



1 **Holocene phototrophic community and anoxia dynamics in**
2 **meromictic Lake Jaczno (NE Poland) using high-resolution**
3 **hyperspectral imaging and HPLC data.**

4 Stamatina Makri¹, Andrea Lami², Luyao Tu¹, Wojciech Tylmann³, Hendrik Vogel⁴, Martin
5 Grosjean¹

6
7 ¹Institute of Geography & Oeschger Centre for Climate Change Research, University of Bern, Hallerstrasse 12,
8 3012 Bern, Switzerland

9 ²ISE-CNR Institute of Ecosystem Study, 50 Largo Tonolli, 28922 Verbania Pallanza, Italy

10 ³Faculty of Oceanography and Geography, University of Gdansk, Bazynskiego 4, PL-80952 Gdansk, Poland

11 ⁴Institute of Geological Sciences & Oeschger Centre for Climate Change Research, University of Bern, 3012,
12 Bern, Switzerland

13 *Correspondence to:* Stamatina Makri (stamatina.makri@giub.unibe.ch)

14

15 **Abstract.** Global spread of hypoxia and altered mixing regimes in freshwater systems is a growing major
16 environmental concern. Climate change and human impact are expected to increasingly deteriorate aquatic
17 ecosystems. The study of processes and drivers of such changes in the past provides a great asset for prevention
18 and remediation in the future. We used a multi-proxy approach combining high-resolution Hyperspectral Imaging
19 (HSI) pigment data, with specific HPLC chlorophylls and carotenoids to examine Holocene trophic state changes
20 and anoxia evolution in meromictic Lake Jaczno, NE Poland. A redundancy analysis RDA including pollen-
21 inferred vegetation cover, temperature and human impacts provides insight into specific conditions and drivers of
22 changing trophic and redox states in the lake. Anoxic and sulfidic conditions established in Lake Jaczno after
23 initial basin infilling 9500 years ago. Until 6700 cal BP, lake trophy was relatively low, water turbidity was high,
24 and green sulfur bacteria (GSB) were abundant within the phototrophic community, suggesting a deep oxic–anoxic
25 boundary and weak stratification. The period between 6700–500 cal BP is characterized by constantly increasing
26 lake production and a gradual shift from GSB to purple sulfur bacteria (PSB), suggesting a shallower oxic–anoxic
27 boundary and pronounced stratification. Yet, the presence of spheroidene and spheroidone in the sediments
28 indicates intermittent anoxia. After 500 cal BP, increasing human impact, deforestation and intensive agriculture
29 promoted lake eutrophication, with a shift to PSB dominance and establishment of permanent anoxia and
30 meromixis. Our study unambiguously documents the legacy of human impact on processes determining
31 eutrophication and anoxia.

32

33 **Keywords:** Paleolimnology, Anoxia, Meromixis, Varved sediments, North–East Europe, Holocene, Sedimentary
34 pigments, Human impact

35 **1. Introduction**

36 Eutrophication and subsequent oxygen depletion have become primary water quality issues for most freshwater
37 and coastal marine ecosystems globally (Schindler, 2006; Jenny et al., 2016a). Rising mean global temperature
38 can potentially worsen lake anoxia by enhancing water stratification and algal blooms (Adrian et al., 2009;



39 Woolway and Merchant, 2019). Higher lake trophy and reducing conditions in anoxic bottom waters can have
40 diverse and profound negative effects on lake ecosystems, such as toxic algal blooms, fish kills, biodiversity loss
41 (Smol, 2010; Battarbee and Bennion, 2012; Makri et al., 2019), and nutrient recycling into the water column from
42 the sediment through redox processes (Gächter, 1987; Tu et al., 2019). Hence, the global spread of hypoxia grows
43 into a major environmental concern.

44

45 Temporal and spatial extents of hypoxia/anoxia are influenced by both biological (aquatic production, organic
46 matter decomposition) and physical (water stratification and lake mixing) factors (Smith and Schindler, 2009;
47 Friedrich et al., 2014; Jenny et al., 2016b). Environmental and climatic effects such as temperature, seasonality
48 and extreme events, catchment vegetation, land use, human impact and nutrient input affect lake production and
49 oxygen supply in the bottom waters.

50

51 Observational data of anoxia and aquatic production cover usually only very short periods, which restricts the
52 understanding of relevant processes and the knowledge of pre-disturbance conditions. This is most relevant for
53 lake management or restoration. Although recent anoxia and eutrophication have been very well studied and
54 understood (Naeher et al., 2013; Friedrich et al., 2014; Jenny et al., 2016a), less is known about the onset, cessation
55 and specific conditions of these changes in the past due to the lack of effective and easily measurable proxies
56 (Friedrich et al., 2014; Makri et al., 2020). More specifically, to assess lake trophy and/or bottom water
57 oxygenation, proxies such as sedimentary pigments (Lami et al., 2000; Leavitt and Hodgson, 2001; Guilizzoni and
58 Lami, 2002), lipid biomarkers (Naeher et al., 2012), diatom (Bennion and Simpson, 2011) and chironomid records
59 (Little et al., 2000), stable isotopes (Pearson and Coplen, 1978), and redox sensitive elements such as Fe, Mn, Mo,
60 V, and U (Naeher et al., 2013; Wirth et al., 2013; Costa et al., 2015), have been extensively used so far.
61 Nonetheless, on long-term Holocene time-scales, most of these proxy records are typically established at a
62 centennial resolution at best.

63

64 Laminated lake sediments are valuable archives of natural and anthropogenic impacts, providing long-term records
65 via various biogeochemical proxies. Sedimentary photosynthetic pigment records can be effectively used to infer
66 both changes in algal composition and lake oxygen conditions (Guilizzoni et al., 1983; Leavitt, 1993; Lami et al.,
67 2000). Chlorophylls, together with their derivatives, and various carotenoids specific to particular groups of algae
68 can be used to reconstruct overall primary production and the composition of past photosynthetic communities
69 (Leavitt and Hodgson, 2001). Pigments such as okenone and isorenieratene, which are specific to phototrophic
70 sulfur bacteria that live in the anoxic sulfidic zones, are regarded as very good indicators of anoxia (Züllig, 1989;
71 Guilizzoni and Lami, 2002). HPLC-inferred pigments have a coarser temporal resolution due to laborious sample
72 preparation and time-consuming HPLC measurements. Scanning Hyperspectral Imaging (HSI), a novel non-
73 destructive method to quantitatively infer the abundance of algal (TChl: chlorophylls and their derivatives) and
74 bacterial pigments (Bphe: bacteriopheophytins *a* and *b*), offers insight into past trophic and oxygen conditions at
75 unprecedented μm -scale (sub-seasonal) resolution (Butz et al., 2015; Makri et al., 2020), but with lower speciation
76 sensitivity.

77 In this study, we use the varved sediment record of Lake Jaczno (NE Poland) to explore the specific conditions
78 and mechanisms of trophic and oxygen state changes in the Holocene, under changing climatic and environmental



79 conditions. Our research has been guided by the following questions: i) Which conditions drove algae dynamics
80 and oxygen state changes in the Holocene before any significant human intervention? ii) How did climate,
81 catchment vegetation and erosional input affected the phototrophic community in the lake and iii) How does the
82 current trophic state and mixing regime of the lake compare with the past? For this, we combined a high-resolution
83 HSI-inferred record of TChl and Bphe, X-ray fluorescence (XRF) elemental data, and an HPLC pigment record at
84 coarser resolution but specific in the analysis. Our dataset was compared with vegetation and temperature
85 reconstruction data to investigate the environmental conditions at times of aquatic primary production and bottom
86 water oxygenation changes. Lake Jaczno provides ideal conditions to answer these questions. It contains an entirely
87 varved Holocene sediment record, which has so far only been analyzed for the last 1700 years for productivity,
88 anoxia (Butz et al., 2016, 2017) and historical land use (Poraj-Górska et al., 2017). Pollen records have revealed
89 that human pressure was low until the 17th century, when landscapes opened and agriculture intensified (Marcisz
90 et al., 2020). This is rare in Europe. Therefore, this site provides a unique opportunity for a long-term Holocene
91 assessment of the natural causes and dynamics of meromixis and hypoxia with limited anthropogenic impact until
92 historic times.

93 2. Study site

94 Lake Jaczno (54°16'25.5" N 22°52'15.9" E, 163 m a.s.l, Fig. 1a) is a small, 26 m deep, exoreic, kettle-hole lake
95 formed sometime after the Weichselian deglaciation ca. 15 ka BP in the Suwałki Lakeland in NE Poland
96 (Krzywicki, 2002). Lake Jaczno has a total surface area of 0.41 km² separated in five distinct basins with narrow
97 sills. It is fed by three permanent inflows (N and W) and one outflow in the south (Fig. 1b). Jaczno is classified as
98 dimictic and mesotrophic (Tylmann et al., 2013), with incomplete mixing or possibly even meromixis during some
99 years (Butz et al., 2016). Butz et al. (Butz et al., 2017) found that anoxic, and even meromictic conditions,
100 established naturally for most of the past 1700 years. Meromixis was interrupted repeatedly following sediment
101 slumping or flood events.

102 Microscopic and geochemical analyses in the sediments of Lake Jaczno have revealed seasonal layers (calcareous
103 biogenic varves) with a complex succession of diatoms and calcite, detrital siliciclastic material (quartz, clays),
104 organic fragments, and finally amorphous organic matter (Tylmann et al., 2013; Butz et al., 2016; Poraj-Górska et
105 al., 2017). The lake is surrounded by steep slopes and gullies with ephemeral or perennial water flow, transporting
106 detrital material to the lake (Fig. 1c).

107

108 The catchment area (ca. 9 km²) is covered by glacial tills, sands and fluvio-glacial deposits. Modern soils are
109 classified as cambisols and podsols in the northern part and ferralic cambisols in the southern part of the catchment.
110 Agricultural lands dominate in the central and northern parts and forests in the southern parts (Fig. 1c). The lake
111 is surrounded by peatlands and forests dominated by birch, alder and spruce (Weisbrodt et al., 2017). The climate
112 of the region is continental with a mean annual temperature of 6.8°C and a mean annual precipitation of 600 mm
113 (Anon, 2017). The lakes in the area typically remain ice covered from December to March (Amann et al., 2014).

114 Archeological investigations in the Suwałki region indicate sparse or only seasonal human occupation during the
115 Mesolithic and Neolithic (10,000–3800 cal BP) (Engel and Sobczak, 2012). Around 2000 cal BP human presence
116 increases in the region with stronghold settlements, animal husbandry and fishing (Kinder et al., 2019). Yet, the
117 area around Lake Jaczno remained isolated from human influences (Marcisz et al., 2020). Pollen and charcoal



118 data, and increased soil erosion indicate extensive forest clearance, forest fires and intensified agriculture,
119 suggesting permanent settlements and higher human impact since 500 cal BP, especially after 150 cal BP (1800
120 CE) (Kinder et al., 2019; Marcisz et al., 2020). The 1970s are marked by a regeneration of forest cover and a
121 significant increase of fertilizer use in agriculture (Poraj-Górska et al., 2017; Kinder et al., 2019), which markedly
122 increased lake primary production (Butz et al., 2016; Poraj-Górska et al., 2017).

123 3. Materials and methods

124 Two parallel cores ca. 12.5 m long were retrieved in September 2017, using a UWITEC piston corer. The coring
125 site was located at the deepest part (24 m water depth) of the lake in the southern basin, which is protected from
126 direct external inputs (Fig. 1b). The cores were split lengthwise and then described following Schnurrenberger et
127 al. (2003) and the Munsell color chart (Munsell Color (Firm), 2010). Flood deposits and slumps were identified
128 based on grain size, mineral content, and sediment structure. First, the core halves were analysed using non-
129 destructive methods and further analytical measurements were performed after subsampling. The sampling interval
130 for LOI, CNS and dry-bulk density analysis was 10 cm (ca. 80-year resolution, discrete sampling). For the HPLC
131 and spectrophotometer analysis, 46 discrete samples (1–2 cm³) were taken every ca. 30–35 cm (ca. 230 years
132 resolution) taking into account the HSI scanning data and optimization for the proxy-to-proxy calibration of the
133 HSI indices with spectrophotometer data (Butz et al., 2015). The top 10 cm (last ca. 50 years) were subsampled
134 continuously every 1 cm.

135

136 The chronology is based on 18 radiocarbon AMS dates on taxonomically identified terrestrial plant macrofossils
137 (Table 1) measured at the Laboratory for Radiocarbon Analysis at the University of Bern. Samples with < 300 µg
138 C were measured using the gas-source input of the MIni CARbon DAting System (Szidat et al., 2014; Zander et
139 al., 2020). The age–depth model was calculated using Bacon (rbacon v. 2.4.2; Blaauw et al., 2020; Blaauw and
140 Christeny, 2011) and the IntCal13 calibration curve (Reimer et al., 2013). Event layers (>3 cm) and slumps were
141 excluded from the age calculation (Fig. 2). According to changes in lithology, we used model parameters that
142 allowed for a higher sedimentation rate in the lowermost 137 cm (Fig. 2).

143

144 XRF scanning was performed at continuous 2 mm steps using an ITRAX µXRF core scanner (exposure time 20 s,
145 30 kV and 50 mA) equipped with a Cr-tube at the University of Bern. The results are given as counts (peak area).
146 From the detected elements, Ti was used as proxy for erosional input from the catchment, Ca as a proxy for
147 endogenic calcium carbonates, Si/Ti as a proxy for biogenic silica, S, Fe, Mn, and Mn/Fe as proxies of changing
148 redox conditions (Koinig et al., 2003; Croudace and Rothwell, 2015).

149

150 Hyperspectral imaging scanning (HSI) was performed on the freshly oxidized core halves using a Specim PFD-
151 CL-65-V10E camera (400 to 1000 nm spectral range; 2.8 nm spectral resolution). We used a spatial resolution of
152 ~68 µm per pixel with a spectral sampling of 1.57 nm. Data were processed using the ENVI software version 5.4
153 (Exelis Visual Information Solutions, Boulder, Colorado) following Butz et al. (2015). The relative absorption
154 band depths (RABDs) were calculated based on spectral endmembers analysis in ENVI. The RABD₆₇₃ (spectral
155 region 590–730 nm) was used to detect chlorophylls and their diagenetic products (Tchl) and served as a proxy
156 for aquatic primary production (Leavitt and Hodgson, 2001). The RABD₈₄₅ (spectral region 790–895 nm) was
157 used to detect bacteriopheophytin *a* and *b* (*Bp**he*) (Butz et al., 2015, 2016), which is a proxy for anoxia and



158 meromixis as described in Makri et al. (2020). Bphe *a* and *b* is produced by anoxygenic phototrophic purple sulfur
159 and non-sulfur bacteria that proliferate in illuminated anoxic habitats (Yurkov and Beatty, 1998; Madigan and
160 Jung, 2009). Green sulfur bacteria produce bacteriochlorophyll *c*, *d* and *e*, which do not absorb in the RABD₈₄₅
161 range. Therefore, HSI-inferred Bphe reflects purple bacteria abundance.

162

163 The spectral indices were calibrated with absolute pigment concentrations of 46 selected sediment samples (1 cm³)
164 measured by spectrophotometry (Shimadzu UV-1800). Pigments were extracted using pure acetone. The
165 supernatant was evaporated under nitrogen, and extracts were subsequently redissolved in 2 ml of pure acetone
166 (method adapted from Schneider et al. 2018). For the calculation of Bphe concentrations, we used the molar
167 extinction coefficient for Bphe *a* by Fiedor et al. (2002). For TChl, we applied the molar extinction coefficient for
168 chlorophylls and chlorophyll derivatives by Jeffrey et al. (1975). The performance of the proxy-to-proxy linear
169 regression models was assessed using the coefficient of determination (R²) and the root mean square error of
170 prediction (RMSEP) (Butz et al., 2015) run in R (R Core Team, 2015). The calibration model for TChl showed an
171 R² of 0.91 (p < 0.001) and a RMSEP ~8 % (Fig. S1a). The calibration model for Bphe showed an R² of 0.95 (p <
172 0.001) and a RMSEP ~6 % (Fig. S1b). The Shapiro–Wilk and the Kolmogorov–Smirnov tests of the residuals
173 show that they are most likely normally distributed, suggesting that inferences can be made with both models.

174

175 HPLC analysis was conducted on the same 46 samples used for the proxy-to-proxy calibration. Chlorophyll,
176 chlorophyll derivatives and carotenoids were measured using ion pairing reverse-phase (Mantoura and Llewellyn,
177 1983; Hurley, 1988). The system used a UV-VIS detector set at 460 nm and 656 nm for carotenoids and
178 chloropigments, respectively. The results were corrected for water content and expressed as nmol g OM⁻¹ (Züllig,
179 1982; Guilizzoni et al., 1983; Lami et al., 1994). According to Jeffrey et al. (2011), Guilizzoni and Lami (2002),
180 chlorophyll *a*, ββ-carotene, pheophytin *a*, and pheophytin *b* are considered as indicators of total algal biomass.
181 Chlorophyll *b* and lutein are associated with green algae. *B*-carotene, dinoxanthin (pyrophytes), diadinoxanthin
182 (siliceous algae), fucoxanthin (diatoms), diatoxanthin (chrysophytes) and alloxanthin (cryptophytes) are related to
183 brown algae. Echinenone and zeaxanthin are associated to blue–green algae, and myxoxanthophyll and
184 canthaxanthin to colonial and filamentous cyanobacteria (Leavitt and Hodgson, 2001). K-myxol (4-keto-myxol-
185 2'-methylpentoside) is associated with N-fixing cyanobacteria (*Anabaena flos-aquae*) (Kosourov et al., 2016).
186 Pheophorbide *a* is considered as indicator of grazing. In the phototrophic bacteria community, BChl *a* is common
187 to all anoxygenic phototrophic purple bacteria. Okenone (*Chromatium sp.*) is associated with purple sulfur bacteria
188 (PSB), whereas spheroidene and spheroidenone (*Rhodospseudomonas sphaeroides*) are related to purple nonsulfur
189 bacteria (PnSB). Both groups are able to oxidize sulfide. Yet, PSB store any S⁰ formed intracellularly, whereas
190 PnSB do so outside the cell (Madigan and Jung, 2009). The main difference between the two groups is that PSB
191 are strong photoautotrophs, whereas PnSB are physiologically versatile and can grow well both phototrophically
192 and in darkness via fermentation or anaerobic respiration (Madigan and Jung, 2009). *R. sphaeroides* are also
193 excellent N-fixing bacteria. Oxygen tolerance varies among species, with *R. sphaeroides* being able to grow under
194 vigorous aeration. Spheroidenone is produced by *R. sphaeroides* only when even small amounts of oxygen are
195 present (Züllig, 1989). Hence, the presence of spheroidenone is used as an indication of better oxygen conditions,
196 whereas the presence of spheroidene with parallel absence of spheroidenone is used as an indication of meromictic
197 conditions (Züllig, 1989; Guilizzoni and Lami, 2002). Isorenieratene is associated with GSB (*Chlorobium sp.*).
198 GSB have low light requirements and can cope with low light availability, occupying deeper layers in stratified



199 lakes (Montesinos et al., 1983). Hence, a dominance of GSB over PSB is used as an indicator of a deeper oxic–
200 anoxic boundary (Montesinos et al., 1983; Itoh et al., 2003).

201

202 Total organic carbon (TOC) was determined by Loss on Ignition (LOI; Heiri et al., 2001). Total carbon (TC) and
203 total nitrogen (TN) were measured with a CNS-Analyzer (Elementar vario EL cube). Total inorganic carbon (TIC)
204 was calculated by the difference between TC and TOC (Enters et al., 2010). The TOC/TN ratio was used to infer
205 changes in OM sources (Meyers, 2003). The lithogenic flux was calculated based on the residual calculation after
206 removing the organic matter and carbonate fraction by LOI.

207

208 Statistical analysis was performed in R (R Core Team, 2015). To define the sedimentary lithotypes we performed
209 an hierarchical unconstrained clustering on the geochemical proxies (XRF data: Ti, Ca, Si/Ti, Si, S, Fe, Mn, Mn/Fe;
210 HSI: TChl, Bphe; TOC, TIC, TN, TOC/TN, Fig. 3) using the Euclidean distance matrix and the ward.D2 clustering
211 method (Murtagh and Legendre, 2014). On the same dataset, we performed a PCA analysis with the samples
212 grouped based on the unconstrained clustering to investigate the relationships between the lithotypes and the
213 geochemical variables (Fig. S2). The data were log transformed and scaled before statistical analysis. We
214 performed a redundancy analysis (RDA) using the Vegan package (Oksanen et al., 2016) in R to relate the pigment
215 matrix i.e HPLC- and HSI-inferred pigment concentrations (Hellinger-transformed variables) to the environmental
216 variables i.e. temperature (Heikkilä and Seppä, 2010), arboreal pollen (AP), non-arboreal pollen (NAP) (Kinder et
217 al., 2019; Marcisz et al., 2020) and lithogenic flux (log transformed variables). The elements were plotted using
218 scaling 2 (see Borcard et al., 2011, pp. 166–167). This analysis was followed by a permutation test in R to test for
219 significance in the redundancy analysis (Legendre and Legendre, 1998; Borcard et al., 2011). The zones of pigment
220 data were defined by constrained clustering using the Bray distance and ward.D2 linkage method in R.

221 4. Results and interpretation

222 4.1 Chronology

223 The age–depth model (Fig. 2) reveals a basal age of ca. 9500 cal BP. The model shows a stationary distribution,
224 matching prior and posterior accumulation rates, and a smooth sediment accumulation as indicated by its memory
225 or variability (Fig. 2). Three radiocarbon samples (Fig. 2, in red) have calibrated ages that do not fit with the 95 %
226 confidence interval. Based on the lithology and the much older ages, these samples were considered as containing
227 reworked carbon and were excluded from the Bacon model. The sediment sequence is entirely laminated
228 throughout the Holocene showing regular continuous sedimentation without any hiatus. In the lowermost section
229 (1257–1120 cm), sedimentation rates (SR) are relatively high (0.5 cm y^{-1}) and the mean age error (95% confidence
230 interval) is ca. ± 160 years. Numerous event layers and slumps characterize the part between 1120 cm and 800 cm
231 where the SR is ca. 0.2 cm y^{-1} and the mean age error is ca. ± 200 years. From 800 cm to the top, the sediment is
232 continuously varved and the SR is 0.1 cm y^{-1} (0.2 cm y^{-1} in the last 500 years). The mean age error in this section
233 is ca. ± 140 years.

234 4.2 Lithotypes and biogeochemical proxies

235 Figure 3 shows the biogeochemical data that defined four sedimentary lithotypes A–D. Fig. S3 (supplementary
236 material) shows the RGB images and the biogeochemical composition of selected close-ups within the sediment



237 sequence. Lithotype A and B appear in segments between ca. 9500–6800 cal BP (Fig. 3). Lithotype A, at the
238 bottom part (9500–9200 cal BP), consists of light greenish grey (GLEY 2 7/2) fine sand and continues with pale
239 yellow (2.5Y 7/3) and grey (2.5Y 5/1) laminations with light greenish grey silty lenticular bedding. This part is
240 characterized by high detrital inputs (Ti, lithogenic flux), moderate carbonate content (Ca, TIC), low production
241 and biogenic silica (HSI-TChl, Si/Ti) and low TOC. Low HSI-Bphe and S, and higher Mn/Fe ratio indicate
242 effective oxygenation of bottom waters. From ca. 9200–8500 cal BP, lithotype B is introduced and is characterized
243 by slightly higher production (HSI-TChl) and biogenic silica (Si/Ti); carbonates (TIC, Ca), TOC and TN contents
244 increase, whereas higher S, Fe and HSI-Bphe, and lower Mn/Fe ratio indicate the development of anoxic (sulfidic)
245 conditions. Between ca. 8500–6800 cal BP, lithotype A continues with varved sediments; starting with biogenic
246 pale yellow (2.5Y 7/3) and grey (2.5Y 5/1) varves with intercalated reddish brown (2.5YR 5/4) and reddish black
247 (2.5YR 2.5/1) laminations rich in clastic material and iron oxides. In the second half of this part, varves are less
248 well-preserved with several intercalated clastic-rich laminations. Based on color, layer thickness, and grain size
249 we interpret these intercalated layers as event (flood) deposits. In this period, lithotype A is characterized by high
250 detrital input (Ti, lithogenic flux); primary production (HSI-TChl) remains unchanged and carbonates (Ca, TIC)
251 show increased variability. Biogenic silica (Si/Ti), TOC and Fe slightly decrease. S decreases, HSI-Bphe is very
252 low or absent and Mn/Fe increases, suggesting better oxygen conditions.

253
254 Lithotype C occurs between ca. 6800–500 cal BP and consists mainly of light grey (2.5YR) and dark grey (2.5YR
255 4/1) fine biogenic varves, with some dispersed event layers that occur only at the beginning of this period until ca.
256 6000 cal BP. This period is characterized by low erosional input (Ti, lithogenic flux), gradually increasing
257 production (HSI-TChl) and TOC content, fluctuating biogenic silica (Si/Ti) and constantly high carbonates content
258 (Ca, TIC). S counts are minimal. HSI-Bphe is mostly present suggesting the development of anoxic conditions in
259 the hypolimnion. Mn/Fe seems to fluctuate, with higher values when HSI-Bphe is lower and vice versa.

260
261 Lithotype D occurs from ca. 500 cal BP to the present and consists of biogenic pale yellow (2.5Y 7/3), grey (2.5Y
262 5/1) and dark grey (2.5Y 4/1) calcareous biogenic varves. This period is characterized by instances of higher
263 detrital input (Ti) and several intercalated event (flood) layers. Mn counts also increase. Primary production (HSI-
264 TChl), TOC and TN reach maximum levels, whereas biogenic silica and carbonates (Ca, TIC) decrease. HSI-Bphe
265 reach maximum values at the top suggesting persistent anoxia in this part. The Mn/Fe ratio and HSI-Bphe show
266 opposite fluctuations indicating phases of better oxygen conditions when Bphe is absent.

267 4.3 HPLC pigment stratigraphy

268 Figure 4 presents the pigment dataset of individual chlorophylls and carotenoids measured by HPLC in the
269 Holocene (Fig. 4a), and for the last 50 years (Fig. 4b). The pigments are grouped according to their taxonomic
270 relation and the zones are defined by constrained clustering, which yielded boundaries that are similar to those of
271 the sediment lithotypes (Fig. 3).

272
273 In zone I (ca. 9500–9200 cal BP), pigment concentrations are very low. Chromophytes are more abundant than
274 green algae, especially cryptophytes (alloxanthin) and chrysophytes (fucoxanthin). Blue–green algae (echinenone,
275 zeaxanthin) are present in low concentrations. Grazing (pheophorbide *a*) is low. In the purple bacteria group,



276 *Chromatium* species (okenone, PSB) are absent, whereas *R. sphaeroides* (spheroidene and spheroidenone, PnSB)
277 are both present in low concentrations. *Chlorobium sp.* (isorenieratene, GSB) are present in traces.

278

279 In zone II (ca. 9200–6700 cal BP), pigment concentrations increase overall. Green algae (chlorophyll *b*, lutein)
280 still have low concentrations, whereas chromophytes (β -carotene) become more abundant especially pyrophytes
281 (dinoxanthin) and chrysophytes (diatoxanthin) that show a distinctive local maximum around 7300 cal BP.
282 Colonial filamentous cyanobacteria (canthaxanthin) appear in this zone. Grazing (pheophorbide *a*) starts
283 increasing around 8300 cal BP. *Chromatium sp.* (okenone, PSB) is mostly absent. *R. sphaeroides* (spheroidene
284 and spheroidenone, PnSB) have moderate concentrations, whereas *Chlorobium sp.* (isorenieratene, GSB) reach a
285 maximum around 7300 cal BP.

286

287 In zone III (6700–500 cal BP), most pigments concentration increase gradually. Green algae (chlorophyll *b*)
288 increase significantly. Chromophytes remain abundant. Diatoms and other siliceous algae (diadinoxanthin,
289 fucoxanthin), and cryptophytes (alloxanthin) show a local maximum around 2000 cal BP. Blue–green algae
290 (echinenone, zeaxanthin) increase gradually. More colonial filamentous cyanobacteria (myxoxanthophyll) appear
291 around 2300 cal BP and, together with zeaxanthin, reach a maximum around 2000 cal BP. N-fixing cyanobacteria
292 (*k*-myxol) appear at ca. 5000 cal BP. *Chromatium sp.* (okenone, PSB) appear in this zone and increase gradually.
293 *R. sphaeroides* (spheroidene and spheroidenone, PnSB) also show a gradual increase, whereas *Chlorobium sp.*
294 (isorenieratene, GSB) decrease to minimum concentrations.

295

296 Zone IV (500 cal BP to present), is characterized by a further gradual increase of most pigments, reaching
297 unprecedented maximum concentrations at the top. In more detail, Fig. 4b shows the distribution of pigment
298 concentrations in the last 50 years. Most pigments reach maximum values around 1997 CE. *Chromatium sp.*
299 (okenone, PSB) have high concentrations, whereas *R. sphaeroides* are present producing only spheroidene and
300 almost no spheroidenone. *Chlorobium sp.* (isorenieratene, GSB) show only trace concentrations around 1997 CE.

301 4.4 The relationships between land use, temperature, and pigment stratigraphy

302 We applied a redundancy analysis (RDA) to examine the response of our HPLC- and HSI-inferred pigment dataset
303 to land use changes (arboreal pollen: AP, and non-arboreal pollen: NAP; Kinder et al., 2019; Marcisz et al., 2020),
304 annual mean temperature variability (Heikkilä and Seppä, 2010) and catchment surface processes (lithogenic flux).
305 Figure 5 shows the RDA ordination output in a triplot with the explanatory variables (in blue) and response (in
306 red) variables, as well as the samples divided into the four distinct zones defined by constrained clustering (see
307 Sect. 4.3). The numerical output shows that the first two axis (RDA 1 20.95 % and RDA 2 11.25 %) explain 36 %
308 of the variation (unadjusted values). The R^2_{adj} for the constrained ordinations suggests that this model explains ca.
309 29 % of the variation in the data. The permutation test on the unconstrained ordinations indicates that the first two
310 axis are significant ($p < 0.001$; Table S1) and represent the data adequately.

311

312 The RDA triplot (Fig. 5) shows that AP and NAP play an important role in the distribution of the pigment data
313 along the first axis (RDA 1). Lithogenic flux and temperature drive pigment variability along the second axis
314 (RDA 2). Lithogenic flux is strongly correlated with siliceous algae (fucoxanthin, diadinoxanthin) and blue–green
315 algae (echinenone), as well as enhanced aquatic primary production (HSI-TChl) by green algae (lutein) and



316 cryptophytes (alloxanthin, β -carotene). Lithogenic flux is clearly anticorrelated with PSB (okenone, HSI-Bphe)
317 and Chl *a*. AP is mainly correlated with GSB pigments (isorenieratene) indicating a deeper oxic–anoxic boundary,
318 and PnSB (spheroidene and spheroidenone) that suggest a more effective oxygenation of the water column. AP is
319 also correlated with variables indicating the presence of chromophyte (brown) algae, pyrophytes (dincoxanthin),
320 chrysophytes (diatoxanthin), as well as some blue–green algae (zeaxanthin). Higher lithogenic input and AP drive
321 pigment variability in zones I and II. Temperature seems to be correlated with higher production of some green
322 algae (Chl *b*, pheophytin *b*), increased cyanobacteria abundance ($\beta\beta$ -carotene, β -carotene) and colonial-
323 filamentous cyanobacteria (myxoxanthophyll, canthaxanthin). Temperature seems to drive pigment variability
324 mainly in zone III. NAP is correlated with PSB production (BChl *a*, HSI-Bphe and okenone), overall higher
325 primary production (Chl *a*, pheophytin *a*), higher grazing (pheophorbide *a*), and N-fixing cyanobacteria (k-myxol).
326 NAP drives pigment variability in zone IV.

327 5. Discussion

328 5.1 Combining sedimentological and biogeochemical data to infer past lake production and bottom 329 water oxygenation

330 The 12.5 m long and almost entirely varved sediment record of Lake Jaczno continuously spans the last ca. 9500
331 cal yr BP (Fig. 2). The chronology is robust and exclusively based on terrestrial macrofossils. The lithology of
332 Lake Jaczno (Fig. 3) revealed the deposition of frequent event layers between 8500–7000 cal BP, which likely
333 reflect a regional catchment/climatic signal as similar features have been observed, for the same period, in the
334 nearby Lake Szurpity (Kinder et al., 2020). The physical characteristics of the catchment favored the transport of
335 lithogenic material into the lake (Fig. 3), thereby possibly affecting the density stratification, light availability, and
336 subsequently the phototrophic community dynamics. A proper assessment of these changes requires high-
337 resolution data that is impossible to reach using HPLC data alone. Yet, the combination of high-resolution (μ -
338 scale) calibrated HSI bulk data for TChl and Bphe, combined with scanning XRF and compound specific HPLC
339 data, provides a unique opportunity for paleoproduction and paleooxygenation reconstructions at sub-seasonal
340 scale for multi-millennial-long records (Butz et al., 2017; Makri et al., 2020). This approach is directly applicable
341 to diverse lacustrine (Butz et al., 2017; Schneider et al., 2018; Makri et al., 2020; Sanchini et al., 2020) and
342 potentially marine environments (Hubas et al., 2011, 2013) with uncertain past redox state changes.

343
344 The calibration of the RABD₆₇₃ and RABD₈₄₅ to absolute pigment concentrations of green pigments (chlorophylls
345 and diagenetic products) and Bphe (*a* and *b*) respectively, revealed robust calibration statistics (Fig. S1,
346 supplementary material) with very low uncertainties (ca. 6–8 %) comparable to other studies (Butz et al., 2017;
347 Schneider et al., 2018; Makri et al., 2020; Sanchini et al., 2020). Between ca. 9200 and 7000 cal BP, the calibration
348 model of the RABD₆₇₃ for green pigments calculates negative concentrations (Fig. 3). This offset can be produced
349 by matrix effects, i.e. the variability of the reflectance of the sediment matrix or substances that absorb in the same
350 range as chlorophylls and their diagenetic products (590–730 nm) (Makri et al., 2020). Interestingly, GSB
351 (isorenieratene) peak between 9200 and 7000 cal BP (Fig. 4). GSB contain bacteriochlorophyll *c*, *d*, and *e* that
352 absorb in the same range as chlorophylls and chlorophyll derivatives (Oren, 2011). This could indicate that a part
353 of the RABD₆₇₃ calibration error may be due to the increased GSB abundance. Nonetheless, the calibration
354 statistics reveal an overall error of less than 8 %.



355 **5.2 Holocene production dynamics and chemocline evolution**

356 The presence of anoxygenic sulfur bacteria throughout our record, combined with chlorophylls, carotenoids and
357 geochemical evidence, suggests that euxinic conditions prevailed in Lake Jaczno for most of the past 9500 years.
358 Nonetheless, the changing composition of photosynthetic sulfur bacteria indicates persisting but variable euxinia.
359 Figure 6 summarizes the Holocene evolution of the relative abundance of PSB, PnSB and GSB, *Chromatium*
360 (*okenone*) and *Clorobium* (*isorenieratene*), the content of spheroidene and spheroidenone pigments produced by
361 *R. sphaeroides*, and the high-resolution calibrated HSI-TChl and Bphe, with respect to lithogenic flux, climate
362 variability (annual mean temperature; Heikkilä and Seppä, 2010) and human impact (land use and vegetation
363 cover; Kinder et al., 2019; Marcisz et al., 2020).

364 **5.2.1 Low trophic levels with a deep oxic–anoxic boundary**

365 In the period from 9500 to 6700 cal BP, which corresponds to pigment zones I and II, the phototrophic bacteria
366 population is dominated by GSB (Fig. 6). A small percentage of PnSB (*R. sphaeroides*) is present and seems to
367 produce both spheroidene and spheroidenone during this time. *Chromatium* (*okenone*, PSB) is almost completely
368 absent. Considering that *R. sphaeroides* produces spheroidenone only when even small amounts of oxygen is
369 present (Züllig, 1989) we suggest that, in this period, euxinic conditions were already present but the strength or
370 extent of anoxia was likely weak. HSI-Bphe that corresponds to purple bacteria is very low. Anoxia is mainly a
371 function of lake stratification and productivity. HSI-TChl, which indicates total primary production, is still at low
372 levels (Fig. 6). Indeed, the stratigraphy of individual pigments indicates low to moderate in-lake production, which
373 mainly consists of chromophyte (brown) algae and some colonial cyanobacteria (*canthaxanthin*) (Fig. 4), which is
374 also confirmed in the RDA analysis (Fig. 5). Brown siliceous algae are well adapted and tolerant algae species that
375 thrive in oligotrophic conditions in symbiosis with other algae species and bacteria (Bird and Kalf, 1986; Wetzel,
376 2001). Similar observations of algae composition were made in Lake Peipsi (Tönno et al., 2019; Estonia) and Lake
377 Lazduny (Sanchini et al., 2020; NE Poland).

378
379 Temperature gradually increased and a closed forest canopy with pine/birch and later elm/hazel/alder persisted in
380 the catchment (Fig. 6) (Galka, 2014). These provided shelter from wind and increased the nutrient pools in the
381 catchment soils (Bajard et al., 2017). The closed forest canopy, combined with the deep and relatively small basin
382 (relative depth 3.01 %), favors the establishment of a naturally anoxic hypolimnion (Zolitschka et al., 2015). Yet,
383 it seems that enhanced permanent stratification was still not established in the lake. This phase of GSB dominance
384 corresponds to a period of high lithogenic flux or high-energy sedimentation (Fig. 3, 6), as confirmed in the RDA
385 analysis (Fig. 5). Turbidity currents and underflows can increase turbidity and nutrient availability, but can also
386 cause sporadic ventilation of bottom waters. Higher suspended matter and/or algal growth would decrease light
387 availability in the oxic–anoxic boundary. Since GSB and PnSB are more tolerant to low light intensities than PSB
388 (Biebl and Pfennig, 1978; Parkin and Brock, 1980; Madigan and Jung, 2009), a dominance of GSB and presence
389 of spheroidene and spheroidenone in the sediments is expected under these conditions. Similar observations were
390 made in Lake Cadagno (Wirth et al., 2013). GSB often inhabit the lowermost part of stratified water bodies due to
391 their efficient light capture (Manske et al., 2005; Imhoff, 2014). Higher abundance of GSB could also indicate a
392 deep oxic–anoxic boundary in the lake (Itoh et al., 2003; Antoniadis et al., 2009).



393 5.2.2 Gradually increasing trophic levels with a shallower oxic–anoxic boundary

394 The period from 6700 to 500 cal BP, which corresponds to pigment zone III, is characterized by a gradual shift in
395 the phototrophic bacterial community to higher PSB abundance, especially after ca. 2000 cal BP (Fig. 6). GSB are
396 present, inhabiting the anoxic layers below PSB and seem to fluctuate as a function of the primary production in
397 the oxic layer (HSI-TChl) and related light availability (Montesinos et al., 1983). When production was higher in
398 the oxic layers, *Chromatium* (okenone, PSB) increased and *Chlorobium* (isorenieratene, GSB) decreased. This is
399 also confirmed by the individual pigment stratigraphy (Fig. 4). Green algae (chlorophyll *b*, lutein) and N-fixing
400 cyanobacteria (k-myxol) increase markedly since ca. 5500 cal BP, indicating a higher lake trophic than before,
401 driven most probably by lake ontogeny and a gradual increase of nutrient availability. The appearance of N-fixing
402 cyanobacteria at that time agrees with this interpretation. Prolonged periods of anoxia leading to intense recycling
403 of phosphorous from the sediments would decrease the N:P ratio in the water column promoting nitrogen fixation
404 by N-fixing algae (Howarth et al., 1999; Vitousek et al., 2002). Similar trends in lake trophic evolution are reported
405 from nearby Lake Szurpily (Kinder et al., 2019) and Lakes Albano and Peipsi (Lami et al., 2000; Guilizzoni and
406 Lami, 2002; Tönno et al., 2019). An increase of *Chromatium* (okenone, PSB) over *Chlorobium* (isorenieratene,
407 GSB) with increasing lake trophic has been reported from other lakes, e.g. Lake Albano in Italy (Lami et al., 1994),
408 Little Round Lake in Canada (Brown et al., 1984), and Lake Hamana in Japan (Itoh et al., 2003). *R. sphaeroides*
409 (PnSB) are also present producing both spheroidene and spheroidenone, suggesting phases of effective aeration of
410 bottom waters (Züllig, 1989).

411
412 The catchment is continuously densely forested and human impact is very low (Fig. 6). The RDA analysis points
413 to a temperature driven pigment variability in this zone, but mainly for cyanobacteria abundance (Fig. 5).
414 Cyanobacteria can benefit from higher water temperature, yet nutrient inputs have in most cases a much stronger
415 and synergetic effect (Lürling et al., 2018). Temperature variability did not seem to have affected lake stratification
416 directly. However, seasonality, precipitation and windiness play an important role in lake circulation and are not
417 reflected in the annual mean temperature variability. Hence, the role of climate may be underestimated. The oxic–
418 anoxic stratification was enhanced in this period but permanent perennial anoxia was still not established as
419 indicated also by the low HSI-Bphe concentrations. The increase in PSB abundance suggests a shallower oxic–
420 anoxic boundary (Itoh et al., 2003). It appears that during most of the Holocene, anoxia was largely influenced by
421 primary production and lithogenic flux. The case of Lake Jaczno is different from e.g. Lake Łazduny (Masurian
422 Lake District, NE Poland; Sanchini et al., 2020), where erosional input is negligible and anoxia was mainly a
423 function of primary production and forest cover.

424 5.2.3 20th century eutrophication, shallow oxic–anoxic boundary and meromixis

425 In the period from 500 cal BP to the present, which corresponds to pigment zone I, phototrophic sulfur bacteria
426 composition changed to an almost complete dominance of purple bacteria (Fig. 6). Between 500–200 cal BP, HSI-
427 Bphe and the absolute concentrations of PSB (okenone,) and PnSB (spheroidene and spheroidenone) are at a
428 minimum, but dominate the phototrophic bacteria community since GSB are completely absent (Fig. 4, 6). Lake
429 production (HSI-TChl) also decreases while lithogenic flux increases (Fig. 6). These suggest an oxic rather than
430 anoxic phase during this period, with some intervals of weak euxinia. Increased Mn accumulation during this time
431 (Fig. 3) supports the indications of rather oxygenated bottom waters.

432



433 Between 200 cal BP to the present, when human impact starts to increase in the catchment (Fig. 5), PSB increase
434 as well. Presence of spheroidene and only trace concentrations of spheroidenone (PnSB) and isorenieratene (GSB)
435 suggest increasing and gradually persisting anoxia. Intensive agriculture in the last 100 years and use of fertilizers
436 increased primary production (HSI-TChl) substantially to unprecedented levels in the 1990s, relative to the
437 Holocene baseline. This is also reflected in the individual pigment stratigraphy (Fig. 4). Bphe reaches maximum
438 levels suggesting persisting anoxia and mostly meromictic conditions in the lake, especially since the 1970s when
439 gradual afforestation in the catchment is observed. This is also supported by the HPLC-inferred composition of
440 phototrophic bacteria (Fig. 3).

441 In this period, the high-resolution HSI-Bphe record indicates that the intervals of lowest AP in the catchment
442 coincide with absence of Bphe, indicating oxic bottom waters. Bphe increases again only when AP and the tree
443 canopy recovers (Fig. 6), with a parallel absence of spheroidenone (PnSB), suggesting meromictic conditions.
444 Butz et al. (Butz et al., 2016, 2017) showed that these intervals of low AP and low or absent Bphe in the sediments
445 were accompanied by strong pulses of terrigenous material from the catchment. The role of human impact with
446 regard to anoxia and interrelated catchment processes (deforestation/afforestation and nutrient inputs) has also
447 been shown in other lakes with diverse timing of human impact onset. For example, Lake Moossee (Makri et al.,
448 2020) and Soppensee (Lotter, 1999) on the Swiss Plateau, Lakes Albano and Nemi in Italy (Guilizzoni et al.,
449 2002), Lake Zazari in Greece (Gassner et al., 2020) with an early Mid-Holocene human impact, and Lake Szurpity
450 (Kinder et al., 2019) in the vicinity of Lake Jaczno with a late human impact, mainly in the last 500 years.

451 6. Conclusions

452 In this study, we used a multiproxy approach, combining high-resolution HSI pigment data with lower resolution
453 HPLC-inferred concentrations of specific algal pigments, and geochemical data to investigate algal community
454 composition and its relationship with aquatic production and water column oxygenation in a 9500-years sediment
455 record from NE Poland. Land use changes, vegetation cover and climate variability were also taken into account.
456 Our aim was to examine factors that determine trophic state changes and lake stratification, in a lake system with
457 stable catchment vegetation and low human impact until very recent times.

458
459 The Holocene sedimentary pigment and geochemical record of Lake Jaczno revealed distinct changes in lake
460 trophy and stratification states, mainly driven by the catchment evolution, lithogenic flux, nutrient input and
461 subsequent increase in primary production. The lake had a first phase (9500–6700 cal BP) of low production that
462 consisted mainly of brown algae in the oxic zone, yet an early immediate establishment of weak euxinic conditions
463 in a deep water column dominated by GSB in its anoxic zone. Increased suspended loads, turbidity currents and
464 underflows seem to have increased turbidity and restricted the proliferation of PSB at the deep oxic–anoxic
465 boundary. Between 6700–500 cal BP, primary production increased gradually with higher contributions of green
466 algae and cyanobacteria, following lake ontogeny in a continuously densely forested catchment. The oxic–anoxic
467 boundary became gradually shallower with a shift from GSB to PSB. The composition of phototrophic bacteria
468 and the presence of spheroidene and spheroidenone (PnSB) in the sediments suggest pronounced yet intermittent
469 euxinia in the lake. Between 500 cal BP to the present, lake trophy increases dramatically, especially in the last
470 100 years, due to intensified human impact. Eutrophication accompanied by catchment deforestation and



471 subsequent afforestation after land abandonment were the main driving forces for the establishment of permanently
472 anoxic and meromictic conditions in the modern lake.

473

474 This study highlights the great potential of calibrated and validated HSI measurements combined with HPLC data.
475 Lake Jaczno provided a rare site to explore the mechanisms that can potentially induce changes in lake mixing,
476 production and sustained bottom water anoxia in times from minimum to intensive human impact, in a naturally
477 stratified lake system. Our findings, together with findings from other lakes across Europe, can greatly expand our
478 understanding on these major environmental problems while providing a tailored toolset for implementing
479 effective remediation techniques in the future.

480 **Data availability**

481 The data will be made available at PANGAEA

482 **Author contributions**

483 **Stamatina Makri:** Investigation, Data Curation, Formal analysis, Writing - Original Draft, Visualization. **Luyao**
484 **Tu:** Investigation, Writing - Review & Editing. **Andrea Lami:** Investigation, Writing - Review & Editing.
485 **Wojciech Tylmann:** Writing - Review & Editing. **Hendrik Vogel:** Writing - Review & Editing, **Martin**
486 **Grosjean:** Conceptualization, Methodology, Writing - Review & Editing, Supervision, Funding acquisition.

487 **Competing interests**

488 The authors declare that they have no conflict of interest.

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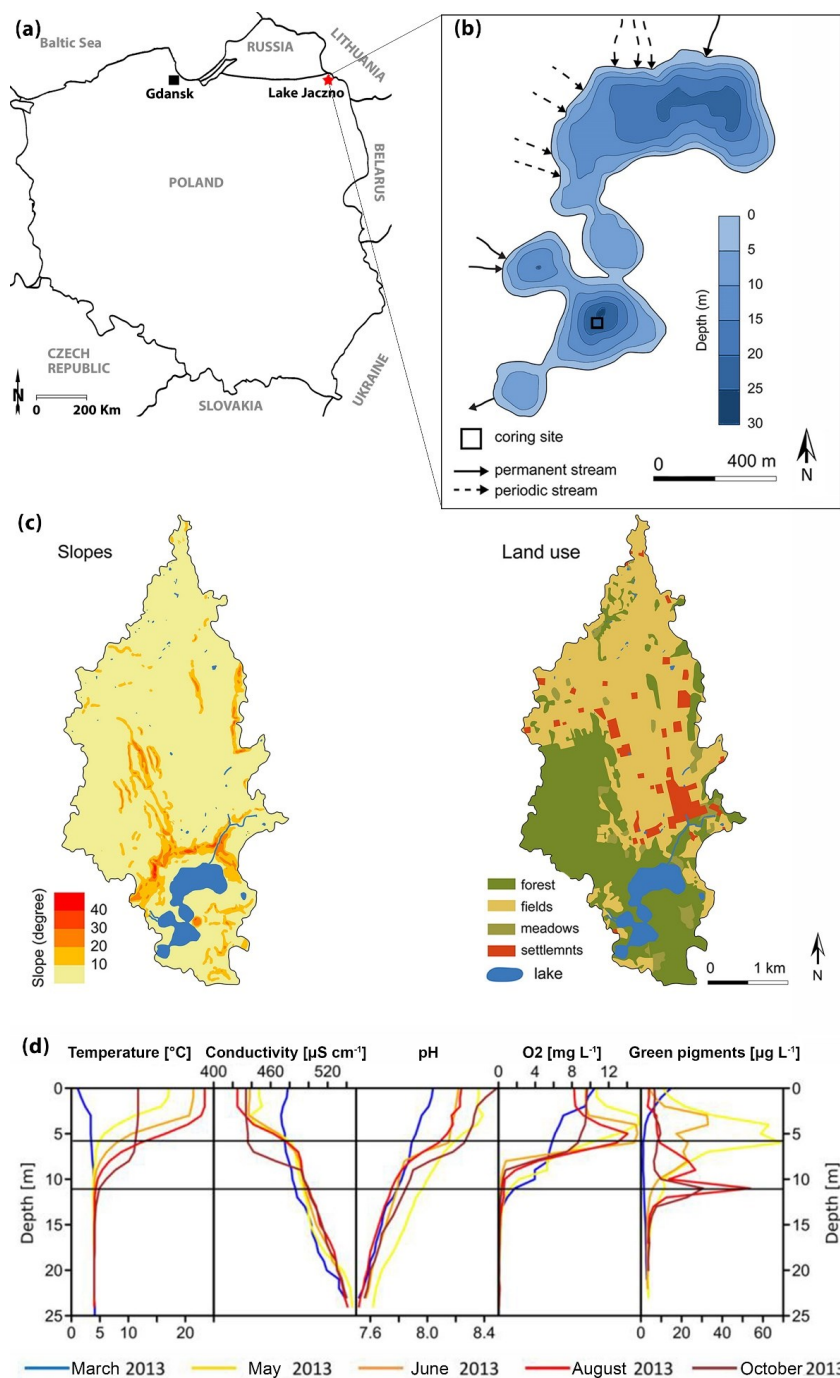
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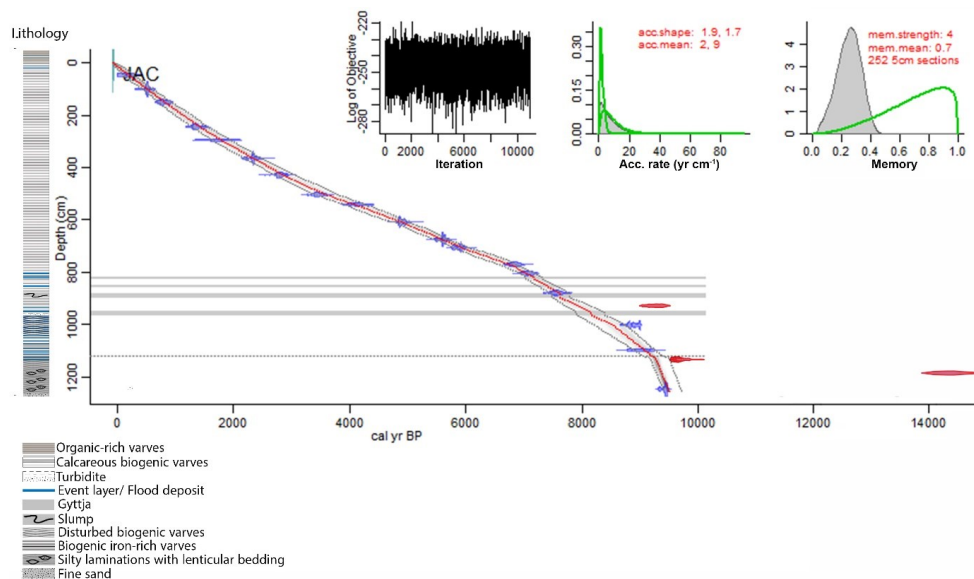
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Figure 1: a) Localization of Lake Jaczno. b) Lake bathymetry (modified from Poraj-Górska et al., 2017) and coring position c) Slopes and land use maps of the catchment (modified from Poraj-Górska et al., 2017) d) Seasonal limnological parameters in 2013 CE (Butz et al., 2016).



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774 **Figure 2: Age depth model and lithology of Lake Jaczno. The red line is the modelled chronology using**
775 **Bacon (Blaauw and Christeny, 2011; Blaauw et al., 2020); the excluded outliers are shown in red. The grey**
776 **dotted lines indicate the 95 % (2σ) probabilities. The grey horizontal areas indicate event layers (>3 cm)**
777 **excluded from the model. The horizontal dashed line marks the boundary of a higher sedimentation rate**
778 **(model parameter). The upper left panel shows the log objective vs. MCMC iteration that indicates a**
779 **stationary distribution. The middle and right panels indicate the distributions (prior in green, posterior in**
780 **grey) for the accumulation rate and the memory, respectively.**

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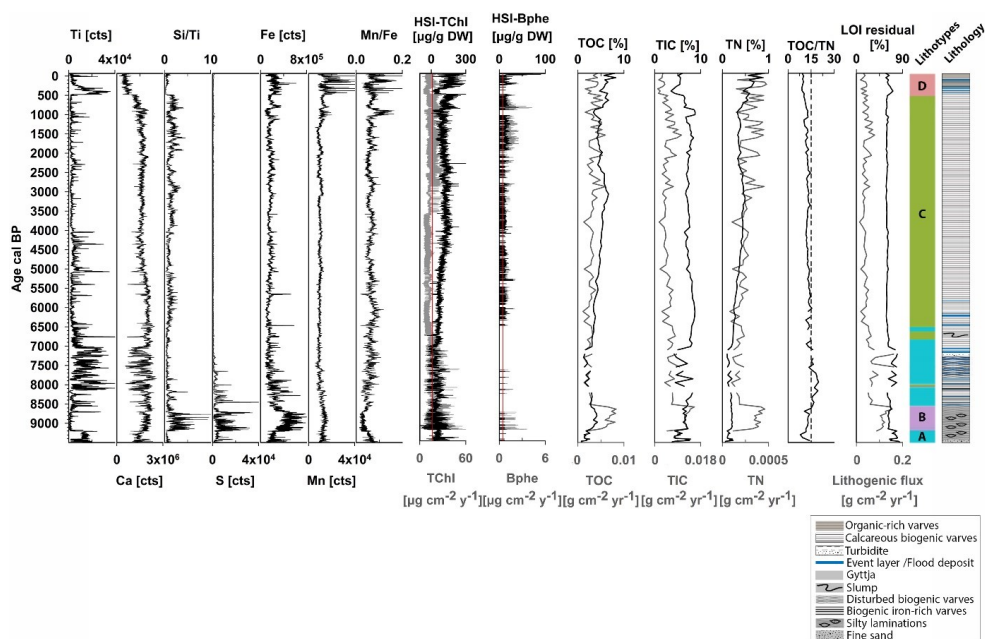
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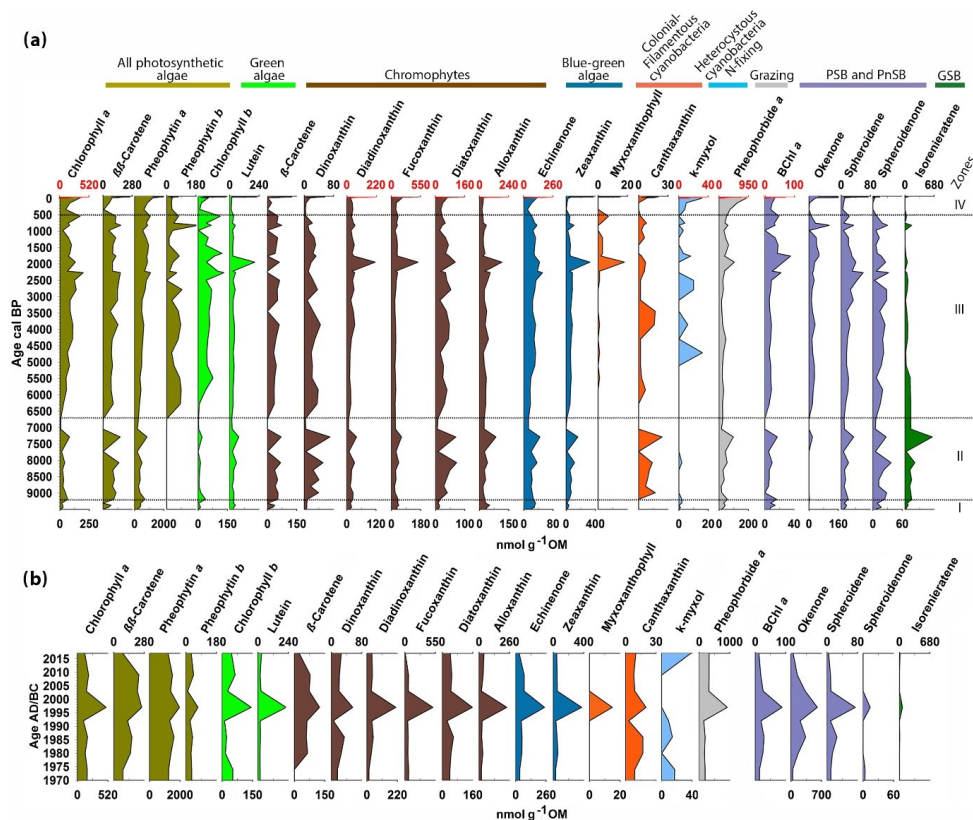
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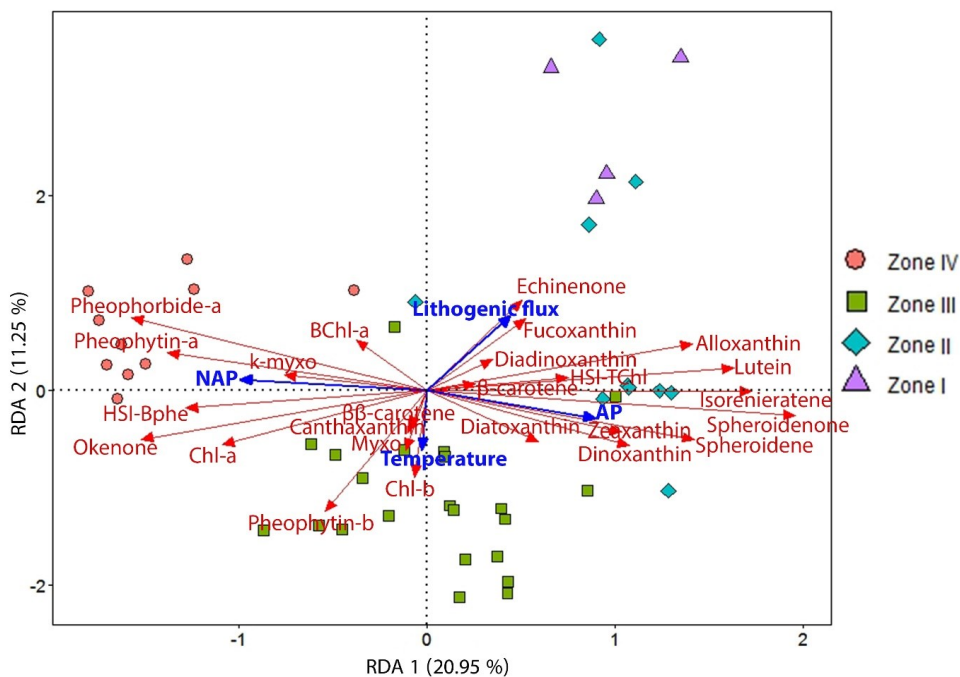
791 **Figure 3: Selected biogeochemical proxies that defined the four sedimentary lithotypes A–D (in different**
792 **color) after unconstrained clustering and PCA analysis (Fig. S2). On the right: sediment lithology based on**
793 **visual examination.**

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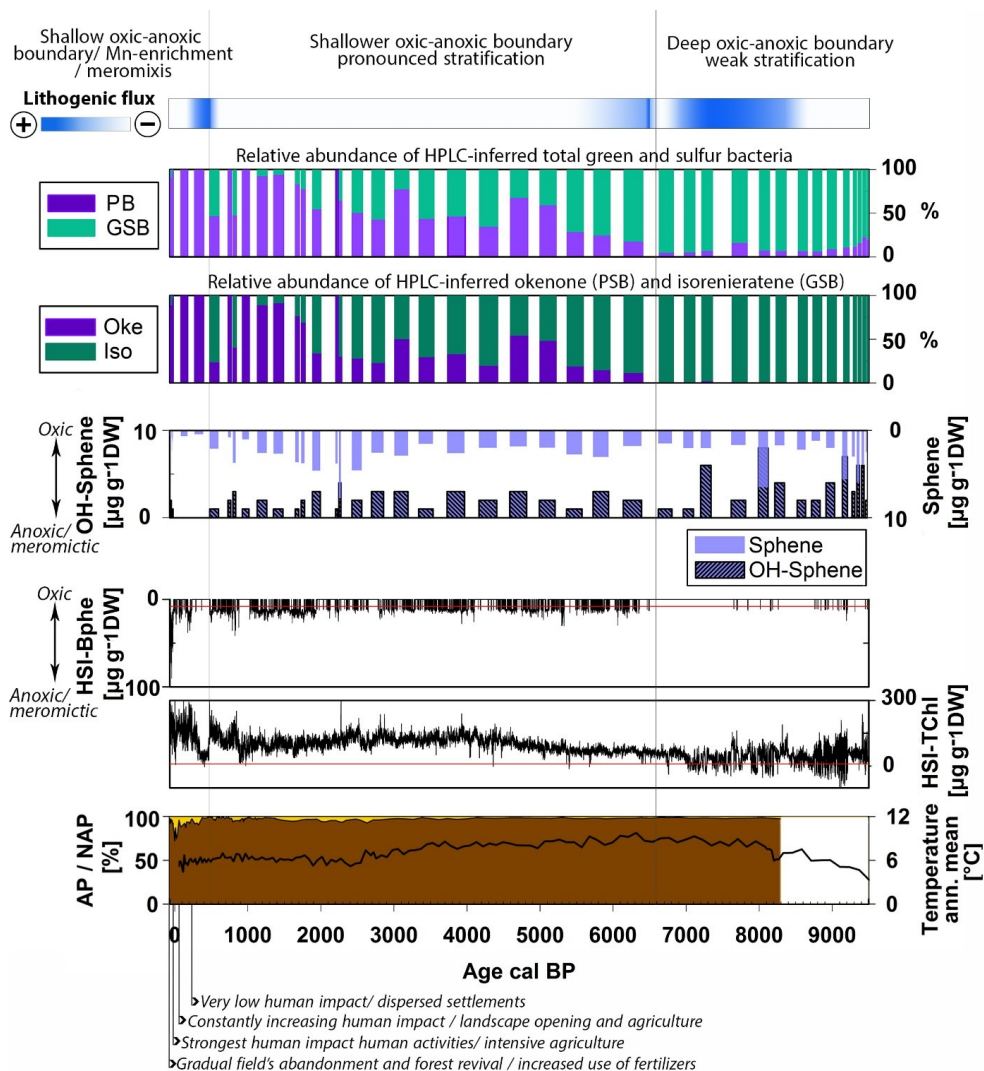


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Figure 4: Chlorophyll, chlorophyll derivatives, carotenoids and bacterial pigments concentrations measured by HPLC a) for the entire Holocene, and b) for the last 50 years. The zones are defined by constrained hierarchical clustering. The different colors indicate different algal groups based on the pigments' taxa affiliation. The occasional red scale on top marks the significantly higher concentrations of these pigments in the last 50 years.



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803 **Fig 5: RDA triplot showing the explanatory variables (AP, NAP, temperature and lithogenic flux) in blue,**
804 **and the response variables (HPLC- and HSI-inferred pigment concentrations) in red. The samples are**
805 **grouped according to the pigment zones (Fig. 4a).**
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808 **Figure 6: Holocene summary of the relative abundance of purple bacteria (sum of PSB and PnSB) and GSB,**
 809 ***Chromatium* (okenone, PSB) and *Clorobium* (isorenieratene, GSB), the content of spheridene and**
 810 **spheridenone pigments produced by *R. sphaeroides* (PnSB), and the high-resolution calibrated HSI-Tchl**
 811 **and HSI-Bphe concentrations. Top: indication of lithogenic flux and general evolution of the chemocline.**
 812 **Bottom: AP/NAP percentages (Kinder et al., 2019; Marcisz et al., 2020) with archaeological evidence of**
 813 **human impact, and the annual mean temperature variability (Heikkilä and Seppä, 2010).**

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818 **Table 1: Radiocarbon age results and calibrated ages. Uncertainties for ^{14}C ages refer to 68 % probabilities**
 819 **(1σ), whereas ranges of calibrated ages refer to 95 % probabilities (2σ). Outlier samples are marked with**
 820 **an asterisk. Indet: indeterminable, dicot: dicotyledonous.**

Sample ID	Material	C mass ($\mu\text{g C}$)	Age ^{14}C BP	Age (cal BP) ^a	Age range (cal BP) ^b	Graphite/ Gas
BE-10957.1.1	<i>Betula alba</i> fruit, woody scale, <i>Pinus sylvestris</i> needle base	104	132±64	142	0–284	gas
BE-10958.1.1	<i>Betula alba</i> fruit scale, woody scale, dicot leaf fragment	219	482±44	755	679–904	graphite
BE-10959.1.1	<i>Pinus sp.</i> periderm, coniferous wood and periderm fragment, <i>Betula alba</i> fruit fragments, coniferous scales	118	872±55	790	694–912	graphite
BE-10960.1.1	<i>Pinus sp.</i> periderm, <i>Betula alba</i> fruit fragments, conifer scale, <i>Pinus sp.</i> periderm, <i>Betula alba</i> fruit fragments	64	1460±67	1364	1283–1522	gas
BE-10961.1.1	<i>Betula alba</i> fruit fragments, conifer scale, <i>Betula alba</i> fruit, semi-charred periderm	19	1781±127	1706	1410–1987	gas
BE-10962.1.1	<i>Alnus glutinosa</i> fruit, <i>Betula alba</i> fruit fragments, needle/leaf indet, male anthere indet	268	2321±39	2341	2180–2458	graphite
BE-10963.1.1	<i>Betula alba</i> fruit fragments, <i>Pinus sp.</i> periderm, male anthere indet, <i>Betula alba</i> fruit fragments, <i>Pinus sp.</i> periderm, conifer scales	73	2677±69	2801	2545–2959	gas
BE-10964.1.1	<i>Betula alba</i> fruit fragments, <i>Pinus sp.</i> periderm, male anthere indet, <i>Betula alba</i> fruit fragments	74	3229±72	3458	3259–3633	gas
BE-10965.1.1	<i>Alnus glutinosa</i> fruit fragment, <i>Betula alba</i> fruit fragments, conifer scales, dicot leaf fragments	130	3758±60	4125	3927–4383	graphite
BE-10966.1.1	Male anthere indet, dicot leaf fragments, conifer scales, indet scale, wood indet	399	4322±36	4887	4836–4972	graphite
BE-10967.1.1	Dicot leaf fragments, indet scale, male anthere indet	428	4860±37	5601	5491–5650	graphite
BE-10968.1.1	Dicot leaf fragments, wood remains	348	5144±39	5906	5753–5989	graphite
BE-10969.1.1	Dicot leaf fragments, indet periderm, wood remains	183	5998±57	6839	6678–6977	graphite



BE-10970.1.1	Deciduous woody scale	365	6153±41	7062	6942–7166	graphite
BE-10971.1.1	<i>Betula alba</i> fruit fragments, <i>Pinus sp.</i> periderm, conifer scale, wood indet	111	6699±79	7567	7439–7674	graphite
*BE-10972.1.1	<i>Pinus sylvestris</i> needle fragments, wood indet	58	8291±99	9279	9029–9475	gas
BE-10973.1.1	Dicot leaf fragments, deciduous periderm, wood indet	996	8018±22	8896	8778–9007	graphite
BE-10974.1.1	<i>Betula alba</i> fruit fragments, <i>Pinus sylvestris</i> needle fragments, dicot leaf fragments	117	8094±91	9019	8651–9287	gas
*BE-10975.1.1	<i>Pinus sylvestris</i> needle fragments, <i>Betula alba</i> fruit fragments, indet periderm	280	8715±54	9677	9548–9887	graphite
*BE-10976.1.1	Dicot. leaf fragments, male anthere indet, conifer needle tip, indet periderm	308	12446±69	14580	14198–14994	graphite
BE-10977.1.1	Dicot leaf fragments, <i>Pinus sp.</i> periderm, woody scale	999	8388±22	9440	9318–9479	graphite

821 ^a Median probability (Stuiver and Reimer, 1993)

822 ^b Calibrated age range with the IntCal 13 calibration curve (Stuiver and Reimer, 1993; Reimer et al.,
 823 2013)

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