Iron homeostasis alterations and risk for akathisia in patients treated with antipsychotics: a systematic review and meta-analysis of cross-sectional studies

Georgios Schoretsanitis, MD, PhD¹; Adriani Nikolakopoulou, PhD²; Daniel Guinart, MD¹; Christoph U.

Correll, MD^{1,3,4,5}; John M. Kane, MD^{1,3,4}

1. The Zucker Hillside Hospital, Department of Psychiatry, Northwell Health, Glen Oaks, New York, USA:

GS, DG, CUC, JMK

2. University of Bern, Institute of Social and Preventive Medicine, Bern, Switzerland: AN

3. The Donald and Barbara Zucker School of Medicine at Hofstra/Northwell, Department of Psychiatry and Molecular Medicine, Hempstead, New York, USA: **CUC, JMK**

4. The Feinstein Institute for Medical Research, Center for Psychiatric Neuroscience, Manhasset, New York, USA: **CUC**, JMK

5. Charité Universitätsmedizin, Department of Child and Adolescent Psychiatry, Berlin, Germany: CUC

Corresponding author: Georgios Schoretsanitis, MD, PhD The Zucker Hillside Hospital 7559 263rd Street Glen Oaks, NY 11004 Email: george.schor@gmail.com Phone: +1 718-470-5914 ORCID: http://orcid.org/0000-0002-3851-4117

Abstract

Iron homeostasis may be implicated in the pathophysiology of antipsychotic-related akathisia. We performed a systematic review in six databases from database inception until 03/2020, conducting a metaanalysis of studies investigating iron metabolism in antipsychotic-treated patients with versus without akathisia. Using a fixed- and a random-effects model, standardized mean difference (SMD) was estimated for levels of iron, ferritin, transferrin and total iron-binding capacity. Meta-regression analyses included sex, age, illness duration and antipsychotic treatment and dose. Subgroup analyses included chronic vs. acute akathisia and different diagnoses. Study quality was assessed using the Newcastle-Ottawa scale. In 10 studies (n=395), compared to non-akathisia patients (n=213), iron levels were lower in patients with akathisia (n=182; fixed-effect model: SMD=-0.49, 95%CI=-0.28,-0.70, p<0.001; random-effects model: SMD=-0.55, 95%CI=-0.14,-0.96, p=0.008). For secondary outcomes, differences were significant regarding lower ferritin levels in patients with akathisia in the fixed-effect model (SMD=-0.32, 95%CI=-0.08,-0.55, p=0.007), but not in the random-effects model (SMD=-0.29, 95%CI=0.20,-0.79, p=0.24). None of the moderators/mediators had a significant effect on the group difference of iron levels. Subgroup analyses reported lower iron levels in both patients with chronic and acute akathisia vs. patients without. Iron levels for schizophrenia patients were lower in the fixed-effect model (SMD=-0.55, 95%CI=-0.23, -0.86, p<0.001), while a trend was observed in the random-effects model (SMD=-0.52, 95%CI=-0.07, -1.12, p=0.08). The studies' quality was overall poor, with one exception. This meta-analysis suggests lower iron levels in akathisia patients, while ferritin differences were significant only in the fixed-effect model. Further data are required to promote the understanding of related pathways.

Keywords: iron metabolism; iron; akathisia; antipsychotic; movement disorders; meta-analysis

1. Introduction

Iron is a fundamental trace element implicated in numerous vital processes of human physiology (Andrews, 1999). Involvement of iron homeostasis disturbances has been hypothesized in the context of the pathophysiology for different mental illnesses (Kim et al., 2018; Percinel et al., 2016; Quinn et al., 2011; Shorter and Wachtel, 2013). In the context of schizophrenia, iron excess may be related to toxicity underpinning cognitive deficits (Cao et al., 2019). In particular, iron deficiency has also received attention in light of the mechanisms underlying motor symptoms associated with antipsychotic treatment. For example, lower iron levels were reported in patients with versus without catatonic symptoms (Peralta et al., 1999), whereas no differences were reported in patients developing antipsychotic-related dystonic reactions or tardive dyskinesia (Chong et al., 2004; Sachdev, 1994; Spina et al., 1994).

Iron homeostasis status markers have been more extensively studied in patients with akathisia, which is a relatively common adverse effect of both first-generation antipsychotics (FGAs) and second-generation antipsychotics (SGAs). In an overview of meta-analyses and Cochrane reviews, prevalences of akathisia ranged from 15.9% with chlorpromazine and 24.1% for haloperidol for FGAs, and from 3.3% with paliperidone and 5.2% with quetiapine to 8.5% with aripiprazole, 11.3% with amisulpride and 14.2% with risperidone for SGAs (Martino et al., 2018). In a meta-analysis of 48 studies of the most recently approved SGAs, the weighted mean incidence of akathisia was 7.7% (95% confidence interval (CI)=6.5-9.1), ranging from 3.9% (95%CI=2.4-6.3) for iloperidone to 6.8% (95%CI=5.1-9.0) for asenapine, 10.0% (95%CI=7.4-13.5) for brexpiprazole, 12.7% (95%CI=10.1-16.1) for lurasidone, and 17.2% (95%CI-13.4-22.1) for cariprazine (Demyttenaere et al., 2019). Several cross-sectional studies reported lower iron levels in patients with versus without antipsychotic-related akathisia (Barton et al., 1990; Brown et al., 1987; Hofmann et al., 2000; Horiguchi, 1991; Kuloglu et al., 2003; Valles et al., 1992), while other studies did not detect significant differences (Barnes et al., 1992; Juncal-Ruiz et al., 2017; Nemes et al., 1991; O'Loughlin et al., 1991; Soni et al., 1993). Discrepancies in these findings may be at least partially due to different demographic and clinical characteristics of study samples (Friedman, 1991). Further, two of these studies also included individuals without psychiatric diagnoses/medication as an additional control group (Kuloglu et al., 2003; Nemes et al., 1991). In one sample, a ranking order with iron levels being higher in healthy controls followed by patients without and finally those with akathisia was observed (Kuloglu et al., 2003), whereas in another sample, no significant intergroup differences were reported (Nemes et al., 1991).

There are two major frameworks linking depletion of iron stores and antipsychotic-related akathisia. One hypothesis explains this association in terms of direct antipsychotic-related effects on iron status (Calarge and Ziegler, 2013; O'Loughlin et al., 1991). Preclinical and clinical evidence have provided hints on psychotropic medications, including antipsychotics, such as aripiprazole and mixed SGAs, affecting levels of factors involved in maintaining iron homeostasis (Bogdan et al., 2015; Chen et al., 2018). For example, in aripiprazole-treated rats, elevated glycoprotein levels were reported, which is implicated in hepcidin response to iron depletion (Bogdan et al., 2015). Moreover, a case-control study assessed trace elements before and after treatment with SGAs in newly diagnosed patients with schizophrenia reporting lower iron levels at baseline compared to healthy controls (Chen et al., 2018). Antipsychotic treatment effects were only reported in males with elevated follow-up iron levels, whereas duration of antipsychotic treatment did not affect iron levels (Chen et al., 2018). Another possibility is that patients with low iron levels are at high-risk of developing akathisia, with pathways still remaining to be elucidated (Sachdev and Loneragan,

1991). A proposed link may encompass the oxidative effects of iron depletion on the dopaminergic system in the basal ganglia (Yanik et al., 2004), in that antipsychotic-related dopamine receptor upregulation leads to decreased inhibitory activity as well as dopamine receptor hypersensitivity (Solmi et al., 2018a; Solmi et al., 2018b). Nevertheless, prospective short-term data have not supported this hypothesis (O'Loughlin et al., 1991; Sachdev and Loneragan, 1991). However, medium-term or long-term antipsychotic-related effects on iron status are poorly understood. As early interest wore off, this area has not received attention in recent years. Conversely, the awareness of the need to identify and treat movement abnormalities in people with schizophrenia and movement disorders associated with antipsychotics has increased (Carbon et al., 2018; Demyttenaere et al., 2019; Martino et al., 2018; Solmi et al., 2018a; Solmi et al., 2018b). Thus, the purpose of this study was to systematically review and meta-analyse data on blood measures of iron status in adults with versus without antipsychotic-associated akathisia.

2. Experimental procedures

2.1. Search strategy

The study was conducted with use of PRISMA guidelines (Hutton et al., 2015) and registered with PROSPERO (registration number CRD42019121376). Studies of iron status and akathisia were identified by searching Embase and Medline using the following strategy: "iron" AND "akathisia". Databases were searched last in January 15, 2019 for publications without language restriction since data inception. References from identified studies were hand-searched for additional studies. An additional search in Scopus, PsycInfo, Googlescholar and Cochrane Library was performed on March 15, 2020.

2.2. Inclusion & exclusion criteria

Type of studies

Included were observational studies in antipsychotic-treated adults reporting on blood concentrations of iron status markers in patients with versus without antipsychotic-related akathisia regardless of the diagnosis and the setting. Case reports were not included.

Types of participants

Antipsychotic-treated adult patients of both sexes with versus without antipsychotic-related akathisia. There were no restrictions with regards to psychiatric diagnosis, setting, and dosage or duration of antipsychotic treatment. Patients receiving antipsychotic monotherapy as well as polypharmacy were considered. Patients with neurological diseases were excluded.

Comparator

Patients exposed to antipsychotics without antipsychotic-related akathisia.

Types of exposure

Any type of akathisia regardless of the assessment method in patients treated with antipsychotics.

Outcomes

The primary outcome was defined as standardized mean difference (SMD) for iron blood concentrations between patients with versus without antipsychotic-related akathisia. Secondary outcomes included SMDs for ferritin, transferrin and TIBC.

Selection of eligible studies was independently performed by two authors (GS and DG). In case of doubt, papers were discussed and consensus was reached. As consensus was reached in all cases, no additional co-author was involved.

2.3. Data extraction

One author (GS) extracted data, which were independently verified by another author (DG). Sample sizes, demographic characteristics, akathisia ratings, daily antipsychotic dosages, iron, ferritin, transferrin concentration as well as total iron-binding capacity (TIBC) values (mean and standard deviation [SD]) were extracted. Before data entry, values were converted to the same unit for each parameter. Weighted means for covariates were computed based on means of subgroups. One study provided levels for a group with tardive akathisia and dyskinesia (Sachdev, 1994). Based on provided data, means were computed for patients exclusively suffering from akathisia symptoms, while SDs of the whole group were adopted. The same strategy for missing SDs was applied in another study, where SDs for the whole group were available (O'Loughlin et al., 1991). In three studies (Barton et al., 1990; Brown et al., 1987; Horiguchi, 1991), data for means and variance measures were extracted from figures using the WebPlotDigitlizer (version 4.2 for Windows); in one of the studies, authors provided a mean value for four blood samples withdrawn within two days (Barton et al., 1990). In two cases, authors were contacted to provide missing data (Barnes et al., 1992; Treloar et al., 1994), but no data were available, given the age of the studies.

2.4. Quality of studies

The Newcastle-Ottawa scale for case-control studies and the modified version for cross-sectional studies were used for quality assessment (Herzog et al., 2013; Wijarnpreecha et al., 2019). For cross-sectional studies, we removed the item "representativeness of the exposed cohort" that we judge related to applicability, and added ascertainment of akathisia as described elsewhere (Li et al., 2016).

2.5. Statistical analysis

Outcomes were compared between groups of patients with versus without akathisia using fixed- and random-effects inverse-variance meta-analyses. Results were summarized using SMD and 95% confidence intervals (CI) and were presented in forest plots. Heterogeneity variance parameter (τ^2) was calculated using the DerSimonian-Laird estimator (DerSimonian and Laird, 1986). We also calculated the I-square (I^2) statistic, which measures the proportion of variability that can be attributed to heterogeneity rather than random error, with an I² >50% representing significant heterogeneity (Higgins et al., 2003). Thereafter, the effects of demographic and clinical parameters, including sex, age, duration of illness as well as antipsychotic treatment and daily dose (in chlorpromazine equivalents; CPZE) were assessed in a meta-regression analysis (Borenstein et al., 2009). We also included two subgroup analyses for the primary outcome, focusing separately on studies of patients i) with chronic or acute akathisia vs. controls, and ii) patients with schizophrenia-spectrum vs. affective disorders. All analyses were performed using the meta package in R for the analyses (Schwarzer et al., 2015).

2.6. Quality of meta-analytical evidence

We rated the quality of evidence according to the Grades of Recommendation, Assessment, Development, and Evaluation (GRADE) Working Group system (Balshem et al., 2011; Guyatt et al., 2008). GRADE allows rating the certainty of estimate for every outcome. We used the GRADEProGDT app for the summary of Findings tables. The following factors were considered: study design, risk of bias, consistency, precision, directness, presence of large, effect, dose–response gradient and publication bias.

3. Results

3.1. Search Results

The electronic data base search yielded 50 articles from Medline, 99 from Embase and two from the fulltext reviewed articles' reference lists (Juncal-Ruiz et al., 2017; Spina et al., 1994). The additional search in Scopus, PsycInfo, Googlescholar and Cochrane Library did not yield any further articles of interest. After removing duplicates 128 articles remained. After exclusion of 103 articles based on title and abstract review, 25 articles were full-text screened, leading to rejection of 15 papers due to lack of iron levels (n=8), investigation of antipsychotic-related movement disorders other than akathisia (n=3), failure to provide/extract data for mean iron levels per study group (n=2), iron supplementation studies (n=1), and case reports (n=1). Ultimately, 10 studies fulfilled all inclusion criteria and were used for data extraction (Supplementary Figure 1).

3.2. Study and patient characteristics

We meta-analyzed 10 cross-sectional studies with 395 patients (mean age=42.2±9.1 years, 34.5% females, 85.6% schizophrenia-spectrum disorders, mean illness duration=18.3±6.3 years), of which 182 patients had akathisia) and 213 did not have akathisia (Table 1). Akathisia was assessed with a rating scale in nine (90%) of the ten studies. The mean duration of antipsychotic treatment was 12.9±5.54 years and the mean daily chlorpromazine equivalent dose was 875.05±271.6 mg/day. Seven (70.0%) of the ten studies adjusted the group comparison for demographic and antipsychotic dose differences (Table 1).

3.3. Primary outcome

In 10 studies (n=395) including 182 patients with and 213 patients without akathisia, iron blood levels were significantly lower in patients with versus without akathisia (fixed-effect model: SMD=-0.49, 95%CI=-0.28, -0.70, p<0.001; random-effects model: SMD=-0.55, 95%CI=-0.14, -0.96, p=0.008) (Figure 1). Heterogeneity was large (l^2 =72%, τ^2 =0.30). Differences did not essentially change after removing the three studies (Horiguchi, 1991; Nemes et al., 1991; O'Loughlin et al., 1991) that did not control for demographic and antipsychotic dose differences between study groups (fixed-effect model: SMD=-0.52, 95%CI=-0.27, -0.76, p<0.001; random-effects model: SMD=-0.51, 95%CI=-0.08, -0.95, p=0.02), while the heterogeneity decreased slightly, but remained substantial (l^2 =67.5%, τ^2 =0.23).

Figure 1 here

3.4. Secondary outcomes

Regarding ferritin levels (studies=7, n=310), the fixed-effect model yielded lower levels in patients with versus without akathisia (SMD=-0.32, 95%CI=-0.08, -0.55, p=0.007), whereas effects were not significant in the random-effects model (SMD=-0.29, 95%CI=-0.79, 0.20, p=0.24). Heterogeneity was substantial (I^2 =77.6%, τ^2 =0.35) (Figure 2).

For transferrin (studies=3, n=134), no significant differences were detected between groups using the fixed-effect model (SMD=0.08, 95%CI=-0.29, 0.45, p=0.67) or the random-effects model (SMD=-0.10, 95%CI=0.67, -0.88, p=0.79). Heterogeneity was large (l^2 =75%, τ^2 =0.35) (Figure 3).

Lastly, for TIBC (studies=6, n=238), no significant differences between patients with versus without akathisia were observed using the fixed-effect model (SMD=0.15, 95%CI=-0.11, 0.41, p=0.26) or the random-effects model (SMD=0.12, 95%CI=-0.29, 0.54, p=0.56) (Figure 4). Heterogeneity was substantial (l^2 =60%, τ^2 =0.16).

"Figures 2,3,4 here

3.5. Meta-regression analysis of iron level differences

The meta-regression for iron concentration differences demonstrated no significant influence for sex (estimated coefficient=-1.73, 95%CI=-4.00, 0.55, =-4.00 p=0.14), age (estimated coefficient=0.03, 95%CI=-0.03, 0.09, p=0.35), CPZE daily dosage (estimated coefficient=0.001, 95%CI=-0.001, 0.004, p=0.36), antipsychotic treatment duration (estimated coefficient=0.12, 95%CI=-0.05, 0.3, p=0.17) and illness duration in years (estimated coefficient=0.03, 95%CI=-0.07, 0.12, p=0.57).

3.6. Subgroup analyses

3.6.1. Iron differences in patients with chronic or acute akathisia vs. control group

In 7 studies (n=265), iron levels were significantly lower in patients with vs. without chronic akathisia (fixed-effect model: SMD=-0.56, 95%Cl=-0.3, -0.81, p<0.001; random-effects model: SMD=-0.66, 95%Cl=-0.08, -1.24, p=0.02). Heterogeneity was high (I^2 =79.6%, τ^2 =0.47). The same pattern was observed for patients with vs. without acute akathisia (SMD=-0.53, 95%Cl=-0.07, -1.0, p=0.02 for both fixed- and random-effects modes in two studies with 86 patients). Heterogeneity was very low (I^2 =0.0%, τ^2 =0.0).

3.6.2. Iron differences in patients with schizophrenia-spectrum disorders

Four studies (n=176) assessed exclusively patients with schizophrenia-spectrum disorders: iron levels were lower in schizophrenia patients with vs. without chronic akathisia (fixed-effect model: SMD=-0.55, 95%CI=-0.23, -0.86, p<0.001), while a trend was observed in the random-effects model (SMD=-0.52, 95%CI=-0.07, -1.12, p=0.08). The heterogeneity was substantial (l^2 =72.0%, τ^2 =0.26). As the rest of study samples had mixed diagnoses and data were provided in aggregate form, we were unable to perform subgroup analyses for any other diagnoses.

3.7. Quality assessment

The assessment of risk of bias was invariably hampered by the scarcity of provided information. Lack of critical information included assessment methods for akathisia as well as iron levels, blinding of akathisia

raters regarding the iron markers' levels, use of parametric tests without assessing normality of iron homeostasis data distributions, and comorbidities that may relate to iron alterations, such as restless leg syndrome (Matthews, 1976). Of studies included for the primary outcome, one was rated as high quality and the rest as poor quality (Supplementary Table S1).

3.8. GRADE assessment

For the primary and the secondary outcomes the certainty of the results was rated as very low; this was mainly due to inconsistency between studies as well as the baseline assumption of low certainty for observational studies (Supplementary Table S2).

4. Discussion

Although akathisia is a relatively common antipsychotic-related adverse event (Demyttenaere et al., 2019; Martino et al., 2018), the pathogenesis of akathisia and other antipsychotic-associated movement disorders remains poorly understood. In that context, early data have provided some insight on potential links between peripheral markers of iron homeostasis and akathisia (Barton et al., 1990; Brown et al., 1987; O'Loughlin et al., 1991). This meta-analysis provides preliminary evidence that lower iron blood levels were associated with antipsychotic-related akathisia in comparison to patients without akathisia. Based on animal models (Ashkenazi et al., 1982), it has been hypothesized that iron levels are linked to akathisia symptoms via central dopaminergic transmission (Kim et al., 2018). However, the direction of this association is unclear. In other words, it is unclear if antipsychotic-related antidopaminergic effects lead to decreased dopaminergic activity that is then associated with iron depletion, or whether baseline iron supplementation may be indicated in the acute treatment phase, likely in the subgroup of patients at highest risk for akathisia, who could be the subgroup with pathological iron homeostasis. However, clinical data remain highly limited (Cotter and O'Keeffe, 2007; Gold and Lenox, 1995) and additional studies are needed to substantiate this hypothesis.

Based on the meta-regression analyses, none of the available moderators or mediators essentially contributed to the heterogeneity of iron blood levels. However, one study reported higher iron levels in males with akathisia compared to females (Nemes et al., 1991). Apart from physiological differences, a previous study reported antipsychotic-related iron level decrease in men, but not in women over a period of 8 weeks (Chen et al., 2018), in contrast to the evidence of Nemes and co-workers (Nemes et al., 1991), although Chen et al did not assess akathisia (Chen et al., 2018). Thus, sex-related patterns of antipsychotic-effects on iron homeostasis and the risk for akathisia need to further be investigated.

Previous literature has suggested dose-dependent patterns for akathisia (Salem et al., 2017). However, our evidence did not detect a differential antipsychotic dose-dependent effect on iron level patterns in akathisia patients. Nevertheless, as studies did not specify the various types of antipsychotic treatment, no further conclusions on this finding can be drawn. Moreover, not all studies reported these data, and the included dosages were daily averages and not cumulative dosages. Cumulative dosages might be a more suitable surrogate when addressing antipsychotic-dose effects (Modestin et al., 2000), but data were not available. Future trials addressing antipsychotic dose-effects will also need to stratify for the profiles

of patients, given the higher prevalence of iron deficiency in older aged people and in women (Coad and Pedley, 2014; Fairweather-Tait et al., 2014).

Inferences on iron status also require concurrent measurement of additional markers to iron measurements, such as ferritin, transferrin and TIBC (Wick et al., 2011). Nevertheless, differences in ferritin, transferrin and TIBC were not significant between patients with versus without akathisia, but data were limited and we cannot exclude a type II error. Thus, it is premature to draw conclusions on the relationship between akathisia and iron transport or storage in patients with akathisia. Patients with akathisia displayed lower ferritin levels compared to patients without akathisia, a difference that was only statistically significant in the fixed-effect model, but not significant in the random-effects model. Although the notion of decreased ferritin levels would tie in well with the concept of latent or acute iron store depletion (Hofmann et al., 2000), our findings marginally support that relationship. Specifically, ferritin differences between patients with and without akathisia reached statistical significance only in one of the included studies (Kuloglu et al., 2003). Kuloglu and associates provided some discussion on that finding using the long-term inpatient status of patients with akathisia as a potential explanation (Kuloglu et al., 2003). More data for ferritin are necessary to provide a clearer overview on iron storage in patients with akathisia.

Evidence for the involvement of iron transport in patients with akathisia, as reflected by patterns for transferrin and TIBC levels, were not particularly supportive, as findings were negative; yet the number of studies and patients were the smallest in these comparisons. Moreover, the large heterogeneity raises concerns when interpreting these findings, indicating that other factors not accounted for in our analyses are related to akathisia and iron homeostasis. For example, this heterogeneity may be linked to the variance of the analytical methods applied for the measurement of the iron status markers, but may also relate to other biological and clinical correlates of iron homeostasis and akathisia.

Results of this study need to be interpreted within its limitations. First, these include foremost the still small number of studies and patients that could be included in this meta-analysis. Since the significant finding regarding lower iron levels in patients with akathisia was observed in the analysis with the largest number of studies and patients, and the finding of lower ferritin levels at least in the fixed-effect model was found in the second largest number of studies and patients, we cannot exclude a type II error for the negative random-effects model findings in transferrin, ferritin and TIBC. Second, antipsychotic treatment that has differential risk for akathisia (Demyttenaere et al., 2019; Martino et al., 2018) was not detailed. Third, results were highly heterogeneous, very likely reflecting heterogeneous samples, such as in terms of antipsychotic doses, as well as methodological heterogeneity, limiting the certainty of the meta-analytic evidence. Fourth, information was mostly missing to grade the studies as high quality, an assessment only achieved by one of the ten studies. Fifth, we restricted our analyses to cross-sectional studies, but followup data on iron metabolism status in relationship to akathisia status in antipsychotic-treated patients are unfortunately too scarce to be meta-analyzed (Juncal-Ruiz et al., 2017; Sachdev and Loneragan, 1991). Sixth, three of the ten studies did not adjust the group comparisons for demographic and antipsychotic dose differences between study groups. Seventh, a large study that did not replicate the iron differences between patients with versus without akathisia could not be included in the meta-analysis due to failure to extract meta-analyzable data (Barnes et al., 1992). The authors were contacted, but data were no longer available, given the age of the study. Eighth, due to lack of specific data for any other diagnostic group,

subgroup analyses by diagnosis were only possible for schizophrenia-spectrum disorders. Finally, the number of moderator and mediator variables that could be examined in the meta-regression was small. In fact, one of the included studies did not report using any akathisia rating scale to group patients into those with vs. without akathisia (Horiguchi, 1991). However, although, clearly, more data are needed, the current meta-analysis is the first to comprehensively assess the relationship between iron metabolism markers and akathisia status in antipsychotic-treated patients, indicating that deficient iron levels may be implicated in the pathophysiology of akathisia.

In summary, results of this first meta-analysis of the relationship between iron homeostasis markers and akathisia in antipsychotic-treated patients emphasizes the complexity of the potential role of iron homeostasis in the risk for akathisia. Patients experiencing akathisia may have lower iron levels compared to patients without akathisia. Available data for other iron markers are currently still too limited to make conclusive statements. No differences were reported for transferrin and TIBC, whereas for ferritin one of the applied models reported lower levels in patients with akathisia. Cross-sectional and, especially, prospective studies focusing on the dynamic course of the interaction between iron status alterations and akathisia are required to shed additional light on the interaction between iron homeostasis and akathisia risk in antipsychotic-treated patients. Future research also needs to put more emphasis on unravelling the role of relevant clinical factors, such as acute versus chronic akathisia, duration of antipsychotic-exposure, antipsychotic type and dose. A better understanding of these factors and of biological underpinning could contribute to predicting, preventing, and potentially treating antipsychotic-related akathisia.

References

Andrews, N.C., 1999. Disorders of iron metabolism. The New England journal of medicine 341, 1986-1995.

Ashkenazi, R., Ben-Shachar, D., Youdim, M.B., 1982. Nutritional iron and dopamine binding sites in the rat brain. Pharmacology, biochemistry, and behavior 17 Suppl 1, 43-47.

Balshem, H., Helfand, M., Schunemann, H.J., Oxman, A.D., Kunz, R., Brozek, J., Vist, G.E., Falck-Ytter, Y., Meerpohl, J., Norris, S., Guyatt, G.H., 2011. GRADE guidelines: 3. Rating the quality of evidence. Journal of clinical epidemiology 64, 401-406.

Barnes, T.R., Halstead, S.M., Little, P.W., 1992. Relationship between iron status and chronic akathisia in an in-patient population with chronic schizophrenia. The British journal of psychiatry : the journal of mental science 161, 791-796.

Barton, A., Bowie, J., Ebmeier, K., 1990. Low plasma iron status and akathisia. Journal of neurology, neurosurgery, and psychiatry 53, 671-674.

Bogdan, M., Silosi, I., Surlin, P., Tica, A.A., Tica, O.S., Balseanu, T.A., Rauten, A.M., Camen, A., 2015. Salivary and serum biomarkers for the study of side effects of aripiprazole coprescribed with mirtazapine in rats. International journal of clinical and experimental medicine 8, 8051-8059.

Borenstein, M., Hedges, L.V., Higgins, J.P., Rothstein, H.R., 2009. Meta-Regression, in: Borenstein, M., Hedges, L.V., Higgins, J.P., Rothstein, H.R. (Eds.), Introduction to Meta-Analysis. John Wiley & Sons. Brown, K.W., Glen, S.E., White, T., 1987. Low serum iron status and akathisia. Lancet 1, 1234-1236. Calarge, C.A., Ziegler, E.E., 2013. Iron deficiency in pediatric patients in long-term risperidone treatment. Journal of child and adolescent psychopharmacology 23, 101-109.

Cao, B., Yan, L., Ma, J., Jin, M., Park, C., Nozari, Y., Kazmierczak, O.P., Zuckerman, H., Lee, Y., Pan, Z., Brietzke, E., McIntyre, R.S., Lui, L.M.W., Li, N., Wang, J., 2019. Comparison of serum essential trace metals between patients with schizophrenia and healthy controls. Journal of trace elements in medicine and biology : organ of the Society for Minerals and Trace Elements 51, 79-85.

Carbon, M., Kane, J.M., Leucht, S., Correll, C.U., 2018. Tardive dyskinesia risk with first- and secondgeneration antipsychotics in comparative randomized controlled trials: a meta-analysis. World psychiatry : official journal of the World Psychiatric Association 17, 330-340.

Chen, X., Li, Y., Zhang, T., Yao, Y., Shen, C., Xue, Y., 2018. Association of Serum Trace Elements with Schizophrenia and Effects of Antipsychotic Treatment. Biological trace element research 181, 22-30. Chong, S.A., Mythily, Remington, G., 2004. Tardive dyskinesia and iron status. Journal of clinical psychopharmacology 24, 235-236.

Coad, J., Pedley, K., 2014. Iron deficiency and iron deficiency anemia in women. Scandinavian journal of clinical and laboratory investigation. Supplementum 244, 82-89; discussion 89.

Cotter, P.E., O'Keeffe, S.T., 2007. Improvement in neuroleptic-induced akathisia with intravenous iron treatment in a patient with iron deficiency. Journal of neurology, neurosurgery, and psychiatry 78, 548. Demyttenaere, K., Detraux, J., Racagni, G., Vansteelandt, K., 2019. Medication-Induced Akathisia with Newly Approved Antipsychotics in Patients with a Severe Mental Illness: A Systematic Review and Meta-Analysis. CNS drugs 33, 549-566.

DerSimonian, R., Laird, N., 1986. Meta-analysis in clinical trials. Controlled clinical trials 7, 177-188. Fairweather-Tait, S.J., Wawer, A.A., Gillings, R., Jennings, A., Myint, P.K., 2014. Iron status in the elderly. Mechanisms of ageing and development 136-137, 22-28.

Friedman, E.H., 1991. Serum iron and akathesia. Biological psychiatry 30, 1064-1065.

Gold, R., Lenox, R.H., 1995. Is there a rationale for iron supplementation in the treatment of akathisia? A review of the evidence. The Journal of clinical psychiatry 56, 476-483.

Guyatt, G.H., Oxman, A.D., Vist, G.E., Kunz, R., Falck-Ytter, Y., Alonso-Coello, P., Schunemann, H.J., Group, G.W., 2008. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. Bmj 336, 924-926.

Herzog, R., Alvarez-Pasquin, M.J., Diaz, C., Del Barrio, J.L., Estrada, J.M., Gil, A., 2013. Are healthcare workers' intentions to vaccinate related to their knowledge, beliefs and attitudes? A systematic review. BMC public health 13, 154.

Higgins, J.P., Thompson, S.G., Deeks, J.J., Altman, D.G., 2003. Measuring inconsistency in meta-analyses. BMJ 327, 557-560.

Hofmann, M., Seifritz, E., Botschev, C., Krauchi, K., Muller-Spahn, F., 2000. Serum iron and ferritin in acute neuroleptic akathisia. Psychiatry research 93, 201-207.

Horiguchi, J., 1991. Low serum iron in patients with neuroleptic-induced akathisia and dystonia under antipsychotic drug treatment. Acta psychiatrica Scandinavica 84, 301-303.

Hutton, B., Salanti, G., Caldwell, D.M., Chaimani, A., Schmid, C.H., Cameron, C., Ioannidis, J.P., Straus, S., Thorlund, K., Jansen, J.P., Mulrow, C., Catala-Lopez, F., Gotzsche, P.C., Dickersin, K., Boutron, I., Altman, D.G., Moher, D., 2015. The PRISMA extension statement for reporting of systematic reviews incorporating network meta-analyses of health care interventions: checklist and explanations. Annals of internal medicine 162, 777-784.

Juncal-Ruiz, M., Ramirez-Bonilla, M., Gomez-Arnau, J., Ortiz-Garcia de la Foz, V., Suarez-Pinilla, P., Martinez-Garcia, O., Neergaard, K.D., Tabares-Seisdedos, R., Crespo-Facorro, B., 2017. Incidence and risk factors of acute akathisia in 493 individuals with first episode non-affective psychosis: a 6-week randomised study of antipsychotic treatment. Psychopharmacology 234, 2563-2570.

Kim, S.W., Stewart, R., Park, W.Y., Jhon, M., Lee, J.Y., Kim, S.Y., Kim, J.M., Amminger, P., Chung, Y.C., Yoon, J.S., 2018. Latent Iron Deficiency as a Marker of Negative Symptoms in Patients with First-Episode Schizophrenia Spectrum Disorder. Nutrients 10.

Kuloglu, M., Atmaca, M., Ustundag, B., Canatan, H., Gecici, O., Tezcan, E., 2003. Serum iron levels in schizophrenic patients with or without akathisia. European neuropsychopharmacology : the journal of the European College of Neuropsychopharmacology 13, 67-71.

Li, L., Li, S., Liu, J., Deng, K., Busse, J.W., Vandvik, P.O., Wong, E., Sohani, Z.N., Bala, M.M., Rios, L.P., Malaga, G., Ebrahim, S., Shen, J., Zhang, L., Zhao, P., Chen, Q., Wang, Y., Guyatt, G.H., Sun, X., 2016. Glucagon-like peptide-1 receptor agonists and heart failure in type 2 diabetes: systematic review and meta-analysis of randomized and observational studies. BMC cardiovascular disorders 16, 91. Martino, D., Karnik, V., Osland, S., Barnes, T.R.E., Pringsheim, T.M., 2018. Movement Disorders Associated With Antipsychotic Medication in People With Schizophrenia: An Overview of Cochrane Reviews and Meta-Analysis. Canadian journal of psychiatry. Revue canadienne de psychiatrie, 706743718777392.

Matthews, W.B., 1976. Letter: Iron deficiency and restless legs. British medical journal 1, 898. Modestin, J., Stephan, P.L., Erni, T., Umari, T., 2000. Prevalence of extrapyramidal syndromes in psychiatric inpatients and the relationship of clozapine treatment to tardive dyskinesia. Schizophrenia research 42, 223-230.

Nemes, Z.C., Rotrosen, J., Angrist, B., Peselow, E., Schoentag, R., 1991. Serum iron levels and akathisia. Biological psychiatry 29, 411-413.

O'Loughlin, V., Dickie, A.C., Ebmeier, K.P., 1991. Serum iron and transferrin in acute neuroleptic induced akathisia. Journal of neurology, neurosurgery, and psychiatry 54, 363-364.

Peralta, V., Cuesta, M.J., Mata, I., Serrano, J.F., Perez-Nievas, F., Natividad, M.C., 1999. Serum iron in catatonic and noncatatonic psychotic patients. Biological psychiatry 45, 788-790.

Percinel, I., Yazici, K.U., Ustundag, B., 2016. Iron Deficiency Parameters in Children and Adolescents with Attention-Deficit/Hyperactivity Disorder. Child psychiatry and human development 47, 259-269.

Quinn, C., Uzbeck, M., Saleem, I., Cotter, P., Ali, J., O'Malley, G., Gilmartin, J.J., O'Keeffe, S.T., 2011. Iron status and chronic kidney disease predict restless legs syndrome in an older hospital population. Sleep medicine 12, 295-301.

Sachdev, P., 1994. Tardive akathisia, tardive dyskinesia, and serum iron status. Journal of clinical psychopharmacology 14, 147-149.

Sachdev, P., Loneragan, C., 1991. Acute drug-induced akathisia is not associated with low serum iron status. Psychopharmacology 103, 138-139.

Salem, H., Nagpal, C., Pigott, T., Teixeira, A.L., 2017. Revisiting Antipsychotic-induced Akathisia: Current Issues and Prospective Challenges. Current neuropharmacology 15, 789-798.

Schwarzer, G., Carpenter, J.R., Rücker, G., 2015. Meta-Analysis with R. Springer, Heidelberg. Shorter, E., Wachtel, L.E., 2013. Childhood catatonia, autism and psychosis past and present: is there an 'iron triangle'? Acta psychiatrica Scandinavica 128, 21-33.

Solmi, M., Pigato, G., Kane, J.M., Correll, C.U., 2018a. Treatment of tardive dyskinesia with VMAT-2 inhibitors: a systematic review and meta-analysis of randomized controlled trials. Drug design, development and therapy 12, 1215-1238.

Solmi, M., Pigato, G.G., Roiter, B., Guaglianone, A., Martini, L., Fornaro, M., Monaco, F., Carvalho, A.F., Stubbs, B., Veronese, N., Correll, C.U., 2018b. Prevalence of Catatonia and Its Moderators in Clinical Samples: Results from a Meta-analysis and Meta-regression Analysis. Schizophrenia bulletin 44, 1133-1150.

Soni, S.D., Tench, D., Routledge, R.C., 1993. Serum iron abnormalities in neuroleptic-induced akathisia in schizophrenic patients. The British journal of psychiatry : the journal of mental science 163, 669-672. Spina, E., Ancione, M., Di Rosa, A.E., Artemisia, A., Natoli, C., Caputi, A.P., 1994. Iron status in schizophrenic patients with acute neuroleptic-induced dystonic reactions. Progress in neuro-psychopharmacology & biological psychiatry 18, 891-898.

Treloar, A.J., Crook, M.A., Tutt, P., White, D.P., Philpot, M.P., 1994. Iron status, movement disorders, and acute phase response in elderly psychiatric patients. Journal of neurology, neurosurgery, and psychiatry 57, 208-210.

Valles, V., Guillamat, R., Vilaplana, C., Duno, R., Almenar, C., Almenar, C., 1992. Serum iron and akathisia. Biological psychiatry 31, 1174-1175.

Wick, M., Pinggera, W., Lehmann, P., 2011. Clinical Aspects and Laboratory – Iron Metabolism, Anemias. Concepts in the anemias of malignancies and renal and rheumatoid diseases. Springer, Wien New York. Wijarnpreecha, K., Werlang, M., Panjawatanan, P., Kroner, P.T., Cheungpasitporn, W., Lukens, F.J., Pungpapong, S., Ungprasert, P., 2019. Association between sarcopenia and hepatic encephalopathy: A systematic review and meta-analysis. Annals of hepatology.

Yanik, M., Kocyigit, A., Tutkun, H., Vural, H., Herken, H., 2004. Plasma manganese, selenium, zinc, copper, and iron concentrations in patients with schizophrenia. Biological trace element research 98, 109-117.

Role of funding source

None.

Contributors

Participated in research design: GS, AN, DG, CUC, JMK

Performed data analysis: GS, AN

Wrote or contributed to the writing and critical revisions of the manuscript: GS, AN, DG, CUC, JMK

Conflicts of interest

Drs. Schoretsanitis, Nikolakopoulou and Guinart have nothing to disclose.

Dr. Correll has been a consultant and/or advisor to or has received honoraria from Alkermes, Allergan, Angelini, Boehringer-Ingelheim, Gedeon Richter, Gerson Lehrman Group, Indivior, IntraCellular Therapies, Janssen/J&J, LB Pharma, Lundbeck, MedAvante-ProPhase, Medscape, Merck, Neurocrine, Noven, Otsuka, Pfizer, Recordati, Rovi, Servier, Sumitomo Dainippon, Sunovion, Supernus, Takeda, and Teva. He has provided expert testimony for Bristol-Myers Squibb, Janssen, and Otsuka. He served on a Data Safety Monitoring Board for Boehringer-Ingelheim, Lundbeck, Rovi, Supernus, and Teva. He received royalties from UpToDate and grant support from Janssen and Takeda. He is also a shareholder of LB Pharma.

Dr. Kane has been a consultant for or received honoraria from Alkermes, Dainippon Sumitomo, Eli Lilly, Forum, Allergan, Genentech, H. Lundbeck, Intracellular Therapies, Janssen Pharmaceutica, Johnson and Johnson, LB Pharmaceuticals, Merck, Minerva, Neurocrine, Otsuka, Pierre Fabre, Reviva, Roche, Sunovion, Takeda and Teva. He has received grant support from Otsuka, Lundbeck and Janssen. He has participated in advisory boards for Alkermes, Dainippon Sumitomo, Intracellular Therapies, Lundbeck, Neurocrine, Otsuka, Pierre Fabre, Takeda, and Teva. He is a Shareholder in Vanguard Research Group and LB Pharmaceuticals, Inc.

Acknowledgements

Authors are particularly grateful to Dr. K. P. Ebmeier, Oxford University, Oxford, UK and Dr. C. Ermis, Dokuz Eylul University, Department of Child and Adolescent Psychiatry, Izmir, Turkey for their valuable feedback during preparation of this manuscript. Authors are also very grateful to Dr. C. Gastaldon, WHO Collaborating Centre for Research and Training in Mental Health and Service Evaluation, Department of Neuroscience, Biomedicine and Movement Sciences, Section of Psychiatry, University of Verona, Verona, Italy for her expertise in statistics.

	246.8 (66.29) 260.72	(177.73) 314.36 (59.35) 281.39 (37.73)	đ	£	Å	đ	404.3 (20.91) 390.15 (15.16)
	호 호	a a	đ	đ	đ	đ	225.13 (13.34) 217.28 (16.36)
	43.76 (25.79) 73.07		56.94 (39.54)	102.13 (87.73)	đ	đ	53.37 (7.92) 225.13 65.94 217.28 (11.08) (16.36)
Iron blood levels (SD), mg/dl	100.1 (35.3) 91.2 (30.2)	58.1 (26.3) 103.6 (40.1)	88.52 (40)	114.74 (55.1)	79.6 (28.7) ⁴ NP	126.9 (20.2) NP	68.9 (9.2) 81.3 (10.6)
Group Iron bl Comparison levels adjusted for mg/dl demo- graphic and CPZE group differences	Yes	Yes	Yes		£		Yes
	942.5 999.0	387 (241.6) 361 (477 3)	1031 (651)	818 (517)	đ	đ	1013 (241) 987 (278)
	18.5 (4.8) 16.1	(1.5) 17.0 18.4 18.4		đ	64	5.8	14.2 (1.91) 13.8 (2.21)
Diagnosis Duration of illness (SD), years	27.8 24.4	25.0 (10.1) 21.4 (11.9)		56.5% SZ, 7.9 (6.5) NP 39.1% SA, 4.3% AD, 3%	£	R	17.4 (2.8) 17.6 (5.1)
Diagnosis	đ	đ	54.5% SZ, 30.3% SA, 12.1% AD	56.5% SZ, 39.1% SA, 4.3% AD, 3%		94.1% SZ, 5.9% AD	100% SZ
°. °+	33.3 33.3	61.5 61.5	63.6	43.5	11.1 ⁴	0	46.7 46.7
Age, years (SD)	1 51.5 51.8	60.1 (11.9) 57 (9.0)	38.5 (14.5)	34 (9.6)	334	29	42.0 (4.1) 39.8 (4)
Akathisia Age, rating year scale (SD)	Akathisia rating scale ²	Akathisia rating scale ²	HAS ³		đ		BARS ⁵
c	15 15	13 13	33	33	10	1	8 8
Author, year Total n Group n Akathisia Age, % ² rating years scale (SD)	Chronic akathisia Non-	akaunisia Chronic akathisia Non- akathisia	Acute akathisia	Non- akathisia	Chronic akathisia	Non- akathisia or -dvstonia	Chronic akathisia Non- akathisia
Total n	30	26	56		72		60
Author, year Total n Group	Barton, 1990	Brown, 1987 26	Hofmann, 2000		Horiguchi, 1991		Kuloglu, 2003

 Table 1. Characteristics of included studies (in alphabetical order)

ia 44.8 4 92.5 ST 19.5 NP 1277 No 83.3 (37.1) 116.3 (80.5) 44.8 4 19.4 NP 817 (96.4) 78.8 (27.3) 93.5 (73.1) (12.1) 10.4 NP NP NP NP NP 78.8 (27.3) 93.5 (73.1) (12.1) 11.4 NP NP NP NP NP NP NP NP NP NP NP NP NP 112.8 NP (12.1) 10% SZ NP 14.3 733.2 Yes 99.05 (40.8) 115.06 (10.2) 33.7 100% SZ NP 14.3 530.2 93.3 (38.5) 139.5 38.7 33.3 NP 14.3 530.2 (90.18) 115.06 (115.4) (10.2) 33.7 100% SZ NP 14.3 530.2 (91.3) (135.4) (14.7) 38.7 33.7 100% SZ NP NP NP Yes 93.38 (37.4) 71.4 (74.7) NP NP NP NP		rating years scale (SD)			of illness (SD), years	of treat- ment ¹ (SD), years	dosage (SD)	Comparison levels adjusted for mg/dl demo- graphic and CPZE group differences	levels (SD), mg/dl	blood levels (SD), ng/ml	blood levels (SD), mg/dl	(SD), μg/dl
Nor. 25 scale ² 4.8 4 9.4 9.4 19.4 NP NP 78.8 (27.3) 9.55 (73.1) adathisia 6 Adathisia NP	50 Chronic Akathisia	sia .	4	92% SZ 8% BD	19.5 (11.1)	đ	1277 (1333)	No	83.3 (37.1)	116.3 (80.5)	dN	£
Optime Acute 6 Akathisia N	isia		4		19.4 (12.4)	đ	817 (964)		78.8 (27.3)	93.5 (73.1)	NP	đ
Non- 24 scale ² N <	30 Acute akathisia	sia	đ	56.7% BD.	R N	đ	đ	No	112.8	đ	200.0	đ
(10.2) 33.7 100% 57 NP 14.3 723.2 Yes 99.05 (40.8) 115.06 akathisia (10.2) 38.7 33.3 NP 14.3 500.2 98.33 (38.5) 139.55 Non- 30 38.7 33.3 NP 14.3 500.2 98.33 (38.5) 139.55 Non- 38.7 84.4) (10.2) 33.7 100% 52 22.8 NP NP 96.33 (38.5) 139.55 1993 44 Chronic 22 Akathisia NP NP NP 97.33 (38.5) 139.55 1993 44 Chronic 14 Akathisia NP NP NP NP 14.43 71.4 (74.7) 1992 28 Chronic 14 Akathisia NP NP NP NP 98.33 (34.9) 51.6 (51.1) Non- 21 Akathisia Tating NP NP NP NP NP NP NA Non- 14 </td <td>isia</td> <td></td> <td>đ</td> <td>43.3% SZ</td> <td>đ</td> <td>AN</td> <td>A</td> <td></td> <td>128.99</td> <td>đ</td> <td>260.0</td> <td>₽</td>	isia		đ	43.3% SZ	đ	AN	A		128.99	đ	260.0	₽
	44 Tardive akathisia		35.7	100% SZ	đ	14.3 (8.4)	723.2 (596.5)	Yes	99.05 (40.8)	115.06 (115.4)	311.43 (50.0)	đ
1993 44 Chronic 22 Akathisia NP NP Yes 87.68 (37.4) 71.4 (74.7) Non- 22 scale ² NP NP NP Yes 87.68 (37.4) 51.6 (51.1) Non- 22 scale ² NP NP NP Yes 87.68 (37.4) 51.6 (51.1) Non- 22 scale ² NP NP NP Yes 84.88 (27.9) NP Akathisia 14 Akathisia NP NP NP NP Yes 84.88 (27.9) NP Non- 14 Scale ² NP NP NP NP NP Yes 84.88 (27.9) NP Non- 14 Scale ² NP NP NP NP Yes 84.88 (27.9) NP Akathisia 182 Akathisia 34.5 Sz: 18.32 13.95 84.44 Non- 213 rating 42.164.91 85.6% 45.3 42.71.6 NN 44.4 Non- 213 rating 42.71.6 NN <td>nisia or cinesia</td> <td>38.7 (8.4)</td> <td>33.3</td> <td></td> <td>đ</td> <td>14.3 (9.2)</td> <td>620.2 (801.8)</td> <td></td> <td>98.33 (38.5)</td> <td>139.5 (193.3)</td> <td>320.0 (100.0)</td> <td>£</td>	nisia or cinesia	38.7 (8.4)	33.3		đ	14.3 (9.2)	620.2 (801.8)		98.33 (38.5)	139.5 (193.3)	320.0 (100.0)	£
Non- 1972 28 Non- No- 109.46 NP	44 Chronic akathisia	sia	dz g	100% SZ	22.8 (14.9)			Yes	87.68 (37.4)	71.4 (74.7)	e g	330.75 (51.29)
5, 1992 28 Chronic 14 Akathisia NP NP NP Yes 84.88 (27.9) NP akathisia rating Non- 14 scale ² NP NP NP Yes 84.88 (27.9) NP Non- 14 scale ² NP NP NP NP NP 109.46 NP akathisia 182 Akathisia 34.5 52: 18.32 12.9 875.05 Yes: N=3; 93.34 84.4 Non- 213 rating 42.16±9.1 85.6% ±6.3 ±5.54 ±271.6 No: N=7 ±18.21 ±31.77 Non- 213 rating 42.16±9.1 85.6% ±6.3 ±5.54 ±271.6 No: N=7 ±18.21 ±31.77 akathisia scale: NP: NP: NP: NP: NP: At.4 Non- 213 rating 42.16±9.1 85.6% ±6.3 ±5.74 ±271.6 No: N=7 ±18.21 ±31.77 M= 6; NP: NP: NP: NP: NP:	isia		ż			ż	ż		(2.+c) 70.cz	(1.10) 0.10	È	(11.84)
Non- 14 scale ² NP NP NP NP 109.46 NP akathisia akathisia 182 Akathisia 182 Akathisia (40.2) 395 Akathisia 182 Akathisia 34.5 5.2: 18.32 12.9 875.05 Yes: N=3; 93.34 84.4 Non- 213 rating 42.16±9.1 85.6% ±6.3 ±5.54 ±271.6 No: N=7 ±18.21 ±31.77 akathisia scale: NP: NP: NP: NP: 440.23 ±31.77 BARS: BD: NP: NP: Acthisia ±5.54 ±271.6 No: N=7 ±31.77 Mathisia scale: 13.9% BD: NP: NP: Acthisia ±31.77 M = 6; 13.9% BD: NP: NP: NP: NE: ±31.77 M = 2; 6.05% MD: NP: ND: ND: ±31.77	Chronic akathisia	sia	£	100% SZ	đ	đ	đ	Yes	84.88 (27.9)	AP	dN	289.96 (60.34)
395 Akathisia 182 Akathisia 34.5 52: 18.32 12.9 875.05 Yes: N=3; 93.34 84.4 Non- 213 rating 42.16±9.1 85.6% ±6.3 ±5.54 ±271.6 No: N=7 ±18.21 ±31.77 akathisia scale: NP: NP: NP: N = 6; 13.9% BARS: BD: N = 2; 6.05% HAS: SA: 5.8% N = 1: AD: 2.0%	isia		đ		đ	đ	đ		109.46	đ	đ	313.43
213 rating 42.16±9.1 85.6% ±6.3 ±5.54 ±271.6 No: N=7 ±31.77 isia scale: NP: NP: ±13.9% N = 6; 13.9% BD: BD: N = 2; 6.05% M = 2; 6.05% N = 1: 58% N = 1: 20%	Akathisia	Akathisia	34.5	SZ:	18.32	12.9	875.05	Yes: N-=3;	93.34	84.4	258.75	321.03
	iisia		9.1	85.6% NP:	±6.3	± 5.54	±271.6	No: N = 7	±18.21	± 31.77	±50.5	±47.36
		N = 6;		13.9%								
		BARS:		BD:								
		N = 2; UAC:		%CU.9								
		N = 1		AD: 7.0%								
		NP:		Other:								
-		N = 1		0.6%								

Figure 1. Iron blood levels (mg/dL) in patients with vs. without akathisia.

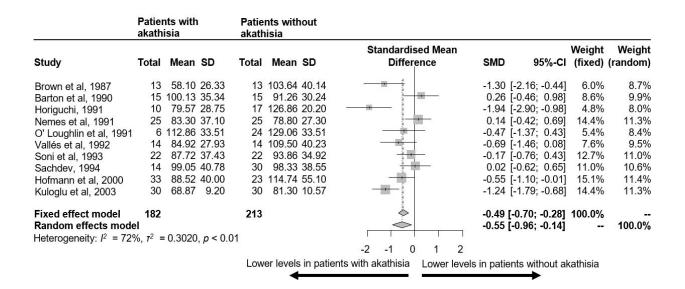


Figure 2. Ferritin blood levels (ng/mL) in patients with vs. without akathisia.

	Patie akatł	nts with nisia	י	Patier akathi	nts with isia	out					
Study	Total	Mean	SD	Total	Mean	SD	Standardised Mean Difference	SMD	95%-CI	Weight (fixed)	Weight (random)
Brown et al. 1987	13	136.20	202.13	13	83.73	82.37	 	0.33	[-0.45; 1.10]	9.0%	12.8%
Barton et al. 1990	15	43.76	25.79	15	73.07	42.63			[-1.56; -0.06]		13.1%
Nemes et al, 1991	25	116.30	80.50	25	93.50	73.10		0.29	[-0.27; 0.85]	17.4%	15.1%
Soni et al. 1993	22	71.40	74.70	22	51.60	51.10		0.30	[-0.29; 0.90]	15.3%	14.7%
Sachdev, 1994	14	115.06	115.40	30	139.50	193.30		-0.14	[-0.77; 0.50]	13.4%	14.2%
Hofmann et al, 2000	33	56.94	39.54	23	102.13	87.73		-0.70	[-1.25; -0.15]	17.9%	15.1%
Kuloglu et al, 2003	30	53.37	7.92	30	65.94	11.08		-1.29	[-1.85; -0.73]	17.3%	15.0%
Fixed effect model	152			158			\diamond	-0.32	[-0.55; -0.09]	100.0%	
Random effects mod	lel						\sim	-0.29	[-0.79: 0.20]		100.0%
Heterogeneity, $l^2 = 78^{\circ}$	$\% T^2 =$	0 3548	p < 0.01								
	, .		F 0.01				-1.5 -1 -0.5 0 0.5 1 1.5				
				Lov	ver level	s in patie	nts with akathisia Lower levels	in patie	nts without ak	athisia	

Figure 3. Transferrin blood levels (mg/dL) in patients with vs. without akathisia.

	Patier akath	nts with isia		Patie akath	nts wit nisia	hout					
Study	Total	Mean	SD	Total	Mean	SD	Standardised Mean Difference	SMD	95%-CI	Weight (fixed)	Weight (random)
O'Loughlin et al, 1991	6	200.00	60.00	24	260.00	60.00		-0.97 [-	1.91; -0.04]	15.5%	27.4%
Sachdev, 1994	14	311.43	50.00	30	320.00	100.00		-0.10 [-	0.73; 0.54]	33.5%	34.8%
Kuloglu et al, 2003	30	225.13	13.34	30	217.28	16.36		0.52 [0.00; 1.03]	50.9%	37.8%
Fixed effect model	50			84				0.08 [-	0.29; 0.45]	100.0%	
Random effects mode Heterogeneity, $l^2 = 75\%$).3484.	p = 0.02	1				-0.10 [-	0.88; 0.67]		100.0%
	.,						-1.5 -1 -0.5 0 0.5 1 1.5				
				Lov	ver leve	ls in patie	ts with akathisia Lower levels	in patien	ts without al	kathisia	

Figure 4. Total iron-binding capacity (TIBC) (μ g/dL) in patients with vs. without akathisia.

	Patier akath	nts with isia	Ú.	Patie akath	nts with isia	out					
Study	Total	Mean	SD	Total	Mean	SD	Standardised Mean Difference	SMD	95%-CI	Weight (fixed)	Weight (random)
Brown et al, 1987	13	314.36	59.35	13	281.39	37.73		- 0.64 [-0.15; 1.43]	10.6%	13.9%
Barton et al, 1990	15	246.80	66.29	15	260.72	49.61		-0.23	-0.95: 0.49]	12.9%	15.3%
Nemes et al. 1991	25	282.00	40.70	25	292.90	34.10		-0.29	-0.84: 0.271	21.5%	18.7%
Vallés et al, 1992	14	289.96	60.34	14	313.43	48.61			-1.17; 0.33]	11.9%	14.7%
Soni et al, 1993	22	330.75	51.29	22	323.49	11.84			-0.40: 0.781	19.0%	17.9%
Kuloglu et al, 2003	30	404.30	20.91	30	390.15	15.16		0.76	0.24; 1.29]	24.1%	19.5%
Fixed effect model	119			119				0.15 [-0.11; 0.41]	100.0%	
Random effects mod	lel							0.12	0.29; 0.54]		100.0%
Heterogeneity, / = 60	%, τ = (0.1566,	p = 0.03	1			1 1 1 1 1				
							-1 -0.5 0 0.5 1				
				Low	er levels	in patien	s with akathisia Lower levels	in patier	nts without a	kathisia	