

ARTICLE

Open Access

On the relationship of first-episode psychosis to the amphetamine-sensitized state: a dopamine $D_{2/3}$ receptor agonist radioligand study

Ana Weidenauer¹, Martin Bauer^{1,2}, Ulrich Sauerzopf¹, Lucie Bartova¹, Lukas Nics³, Sarah Pfaff³, Cecile Philippe³, Neydher Berroterán-Infante³, Verena Pichler³, Bernhard M. Meyer¹, Ulrich Rabl¹, Patrick Sezen¹, Paul Cumming^{4,5}, Thomas Stimpfl⁶, Harald H. Sitte⁷, Rupert Lanzenberger¹, Nilufar Mossaheb⁸, Alexander Zimprich⁹, Pablo Rusjan¹⁰, Georg Dorffner¹¹, Markus Mitterhauser^{3,12}, Marcus Hacker³, Lukas Pezawas¹, Siegfried Kasper¹, Wolfgang Wadsak³, Nicole Praschak-Rieder¹ and Matthäus Willeit¹

Abstract

Schizophrenia is characterized by increased behavioral and neurochemical responses to dopamine-releasing drugs. This prompted the hypothesis of psychosis as a state of “endogenous” sensitization of the dopamine system although the exact basis of dopaminergic disturbances and the possible role of prefrontal cortical regulation have remained uncertain. To show that patients with first-episode psychosis release more dopamine upon amphetamine-stimulation than healthy volunteers, and to reveal for the first time that prospective sensitization induced by repeated amphetamine exposure increases dopamine-release in stimulant-naïve healthy volunteers to levels observed in patients, we collected data on amphetamine-induced dopamine release using the dopamine $D_{2/3}$ receptor agonist radioligand [¹¹C]-(+)-PHNO and positron emission tomography. Healthy volunteers ($n = 28$, 14 female) underwent a baseline and then a post-amphetamine scan before and after a mildly sensitizing regimen of repeated oral amphetamine. Unmedicated patients with first-episode psychosis ($n = 21$; 6 female) underwent a single pair of baseline and then post-amphetamine scans. Furthermore, T1 weighted magnetic resonance imaging of the prefrontal cortex was performed. Patients with first-episode psychosis showed larger release of dopamine compared to healthy volunteers. After sensitization of healthy volunteers their dopamine release was significantly amplified and no longer different from that seen in patients. Healthy volunteers showed a negative correlation between prefrontal cortical volume and dopamine release. There was no such relationship after sensitization or in patients. Our data in patients with untreated first-episode psychosis confirm the “endogenous sensitization” hypothesis and support the notion of impaired prefrontal control of the dopamine system in schizophrenia.

Introduction

Several lines of evidence demonstrate increased sub-cortical dopamine (DA) transmission in psychotic

patients with schizophrenia (SCZ). Positron emission tomography (PET) studies show increased dopamine synthesis capacity and heightened behavioral and neurochemical responses towards DA-releasing compounds^{1–6}. The common mechanism of action of all antipsychotic drugs—reducing DA transmission at postsynaptic $D_{2/3}$ receptors—confirms the key role of DA signaling in psychosis⁷. While the pathophysiological basis of DA dysfunction in SCZ remains unknown, current versions of

Correspondence: Matthäus Willeit (matthaeus.willeit@meduniwien.ac.at)

¹Department of Psychiatry and Psychotherapy, Division of General Psychiatry, Medical University of Vienna, Währinger Gürtel 18-20, 1090 Vienna, Austria

²Department of Clinical Pharmacology, Medical University of Vienna, Vienna, Austria

Full list of author information is available at the end of the article.

These authors contributed equally: Ana Weidenauer, Martin Bauer

© The Author(s) 2020



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

the DA theory of SCZ⁸ posit that upstream pathogenic factors converge on subcortical DA pathways to mediate the expression and intensity of psychotic symptoms^{9–11}.

Sensitization denotes a process by which repeated exposure to a stimulus induces a progressive increase in responses to the very same stimulus^{12,13}. When repeatedly administered, *D*-amphetamine (AMPH) induces behavioral sensitization and a progressive amplification in AMPH-induced DA release^{14–16}. Since psychotic patients show elevated responses to AMPH without any prior drug exposure¹, psychosis has been conceptualized as a state of “endogenous sensitization”^{10,17,18}. The prefrontal cortex (PFC), origin of reciprocal regulatory connections to subcortical DA neurons¹⁹, has often been found to be structurally and functionally impaired in SCZ^{20,21}. Earlier studies have shown particularly strong relationships between subcortical DA metabolism and the left-hemispheric dorso-lateral PFC (DLPFC) and inferior frontal gyrus (IFG) in SCZ and subjects at-risk mental state for SCZ^{22,23}. In order to collect experimental evidence supporting this concept, we used PET and the DA $D_{2/3}$ receptor agonist radioligand (+)-4-propyl-3,4,4a,5,6,10b-hexahydro-2*H*-naphtho[1,2-*b*][1,4]oxazin-9-ol ($[^{11}\text{C}]\text{-}(+)\text{-PHNO}$)²⁴ for measuring AMPH-induced changes in $D_{2/3}$ receptor binding, semi-quantitative index of DA release, in drug-naïve patients with first-episode psychosis (FEP). Healthy volunteers (HV) were studied before and after exposure to a mildly sensitizing regime of repeated AMPH administration. In order to identify upstream pathogenic mechanisms of psychotic hyperdopaminergia, we analyzed volumetric parameters in the PFC for their relationship to indices of subcortical DA release.

Materials and methods

All procedures of this study (Clinical Trial Registry: EUDRACT 2010-019586-29) were approved by the ethics committee of the Medical University of Vienna and pertinent federal regulatory authorities. After a test-retest phase ensuring reliability of local $[^{11}\text{C}]\text{-}(+)\text{-PHNO}$ PET imaging procedures (six male HV), 42 HV and 29 anti-psychotic-naïve (or minimally exposed) patients with FEP capable of providing informed consent were recruited between 2013 and 2017; HV were required to be of good health based on physical examination, history, ECG, and laboratory results. Exclusion criteria comprised any intake of drugs of abuse except nicotine, caffeine, and alcohol (occasional use only), five or more stimulant exposures lifetime, psychiatric disorders (evaluated with the DSM-IV based M.I.N.I. questionnaire²⁵), having a first-degree relative with schizophrenia or bipolar disorder or having any contraindications against receiving a PET, MRI or *d*-amphetamine; in addition, FEP patients were required to have a minimum Positive and Negative Symptom Scale

(PANSS^{26,27}) score of 55 with >3 on at least two PANSS psychosis items or >4 on one psychosis item; no or minimal lifetime exposure to antipsychotics; no lifetime exposure to antipsychotic depot preparations; no antipsychotics within two weeks prior to scanning (for details see Supplemental Material). Diagnoses of FEP in SCZ according to DSM-IV were independently made by at least two experienced psychiatrists (N.M.; N.P.-R., S.K., M. W.). Three patients were previously exposed to olanzapine, aripiprazole, or quetiapine. Antipsychotics had been discontinued at least two months prior to inclusion without attaining a predefined threshold of two treatment weeks or lifetime exposure up to 50 mg haloperidol-equivalent. Three patients had a history of antidepressant treatment discontinued at least two months prior to inclusion. During the study, eleven patients required symptomatic treatment for psychomotor agitation or insomnia (lorazepam 1–10 mg or zolpidem 10 mg per day). Data sets (complete or partial) of 28 + 6 HV and 21 FEP patients entered final analysis (see Supplemental Material and Supplementary Table 1 for full details).

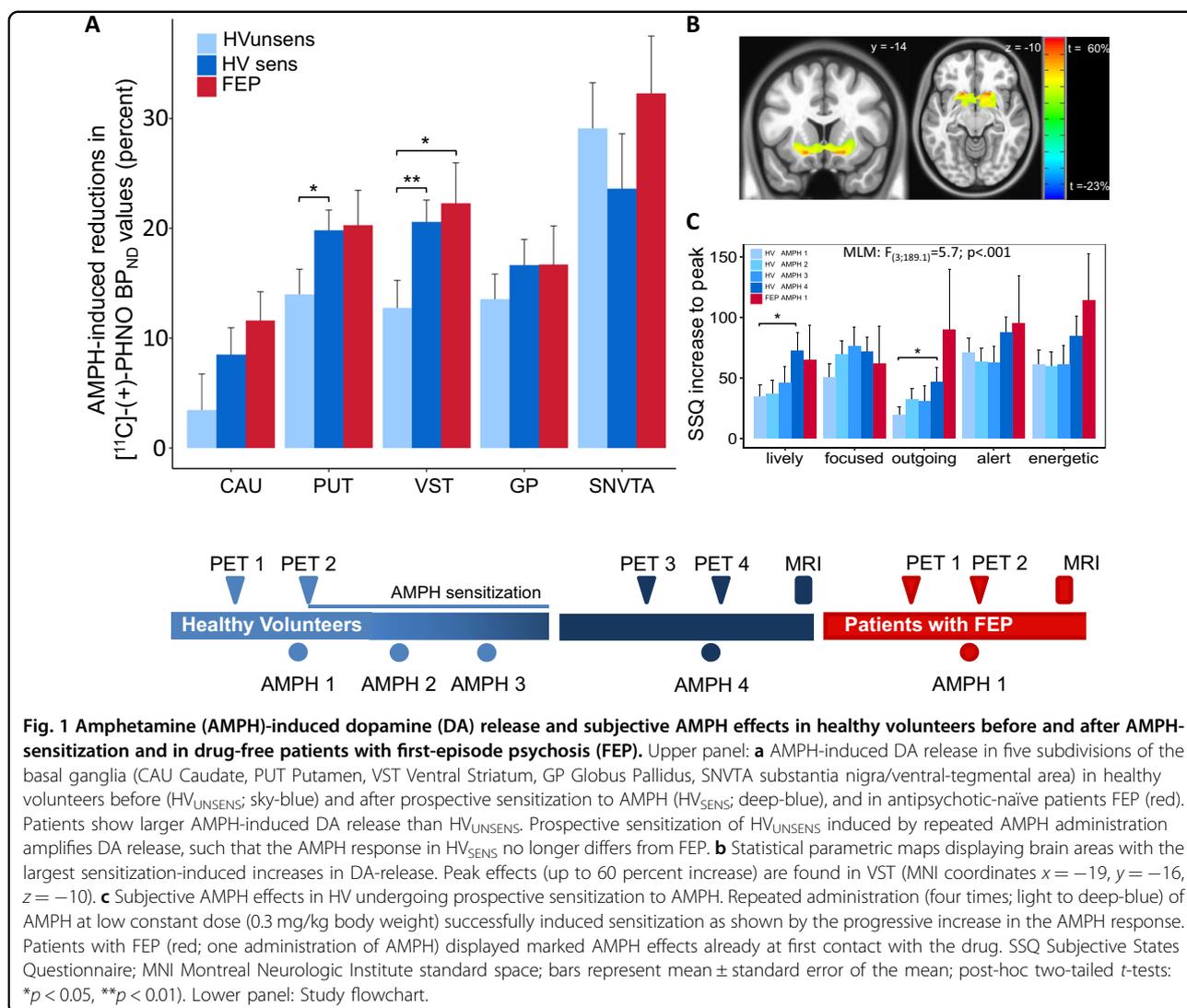
Study setup and prospective sensitization

First, drug-naïve HV (HV_{UNSENS}) underwent an AMPH-free $[^{11}\text{C}]\text{-}(+)\text{-PHNO}$ PET scan (baseline₁, PET₁). On a separate day less than five 5 days apart, a second scan (PET₂) was performed 90–120 min after oral administration of 0.4 mg/kg body weight AMPH (Attention[®], MEDICE Arzneimittel GmbH, Iserlohn, D), a time point at which subjective effects are peaking and blood levels are still rising (blood levels are highest after 3–4 h²⁸). The AMPH dose was chosen according to earlier studies showing that dosages between 0.3 and 0.5 mg/kg body weight induce reliable reductions in $[^{11}\text{C}]\text{-}(+)\text{-PHNO}$ BP_{ND} values^{29,30}. The dose corresponds to a low to medium dose used for treating attention deficit-hyperactivity disorder in children. Administration of AMPH at this same dose was repeated two more times at intervals of two days. Two to four weeks thereafter, the now sensitized HV (HV_{SENS}) underwent another AMPH-free scan (baseline₂, PET₃), and within five days, a fourth scan (PET₄) preceded by AMPH dose four (Fig. 1, lower panel).

Patients with FEP received one baseline scan (PET₁) and one AMPH-scan (PET₂; protocol as above). AMPH led to temporary increases in heart rate and blood pressure (Supplementary Fig. 1). Occasionally, participants reported mild headache and insomnia the night after AMPH administration. Neither HV nor patients experienced any serious AMPH-related adverse events.

$[^{11}\text{C}]\text{-}(+)\text{-PHNO}$ PET and MR imaging

$[^{11}\text{C}]\text{-}(+)\text{-PHNO}$ was synthesized as described earlier³¹. Quality control was in accordance with European



Pharmacopoeia. PET images were acquired on a GE Advance scanner (General Electric Medical Systems, Milwaukee, WI). Emission data were acquired over 90 min after bolus-injection of 309 [81] MBq (mean [SD]) [^{11}C]-(+)-PHNO. Raw data were reconstructed by filtered-back projection to yield dynamic images in 15 consecutive one-minute frames followed by 15 five-minute frames. With exception of two patients who chose to terminate their participation early, all subjects underwent T1 and proton density (PD) weighted 3 T magnetic resonance (MR) imaging (see Supplemental Material).

Behavioral and hormonal measurements

Subjective AMPH effects were recorded using the drug effects questionnaire³² and the subjective states questionnaire (SSQ)³³. The PANSS scale²⁶ was administered by certified raters for measuring baseline psychopathology

in FEP patients, the brief psychiatric rating scale (BPRS³⁴) was used to measure AMPH-induced changes in psychopathology. Blood for serum AMPH levels was collected at the beginning of PET scans. Heart rate, blood pressure (systolic and diastolic were determined repeatedly before and during PET scans (see Supplemental Material).

Image analysis

Analysis in regions of interest

Frame-wise motion correction and co-registration of attenuation-corrected average PET images to T1-weighted MRIs was performed using AFNI software. PET images of two FEP patients who did not undergo MR imaging (see above) were co-registered to normalized [^{11}C]-(+)-PHNO template images (one each for no-intervention and AMPH scans) created by averaging spatially normalized PET images (early and late low-contrast

frames omitted) of 15 HV. Individually optimized bilateral regions of interest (ROIs) were obtained for the caudate nucleus (CAU), putamen (PUT), ventral striatum (VST), and cortical cerebellum (CER) using the automated image analysis software ROMI³⁵ as described elsewhere³⁰. Since automated algorithms provided no satisfactory ROI delineation in globus pallidus (GP) and substantia nigra/ventral tegmental area (SNVTA), GP and SN/VTA were delineated manually by a single rater on individual PD MR images (fused with PET images for aiding delineation of SNVTA). ROI delineation was evaluated independently by a second rater visually controlling for anatomical fit and assessing outliers and time drift in ROI sizes. Decay-corrected time–activity curves (TACs) were extracted from the dynamic sequence. The simplified reference tissue model (SRTM2)^{36,37} implemented in PMOD software (Version 3.6; PMOD Technologies Ltd, Zurich, Switzerland) was used to derive binding potential (BP_{ND}) values in each ROI. Cerebellar cortex (CER) avoiding midline structures served as reference region since it is virtually devoid of DA D_{2/3} receptors in humans^{38–40}. Relative AMPH-induced change in [¹¹C]-(+)-PHNO BP_{ND} binding was calculated as $[(BP_{ND\ baseline} - BP_{ND\ AMPH}) / BP_{ND\ baseline} * 100]$ (ΔBP_{ND} ; for the sake of simplicity henceforth designated “DA release”).

Parametric analysis

Voxel-wise BP_{ND} maps were calculated using the PMOD 3.6 SRTM basis function implementation⁴¹. TACs previously derived in CER (low-binding) and VST (high-binding) were used to optimize iterative model fitting procedures. Effects of AMPH and AMPH-sensitization were analyzed using the AFNI programs 3dttest++ and 3dLME.

Volumetric analyses

T1-weighted MR images were processed using FreeSurfer 6.0 software (<http://surfer.nmr.mgh.harvard.edu/>). Cortical gray voxels were allocated to a set of regions predefined in the Destrieux atlas⁴² using a Bayesian algorithm. Skull-stripping, gross accuracy of delineation, and surface modeling were quality-controlled by visual inspection of processed images.

Statistical analysis

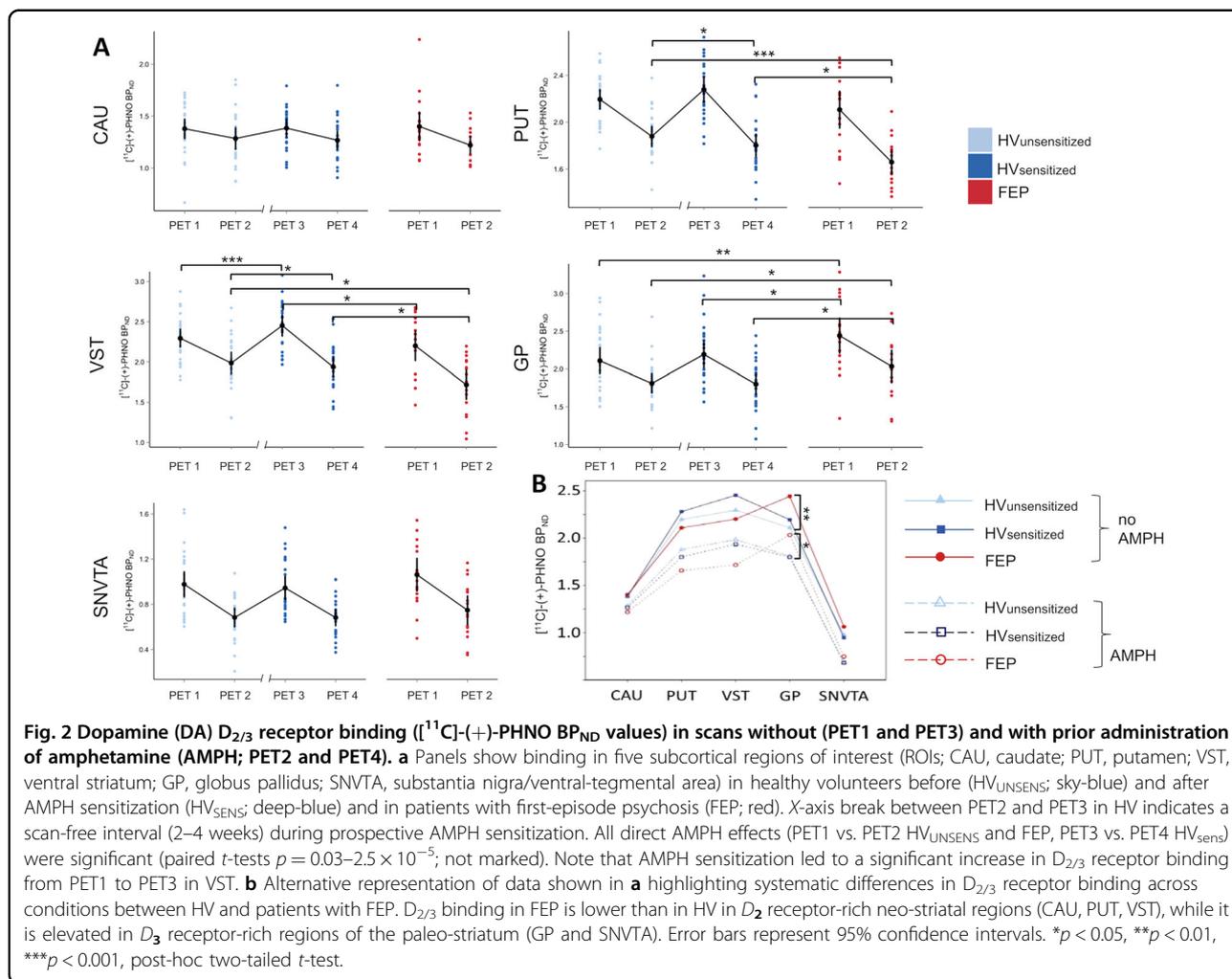
The sample size of this study was planned according to data collected with antagonist radioligands in sensitization and patients with schizophrenia^{3–5,15}. Statistical analysis of [¹¹C]-(+)-PHNO BP_{ND} values obtained in the ROI-based analysis of [¹¹C]-(+)-PHNO binding was carried out using mixed linear models (MLM) as implemented in SPSS 24.0 (SPSS 24.0; IBM Corp., Armonk, NY) and R 12.0.1 (R Foundation for Statistical Computing, Vienna, Austria; <https://www.r-project.org>) software.

Analysis of differences in DA release (ΔBP_{ND}) between HV before and after sensitization was carried out using ΔBP_{ND} as dependent variable and group (FEP vs. HV_{UNSENS}, and FEP vs. HV_{SENS}, respectively) as fixed variable; ROI was entered as random variable. For analyzing receptor binding, BP_{ND} was entered as dependent variable, subject status (HV_{UNSENS}, HV_{SENS}, FEP) as fixed between-group factor, scan condition (AMPH yes/no) as fixed repeated factor, and ROI as fixed or random repeated factor. MLM analyses testing the effects of covariates (PFC volume parameters) were carried out analogously. All main effects and all relevant interactions were entered into the respective models. The subject-identifier variable was entered as random factor where appropriate. Two-tailed *t*-tests (paired where appropriate) were used for post-hoc tests after assuring normal distribution. Correlations were calculated using Pearson product moment (*r*) or, if appropriate, Spearman rank sum (*rho*) correlation coefficients. Results confirming the main a-priori study hypotheses (Figs. 1, 2) were not corrected for multiple comparisons. More exploratory results (Figs. 3–5) were carried out using ROI-based and parametric methods (voxel-wise maps for PET images, vertex-wise analysis of MR images) for independent confirmation. Results of the ROI-based analysis on the relationship between PANSS items and BP_{ND} values (Fig. 3; 30 PANSS items, 5 ROIs, two hemispheres and on/off AMPH) and the relationship between regional cortical volumes as implemented in FreeSurfer 6.0 software⁴² and ΔBP_{ND} (Fig. 4; 33 cortical regions, 5 ROIs, two hemispheres; three groups) were Bonferroni corrected, resulting in adjusted significance levels of $p_{corr1} = 0.00008$ and $p_{corr2} = 0.00005$, respectively.

Results

DA release in FEP and sensitization

The concept of “endogenous sensitization” in psychosis¹⁷ predicts that the difference in AMPH-induced DA release between FEP patients and HV should disappear or substantially diminish after HV are sensitized to AMPH. Analysis of AMPH-induced changes in [¹¹C]-(+)-PHNO binding (ΔBP_{ND}) showed significant differences between unsensitized HV and FEP (HV_{UNSENS} vs. FEP: $F_{(1,184.04)} = 11.6$, $p = .0008$; Fig. 1a). After HV were sensitized, groups were no longer significantly different (HV_{SENS} vs. FEP: $F_{(1,175.6)} = 0.99$, $p = .32$; Fig. 1a, Supplementary Table 2). Including alcohol and nicotine consumption and sex as covariates into the model did not relevantly alter results (HV_{UNSENS} vs. FEP: $F_{(1,139.9)} = 11.6$, $p = 0.0008$, see Supplemental Material for details). In good agreement with the ROI-based analysis, parametric ΔBP_{ND} maps showed the most robust effects of sensitization in the VST (Fig. 1b). Indicating behavioral sensitization in HV_{SENS}, repeated AMPH-administration induced progressive enhancement of subjective AMPH



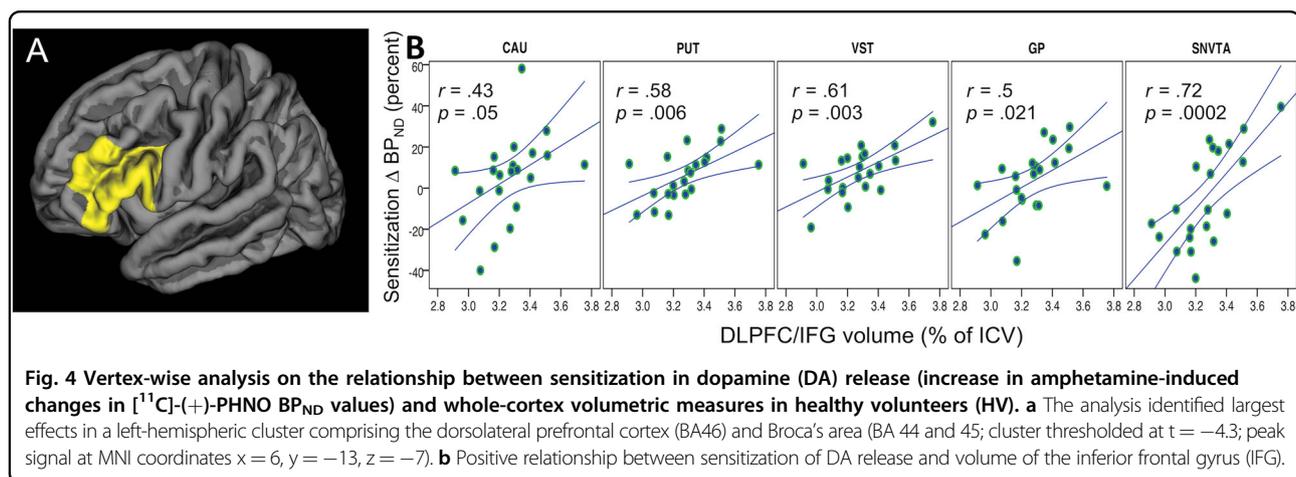
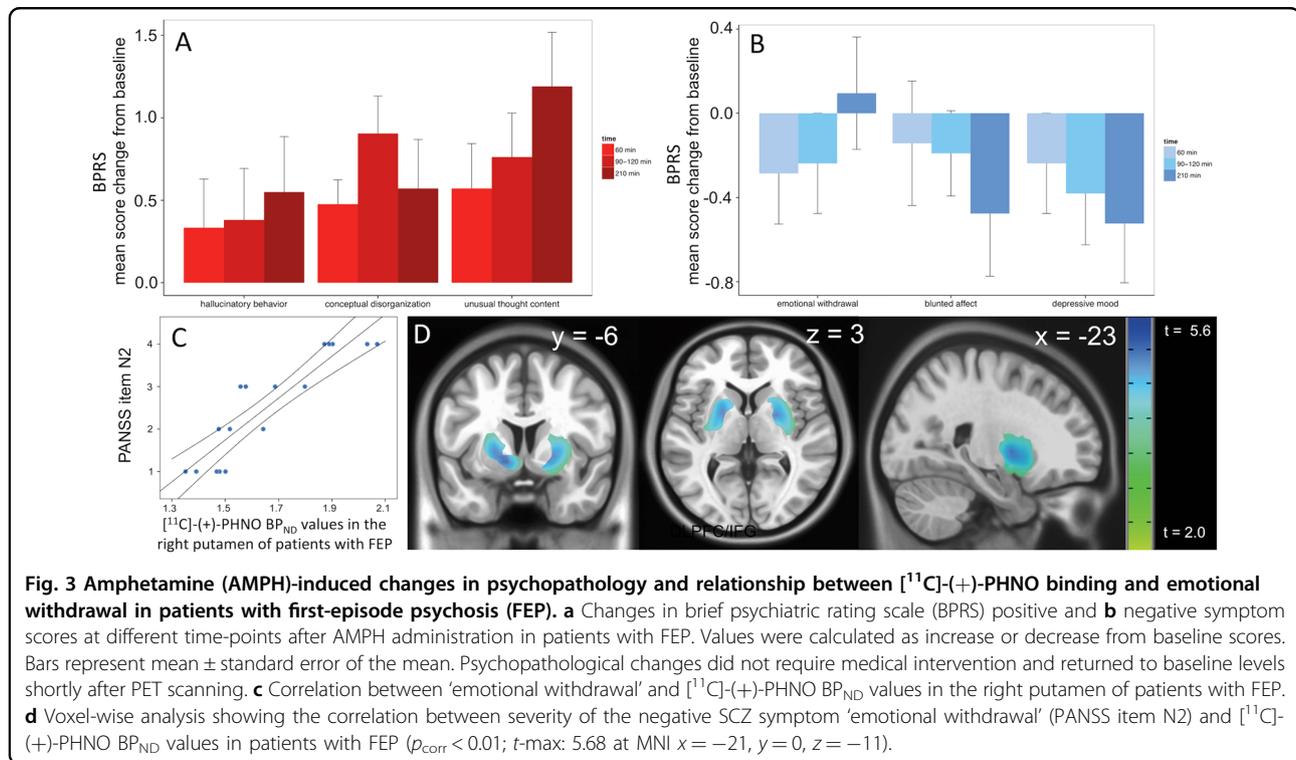
effects up to levels observed in FEP (Fig. 1c, Supplementary Fig. 2).

Analysis of group differences and AMPH-effects at the level of $D_{2/3}$ receptor binding before sensitization ($[^{11}C]$ (+)-PHNO BP_{ND} values by group, condition, and ROI) showed significant two-way interactions between group and condition (HV_{UNSENS}/FEP * no-AMPH/AMPH: $F_{(1;305.7)} = 9.9$, $p = .009$), group and ROI ($F_{(4;140)} = 10.7$, $p = 1.3 \times 10^{-7}$), and a significant three-way interaction between group, condition, and ROI ($F_{(8;140)} = 3.0$, $p = .004$).

The same model, when applied to FEP and HV after sensitization, did no longer show a significant two-way interaction between group and condition ($F_{(1;275.8)} = 0.6$, $p = 0.42$). However, the group*ROI ($F_{(4;123)} = 11.2$, $p = 9.4 \times 10^{-8}$), and a group*condition* ROI interactions remained significant, ($F_{(8;123.1)} = 5.5$, $p = 5 \times 10^{-5}$; Fig. 2a, b). At least in part, this is due to the fact that irrespective of

AMPH pretreatment, $D_{2/3}$ receptor binding was consistently lower in FEP than in HVs in neo-striatal ROIs (CAU, PUT, VST) but higher in GP (and trend-wise also in SNVTA; Fig. 2b; Supplementary Table 3). Baseline $D_{2/3}$ receptor binding and sensitization showed a significant positive correlation for all regions in HV (see Supplementary Fig. 3). An analysis of functional subdivisions of the basal ganglia as previously published did not reflect our findings (Supplementary Fig. 4).

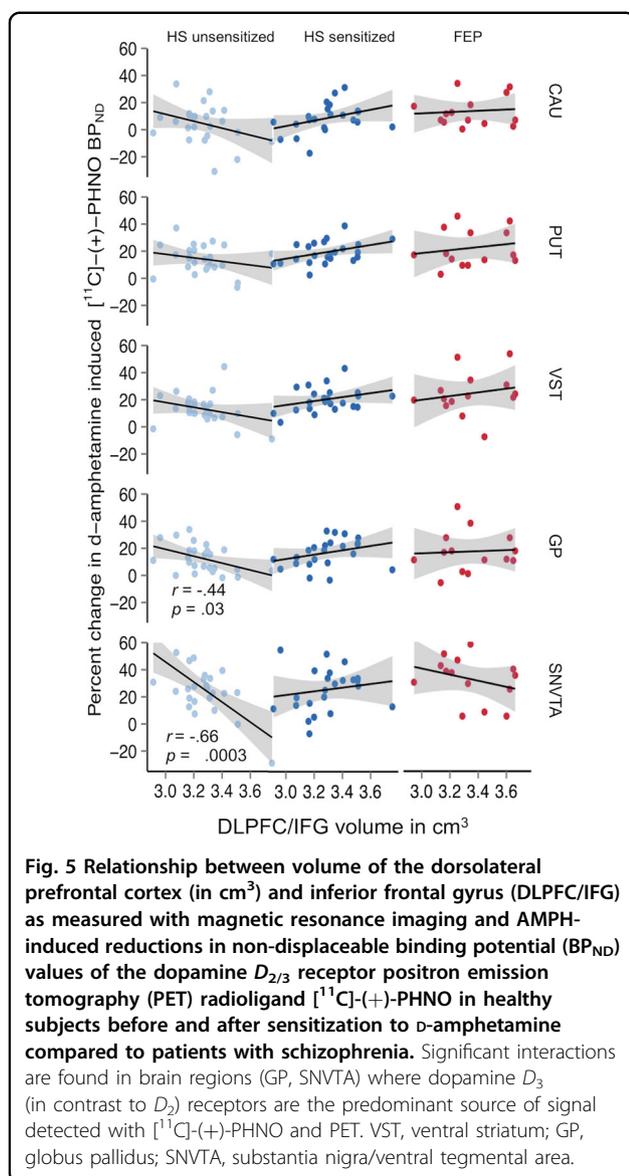
Indices of DA release were positively correlated with behavioral AMPH effects (see Supplemental Material). It is known that benzodiazepines might interact with dopamine neurotransmission⁴³. Thus we compared DA release between patients receiving benzodiazepines and those who did not require sedation, we found no significant difference (16 available datasets for calculating DA release, 7 not receiving lorazepam, 9 receiving lorazepam; two-sided *t*-test $p > 0.05$ for all ROIs).



AMPH effects on psychopathology

With a mean [SD] PANSS score of 82.2 [6.9] (subscales for positive, negative, and general SCZ symptoms: 21.4 [6.8], 20.1 [6.1], and 40.8 [9.4]), FEP patients were suffering from acute psychosis of moderate to marked severity at the time of PET scanning. Administration of AMPH prompted a temporary increase in psychotic symptoms. Symptoms returned to baseline after 3–4 h without specific intervention. While positive symptoms of SCZ (in particular hallucinations, delusions, and thought-disorder) increased with AMPH, negative (especially blunted affect and emotional withdrawal) and affective

symptoms (depressed mood) improved (Fig. 3a, b; Supplementary Table 4). In contrast to earlier studies^{3–6,44–46} but in line with a recent study using the $\text{D}_{2/3}$ receptor agonist radioligand $[^{11}\text{C}]N\text{-propyl-apomorphine}$ ($[^{11}\text{C}] \text{NPA}^{47}$), we did not observe significant relationships between positive symptoms and DA release. However, there were significant relationships between $[^{11}\text{C}](+)\text{-PHNO}$ BP_{ND} values in the AMPH condition and negative symptoms. Correlations were strongest in PUT and driven mainly by the PANSS item N2 (“emotional withdrawal”; right PUT: $\rho = 0.93$, $p = 7.7 \times 10^{-8}$; Fig. 3c, d; Supplementary Fig. 5). Typical behavioral effects of



AMPH in healthy volunteers included increased production of speech, extraversion, alertness, and enhanced energy.

PFC volume and subcortical DA release

Dysfunctional PFC-DA interactions are prime candidates for “upstream” pathogenic mechanisms underlying subcortical hyperdopaminergia in patients with SCZ. Furthermore, brain circuits involved in cognitive functions encompassing the PFC are often found to be compromised in patients with SCZ⁴⁸. Thus we derived volumetric parameters from MR images and, in a first step, analyzed the relationship between indices of DA release and overall PFC volume. Consistent with inhibition of DA release by the PFC^{19,22}, we observed an inverse

relationship between PFC volume and DA release in AMPH naïve $\text{HV}_{\text{UNSENS}}$. After AMPH sensitization, the association was lost or even reversed and more resembled the pattern observed in FEP patients. An anatomically unbiased vertex-wise analysis of the relationship between sensitization of DA release in SNVTA ($\Delta\text{BP}_{\text{ND-SENS}} - \Delta\text{BP}_{\text{ND-UNSENS}}$) and whole-cortex volumetric parameters identified a left-hemispheric cluster spanning the boundaries of middle and inferior frontal gyrus as main driver of the whole-PFC signal (Fig. 4a, b). In a next step, we analyzed correlations between DA release and volumes of individual PFC sub-regions as implemented into Freesurfer 6.0 software according to anatomical and functional priors⁴². In perfect agreement with the vertex-wise analysis, we observed the by far strongest correlations in three anatomically adjacent left-hemispheric regions: Brodmann areas (BA) 46 (dorsolateral prefrontal cortex; DLPFC), and BA 44 and BA 45, or pars opercularis and triangularis of the left inferior frontal gyrus (IFG), together forming “Broca’s area”⁴⁹.

Plotting the correlations between volume of these regions (henceforth referred to as *DLPFC/IFG*) and DA release in the five subcortical ROIs (CAU, PUT, VST, GP, and SNVTA in rostral-caudal anatomical order) revealed a remarkable “herring bone”-like pattern (Fig. 5). This pattern is shaped by negative regression lines—indicating *DLPFC/IFG*-inhibition of DA release in $\text{HV}_{\text{UNSENS}}$ —and positive regression lines observed after the same subjects had undergone AMPH sensitization ($\text{HV}_{\text{UNSENS}}$ vs. HV_{SENS} vs. FEP: $F_{(2,278.9)} = 10.14$, $p = .00005$). We did not observe any significant relationships between subcortical DA release and cortical volumes in FEP (Fig. 5). To account for possible changes in brain volume induced by AMPH sensitization, we derived volumetric parameters in an independent group of AMPH naïve, sex and age matched HVs. There were no significant differences between our group of HV_{SENS} and the independent HV group (Supplementary Fig. 6). Although of restricted power, these results argue against gross volumetric effects arising from our sensitization procedure.

Discussion

This study is the first to directly compare AMPH-induced DA release in patients with FEP to DA release in HV before and after prospective AMPH sensitization. Our data confirm the main hypothesis of this work: subcortical DA transmission in patients with FEP is in a state of “endogenous sensitization” towards AMPH. They thus replicate and extend the results of earlier studies that have used $D_{2/3}$ receptor antagonist-radioligands for studying DA release in SCZ^{3–6} and the effects of AMPH sensitization in HV¹⁵. In contrast to these studies, we found no relationship between DA release and change in positive symptoms of SCZ. However, we observed a significant

correlation between [^{11}C]-(+)-PHNO BP_{ND} values after AMPH in the putamen and negative symptoms, in particular emotional withdrawal. Although the interpretation of this finding is not straightforward, the finding was highly significant and is somewhat supported another [^{11}C]-(+)-PHNO study⁵⁰ that found a negative relationship between stress-induced DA release in SNVTA and negative symptoms in individuals with clinical high risk for schizophrenia (and at trend level, also patients with FEP). Together, these results corroborate the notion that reduced DA transmission is a relevant element in the pathogenesis of negative schizophrenic symptoms.

Our results are at odds with those of a recent study using the $\text{D}_{2/3}$ receptor agonist radioligand [^{11}C]NPA⁴⁷. In contrast to our results and those obtained with $\text{D}_{2/3}$ antagonist radioligands^{3–6}, the study by Frankle et al. study did not find enhanced AMPH-induced DA release in schizophrenia. Besides differences in clinical variables (as for example a smaller proportion of medication-naïve patients in the Frankle et al. study), the discrepancy could also reflect different properties of the two radioligands. Since the method used in both studies does not allow for reliably distinguishing between the absolute number of receptors and receptor affinity (including changes in apparent affinity induced by competition with endogenous DA⁵¹), it remains open how these parameters exactly relate to each other in dopaminergic dysfunction of SCZ⁵².

Relationships between volumetric and functional data suggest that AMPH-induced DA release in HV at first exposure to the drug is under inhibitory control of the left-hemispheric *DLPFC/IFG*. At the same time, AMPH sensitization increased DA release to a greater extent in subjects with larger *DLPFC/IFG* volumes. This relationship was particularly pronounced in GP and SNVTA (Fig. 4), where the [^{11}C]-(+)-PHNO signal is predominantly reflecting binding to DA D_3 rather than D_2 receptor subtypes^{53–55}. Since it is unlikely, according to our data obtained in an independent HV sample, that AMPH sensitization had induced major volumetric changes, the “herring bone” pattern most likely reflects a true functional shift in control of subcortical DA release by the *DLPFC/IFG* from inhibition in $\text{HV}_{\text{UNSENS}}$ towards facilitation in HV_{SENS} . This interpretation is supported by rodent data showing that expression of the sensitized state is critically dependent on the integrity of PFC neuronal tissue and its interaction with subcortical D_3 receptors⁵⁶.

In our study, prospective sensitization of HV led to an increase in DA release to the magnitude observed in FEP. *DLPFC/IFG* volumes, while showing strong correlations to DA release in $\text{HV}_{\text{UNSENS}}$, did not show any significant relationship to DA release in patients with FEP (Fig. 4). While we cannot rule out that this is due to lack of power, the very same region was found to show abnormal cortical

folding in SCZ patients with a mean illness duration of two decades and extensive exposure to antipsychotics⁵⁷. Thus, our data support the long-standing conjecture that the hyperdopaminergic state in SCZ is directly related to a dysfunction in top-down control of subcortical DA transmission by the PFC^{22,58}, and they are consistent with the notion that antipsychotics act by rectifying the consequences of upstream pathogenic factors, which persist despite antipsychotic treatment.

Antipsychotic-sensitive hyperlocomotion is a standard measure for sensitization in rodents and a widely accepted animal model of SCZ. Although not formally quantified in this study, the most evident behavioral effect of AMPH in our participants was an increase in quantitative speech production. Thus, while expressing on a motor level in rodents, enhanced DA transmission in human AMPH-sensitive brain circuits seems to affect primarily neuronal functions involved in the processing of language and speech, making speech being the human equivalent to rodent hyperlocomotion. Alterations in cytoarchitecture introduced into the rodent PFC by disrupting the cytoskeleton of dendritic spines increase subcortical DA levels and induce hyperlocomotion via direct projections between the PFC and SNVTA⁵⁹. Similar alterations in the *DLPFC/IFG* of patients with SCZ may cause dysfunctions in hierarchical control within *DLPFC/IFG*–SNVTA–striato-thalamic loops and may be involved in mediating the autonomous (i.e., not subject to will) production of language in auditory verbal hallucinations of patients with SCZ. Simultaneously activated by certain grammatical constructs⁶⁰, altered interactions between the *DLPFC/IFG* and the evolutionarily ancient SNVTA may also be involved in generating the fascinating and inherently grammatical symptom-complex of schizophrenic “ego disturbances”.

As prospective AMPH sensitization in HV successfully mimics the endophenotype of increased DA release observed in “psychotic sensitization”, our data prompt the question why the endophenotype associates with psychotic symptoms in SCZ but is occurring—unless re-exposed to AMPH—without any relevant behavioral effects in HVs. Symptom-provocation studies⁶¹ and ecological evidence strongly suggest this to be a matter of dose and intensity of exposure: While an escalation of the AMPH dose induces transient psychotic symptoms in psychiatrically healthy subjects⁶¹, subjects with stimulant-use disorders indeed exhibit disproportionately high rates of psychotic disorders⁶². Interestingly, subjects abusing methamphetamine or cocaine exhibit increased [^{11}C]-(+)-PHNO binding in the GP^{63,64}. What seems to differentiate AMPH-induced from “endogenous psychotic” sensitization in our data—higher [^{11}C]-(+)-PHNO BP_{ND} values in GP—may just be a matter of dose and length of exposure to AMPH.

An inherent limitation of our method is that it does not discriminate the effects of extracellular DA levels from changes in maximal $D_{2/3}$ receptor binding capacity. In addition, although more sensitive towards fluctuations in extracellular DA, the differences between HVs and FEP patients observed with the agonist radioligand [^{11}C]-(+)-PHNO are by no means larger than those observed with antagonist radioligands^{1–6}. However, this could in part also be a consequence of increased $D_{2/3}$ receptor occupancy due to higher baseline extracellular DA levels in patients with FEP (Fig. 3b). Another limitation might be the fact that some patients required benzodiazepines due to agitation or anxiety. Although there was no significant difference between medicated and unmedicated patients we cannot rule out the possibility that this might be a confounding factor. Furthermore, although sex did not play a role when entered as a covariate, the imbalance between male and female patients in the FEP sample has to be mentioned as a limitation. Lastly, our design has the limitation that we did not expose, mainly for ethical reasons, our FEP patients to a sensitizing regime of repeated AMPH. However, rarely cited earlier work by Strakowski et al.⁶⁵ has shown that FEP patients fail to show a progressive enhancement in the response to repeated AMPH. This suggests that FEP patients, at least at the level of behavior, are already maximally sensitized. Our data support and extend this finding to a neurochemical level. In summary, we feel confident that our work provides the first direct experimental proof for the hypothesis that the pathogenic substrate underlying psychosis in SCZ is indeed a state of “endogenous sensitization” in subcortical DA systems.

Acknowledgements

This study was funded by the Austrian Science Fund FWF [Grant No. P23585-B09], the Anniversary Fund of the Austrian National Bank [Grant No. 16723], the Medical Scientific Fund of the Mayor of Vienna (Medizinisch-Wissenschaftlichen Fonds des Bürgermeisters der Bundeshauptstadt Wien) [Grant No. 15189], and the Vienna Science and Technology Fund (WWTF) [Grant No. CS15-033], all granted to M.W., and the Austrian Science Funds FWF [Grant No. F3506-B11] granted to H.H.S.

Author details

¹Department of Psychiatry and Psychotherapy, Division of General Psychiatry, Medical University of Vienna, Währinger Gürtel 18-20, 1090 Vienna, Austria. ²Department of Clinical Pharmacology, Medical University of Vienna, Vienna, Austria. ³Department of Biomedical Imaging and Image Guided Therapy, Medical University of Vienna, Vienna, Austria. ⁴School of Psychology and Counseling and IHLI, Queensland University of Technology, Brisbane, Australia. ⁵Department of Nuclear Medicine, Inselspital, Bern University Hospital, University of Bern, Bern, Switzerland. ⁶Department of Laboratory Medicine, Medical University of Vienna, Vienna, Austria. ⁷Institute of Pharmacology, Medical University of Vienna, Vienna, Austria. ⁸Department of Psychiatry and Psychotherapy, Division of Social Psychiatry, Medical University of Vienna, Vienna, Austria. ⁹Department of Neurology, Medical University of Vienna, Vienna, Austria. ¹⁰Research Imaging Centre, Centre for Addiction and Mental Health, University of Toronto, Toronto, Canada. ¹¹Center for Medical Statistics, Informatics and Intelligent Systems, Medical University of Vienna, Vienna, Austria. ¹²Ludwig Boltzmann Institute Applied Diagnostics, Vienna, Austria

Author contributions

Drs. Weidenauer, Sauerzopf, Bauer, and Willeit had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Concept and design: Willeit, Bauer. Acquisition, analysis, or interpretation of data: All authors. Drafting of the manuscript: Weidenauer, Bauer, Sauerzopf, Praschak-Rieder, Willeit. Critical revision of the manuscript for important intellectual content: All authors. Statistical analysis: Dorffner, Weidenauer, Bauer, Sauerzopf, Rusjan, Willeit. Planning, analysis and interpretation of radiochemistry, laboratory and genetic analyses, structural MR imaging: Pichler, Pfaff, Berrotéran-Infante, Nics, Philippe, Wadsak, Mitterhauser, Sitte, Mitterhauser, Zimprich, Rusjan, Stimpfl, Pezawas, Meyer, Sezen, Rabl.

Conflict of interest

Without relevance to this work, Matthäus Willeit declares to having received speaker honoraria and consulting fees from Janssen-Cilag Pharma GmbH, Austria. Without relevance to this work, Wolfgang Wadsak declares to having received speaker honoraria from GE Healthcare, research grants from Ipsen Pharma, Eckert-Ziegler AG, Scintomics and ITG. WW is a part time employee of CBmed Ltd (Center for Biomarker Research in Medicine, Graz, Austria). Without relevance to this work Georg Dorffner is employee and shareholder of The Siesta Group GmbH, a clinical trial service provider in the area of electrophysiological measurements such as EEG. Harald Sitte received grants/research support, consulting fees and/or honoraria from AbbVie, Aesca, Amgen, Astellas, AstraZeneca, Astropharma, Chiesi, Gebro, IMH, IIR, Janssen-Cilag Lundbeck, MSD, Mundipharma, Pfizer, Ratiopharm, Roche, Sandoz, Serumwerk Bernburg, Shire, Vertex and is a member of advisory boards of Amgen, Chieri and Sanofi-Aventis. Marcus Hacker received consulting fees and/or honoraria from Bayer Healthcare BMS, Eli Lilly, EZAG, GE Healthcare, Ipsen, ITM, Janssen, Roche, Siemens Healthineers. Rupert Lanzenberger received travel grants and/or conference speaker honoraria from Shire, AstraZeneca, Lundbeck A/S, Dr. Willmar Schwabe GmbH, Orphan Pharmaceuticals AG, Janssen-Cilag Pharma GmbH, and Roche Austria GmbH. Siegfried Kasper received grants/research support, consulting fees and/or honoraria within the last three years from Angelini, AOP Orphan Pharmaceuticals AG, Celegne GmbH, Eli Lilly, Janssen-Cilag Pharma GmbH, KRKA-Pharma, Lundbeck A/S, Mundipharma, Neuraxpharm, Pfizer, Sanofi, Schwabe, Servier, Shire, Sumitomo Dainippon Pharma Co. Ltd. and Takeda. Ana Weidenauer, Martin Bauer, Ulrich Sauerzopf, Lucie Bartova, Lukas Nics, Sarah Pfaff, Cecile Phelippe, Neydher Berrotéran-Infante, Verena Pichler, Bernhard Meyer, Ulrich Rabl, Patrick Sezen, Paul Cumming, Thomas Stimpfl, Nilufar Mossaheb, Alexander Zimprich, Pablo Rusjan, Markus Mitterhauser, Lukas Pezawas, and Nicole Praschak-Rieder have no conflict of interest to declare.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary Information accompanies this paper at (<https://doi.org/10.1038/s41398-019-0681-5>).

Received: 2 September 2019 Revised: 1 October 2019 Accepted: 1 November 2019

Published online: 08 January 2020

References

- Lieberman, J. A., Kane, J. M. & Alvir, J. Provocative tests with psychostimulant drugs in schizophrenia. *Psychopharmacology* **91**, 415–433 (1987).
- Fusar-Poli, P. & Meyer-Lindenberg, A. Striatal presynaptic dopamine in schizophrenia, part II: meta-analysis of [(18)F]/(11)C]-DOPA PET studies. *Schizophrenia Bull.* **39**, 33–42 (2013).
- Laruelle, M. et al. Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc. Natl Acad. Sci. USA* **93**, 9235–9240 (1996).
- Breier, A. et al. Schizophrenia is associated with elevated amphetamine-induced synaptic dopamine concentrations: evidence from a novel positron emission tomography method. *Proc. Natl Acad. Sci. USA* **94**, 2569–2574 (1997).

5. Abi-Dargham, A. et al. Increased striatal dopamine transmission in schizophrenia: confirmation in a second cohort. *Am. J. Psychiatry* **155**, 761–767 (1998).
6. Laruelle, M., Abi-Dargham, A., Gil, R., Kegeles, L. & Innis, R. Increased dopamine transmission in schizophrenia: relationship to illness phases. *Biol. Psychiatry* **46**, 56–72 (1999).
7. Howes, O. D. et al. The nature of dopamine dysfunction in schizophrenia and what this means for treatment. *Arch. Gen. Psychiatry* **69**, 776–786 (2012).
8. van Rossum, J. M. The significance of dopamine-receptor blockade for the mechanism of action of neuroleptic drugs. *Arch. Internationales de Pharmacodynamie et de Therapie* **160**, 492–494 (1966).
9. Kapur, S. Psychosis as a state of aberrant salience: a framework linking biology, phenomenology, and pharmacology in schizophrenia. *Am. J. Psychiatry* **160**, 13–23 (2003).
10. Howes, O. D. & Kapur, S. The dopamine hypothesis of schizophrenia: version III—the final common pathway. *Schizophr. Bull.* **35**, 549–562 (2009).
11. Winton-Brown, T. T., Fusar-Poli, P., Ungless, M. A. & Howes, O. D. Dopaminergic basis of salience dysregulation in psychosis. *Trends Neurosci.* **37**, 85–94 (2014).
12. Pinsker, H. M., Hening, W. A., Carew, T. J. & Kandel, E. R. Long-term sensitization of a defensive withdrawal reflex in *Aplysia*. *Science* **182**, 1039–1042 (1973).
13. Sulzer, D. How addictive drugs disrupt presynaptic dopamine neurotransmission. *Neuron* **69**, 628–649 (2011).
14. Pierce, R. C. & Kalivas, P. W. Amphetamine produces sensitized increases in locomotion and extracellular dopamine preferentially in the nucleus accumbens shell of rats administered repeated cocaine. *J. Pharmacol. Exp. Ther.* **275**, 1019–1029 (1995).
15. Boileau, I. et al. Modeling sensitization to stimulants in humans: an [11C]raclopride/positron emission tomography study in healthy men. *Arch. Gen. Psychiatry* **63**, 1386–1395 (2006).
16. Steinkellner, T. et al. In vivo amphetamine action is contingent on alphaCaMKII. *Neuropsychopharmacology* **39**, 2681–2693 (2014).
17. Laruelle, M. The role of endogenous sensitization in the pathophysiology of schizophrenia: implications from recent brain imaging studies. *Brain Res. Rev.* **31**, 371–384 (2000).
18. Weidenauer, A. et al. Making sense of: sensitization in schizophrenia. *Int. J. Neuropsychopharmacol.* **20**, 1–10 (2017).
19. Pycocock, C. J., Kenwin, R. W. & Carter, C. J. Effect of lesion of cortical dopamine terminals on subcortical dopamine receptors in rats. *Nature* **286**, 74–76 (1980).
20. Falkai, P. et al. Kraepelin revisited: schizophrenia from degeneration to failed regeneration. *Mol. Psychiatry* **20**, 671–676 (2015).
21. Guo, J. Y., Ragland, J. D. & Carter, C. S. Memory and cognition in schizophrenia. *Mol. Psychiatry* **24**, 633–642 (2019).
22. Meyer-Lindenberg, A. et al. Reduced prefrontal activity predicts exaggerated striatal dopaminergic function in schizophrenia. *Nat. Neurosci.* **5**, 267–271 (2002).
23. Fusar-Poli, P. et al. Abnormal prefrontal activation directly related to pre-synaptic striatal dopamine dysfunction in people at clinical high risk for psychosis. *Mol. Psychiatry* **16**, 67–75 (2011).
24. Wilson, A. A. et al. Radiosynthesis and evaluation of [11C](+)-4-propyl-3,4,4a,5,6,10b-hexahydro-2H-naphtho[1,2-b][1,4]oxazin-9-ol as a potential radiotracer for in vivo imaging of the dopamine D2 high-affinity state with positron emission tomography. *J. Med. Chem.* **48**, 4153–4160 (2005).
25. Sheehan, D. V. et al. The Mini-International Neuropsychiatric Interview (M.I.N.I.): the development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. *J. Clin. Psychiatry* **59**, 22–33 (1998).
26. Kay, S. R., Fiszbein, A. & Opler, L. A. The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr. Bull.* **13**, 261–276 (1987).
27. Muller, M. J., Rossbach, W., Davids, E., Wetzels, H. & Benkert, O. [Evaluation of standardized training for the “Positive and Negative Syndrome Scale” (PANSS)]. *Der Nervenarzt* **71**, 195–204 (2000).
28. Brauer, L. H. & de Wit, H. Subjective responses to d-amphetamine alone and after pimozide pretreatment in normal, healthy volunteers. *Biol. Psychiatry* **39**, 26–32 (1996).
29. Shotbolt, P. et al. Within-subject comparison of the sensitivity of [11C](+)-PHNO and [11C]raclopride to amphetamine induced changes in endogenous dopamine in healthy human volunteers. *Neuroimage* **52**(Supplement 1), S42–S43 (2010).
30. Willeit, M. et al. First human evidence of d-amphetamine induced displacement of a D2/3 agonist radioligand: A [11C](+)-PHNO positron emission tomography study. *Neuropsychopharmacology* **33**, 279–289 (2008).
31. Rami-Mark, C. et al. Reliable set-up for in-loop (1)(1)C-carboxylations using Grignard reactions for the preparation of [carbonyl-(1)(1)C]WAY-100635 and [(1)(1)C](+)-PHNO. *Appl. Radiat. Isot.* **82**, 75–80 (2013).
32. White, T. L., Justice, A. J. & de Wit, H. Differential subjective effects of D-amphetamine by gender, hormone levels and menstrual cycle phase. *Pharm. Biochem. Behav.* **73**, 729–741 (2002).
33. Justice, A. J. & de Wit, H. Acute effects of estradiol pretreatment on the response to d-amphetamine in women. *Neuroendocrinology* **71**, 51–59 (2000).
34. Mombour, W., Kockott, G. & Fliege, K. The use of the brief psychiatric rating scale (BPRS) by overall and gorham for the diagnosis of acute paranoid psychoses: evaluation of a german translation of the BPRS (author’s transl). *Pharmakopsychiatr. Neuropsychopharmacol.* **8**, 279–288 (1975).
35. Rusjan, P. et al. An automated method for the extraction of regional data from PET images. *Psychiatry Res.* **147**, 79–89 (2006).
36. Wu, Y. & Carson, R. E. Noise reduction in the simplified reference tissue model for neuroreceptor functional imaging. *J. Cereb. Blood Flow. Metab.* **22**, 1440–1452 (2002).
37. Ginovart, N. et al. Positron emission tomography quantification of [11C](+)-PHNO binding in the human brain. *J. Cereb. Blood Flow. Metab.* **27**, 857–871 (2007).
38. Camps, M., Cortes, R., Gueye, B., Probst, A. & Palacios, J. M. Dopamine receptors in human brain: autoradiographic distribution of D2 sites. *Neuroscience* **28**, 275–290 (1989).
39. Hall, H. et al. Autoradiographic localization of extrastriatal D2-dopamine receptors in the human brain using [125I]epidepride. *Synapse* **23**, 115–123 (1996).
40. Levant, B. Differential distribution of D3 dopamine receptors in the brains of several mammalian species. *Brain Res.* **800**, 269–274 (1998).
41. Gunn, R. N., Lammertsma, A. A., Hume, S. P. & Cunningham, V. J. Parametric imaging of ligand-receptor binding in PET using a simplified reference region model. *NeuroImage* **6**, 279–287 (1997).
42. Destrieux, C., Fischl, B., Dale, A. & Halgren, E. Automatic parcellation of human cortical gyri and sulci using standard anatomical nomenclature. *NeuroImage* **53**, 1–15 (2010).
43. Vilkman, H. et al. The effects of lorazepam on extrastriatal dopamine D(2/3)-receptors-A double-blind randomized placebo-controlled PET study. *Psychiatry Res.* **174**, 130–137 (2009).
44. Farde, L., Hall, H., Pauli, S. & Halldin, C. Variability in D2-dopamine receptor density and affinity: a PET study with [11C]raclopride in man. *Synapse* **20**, 200–208 (1995).
45. Abi-Dargham, A. et al. Prefrontal dopamine D1 receptors and working memory in schizophrenia. *J. Neurosci.* **22**, 3708–3719 (2002).
46. Abi-Dargham, A. et al. Increased baseline occupancy of D2 receptors by dopamine in schizophrenia. *Proc. Natl Acad. Sci. USA* **97**, 8104–8109 (2000).
47. Frankle, W. G. et al. Amphetamine-induced striatal dopamine release measured with an agonist radiotracer in schizophrenia. *Biol. Psychiatry* **83**, 707–714 (2018).
48. Driesen, N. R. et al. Impairment of working memory maintenance and response in schizophrenia: functional magnetic resonance imaging evidence. *Biol. Psychiatry* **64**, 1026–1034 (2008).
49. Broca, P. Remarques sur le siège de la faculté du langage articulé, suivies d’une observation d’aphémie (perte de la parole). *Bull de la Société d’anatomie* **6**, 330–357 (1861).
50. Tseng, H. H. et al. Nigral stress-induced dopamine release in clinical high risk and antipsychotic-naïve schizophrenia. *Schizophr. Bull.* **44**, 542–551 (2018).
51. Ginovart, N., Wilson, A. A., Houle, S. & Kapur, S. Amphetamine pretreatment induces a change in both D2-receptor density and apparent affinity: a [11C]raclopride positron emission tomography study in cats. *Biol. Psychiatry* **55**, 1188–1194 (2004).
52. Slifstein, M. & Abi-Dargham, A. Is it pre-synaptic or postsynaptic? Imaging striatal dopamine excess in schizophrenia. *Biol. Psychiatry* **83**, 635–637 (2018).
53. Graff-Guerrero, A. et al. Blockade of [11C](+)-PHNO binding in human subjects by the dopamine D3 receptor antagonist ABT-925. *Int. J. Neuropsychopharmacol.* **13**, 273–287 (2010).
54. Rabiner, E. A. et al. In vivo quantification of regional dopamine-D3 receptor binding potential of (+)-PHNO: Studies in non-human primates and transgenic mice. *Synapse* **63**, 782–793 (2009).
55. Tziortzi, A. C. et al. Imaging dopamine receptors in humans with [11C](+)-PHNO: dissection of D3 signal and anatomy. *NeuroImage* **54**, 264–277 (2011).

56. Guillin, O. et al. BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. *Nature* **411**, 86–89 (2001).
57. Wisco, J. J. et al. Abnormal cortical folding patterns within Broca's area in schizophrenia: evidence from structural MRI. *Schizophr. Res.* **94**, 317–327 (2007).
58. Davis, K. L., Kahn, R. S., Ko, G. & Davidson, M. Dopamine in schizophrenia: a review and reconceptualization. *Am. J. Psychiatry* **148**, 1474–1486 (1991).
59. Kim, I. H. et al. Spine pruning drives antipsychotic-sensitive locomotion via circuit control of striatal dopamine. *Nat. Neurosci.* **18**, 883–891 (2015).
60. Newman, A. J., Supalla, T., Hauser, P., Newport, E. L. & Bavelier, D. Dissociating neural subsystems for grammar by contrasting word order and inflection. *Proc. Natl Acad. Sci. USA* **107**, 7539–7544 (2010).
61. Angrist, B. M. & Gershon, S. The phenomenology of experimentally induced amphetamine psychosis—preliminary observations. *Biol. Psychiatry* **2**, 95–107 (1970).
62. Yui, K., Ikemoto, S., Ishiguro, T. & Goto, K. Studies of amphetamine or methamphetamine psychosis in Japan: relation of methamphetamine psychosis to schizophrenia. *Ann. N. Y. Acad. Sci.* **914**, 1–12 (2000).
63. Boileau, I. et al. Higher binding of the dopamine D3 receptor-preferring ligand [¹¹C]-(+)-propyl-hexahydro-naphtho-oxazin in methamphetamine polydrug users: a positron emission tomography study. *J. Neurosci.* **32**, 1353–1359 (2012).
64. Worhunsky, P. D. et al. Regional and source-based patterns of [¹¹C]-(+)-PHNO binding potential reveal concurrent alterations in dopamine D2 and D3 receptor availability in cocaine-use disorder. *NeuroImage* **148**, 343–351 (2017).
65. Strakowski, S. M., Sax, K. W., Setters, M. J., Stanton, S. P. & Keck, P. E. Jr. Lack of enhanced response to repeated d-amphetamine challenge in first-episode psychosis: implications for a sensitization model of psychosis in humans. *Biol. Psychiatry* **42**, 749–755 (1997).