettinger MD, Michaelsen JC, Verdin JP, Brown ME, Barlow M, Hoell A (2008) Warming of the Indian Ocean threatens eastern and southern African food security but could be mitigated by agricultural development. Proc Natl Acad Sci USA 105(32):11081–11086. https://doi.org/10.1073/pnas.0708196105
- Gegios A, Amthor R, Maziya-Dixon B, Egesi C, Mallowa S, Nungo R, Gichuki S, Mbanaso A, Manary MJ (2010) Children consuming cassava as a staple food are at risk for inadequate zinc, iron, and

vitamin A intake. Plant Food Hum Nutr 65(1):64–70. https://doi. org/10.1007/s11130-010-0157-5

- Gemede HF, Ratta N, Haki GD, Woldegiorgis AZ (2014) Nutritional quality and health benefits of okra (*Abelmoschus Esculentus*): a review. Global J Med Res 14:5
- Getinet A, Sharma SM (1996) *Niger, Guizotia abyssinica* (L. f.) Cass. Institute of Plant Genetics and Crop Plant Research, Gatersleben/ International Plant Genetic Resources Institute, Rome
- Ghatak A, Chaturvedi P, Nagler M, Roustan V, Lyon D, Bachmann G, Postl W, Schröfl A, Desai N, Varshney RK, Weckwerth W (2016) Comprehensive tissue-specific proteome analysis of drought stress responses in *Pennisetum glaucum* (L.) R. Br. (Pearl millet). J Proteomics 143:122–135
- Girma D, Cannarozzi G, Weichert A, Tadele Z (2018) Genotyping by sequencing reasserts the close relationship between tef and its putative wild *Eragrostis progenitors*. Diversity 10(2):17
- Godfray HC, Beddington JR, Crute IR, Haddad L, Lawrence D, Muir JF, Pretty J, Robinson S, Thomas SM, Toulmin C (2010) Food security: the challenge of feeding 9 billion people. Science 327(5967):812–818. https://doi.org/10.1126/science.1185383
- Gomez MA, Lin ZD, Moll T, Chauhan RD, Hayden L, Renninger K, Beyene G, Taylor NJ, Carrington JC, Staskawicz BJ, Bart RS (2018) Simultaneous CRISPR/Cas9-mediated editing of cassava eIF4E isoforms nCBP-1 and nCBP-2 reduces cassava brown streak disease symptom severity and incidence. Plant Biotechnol J. https://doi.org/10.1111/pbi.12987
- Gordon E (2014) Teff, Ethiopia's nutritious grain. The Washington Post. https://www.washingtonpost.com/lifestyle/wellness/ teff-ethiopias-nutritious-grain/2014/04/08/0954563e-b9ab-11e3-9a05-c739f29ccb08_story.html?noredirect=on&utm_ term=.9adcb4fb741f. Accessed 7 June 2019
- Gupta SM, Arora S, Mirza N, Pande A, Lata C, Puranik S, Kumar J, Kumar A (2017) Finger millet: a "Certain" Crop for an "Uncertain" future and a solution to food insecurity and hidden hunger under stressful environments. Front Plant Sci. https://doi. org/10.3389/fpls.2017.00643
- Hatakeyama M, Aluri S, Balachadran MT, Sivarajan SR, Patrignani A, Grüter S, Lucy Poveda L, Shimizu-Inatsugi R, Baeten J, Francoijs KJ, Nataraja KN, Reddy YAN, Phadnis S, Ravikumar RL, Schlapbach R, Sreeman SM, Shimizu KK (2017) Multiple hybrid de novo genome assembly of finger millet, an orphan allotetraploid crop. DNA Res. https://doi.org/10.1093/dnares/ dsx036 (dsx036)
- Heslop-Harrison JS, Schwarzacher T (2007) Domestication, genomics and the future for banana. Ann Bot-London 100(5):1073–1084. https://doi.org/10.1093/Aob/Mcm191
- Hittalmani S, Mahesh HB, Shirke MD, Biradar H, Uday G, Aruna YR, Lohithaswa HC, Mohanrao A (2017) Genome and transcriptome sequence of finger millet (*Eleusine coracana* (L.) Gaertn.) provides insights into drought tolerance and nutraceutical properties. BMC Genomics. https://doi.org/10.1186/s12864-017-3850-z
- Hummel AW, Chauhan RD, Cermak T, Mutka AM, Vijayaraghavan A, Boyher A, Starker CG, Bart R, Voytas DF, Taylor NJ (2018) Allele exchange at the EPSPS locus confers glyphosate tolerance in cassava. Plant Biotechnol J 16(7):1275–1282. https:// doi.org/10.1111/pbi.12868
- ICARDA (2015) ICARDA's seed retrieval mission from Svalbard Seed Vault http://www.icarda.org/update/icarda%E2%80%99s-seedretrieval-mission-svalbard-seed-vault. Accessed 31 Jan 2019
- Jeffrey J (2015) Will Ethiopia's teff be the next 'super grain'? BBC Business. https://www.bbc.com/news/business-32128441. Accessed 7 June 2019
- Jia GQ, Huang XH, Zhi H, Zhao Y, Zhao Q, Li WJ, Chai Y, Yang LF, Liu KY, Lu HY, Zhu CR, Lu YQ, Zhou CC, Fan DL, Weng QJ, Guo YL, Huang T, Zhang L, Lu TT, Feng Q, Hao HF, Liu HK, Lu P, Zhang N, Li YH, Guo EH, Wang SJ, Wang SY, Liu

🖄 Springer

JR, Zhang WF, Chen GQ, Zhang BJ, Li W, Wang YF, Li HQ, Zhao BH, Li JY, Diao XM, Han B (2013) A haplotype map of genomic variations and genome-wide association studies of agronomic traits in foxtail millet (*Setaria italica*). Nat Genet 45(8):957. https://doi.org/10.1038/ng.2673

- Kamadi G (2014) Plant breeders to boost Africa's indigenous crops. https://www.un.org/africarenewal/magazine/august-2014/ plant-breeders-boost-africa%E2%80%99s-indigenous-crops. Accessed 9 Jan 2019
- Kamies R, Farrant JM, Tadele Z, Cannarozzi G, Rafudeen MS (2017) A proteomic approach to investigate the drought response in the orphan crop Eragrostis tef. Proteomes 5(4):32
- Kebede W, Tolossa K, Hussein N, Fikre T, Genet Y, Bekele A, Gebremeskel K, Fentahun A, Daba H, Plaza-Wüthrich S, Blösch R, Chanyalew S, Asefa K, Tadele Z (2018) Tef (*Era-grostis tef*) Variety Tesfa. Ethiopian Journal of Agricultural Sciences 28(2):107–112
- Ketema S (1997) Tef, *Eragrostis tef* (Zucc.) Trotter. Institute of Plant Genetics and Crop Plant Research, Gatersleben/International Plant Genetic Resources Institute, Rome
- Kholová J, Hash CT, Kumar PL, Yadav RS, Kocová M, Vadez V (2010) Terminal drought-tolerant pearl millet [*Pennisetum* glaucum (L.) R. Br.] have high leaf ABA and limit transpiration at high vapour pressure deficit. J Exp Botany 61(5):1431–1440
- Kudapa H, Garg V, Chitikineni A, Varshney RK (2018) The RNA-Seq-based high resolution gene expression atlas of chickpea (*Cicer arietinum* L.) reveals dynamic spatio-temporal changes associated with growth and development. Plant, Cell Environ 41(9):2209–2225. https://doi.org/10.1111/pce.13210
- Kumar S, Gupta P, Barpete S, Sarker A, Amri A, Mathur PN, Baum M (2013) Grasspea. In: Singh M, Upadhyaya HD, Bisht IS (eds) Genetic and genomic resources of Grain Legume improvement. Elsevier, Amsterdam, pp 269–292
- Kumar A, Sharma D, Tiwari A, Jaiswal JP, Singh NK, Sood S (2016) Genotyping-by-sequencing analysis for determining population structure of finger millet germplasm of diverse origins. Plant Genome. https://doi.org/10.3835/plantgenome2015.07.0058
- Léder I (2004) Sorghum and millets. In: Füleky G (ed) Cultivated plants primarily as food sources. UNESCO, Eolss Publishers, Oxford
- Lemmon ZH, Reem NT, Dalrymple J, Soyk S, Swartwood KE, Rodriguez-Leal D, Van Eck J, Lippman ZB (2018) Rapid improvement of domestication traits in an orphan crop by genome editing. Nature Plants 4(10):766–770. https://doi.org/10.1038/ s41477-018-0259-x
- Li X, Siddique KHM (2018) Future smart food—rediscovering hidden treasures of neglected and underutilized species for zero hunger in Asia. Food and Agriculture Organization of the United Nations Bangkok, Rome
- Li YL, Ruperao P, Batley J, Edwards D, Khan T, Colmer TD, Pang JY, Siddique KHM, Sutton T (2018) Investigating drought tolerance in chickpea using genome-wide association mapping and genomic selection based on whole-genome resequencing data. Front Plant Sci 9:190. https://doi.org/10.3389/ fpls.2018.00190
- Linares OF (2002) African rice (*Oryza glaberrima*): history and future potential. Proc Natl Acad Sci USA 99(25):16360–16365. https:// doi.org/10.1073/pnas.252604599
- Lugar (2019) Orphan crops: the Lugar Center. http://www.thelugarce nter.org/ourwork-Orphan-Crops.html. Accessed 10 Jan 2019
- Mabaya E, Emana B, Mulugeta F, Mugoya M (2017) Ethiopia Brief 2017: The African Seed Access Index. TASAI. https://tasai.org/ Accessed 7 Apr 2019
- Mahato AK, Sharma AK, Sharma TR, Singh NK (2018) An improved draft of the pigeonpea (*Cajanus cajan* (L.) Millsp.) genome. Data Brief 16:376–380. https://doi.org/10.1016/j.dib.2017.11.066

- Massawe FJ, Mayes S, Cheng A, Chai HH, Cleasby P, Symonds R, Ho WK, Siise A, Wong QN, Kendabie P, Yanusa Y, Jamalluddin N, Singh A, Azman R, Azam-Ali SN (2015) The potential for underutilised crops to improve food security in the face of climate change. Agric Clim Change 29:140–141. https://doi. org/10.1016/j.proenv.2015.07.228
- Mir NA, Riar CS, Singh S (2018) Nutritional constituents of pseudo cereals and their potential use in food systems: a review. Trends Food Sci Tech 75:170–180. https://doi.org/10.1016/j. tifs.2018.03.016
- Mollins J (2017) Smallholder farmers to gain from targeted CRISPR-Cas9 crop breeding. https://www.cimmyt.org/smallholder-farme rs-to-gain-from-targeted-crispr-cas9-breeding/. Accessed 9 Jan 2019
- Moumouni KH, Kountche BA, Jean M, Hash CT, Vigouroux Y, Haussmann BIG, Belzile F (2015) Construction of a genetic map for pearl millet, *Pennisetum glaucum* (L.) R. Br., using a genotyping-by-sequencing (GBS) approach. Mol Breeding. https://doi. org/10.1007/s11032-015-0212-x
- Muller C, Cramer W, Hare WL, Lotze-Campen H (2011) Climate change risks for African agriculture. Proc Natl Acad Sci USA 108:4313–4315. https://doi.org/10.1073/pnas.1015078108
- Munoz-Amatriain M, Mirebrahim H, Xu P, Wanamaker SI, Luo M, Alhakami H, Alpert M, Atokple I, Batieno BJ, Boukar O, Bozdag S, Cisse N, Drabo I, Ehlers JD, Farmer A, Fatokun C, Gu YQ, Guo YN, Huynh BL, Jackson SA, Kusi F, Lawley CT, Lucas MR, Ma Y, Timko MP, Wu J, You F, Barkley NA, Roberts PA, Lonardi S, Close TJ (2017) Genome resources for climateresilient cowpea, an essential crop for food security. Plant J 89(5):1042–1054. https://doi.org/10.1111/tpj.13404
- Murage AW, Midega CAO, Pittchar JO, Pickett JA, Khan ZR (2015) Determinants of adoption of climate-smart push-pull technology for enhanced food security through integrated pest management in eastern Africa. Food Secur 7(3):709–724. https://doi. org/10.1007/s12571-015-0454-9
- NRC (1996) Lost crops of Africa: grains. National Research Council (NRC), vol 1. National Academy Press, Washington, DC
- NRC (2006) Lost Crops of Africa: vegetables. National Research Council (NRC), vol 2. National Academies Press, Washington, DC
- NRC (2008) Lost crops of Africa: fruits. National Research Council (NRC), vol 3. National Academies Press, Washington, DC
- O'Connor A (2016) Is teff the new super grain? The New York Times. https://well.blogs.nytimes.com/2016/08/16/is-teff-the-new-super -grain/. Accessed 7 June 2019
- Odipio J, Alicai T, Ingelbrecht I, Nusinow DA, Bart R, Taylor NJ (2017) Efficient CRISPR/Cas9 genome editing of Phytoene desaturase in Cassava. Front Plant Sci. https://doi.org/10.3389/ fpls.2017.01780
- Okpala B (2015) Benefits of cocoyam (Taro: Colocasia esculenta). https://globalfoodbook.com/benefits-of-cocoyam. Accessed 9 Jan 2019
- Olango TM, Tesfaye B, Catellani M, Pe ME (2014) Indigenous knowledge, use and on-farm management of enset (*Ensete ventricosum* (Welw.) Cheesman) diversity in Wolaita, Southern Ethiopia. J Ethnobiol Ethnomed 10:41. https://doi. org/10.1186/1746-4269-10-41
- Padulosi S (2017) Bring NUS back on the table. GREAT Insights 6(4):21–22
- Pal D, Mishra P, Sachan N, Ghosh AK (2011) Biological activities and medicinal properties of *Cajanus cajan* (L) Millsp. J Adv Pharm Technol Res 2(4):207–214. https://doi. org/10.4103/2231-4040.90874
- Parween S, Nawaz K, Roy R, Pole AK, Suresh BV, Misra G, Jain M, Yadav G, Parida SK, Tyagi AK, Bhatia S, Chattopadhyay D (2015) An advanced draft genome assembly of a desi type

chickpea (*Cicer arietinum* L.). Sci Rep UK 5:12806. https:// doi.org/10.1038/srep12806

- Pearce F (2013) Executive perspective: a call to remember the forgotten crops. https://blogs.thomsonreuters.com/sustainabi lity/2013/12/20/executive-perspective-call-remember-forgo tten-crops/. Accessed 10 Jan 2019
- Provost C, Jobson E (2014) Move over quinoa, Ethiopia's teff poised to be next big super grain. The Guardian. https://www.thegu ardian.com/global-development/2014/jan/23/quinoa-ethiopiateff-super-grain. Accessed 7 June 2019
- Rabbi I, Hamblin M, Gedil M, Kulakow P, Ferguson M, Ikpan AS, Ly D, Jannink JL (2014) Genetic mapping using genotypingby-sequencing in the clonally propagated cassava. Crop Sci 54(4):1384–1396. https://doi.org/10.2135/cropsci2013.07.0482
- Rabbi IY, Kulakow PA, Manu-Aduening JA, Dankyi AA, Asibuo JY, Parkes EY, Abdoulaye T, Girma G, Gedil MA, Ramu P, Reyes B, Maredia MK (2015) Tracking crop varieties using genotyping-by-sequencing markers: a case study using cassava (*Manihot esculenta* Crantz). BMC Genet 16:115
- Raheem D (2011) The need for agro-allied industries to promote food security by value addition to indigenous African food crops. Outlook Agric 40(4):343–349. https://doi.org/10.5367/ Oa.2011.0063
- Sanchez AC (2018) Baobabs are Africa's oldest and most beautiful trees, but they're under threat. https://www.independent.co. uk/environment/baobab-trees-africa-dying-oldest-under-threa t-a8404136.html. Accessed 29 Jan 2019
- Sanginga N, Lyasse O, Singh BB (2000) Phosphorus use efficiency and nitrogen balance of cowpea breeding lines in a low P soil of the derived savanna zone in West Africa. Plant Soil 220(1-2):119–128
- Sarr B (2012) Present and future climate change in the semi-arid region of West Africa: a crucial input for practical adaptation in agriculture. Atmos Sci Lett 13(2):108–112. https://doi.org/10.1002/ Asl.368
- SFSA (2019) Syngenta foundation for sustainable agriculture. https:// www.syngentafoundation.org/. Accessed 3 Feb 2019
- Sharma D, Tiwari A, Sood S, Jamra G, Singh NK, Meher PK, Kumar A (2018) Genome wide association mapping of agro-morphological traits among a diverse collection of finger millet (*Eleusine coracana* L.) genotypes using SNP markers. PLoS One 13(8):e0199444. https://doi.org/10.1371/journal.pone.0199444
- Shongwe ME, van Oldenborgh GJ, van den Hurk B, van Aalst M (2011) Projected changes in mean and extreme precipitation in Africa under global warming. Part II: East Africa. J Climate 24(14):3718–3733. https://doi.org/10.1175/2010jcli2883.1
- Spaenij-Dekking L, Kooy-Winkelaar Y, Koning F (2005) The Ethiopian cereal tef in celiac disease. N Engl J Med 353:1748–1749. https://doi.org/10.1056/NEJMc051492
- Stephenson K, Amthor R, Mallowa S, Nungo R, Maziya-Dixon B, Gichuki S, Mbanaso A, Manary M (2010) Consuming cassava as a staple food places children 2–5 years old at risk for inadequate protein intake, an observational study in Kenya and Nigeria. Nutr J 9:9
- Tadele Z (ed) (2009a) New approaches to plant breeding of orphan crops in Africa. Stämpfli AG, Bern
- Tadele Z (2009b) Role of orphan crops in enhancing and diversifying food production in Africa. Afr Technol Dev Forum J 6(3/4):9–15
- Tadele Z (2009c) Who is who in African orphan crops research, information and development. Afr Technol Dev Forum J 6(3/4):72–81
- Tadele Z (2014) Role of crop research and development in food security of Africa. Int J Plant Biol Res 2:1019
- Tadele Z (2017) Raising crop productivity in Africa through intensification. Agron J 7:22
- Tadele Z (2018) African orphan crops under abiotic stresses: challenges and opportunities. Scientifica 2018:1451894

- Tadele Z (2019) Challenges of food security for orphan crops. Encycl Food Secur Sustain 1:403–408
- Tadele Z, Assefa K (2012) Increasing food production in Africa by boosting the productivity of understudied crops. Agronomy 2(4):240–283
- Varshney RK, Saxena RK, Upadhyaya HD, Khan AW, Yu Y, Kim C, Rathore A, Kim D, Kim J, An S, Kumar V, Anuradha G, Yamini KN, Zhang W, Muniswamy S, Kim JS, Penmetsa RV, von Wettberg E, Datta SK (2017a) Whole-genome resequencing of 292 pigeonpea accessions identifies genomic regions associated with domestication and agronomic traits. Nat Genet 49(7):1082. https ://doi.org/10.1038/ng.3872
- Varshney RK, Shi CC, Thudi M, Mariac C, Wallace J, Qi P, Zhang H, Zhao YS, Wang XY, Rathore A, Srivastava RK, Chitikineni A, Fan GY, Bajaj P, Punnuri S, Gupta SK, Wang H, Jiang Y, Couderc M, Katta MAVSK, Paudel DR, Mungra KD, Chen WB, Harris-Shultz KR, Garg V, Desai N, Doddamani D, Kane NA, Conner JA, Ghatak A, Chaturvedi P, Subramaniam S, Yadav OP, Berthouly-Salazar C, Hamidou F, Wang JP, Liang XM, Clotault J, Upadhyaya HD, Cubry P, Rhone B, Gueye MC, Sunkar R, Dupuy C, Sparvoli F, Cheng SF, Mahala RS, Singh B, Yadav RS, Lyons E, Datta SK, Hash CT, Devos KM, Buckler E, Bennetzen JL, Paterson AH, Ozias-Akins P, Grando S, Wang J, Mohapatra T, Weckwerth W, Reif JC, Liu X, Vigouroux Y, Xu X (2017b) Pearl millet genome sequence provides a resource to improve agronomic traits in arid environments. Nat Biotechnol 35(10):969. https://doi.org/10.1038/nbt.3943
- Verma M, Kumar V, Patel RK, Garg R, Jain M (2015) CTDB: an integrated Chickpea Transcriptome database for functional and applied genomics. PLoS One 10(8):0136880. https://doi.org/10.1371/journal.pone.0136880
- WheatAtlas (2019) Released wheat varieties. http://beta.wheatatlas.org/ varieties. Accessed 30 Jan 2019

- Williams JT, Haq N (2000) Global research on underutilised crops: an assessment of current activities and proposals for enhanced cooperation. Southampton, UK
- Xiong H, Shi A, Mou B, Qin J, Motes D, Lu W, Ma J, Weng Y, Yang W, Wu D (2016) Genetic diversity and population structure of cowpea (Vigna unguiculata L. Walp). PLoS One 11(8):e0160941. https://doi.org/10.1371/journal.pone.0160941
- Yan ZY, Spencer PS, Li ZX, Liang YM, Wang YF, Wang CY, Li FM (2006) *Lathyrus sativus* (grass pea) and its neurotoxin ODAP. Phytochemistry 67(2):107–121. https://doi.org/10.1016/j.phyto chem.2005.10.022
- Ye-Ebiyo Y, Pollack RJ, Spielman A (2000) Enhanced development in nature of larval *Anopheles arabiensis* mosquitoes feeding on maize pollen. Am J Trop Med Hyg 63(1–2):90–93
- Yematawa Z, Muzemil S, Ambachew D, Tripathi L, Tesfaye K, Chala A, Farbos A, O'Neill P, Moore K, Grant M, Studholme DJ (2018) Genome sequence data from 17 accessions of *Ensete ventrico-sum*, a staple food crop for millions in Ethiopia. Data in Brief 18:285–293. https://doi.org/10.1016/j.dib.2018.03.026
- Zhang SK, Chen X, Lu C, Ye JQ, Zou ML, Lu KD, Feng SB, Pei JL, Liu C, Zhou XC, Ma PA, Li ZG, Liu CJ, Liao Q, Xia ZQ, Wang WQ (2018) Genome-wide association studies of 11 agronomic traits in cassava (*Manihot esculenta* Crantz). Front Plant Sci. https://doi.org/10.3389/fpls.2018.00503

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.