An essential novel component of the noncanonical mitochondrial outer membrane protein import system of trypanosomatids

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INTRODUCTION

The acquisition of a bacterial endosymbiont by a primitive host cell and its subsequent conversion into the mitochondrion marks one of the most important transitions in biology—the advent of the eukaryotic cell (Embley and Martin, 2006). A defining aspect of mitochondria is their capability to import proteins from the cytosol, a process that is driven by a set of characteristic protein translocases (Neupert and Herrmann, 2007; Lithgow and Schneider, 2010; Schmidt et al., 2010; Endo et al., 2011).

In the outer membrane we find two such translocases. The most important one—the general entry gate for essentially all imported proteins—is the translocase of the outer mitochondrial membrane (TOM). It consists of the pore-forming core subunit Tom40 (Hill et al., 1998; Ahting et al., 2001), a β-barrel–structured protein, protein import receptors (such as Tom70, Tom20, and Tom22 in yeast), and of a number of smaller subunits. The other translocase in the outer membrane is the sorting and assembly machinery (SAM), whose function is the insertion of β-barrel proteins. Its core subunit is Sam50, which is itself a β-barrel protein (Chacinska et al., 2009).

Extensive studies have revealed that most components and much of the architecture of the TOM and the SAM are conserved between yeast and mammals (Hoogenraad et al., 2002; Dolezal et al., 2006; Schneider et al., 2008; Hewitt et al., 2011). However a more global analysis shows that only the core components of the mitochondrial

ABSTRACT

The mitochondrial outer membrane protein Tom40 is the general entry gate for imported proteins in essentially all eukaryotes. Trypanosomatids lack Tom40, however, and use instead a protein termed the archaic translocase of the outer mitochondrial membrane (ATOM). Here we report the discovery of pATOM36, a novel essential component of the trypanosomal outer membrane protein import system that interacts with ATOM. pATOM36 is not related to known Tom proteins from other organisms and mediates the import of matrix proteins. However, there is a group of precursor proteins whose import is independent of pATOM36. Domain-swapping experiments indicate that the N-terminal presequence-containing domain of the substrate proteins at least in part determines the dependence on pATOM36. Secondary structure profiling suggests that pATOM36 is composed largely of α-helices and its assembly into the outer membrane is independent of the sorting and assembly machinery complex. Taken together, these results show that pATOM36 is a novel component associated with the ATOM complex that promotes the import of a subpopulation of proteins into the mitochondrial matrix.

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Abbreviations used: AspRS2, mitochondrial aspartyl-tRNA synthetase; ATOM, translocate of the outer mitochondrial membrane; BN-PAGE, blue native gel electrophoresis; COX4, cytochrome oxidase subunit 4; CYT C, cytochrome C; CYT C1, cytochrome C1; FCS, fetal calf serum.; HA, hemagglutinin; mHSP70, mitochondrial heat shock protein 70; ORF, open reading frame; pATOM36, peripheral subunit of ATOM of 35 kDa; PPR2, pentatricopeptide protein 2; RNAi, RNA interference; SAM, sorting and assembly machinery; TOM, translocate of the outer mitochondrial membrane; VDAC: voltage-dependent anion channel; TrpRS2, mitochondrial tryptophanyl-tRNA synthetase.

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protein translocases are conserved in all eukaryotes. Among these, Sam50 is the most highly conserved: it is found in all groups of eukaryotes and even has a bacterial orthologue, the Omp85-like protein BamA, that functions in inserting β-barrel proteins into the outer bacterial membrane (Bos et al., 2007). Tom40 is also conserved and found in virtually all eukaryotes; however, although the β-barrel structure of Tom40 would indicate a bacterial ancestry, no bacterial orthologue has been identified (Dolezel et al., 2006; Zeth, 2010).

The receptor subunits of the TOM complex are much less conserved. Thus, in plants Tom22 is severely truncated, and a conventional Tom20 is absent (Macasev et al., 2004). Instead we find a structural analogue of Tom20 that is coded in reverse, suggesting that the structural similarity between the mammalian and fungal Tom20 on one side and the plant Tom20 on the other side is due to convergent evolution and not to common descent (Perry et al., 2006). Tom70 appears to be missing in plants (Chan et al., 2006) but has recently been detected in some representatives of the Chromalveolata, which are unrelated to fungi, mammals, and plants, indicating that Tom70 might be more widespread than previously thought (Tsaoisz et al., 2011).

Recent studies in a number of different protozoa revealed interesting but for the most part minor deviations to the canonical protein import systems described in yeast and mammals (Dolezel et al., 2005). It was therefore a surprise that trypanosomatids have a fundamentally different mitochondrial outer membrane translocation machinery. Trypanosomatids lack Tom40 (Pusnik et al., 2009) and instead employ an outer membrane protein translocation channel termed ATOM, for archaic translocase of the outer mitochondrial membrane (Pusnik et al., 2011). Unlike Tom40, ATOM has a bacterial orthologue. It shows similarities to a subgroup of the bacterial Omp85-like protein family that is distinct from the Sam50 orthologue BamA. This suggests that trypanosomatids have retained an archaic outer membrane protein import system that might have preceded the Tom40-based protein import channel.

The discovery of ATOM provided unexpected new insights into the evolution of the mitochondrial outer membrane protein import system and the evolution of eukaryotes in general. Moreover, it raised the question of what other factors might be required for protein translocation across the outer membrane of trypanosomal mitochondria. Using Trypanosoma brucei as an experimental model system, we discovered such a novel factor. This protein is an essential mitochondrial outer membrane protein that is required for import of a large subset but not all matrix proteins and therefore might have a receptor-like function.

RESULTS
A novel mitochondrial outer membrane protein of T. brucei
Mitochondrial protein import is known to be essential in all eukaryotes under all growth conditions (Baker and Schatz, 1991). Whereas oxidative phosphorylation is essential only in the insect stage of T. brucei, mitochondrial protein import is required throughout the life cycle. Outer mitochondrial membrane proteins that are essential for normal growth of both procyclic and bloodstream forms of the parasite are therefore prime candidates to be components of the mitochondrial protein import system. To find such candidates, we purified mitochondrial outer membranes from T. brucei and characterized their protein composition (unpublished data). Within the outer membrane proteome we found a relatively abundant protein encoded by the open reading frame (ORF) Tb927.7.5700, which in a recent global RNA interference (RNAi) analysis (Alsford et al., 2011) was found to be essential throughout the life cycle. The protein has a predicted molecular weight of 45.5 kDa, but global transcriptome analyses of T. brucei (Siegel et al., 2011), biochemical experiments, and phylogenetic profiling against other kinetoplastid genomes indicate that a gene-interna

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To address this question, immunoprecipitations were performed to see whether pATOM36 interacts with ATOM or with SAM50, the only two components of the mitochondrial outer membrane known to be involved in mitochondrial protein import in *T. brucei*.

Gradient-purified mitochondria containing either HA-tagged pATOM36 or HA-tagged SAM50 were solubilized by digitonin and subjected to immunoprecipitation using anti-HA antibodies. The resulting eluates were analyzed for the presence of pATOM36, SAM50, and ATOM using mass spectrometry. Figure 3A shows that, as expected, the tagged proteins are efficiently recovered in the eluate. Moreover, a small amount of ATOM but not of SAM50 is reproducibly coprecipitated in the pATOM36-HA pull down. Consistent with these results, no pATOM36 is detected in the SAM50-HA pull down. Of interest, as in the case of pATOM36-HA, ATOM is also recovered. The experiment was performed in triplicate, and immunoprecipitations using wild-type mitochondria lacking any tagged proteins served as controls. To confirm these results, we repeated the pATOM36-HA immunoprecipitations and analyzed them by immunoblots. Figure 3B shows that a small but significant amount of ATOM is reproducibly recovered in the pellet fraction, whereas only background levels of the highly abundant membrane proteins cytochrome C1 (CYT C1) subunit 4 of cytochrome oxidase (COX4), and VDAC were detected. In summary, these experiments show that a fraction of pATOM36 is stably associated with ATOM but not with SAM50.

Although pATOM36 sequences are predicted to be largely α-helical in nature, none of these α-helices has the hydrophobicity characteristics of “classic” transmembrane helices. It is worth noting that this is also true for other outer membrane proteins, including Tom6 and Tom7 (Allen et al., 2002). To experimentally investigate whether pATOM36 is a β-barrel protein, we used an RNAi cell line allowing inducible ablation of SAM50. The results in Figure 2 show that ablation of SAM50 causes a strong reduction of the levels of the voltage-dependent anion channel (VDAC), as expected, whereas the levels of pATOM36 are not affected. Thus insertion of pATOM36 into the outer membrane does not require SAM50.

**Figure 1:** pATOM36 is an integral protein of the mitochondrial outer membrane. (A) Immunoblots of total (Tot), digitonin-extracted crude cytosolic (Cyt), and mitochondrial (Mit) fractions from a cell line expressing pATOM36 with an HA tag at its C-terminus (pATOM36-HA) probed with anti-HA antibodies (HA). VDAC and elongation factor 1a (EF-1a) serve as mitochondrial outer membrane and cytosolic marker, respectively. (B) Immunoblot of an alkaline carbonate extraction of pATOM36-HA–containing mitochondria. Pe, pellet; Sup, supernatant. The integral membrane protein VDAC and the soluble mHSP70 serve as markers. (C) Top, immunoblot of untreated and proteinase K-treated (50 μg/ml) pATOM36-HA–containing mitochondria. Mitochondrial carrier protein 5 (MCP-5) serves as a loading control. Bottom, immunoblot of isolated mitoplasts (Mp) having a disrupted outer membrane and the intact pATOM36-HA–containing mitochondria (Mit) shown at the top. CYT C and mHSP70 serve as markers for the intermembrane space and the matrix, respectively. (D) Left, import of in vitro–translated, 35S-Met–labeled pATOM36 into isolated mitochondria analyzed by BN-PAGE. Right, immunoblot of pATOM36-HA–containing mitochondrial extract separated on the same BN-PAGE. The main pATOM36 complex is indicated by the arrow. (E) Antibody shift experiments. Left, pATOM36-HA containing digitonin-lysed (+Digi) and intact mitochondria (−Digi) were incubated with polyclonal anti-HA antibodies (+HA, −antibody) and analyzed by BN-PAGE, and analyzed by immunoblots using a monoclonal anti-HA antiserum (HA). Untreated pATOM36-HA (−HA, −antibody) complex is shown on the left as a control. Right, same as shown on the left, but SAM50-HA–expressing mitochondria were used.

**Figure 2:** In vivo assembly of pATOM36 is independent of SAM50. Levels of pATOM36-HA in total cellular extracts are not affected, whereas the levels of the β-barrel protein VDAC declines during tet induction of SAM50 RNAi.
OM protein import in T. brucei abolishes growth of both insect-stage and bloodstream forms of lines. These cell lines show that ablation of the pATOM36 mRNA protein import, we prepared tetracycline (tet)-inducible RNAi cell lines. To directly study whether pATOM36 functions in mitochondrial import of matrix proteins, we performed experiments in which we examined the import of several matrix proteins into wild-type and pATOM36 RNAi cell lines. The results showed that pATOM36 is required for import of these proteins.

RNAi-mediated ablation of a protein import factor might be expected to cause an accumulation of precursor proteins in the cytosol and a concomitant decrease of the mitochondrial localized mature forms. However, previous experiments showed that accumulation of precursor is difficult to detect because mislocalized proteins that accumulate in the cytosol are often rapidly degraded. A decrease in the levels of newly synthesized imported proteins during induction of RNAi, on the other hand, was shown to be a highly sensitive proxy to quantify the inhibition of mitochondrial protein import (Pusnik et al., 2011).

In a first pATOM36 RNAi cell line, we tested import of newly synthesized mHSP70, for which, unlike for the other tested substrates shown later, we can see a time- and tetracycline-dependent accumulation of the precursor protein (Supplemental Figure S1). This indicates that pATOM36 is required for import of the mitochondrial matrix protein mHSP70.

To determine whether pATOM36 is required for import of other matrix proteins, we produced cell lines in which inducible RNAi was combined with inducible expression of the tagged matrix proteins pentatri-copeptide repeat protein 2 (PPR2) and PPR6, which belong to the same protein family but share very little sequence homology (Pusnik et al., 2007), and the mitochondrial tryptophanyl-tRNA (TrpRS2) (Charrière et al., 2006) and aspartyl-tRNA synthetases (AspRS2; Charrière et al., 2009). Each of these substrates was tested in the pATOM36 RNAi cell line, and, as a control, also in cell lines ablated for the general outer membrane import channel ATOM (Pusnik et al., 2011) and the putative inner membrane translocation pore TIM17 (Gentle et al., 2007; Singha et al., 2008), both of which are expected to be required for import of all matrix proteins.

Figure 5 shows that ablation of either ATOM or TIM17 affects import of all tested substrates. Ablation of pATOM36, however, resulted in a selective reduction of newly synthesized PPR2 and PPR6, whereas the levels of TrpRS2 and AspRS2 were not affected. This suggests that pATOM36 is required for import of PPR2 and PPR6 but not for import of TrpRS2 and AspRS2. Moreover, the fact that in the pATOM36 RNAi cell lines, just as in the ATOM and TIM17 RNAi cell lines, the reduction of PPR2 and PPR6 precedes the growth arrest suggests that pATOM36 plays a direct role in import.

Moreover, Supplemental Figure S2 shows that RNAi-mediated ablation of pATOM36 for 48 h affects neither mitochondrial membrane potential nor organellar ATP production (Bochud-Allemann and Schneider, 2002), indicating that the observed inhibition of protein import is not due to a general mitochondrial dysfunction.

**FIGURE 3:** pATOM36 is a peripheral component of the ATOM complex. (A) A 0.15% digitonin lystate of isolated, HA-tagged pATOM36 (left) and HA-tagged SAM50 containing mitochondria (right) were immunoprecipitated using anti-HA antibodies. The corresponding eluates were analyzed for the presence of pATOM36, SAM50, and ATOM using mass spectrometry. The graphs depict the number of evidences that were detected in the eluate of each experiment. The number of unique peptides identified for each protein is depicted on the top of each column. The experiment was performed in triplicate, and immunoprecipitations using wild-type mitochondria lacking any tagged proteins served as controls. pATOM36, SAM50, nor ATOM was recovered in the eluates of wild-type mitochondria. The only exception was ATOM that was detected by a single peptide/evidence in the first replicates of wild-type immunoprecipitates (data not shown). (B) A 0.15% digitonin lysate of isolated, HA-tagged pATOM36–containing mitochondria was immunoprecipitated using anti-HA antibodies and analyzed by immunoblot. Five percent of the total extract (Inp) and 95% of the bound fraction (IP) were analyzed by immunoblot using a monoclonal anti-HA antibody. The same samples were also analyzed with polyclonal ATOM, VDAC, COX4, and CYT C1 antisera. The experiment shown on the left was performed in quadruplicate and quantitated using the Odyssey infrared imaging system (Li-Cor Biosciences, Lincoln, NE). The means of the relative amounts of ATOM, VDAC, COX4, and CYT C1 that were recovered in the bound fraction were calculated. The pATOM36 that was recovered in the pellet was set to 100%. Standard errors are indicated.

**ATOM complex.** A low-affinity, nonstoichiometric, and dynamic interaction with the ATOM complex might be expected if pATOM36 has a receptor function; in yeast, the canonical protein import receptors Tom20 and Tom70 are recovered in the TOM complex only at low-stringency conditions (Meisinger et al., 2001).

**pATOM36 is required for in vivo import of a subset of matrix proteins**

To directly study whether pATOM36 functions in mitochondrial protein import, we prepared tetracycline (tet)-inducible RNAi cell lines. These cell lines show that ablation of the pATOM36 mRNA abolishes growth of both insect-stage and bloodstream forms of T. brucei (Figure 4).
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The requirement of pATOM36 for mitochondrial protein import was confirmed in vitro by comparing protein import into mitochondria isolated from noninduced and induced pATOM36 RNAi cell lines, respectively. Figure 6 shows that import of the in vitro–translated precursor of alcohol dehydrogenase III (Hauser et al., 1996) was greatly reduced in mitochondria isolated from pATOM36-ablated cells. Mitochondria used in this experiment were isolated 48 h after induction of RNAi, which is before the slow-growth phenotype becomes apparent (Figure 4), indicating that it is the impairment of protein import that causes the growth phenotype.

pATOM36 function depends on the N-terminal presequence-containing domain of substrate proteins

The fact that some but not all matrix proteins require pATOM36 to be imported in vivo is surprising. To investigate which determinants in the substrate proteins are responsible for the pATOM36 dependence of protein import, we produced two variants of PPR2 and TrpRS2. In these variants the N-terminal 45 amino acids, which include the presequence, were swapped between the two proteins, resulting in two variants termed prePPR2-TrpRS2 and preTrpRS2-PPR2, respectively. Figure 7 shows that import of TrpRS2 indeed becomes dependent on pATOM36 when its 45 N-terminal amino acids are replaced by the equivalent region of PPR2. However, the converse experiment shows that the 45 N-terminal amino acids of TrpRS2 are not sufficient to make import of PPR2 independent of pATOM36. The two columns on the right of Figure 7 show that both fusion proteins, as expected, still required ATOM and TIM17 for import.

These results show that the interaction of precursor proteins with pATOM36 is of complex nature. Apparently pATOM36 recognizes a signal in the N-terminal 45 amino acids of PPR2, a region that includes the presequence. However, the fact that the N-terminal 45 amino acids of TrpRS2 are not sufficient to abolish the pATOM36 dependence of import indicates that other regions of the protein are also involved in the recognition process. A comparison of the N-terminal regions of TrpRS2 and AspRS2—two substrates that are independent of pATOM36—with the corresponding region of all other tested substrates did not reveal obvious differences.

DISCUSSION

The capability to import proteins from the cytosol is one of the key features that distinguish mitochondrion from the bacterial endosymbiont from which they derive. It allowed the organelle to use proteins whose genes had been transferred to the nucleus, and current models for mitochondrial evolution envisage that the mitochondrial protein import system was established at a very early stage during mitochondrial evolution (Szklarczyk and Huynen, 2010). Consistent with this notion, the core components of the four hetero-oligomeric protein complexes that mediate protein import have been conserved in essentially all eukaryotes (Dolezal et al., 2006; Lithgow and Schneider, 2010; Hewitt et al., 2011). Given that trypanosomatids have a fundamentally different outer membrane preprotein translo- case, termed ATOM (Schneider et al., 2008; Pusnik et al., 2011), we anticipated that 1) other outer membrane proteins would serve as partners to assist ATOM function and 2) these partner proteins would not necessarily be homologous to the TOM subunits of other eukaryotes.

We have now characterized pATOM36, another novel integral outer membrane protein that is required for mitochondrial protein import in T. brucei. Like ATOM, pATOM36 is essential for growth and viability and appears to be restricted to the trypanosomatids.

To allow detection of pATOM36 on immunoblots, most of our biochemical experiments shown in Figures 1–3 relied on a cell line in which one allele of pATOM36 was marked with an HA tag. Because it is possible that the tag alters the properties of pATOM36 we produced a cell line in which also the second allele is HA tagged (Supplemental Figure S3). The resulting cells grew as well as wild-type cells, indicating that C-terminally tagged pATOM36 is functional.

The component parts of the TOM complex

Besides Tom40, the canonical TOM consists of a number of functionally specialized subunits. The small α-helical proteins Tom5, Tom6, and Tom7 are tightly associated with Tom40 and help to stabilize the import pore. In addition, Tom20 and Tom70 function as import receptors, and both of these proteins contain α-helical domains characterized by tetratricopeptide repeat motifs (Endo et al., 2011). In yeast, neither Tom20 nor Tom70 is essential for growth, indicating that they have partly redundant functions: deletion of both Tom20 and Tom70 is lethal (Lithgow et al., 1994). Tom22 functions as both a protein import receptor that cooperates with Tom20 and as a general organizer of the TOM complex. Lack of Tom22 therefore severely interferes with normal growth (van Wilpe et al., 1999).

The import receptor Tom20 is a relatively recent addition to the TOM complex, with the protein found in yeast having homologues in all fungi and animals but not in other organisms (Likić et al., 2005; Dolezal et al., 2006). A quite distinct protein, also called Tom20, is found in the TOM complex in all plants and green algae (Perry et al., 2006). These proteins share no common ancestor, and yet each is
composed of multiple α-helices: seven including the transmembrane domain for the Tom20 from animals and eight including the transmembrane domain for the Tom20 from plants (Abe et al., 2000; Perry et al., 2006). Both the animal Tom20 and the plant Tom20 are responsible for binding the mitochondrial-targeting sequence of substrate mitochondrial precursor proteins. In an evolutionary sense, this “prefilter” that binds mitochondrial-targeting sequences before the core TOM complex was a late addition, evolving after the divergence of the plant and animal lineages (Macasev et al., 2004; Perry et al., 2006). Independently, an equivalent, nonhomologous α-helical architecture evolved in these protein import receptors.

pATOM36 promotes the import of a subset of mitochondrial proteins

Although there is no sequence similarity between pATOM36 and the canonical import receptors Tom20 and Tom70, there are a number of shared features. A fraction of pATOM36 is associated with the ATOM complex (Figure 3)—the outer membrane protein translocase of trypanosomes—just as Tom20 and Tom70 are peripheral components of the canonical TOM complex (Meisinger et al., 2001). Antibody shift experiments (Figure 1E) indicate that pATOM36, similar to Tom20 and Tom70 (Neupert and Herrmann, 2007; Endo et al., 2011), appears to be anchored with its C-terminal part exposed to the cytosol. Finally, and most important, functional analyses show that each of the three proteins is required for import of a subset of but not all proteins into mitochondria.

Whereas the Tom20 import receptor binds simply to the mitochondrial targeting sequence, the Tom70 import receptor binds to multiple regions of its substrate proteins, whether or not they have mitochondrial targeting sequences (Brix et al., 2000; Chan et al., 2006; Yamamoto et al., 2009). This division of labor between the receptors provides a sophisticated means of recognizing the broad diversity of sequence characteristics in the ∼1000 proteins imported into mitochondria. What features in the substrate proteins determine the pATOM36 dependence of import is unknown, but the in vivo results shown in Figure 7 suggest that the N-terminal 45 amino acids contribute to it.

Blue native gel electrophoresis shows that the ATOM complex is ∼700 kDa (Pusnik et al., 2011), suggesting a hetero-oligomeric structure. Because there is a group of matrix proteins whose import does not depend on pATOM36, we hypothesize that additional protein import receptors are likely to exist.
Antibody shift assays

Mitochondria containing either HA-tagged pATOM36 or SAM50, respectively, were isolated under isotonic conditions (Schneider et al., 2007b). For each type of mitochondria two types of experiments were performed. In the first one intact mitochondria were resuspended in import buffer (Hauser et al., 1996) containing 5 mg/ml fatty acid–free bovine serum albumin and 1 μl of anti-HA polyclonal antibody (LabForce, Nunningen, Switzerland) was added. The reaction was incubated for 15 min on ice, and the mitochondria were reisolated. The supernatant was discarded, and the pellet was resuspended in 50 μl of 1.5% digitonin (Sigma-Aldrich, St. Louis, MO; Pusnik et al., 2011) and resolved by 6–16.5% BN-PAGE. In the second case the mitochondrial samples were first solubilized in import buffer containing 1.5% digitonin before it was incubated with 1 μl of the anti-HA polyclonal antibody for 15 min on ice and resolved on the same BN gel. The gel was then transferred to polyvinylidene fluoride membrane (Millipore, Billerica, MA) and analyzed by immunoblotting using monoclonal anti-HA antiserum (Santa Cruz Biotechnology, Santa Cruz, CA).

Immunoprecipitations

Isotonicily isolated mitochondria (240 μg; Hauser et al., 1996) containing C-terminally HA-tagged pATOM36 were solubilized for 15 min on ice in 300 μl of lysis buffer: 20 mM Tris-HCl, pH 7.4, 0.1 mM EDTA, 200 mM KCl containing 0.15% (wt/vol) digitonin, and a protease inhibitor cocktail (Roche Applied Science, Indianapolis, IN). The lysate was cleared by centrifugation (16,000 × g, 4°C), and the supernatant was incubated for 2.5 h at 4°C with 75 μl of a 1:1 slurry of anti-HA agarose (Roche Applied Science). Subsequently, the beads were washed three times in 750 μl of lysis buffer containing 0.1% (wt/vol) digitonin before elution with 60 μl of SDS sample.
buffer. For mass spectrometric analysis (Figure 3A) the samples were precipitated using acetone and tetracyclly digested in 60% (vol/vol) methanol and 20 mM NH₄HCO₃. The resulting peptides were analyzed by nano high-performance liquid chromatography–electrospray tandem mass spectrometry as described (Gebert et al., 2011). For peptide and protein identification, data sets were correlated with the TriTryp Database 3.1 using MaxQuant, version 1.0.13.13 (Cox and Mann, 2008). A false discovery rate of <1% was applied, and the number of evidences given by MaxQuant was used as a quantitative measure.

For immunoblot analysis cell lysates corresponding to 4 μg and eluate corresponding to 76 μg of mitochondria were separated on SDS polyacylamide gels, transferred to nitrocellulose, and analyzed with the indicated antisera (Figure 3B).

In vitro import and assembly of precursor proteins

The 35S-labeled precursor proteins were produced using the in vitro translation kit TT7 Quick for PCR DNA from Promega (Madison, WI). Import of precursor proteins into isotonically isolated mitochondria of T. brucei was done as described (Hauser et al., 1996), except that the import buffer contained 5 mg/ml fatty acid-free bovine serum albumin. In vitro assembly of 35S-labeled pATOM36 into complexes and subsequent analysis on BN-PAGE were done as described for yeast, except that 1.5% (wt/vol) of digitonin was used for solubilization (Stojanovski et al., 2007).

MISCELLANEOUS

Digitonin extractions to produce a crude mitochondrial fraction and ATP production assays were done as described (Schneider et al., 2007a).

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