# Environmental Risk of Pesticides for Fish in Small- and Medium-Sized Streams of Switzerland 

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#### Abstract

This study assessed the acute and chronic risk of pesticides, singly and as mixtures, for fish using comprehensive chemical data of four monitoring studies conducted in small- and medium-sized streams of Switzerland between 2012 and 2018. Pesticides were ranked based on single substance risk quotients and relative contribution to mixture risk. Concentrations of the pyrethroid insecticides, $\lambda$-cyhalothrin, cypermethrin and deltamethrin, and the fungicides, carbendazim and fenpropimorph, posed acute or chronic single substance risks. Risk quotients of eighteen additional pesticides were equal or greater than 0.1 , and thirteen of those contributed $\geq 30 \%$ to mixture risk. Relatively few substances dominated the mixture risk in most water samples, with chronic and acute maximum cumulative ratios never exceeding 5 and 7, respectively. A literature review of toxicity data showed that concentrations of several pesticides detected in Swiss streams were sufficient to cause direct sublethal effects on fish in laboratory studies. Based on the results of our study, we conclude that pesticides detected in Swiss streams, especially pyrethroid insecticides, fungicides and pesticide mixtures, pose a risk to fish health and can cause direct sublethal effects at environmental concentrations. Sensitive life stages of species with highly specialized life history traits may be particularly vulnerable; however, the lack of toxicity data for non-model species currently prevents a conclusive assessment across species.


Keywords: environmental risk assessment; insecticides; fungicides; herbicides; sublethal toxicity; resident fish species

## 1. Introduction

Contamination of surface water bodies with pesticides has been reported around the world [1,2] and attributed primarily to agricultural and, to a lesser extent, urban sources [3]. In Switzerland, several long-term monitoring studies conducted between 2012 and 2018 detected high numbers and concentrations of pesticides in small- and medium-sized streams predominantly flowing through agricultural land [4-6]. Between 100 and 145 pesticides were detected (corresponding to $40-67 \%$ of analytes), with $19-32$ substances exceeding acute or chronic risk thresholds. On average, samples contained mixtures of $30-50$ pesticides. In addition, analyses of pyrethroid insecticides (previously not analyzed) in 2017 and 2018 showed that these substances were also present at concentrations exceeding acute and chronic risk thresholds for aquatic organisms $[7,8]$. Most at risk were aquatic plants and invertebrates, and at times adverse effects on aquatic vertebrates, which includes fish and amphibians, could not be excluded [9,10]. Uncertainties with respect to the risk for fish remain, both with respect to the identification of the most fish-relevant pesticides, mixture risk effects and the potential for toxic effects on fish at environmental concentrations.

Regulatory environmental risk assessment of chemical water contaminants is performed by comparing exposure concentrations (predicted or measured) to predicted no effect concentrations (PNECs) in prospective risk assessment, or environmental quality criteria/standards in retrospective risk assessment. Both types of threshold values are derived by the same method using reliable and relevant ecotoxicity data (i.e., apical effects such as mortality, reduced growth and reproduction), which can be linked to population level effects [11]. Toxicity data for the most sensitive species, typically determined in standard toxicity tests, are identified and subsequently divided by an assessment factor (AF) to determine the risk threshold of each substance. The resulting risk assessment is considered to be protective of all species. In general, invertebrate or plant species are more sensitive to pesticides than fish. However, there are certain pesticides that are toxic to fish at equally or lower concentrations than other organism groups, e.g., ergosterole inhibiting fungicides [12,13].

The standard risk assessment approach largely ignores non-apical sublethal as well as indirect and multiple stressor effects of pesticides despite the fact that environmental concentrations of pesticides are often below lethal concentrations. Several reviews describe that pesticide concentrations clearly below LC50 levels can disrupt physiological, behavioral and hematological parameters in fish, such as anti-oxidant defense systems or energy metabolism. In addition, they can interfere with the neurological and immune systems and cause histopathological changes in barrier organs such as the intestine and gills and in internal organs such as liver and kidney [14-19]. Similarly, mixture effects are rarely taken into account despite their importance under environmental conditions [20-22]. Finally, adverse effects of pesticides to fish may arise not only from direct toxic effects but also through density- and trait-mediated effects [23,24]. Therefore, the risk of pesticides to aquatic organisms in the field may be severely underestimated [25]. Unexpected sublethal effects and the cumulative effects of contaminant mixtures and multiple stressors may, thus, reduce individual fitness and negatively affect populations of non-target species, potentially disrupting complex ecological systems (e.g., [26,27]).

This study focused on assessing the risk of pesticides for fish based on monitoring data from Switzerland, where $75 \%$ of the 55 indigenous fish species known are currently listed as regionally extinct, critically endangered, endangered, vulnerable or near threatened [28]. Among others, physical habitat changes, diseases as well as exposure to chemical contaminants have been named as potential causes [26]. Fish have complex life cycles and behaviors, can live for multiple years, and many species reproduce only once a year. Such traits may render them more vulnerable to chronic and sublethal toxic effects than the fractional spawners, which are typically used in laboratory toxicity tests, especially if exposure to stressors coincides with the presence of sensitive life stages. To gain a better understanding of the vulnerability of indigenous fish to pesticides detected in Swiss surface waters, (i) fish-specific risk thresholds were derived for pesticides detected in Swiss monitoring campaigns, (ii) pesticide concentrations measured in several recent monitoring studies were evaluated using a standard environmental risk assessment (ERA) to identify pesticides of highest concern for fish by ranking them according to single substance risk quotients and their relative contribution to mixture risk, and (iii) available non-apical effect data on fish-relevant pesticides were reviewed and discussed. In addition, we attempted to identify indigenous fish species potentially vulnerable to pesticides based on an established vulnerability concept.

## 2. Materials and Methods

### 2.1. Analytical Data and Sampling Design

Monitoring data on pesticide concentrations in Swiss streams for the years 2012, 2015, 2017 and 2018 are publicly available (SI, Table S1). Detailed information on sampling sites, water sampling and analytical methods is provided by [4-6] and for pyrethroids by $[7,8]$. In 2012, sampling sites were located in five medium-sized streams with catchment areas of $38-105 \mathrm{~km}^{2}$ comprising agricultural as well as urban areas. In 2015 and 2017, sampling
sites were located in five small streams with catchment areas of $1.6-9$ and $0.9-6.7 \mathrm{~km}^{2}$, respectively, dominated by agricultural land use. In 2017, pyrethroid insecticides were analyzed at one site only (Chrümlisbach, Canton Bern). In 2018, six sites in streams with catchment areas of $1.4-34 \mathrm{~km}^{2}$ were sampled and analyzed exclusively for 23 pyrethroids and the organophosphates (OP), chlorpyrifos and chlorpyrifos-methyl. At two sites (Eschelisbach, Canton Thurgau; Weierbach, Canton Basel-Land), samples were collected in both 2015 and 2017; Chrümlisbach (Canton Bern) and Bainoz (Canton Fribourg) were sampled in both 2017 and 2018.

Sampling designs of the four studies varied slightly. In 2012, 14-day time proportional (TP) composite samples were collected March to July and analyzed for 249 agricultural pesticides. In 2015, 12 h TP composite samples were collected March to August and analyzed for 213 agricultural pesticides. During dry periods between discharge events, 12 h samples were pooled corresponding to the length of the low flow period ( 5 d on average) and then analyzed. In 2017, 3.5-day TP composite samples were collected March to October and analyzed for 217 pesticides. In 2018, only 14-day TP composite samples were collected March to September, and chemical analysis focused on pyrethroids and OPs.

### 2.2. Risk Assessment—Single Substances

Fish-specific acute and chronic predicted no effect concentrations ( $_{\text {PNEC }}^{i, f i s h, a c u t e, ~}$ $\mathrm{PNEC}_{i, \text { fish,chronic }}$ ) were derived for each pesticide detected and registered for use in Switzerland at the time of sampling, if sufficient toxicity data were available. Data were obtained from existing data collections of the Swiss Centre for Applied Ecotoxicology (Dübendorf, Switzerland; www.ecotoxcentre.ch), already evaluated for reliability and relevance according to CRED [11]. Additional data were compiled from the literature, primarily from approval reports of the European Food Safety Authority (EFSA, Parma, Italy; https:/ /www. efsa.europa.eu), biocide assessment reports of the European Chemicals Agency (ECHA, Helsinki, Finland; https:/ /echa.europa.eu/de/information-on-chemicals/), draft PNEC dossiers of the European Commission (EC, Ispra, Italy; https: / /ec.europa.eu/food/plant/ pesticides/) and the US EPA OPP Pesticide Ecotoxicity Database (https:/ /ecotox.ipmcenters. org/index.cfm?menuid=7) (SI, Tables S2 and S3). For the derivation of PNEC ${ }_{\mathrm{i}, \text { fish,acute }}$, the concentrations that caused $50 \%$ mortality $\left(\mathrm{LC}_{50}\right)$ within 96 h in the most sensitive fish species were divided by an assessment factor (AF) of 10. For the derivation of PNEC $_{i}$,fish,chronic, , concentrations that caused sublethal toxicity within $\geq 21 \mathrm{~d}$ (in most cases) the no observed effect concentration (NOEC) of the most sensitive fish species was divided by an AF of 10 . This AF was chosen based on the European Union Guideline for the Derivation of Environmental Quality Standards (EQS) [29], where 10 is used as AF when acute and chronic toxicity data for all trophic levels, including the most sensitive, are available. If chronic data are missing, higher AF are used to extrapolate from acute to chronic toxicity. Because only fish data were used in this study and acute as well as chronic data were available, an AF of 10 was considered appropriate.

Fish-specific single substance risk quotients $\left(\mathrm{RQ}_{\mathrm{i}, \text { acute }}\right.$ and $\left.R \mathrm{Q}_{\mathrm{i}, \text { chronic }}\right)$ were calculated by dividing the measured environmental concentrations (MEC) of a substance by the respective $\mathrm{PNEC}_{\text {fish }}$. If $R Q \geq 1$, adverse acute or chronic effects of this substance on fish cannot be excluded. For the calculation of $\mathrm{RQ}_{\mathrm{i}, \mathrm{acute}}, \mathrm{PNEC}_{\mathrm{i}, \text { fish,acute }}$ were compared to concentrations measured in acute exposure scenarios. Because sampling designs differed between studies, we used MECs of the smallest available collection period to determine acute risk. In 2015, collection periods for TP composite samples ranged from 0.5-day (wet periods) to 24-day TP composite samples (dry periods with assumed absence of exposure peaks). In this case, the PNEC $_{i}$,fish,acute was compared to the MEC of each sample analyzed irrespective of the collection period. For 2017 data, $\mathrm{RQ}_{\mathrm{i}, \text { acute }}$ were determined using MEC of 3.5-day TP composite samples based on [30]. For 2015 and 2017 data, we used 14-day time-weighted (TW) average MEC for the calculation of $\mathrm{RQ}_{\mathrm{i}, \text { chronic }}$. Acute risk was not determined for data generated in 2012 and 2018 when only 14-day TP composite samples were collected. For these samples, only the $\mathrm{RQ}_{\mathrm{i}, \text { chronic }}$ was calculated.

### 2.3. Risk Assessment-Mixtures

Mixture risk quotients $\left(\mathrm{RQ}_{\text {mix,acute }}\right.$ and $\left.\mathrm{RQ}_{\text {mix,chronic }}\right)$ were calculated by adding the $\mathrm{RQ}_{\mathrm{i}}$ of all substances detected in the same water sample following current mixture risk assessment approaches [31]. Values below the limit of quantification (LOQ) were substituted with zero. If $\mathrm{RQ}_{\text {mix }} \geq 1$, adverse effects of the pesticide mixture on fish cannot be excluded.

### 2.4. Identification of Fish-Relevant Pesticides

In order to identify the most relevant pesticides to fish in Switzerland, results of the single substance and mixture risk assessments were used to rank individual pesticides. The primary selection criterion for relevance was $R Q_{i} \geq 0.1$. It was chosen because fewer than 10 substances dominated the acute mixture risk based on $\mathrm{PNEC}_{\mathrm{i}, \mathrm{fish}, \text { acute }}$, even though the median number of substances present per water sample ranged from 23 to 42 [6]. If $\mathrm{RQ}_{\mathrm{i}}$ $\geq 0.1$ in at least in one water sample and the contribution to $R Q_{\text {mix }}$ of $\geq 30 \%$ in the same sample, a substance was classified as highly relevant. The threshold of $\geq 30 \%$ contribution to mixture risk was considered representative based on the analysis of maximum cumulative ratios $\left(\mathrm{MCR}=\sum \mathrm{RQ}_{\mathrm{i}} / \mathrm{RQ}_{\mathrm{i}, \max }\right)$, which indicates how many substances are drivers of the mixture risk [32]. In our dataset, the MCR ranged from 1-7 (acute) and 1-5 (chronic) (SI, Figure S1). To identify the most relevant pesticides, we chose a conservative MCR of 3 (equivalent to ca. $30 \%$ contribution per substance) as our secondary relevance criterion. These criteria were also applied to 2018 data, although analytes comprised only a limited set of insecticides, 23 pyrethroids and 2 OPs, chlorpyrifos and chlorpyrifos-methyl.

### 2.5. Review of Non-Apical Sublethal Effect Data

Publicly available data on sublethal effects were reviewed for fish-relevant substances during the summer of 2018 using Web of Science and Google Scholar with the following search terms: [compound], [endpoint or effect] and [fish].

### 2.6. Vulnerability of Indigenous Fish Species

Our approach to assess the vulnerability of fish species indigenous to Switzerland was based on the principle of vulnerability conceptualized by [33] and applied by [34]. First, we identified species particularly at risk of exposure to pesticides ("external exposure"). The occurrence of fish in habitats most likely to contain pesticides at concentrations of concern was determined with the help of Swiss fish monitoring data recorded by the National Surface Water Quality Observation program (NAWA TREND) [35]. Subsequently, we evaluated the "intrinsic species sensitivity" by ranking available acute toxicity data ( $\mathrm{LC}_{50}$ ) for different fish species using species sensitivity distributions (SSDs), if reliable data were available for at least 5 species [36]. This was the case for the fungicides azoxystrobin and carbendazim, the insecticides chlorpyrifos, diazinon, cypermethrin and $\lambda$-cyhalothrin, as well as the herbicides linuron and diuron. SSD graphs were produced using R 3.5.3 [37].

## 3. Results and Discussion

### 3.1. Single Substance Risk

The fish-specific risk assessment revealed a total of 23 relevant pesticides with $\mathrm{RQi} \geq 0.1$. Table 1 provides a summary of results on these pesticides and their risk levels in different monitoring years, as well as the number of sites and days (across monitoring years) where exceedances occurred. All but five of these pesticides were classified as highly relevant, i.e., they contributed at least $30 \%$ to the mixture risk of the respective sample. Detailed information is shown in SI Table S4.

Concentrations of five pesticides ( $\lambda$-cyhalothrin, cypermethrin, deltamethrin, carbendazim, fenpropimorph) exceeded their respective PNEC (i.e., $\mathrm{RQ}_{\mathrm{i}} \geq 1$ ) at least once and, thus, posed a risk as individual substances. Among insecticides, the pyrethroid $\lambda$-cyhalothrin posed the highest risk in 2017 with an $\mathrm{RQ}_{\mathrm{i}} \geq 1$ for both acute and chronic exposures. In 2018 (when data were limited to 14-day average concentrations and, thus, only chronic risk was assessed), two other pyrethroids, cypermethrin and deltamethrin, were detected at concentrations exceeding the chronic risk threshold $\left(R Q_{i, c h r o n i c} \geq 1\right)$. However, the 14-day MEC of deltamethrin ( $77 \mathrm{ng} / \mathrm{L}$ ) not only exceeded the $\mathrm{PNEC}_{\mathrm{i}, \text { fish,chronic }}$ by a factor of 45 (SI, Table S4), but also its acute risk threshold $\left(\mathrm{PNEC}_{\mathrm{i}, \mathrm{fish}, \mathrm{acute}}=15 \mathrm{ng} / \mathrm{L}\right)$ by a factor of 5 (SI, Table S4). In long-term laboratory studies, the 14-day MEC of this substance caused apical sublethal effects in fish, such as reduced growth (SI, Table S6). Based on runoff models for pyrethroids [38-40], it is possible, however, that one or more short ( $<1$ day) pulses of very high deltamethrin concentrations occurred during the sampling period in 2018. Such brief peak concentrations of pesticides may greatly exceed concentrations detected in 3.5-day composite samples [41]. It is, therefore, possible that deltamethrin exceeded acute effect concentrations for fish, which demonstrates the severe environmental risk this substance might pose.

Besides the three pyrethroids, two fungicides, carbendazim and fenpropimorph, posed single substance risks in 2017 (carbendazim: acute and chronic, fenpropimorph: chronic). Although carbendazim was already banned in Switzerland in 2016, it was permitted for use until 2018, which explains its presence in water samples. Both fungicides were also detected at lower concentrations in previous monitoring studies (fenpropimorph in 2012, carbendazim in 2015) at multiple sites. In June 2017, tissue samples of fish (Leuciscus cephalus, a European cyprinid species) collected from the river Urtenen (BE) contained fenpropimorph ( $c_{\max }=20 \mu \mathrm{~g} / \mathrm{kg}$; Tamara Diethelm, WWF Schweiz, Zurich, Switzerland; personal communication). The river Urtenen is geographically close to Chrümlisbach, the monitoring site where high water concentrations of this compound were detected in 2017. Overall, the pattern seen suggests that water contamination by these two fungicides was widespread.

Another eighteen pesticides, among them four insecticides, nine fungicides and five herbicides, exceeded the fish-specific $\mathrm{RQ}_{\mathrm{i}}$ (chronic or acute) of 0.1 , most of them with regard to chronic risk. All insecticides were categorized as highly relevant. The OP insecticide, chlorpyrifos, which was not measured in 2012, exceeded both the acute and chronic $\mathrm{RQ}_{\mathrm{i}}$ of 0.1 in 2015 and 2017 and the chronic $\mathrm{RQ}_{\mathrm{i}}$ of 0.1 in 2018 at multiple sampling sites. As of May 2021, both chlorpyrifos and chlorpyrifos-methyl are no longer authorized for use in the EU and Switzerland (https:/ /eur-lex.europa.eu/legal-content/EN/TXT/PDF/; https: / /www.psm.admin.ch/de/wirkstoffe/bs/C, both accessed on 28 January 2021). Another OP insecticide, diazinon, banned in Switzerland since 2011 [42], had a chronic $\mathrm{RQ}_{\mathrm{i}} \geq 0.1$ in 2012 and 2015, but not anymore in 2017. Among five herbicides categorized as relevant based on chronic risk, four were highly relevant. S-metolachlor was detected at $R Q_{i} \geq 0.1$ during a considerable part of the monitoring period, for 126 days in 2012 and 56 days in 2015, but not in 2017. Linuron was detected at chronic $R Q_{i} \geq 0.1$ in all three years monitored, while diuron and aclonifen were detected at this level in 2015 only.

Table 1. Pesticides posing (A) a single substance risk of $R Q_{i} \geq 1$ (acute or chronic); (B) a single substance risk of $R Q_{i} \geq 0.1$ and $<1$ (acute or chronic) in at least one water sample based on monitoring data collected in Switzerland (2012-2018). Substances marked with * contributed at least $30 \%$ to mixture risk in the respective sample. Total days of exceedance per monitoring year and number of sites are provided in brackets. IN: insecticide; FU: fungicide; HE: herbicide; -: $\mathrm{RQ}_{\mathrm{i}}<0.1$; NA: substance was not analyzed.

| (A) <br> Substances with $\mathrm{RQ}_{\mathrm{i}} \geq \mathbf{1}$ | Total Days of Exceedance (Number of Sites Affected) |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $R Q_{i}$ | ${ }_{\text {Chronic }}{ }^{1} 2012$ | 2015 |  | $2017{ }^{\text {a }}$ |  | $2018^{b}$ <br> Chronic |
|  |  |  | Acute ${ }^{2}$ | Chronic | Acute | Chronic |  |
| $\lambda$-Cyhalothrin * (IN) | $\geq 0.1$ | NA | NA | NA | 10.5 (1) | 28 (1) | 42 (2) |
|  | $\geq 1.0$ |  |  |  | 3.5 (1) | 14 (1) |  |
| Cypermethrin * (IN) | $\geq 0.1$ | NA | NA | NA | - | 14 (1) | 42 (1) |
|  | $\geq 1.0$ |  |  |  |  |  | 14 (1) |
| Deltamethrin * (IN) | $\geq 0.1$ | NA | NA | NA | - | - | 28 (1) |
|  | $\geq 1.0$ |  |  |  |  |  | 14 (1) |
| Carbendazim * (FU) | $\geq 0.1$ | - | 62(4) | 40.5 (1) | 77 (3) | 66.5 (3) | NA |
|  | $\geq 1.0$ |  |  |  | 3.5 (1) |  |  |
| Fenpropimorph * (FU) | $\geq 0.1$ | 53 (2) | - | - | - | 28 (1) | NA |
|  | $\geq 1.0$ |  |  |  |  | 14 (1) |  |

(B)

Substances with
$\mathrm{RQ}_{\mathrm{i}} \geq 0.1$ and $<1$

| Chlorpyrifos * (IN) | $\geq 0.1$ | NA | $11(3)$ | $70(3)$ | $3.5(1)$ | $63(3)^{\text {c }}$ | $84(3)$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Chlorpyrifos-methyl * (IN) | $\geq 0.1$ | NA | $0.5(1)$ | - | - | - | - |
| Diazinon * (IN) | $\geq 0.1$ | $70(2)$ | - | $28(1)$ | - | - | NA |
| Permethrin * (IN) | $\geq 0.1$ | NA | NA | NA | - | - | N (1) |
| S-Metolachlor * (HE) | $\geq 0.1$ | $126(3)$ | - | $56(1)$ | - | NA |  |
| Linuron * (HE) | $\geq 0.1$ | $14(1)$ | - | $79.5(3)$ | - | $24.5(1)$ | NA |
| Diuron (HE) | $\geq 0.1$ | - | - | $69.5(1)$ | - | NA | NA |
| Aclonifen (HE) | $\geq 0.1$ | - | - | $14(1)$ | NA | NA | NA |
| Pendimethalin * (HE) | $\geq 0.1$ | NA | NA | NA | - | $14(1)$ | NA |
| Epoxiconazole (FU) | $\geq 0.1$ | $42(2)$ | - | $54.5(1)$ | - | $52.5(2)$ | NA |
| Tebuconazole (FU) | $\geq 0.1$ | - | - | - | - | $14(1)$ | NA |
| Fluopyram * (FU) | $\geq 0.1$ | NA | - | $40(1)$ | - | $14(1)$ | NA |
| Flusilazole (FU) | $\geq 0.1$ | - | - | $26.5(1)$ | - | NA |  |
| Pyraclostrobin * (FU) | $\geq 0.1$ | $28(1)$ | - | - | - | NA |  |
| Spiroxamine * (FU) | $\geq 0.1$ | - | - | $28(1)$ | - | - | NA |
| Azoxystrobin (FU) | $\geq 0.1$ | - | - | - | - | $14(1)$ | NA |
| Fluazinam * (FU) | $\geq 0.1$ | - | - | - | - | $14(1)$ | NA |
| Fludioxonil (FU) | $\geq 0.1$ | - | - | - | - | NA |  |

${ }^{1}$ Chronic refers to time-proportional two-week composite samples $(2012,2018)$ and time-weighted averages over 14 day $(2015,2017)$, respectively; ${ }^{2}$ acute refers to: 0.5 -day composite samples (up to 24 -day during dry periods, 2015) and 3.5-day composite samples (2017); ${ }^{\text {a }}$ pyrethroids were analyzed at only 1 of 5 sites in 2017; ${ }^{\text {b }}$ monitoring in 2018 was focused exclusively on 23 pyrethroid insecticides and chlorpyrifos/-methyl, ${ }^{c}$ samples collected from one site (Chrümlisbach, BE) in 2017 were additionally analyzed for chlorpyrifos with a lower LOQ as part of a special study; the period of exceedance based on that dataset was 91 days.

The risk of different pesticides determined for fish in Swiss streams varied from year to year, which is primarily a consequence of differences in application rates, weather conditions and changes in substance authorization status. Another important factor is the analytical capability and capacity. Monitoring and risk assessment results of pyrethroid insecticides in 2017 and 2018 highlight the importance of reducing so-called "blind spots" in the analytical data. In order to monitor this group of extremely toxic, hydrophobic insecticides, it is necessary to have analytical methods with limits of quantification (LOQ) in the picogram per liter range $[7,33]$. Once pyrethroid data became available in Switzerland, four pyrethroid insecticides were categorized as highly fish-relevant contaminants in our study, and all but one (permethrin) posed a single substance risk.

### 3.2. Mixture Risk

Pesticide mixtures presented a chronic or acute risk for fish at one or more sampling sites in each monitoring year, primarily in late spring and late summer (Figures 1 and 2; SI Table S4). The acute mixture risk level for fish was exceeded on two occasions in 2017 (Figure 1). The peak in May was driven by the pyrethroid $\lambda$-cyhalothrin ( $>99 \%$, in green), while carbendazim was the main driver (99\%) in mid-July 2017.


Figure 1. Acute mixture risk quotients $\left(\mathrm{RQ}_{\text {mix, acute }}\right)$ for fish in composite water samples from small streams in Switzerland in 2015 and 2017. Two thousand and fifteen: acute exposure scenario (ExpSc) based on 0.5-24 day time-proportional (TP) composite samples; 2017: acute ExpSc based on 3.5 day TP composite samples; pyrethroids were analyzed at only 1 of 5 sites (Chrümlisbach, $B E$ ). $R_{\text {mix }}$ (Chrümlisbach, $B E$, pyrethroids) is based solely on data for pyrethroids. Gaps indicate periods when no samples were collected; the dashed line indicates $R Q=1$, and the dotted line indicates $R Q=0.1$.


Figure 2. Cont.


Figure 2. Chronic mixture risk quotient $\left(\mathrm{RQ}_{\mathrm{mix}, \text { chronic }}\right)$ for fish in water samples from small- and mid-size streams in Switzerland in 2012-2018. Two thousand and twelve and 2018: chronic ExpSc based on 14 day TP composite samples (except 16-27 April 2012: 11 day); in 2017 (Chrümlisbach, BE Pyrethroids) and 2018, $\mathrm{RQ}_{\text {mix }}$ is based solely on data for pyrethroids and chlorpyrifos/-methyl. Two thousand and fifteen and 2017: chronic ExpSc based on 14 day time-weighted average concentrations. Gaps indicate periods when no samples were taken; the dashed line indicates $R Q=1$, and the dotted line indicates $R Q=0.1$.

Relatively few substances dominated the mixture risk in most water samples, with the chronic and acute MCRs never exceeding 5 and 7, respectively (SI, Figure S1). The fungicide fenpropimorph was responsible for $84 \%$ of $\mathrm{RQ}_{\text {mix,chronic }}$ in April 2012 (Limpach, SO). The pesticides responsible for most of the chronic risk seen in June 2012 (Surb, AG) were the fungicides fenpropimorph (35\%), epoxiconazole ( $14 \%$ ) and flusilazole ( $10 \%$ ) and the herbicide S-metolachlor (17\%). The herbicides, S-metolachlor (67\%) and linuron (23\%), dominated the chronic mixture risk in late April 2015 (Weierbach, BL).

Analyses of water samples collected from Chrümlisbach (BE) in late April/early May 2017 (pyrethroids and chlorpyrifos/-methyl analyzed separately [6,7]) revealed that the pyrethroid $\lambda$-cyhalothrin $\left(91 \%\right.$ of $\left.\mathrm{RQ}_{\text {mix,chronic }}=2.7\right)$ and the fungicide fenpropimorph $\left(96 \%\right.$ of $\left.R Q_{\text {mix, chronic }}=13.1\right)$ were largely responsible for the high chronic risk. This was the only site where pyrethroid insecticides were analyzed in 2017. In 2018, monitoring of six sites was focused exclusively on pyrethroid insecticides, chlorpyrifos and chlorpyrifosmethyl. The chronic risk seen in late May/early June at Beggingerbach (SH) was driven almost entirely by cypermethrin ( $96 \%$ ). In late summer, a peak in deltamethrin concentrations (99.8\%) presented a very high risk to fish (acute and chronic) at Le Bainoz (FR). These results show that it is extremely important to monitor pyrethroid insecticides in surface waters. If not, the risk analysis would miss some of the most important environmental toxicants for fish as well as aquatic invertebrates.

### 3.3. Sublethal Effects

Major uncertainties exist in risk assessment with regard to the significance of the longterm, low-level chronic mixture risk ( $\mathrm{RQ} \geq 0.1$ and $<1$ ) as observed at most sampling sites in this study (Figure 2). Between early March and mid-October 2017, for example, chronic mixture risk was elevated for $>100$ days at four of five monitoring sites, interspersed by short periods of acute or chronic risk $\geq 1$. Information on the effects of such long-term exposures to pesticides is extremely scarce.

Sublethal effects can compromise individual fitness and ultimately lead to a decline of fish abundance [19,43]; however, conventional approaches to environmental risk assessment generally ignore data on non-apical sublethal effects. The risk to resident organisms exposed to low concentrations of pesticides for long periods of time may, therefore, be underestimated. Such effects include cellular effects resulting from the primary modes of action (MOA) of a pesticide or (often unknown) side effects. For pesticides categorized as highly fish-relevant in this study (marked * in Table 1), such primary MOA include the inhibition of cellular processes and components, i.e., sterole biosynthesis, respiration, inhibition of acetyl cholinesterase (AChE) and photosystem II, and the modulation of ion channels (SI, Table S5). Many of these cellular functions are conserved across organism groups; however, even herbicides and fungicides, which primarily target cellular mechanisms only present in plants or fungi, can affect fish via mechanism unrelated to their primary MOA [44]. Our analysis shows that neurotoxic insecticides, primarily pyrethroids, are of highest concern with regard to fish health. This finding corroborates conclusions of previous analyses [18,45,46]. Aside from affecting the nervous system of fish, pyrethroids have been shown to act as endocrine disruptors [47], genotoxicants [48] and immunotoxicants [49].

Figure 3 provides an overview of how MEC detected in Switzerland compared to regulatory toxicity thresholds $\left(\mathrm{LC}_{50}, \mathrm{NOEC}\right)$ as well as non-apical sublethal effect concentrations at or below the MEC of selected fish-relevant pesticides. While known $\mathrm{LC}_{50^{\prime}} \mathrm{s}$ for fish are generally well above MECs, maximum pesticide concentrations detected in Switzerland have been reported to cause non-apical sublethal effects in fish under laboratory conditions (SI Table S6 and references therein). It is important to note, however, that most reported chronic and sublethal effect data are derived from test exposures longer than 3.5 d. Nevertheless, sublethal effects can already occur after brief exposures to environmentally relevant pesticide concentrations (SI, Table S6 and references therein). For example, the MEC of $\lambda$-cyhalothrin ( $31 \mathrm{ng} / \mathrm{L}$ in a 3.5 d composite sample) detected in 2017 exceeded the concentration that caused DNA damage in mosquitofish (Gambusia affinis) exposed for 2 d [50]. Similarly, the organophosphate insecticides, chlorpyrifos and diazinon, affected cellular homeostasis, olfactory function and behavior of fish after a few days of exposure at MECs detected in Swiss creeks [51-54]. The herbicide, linuron, can induce stress in brown trout exposed for 4 d at environmental concentrations [55]. Brief ( 24 h ) exposure to a relatively low concentration ( $5 \mu \mathrm{~g} / \mathrm{L}$ ) of diuron negatively affected the grouping behavior of goldfish [56], a response important for predator avoidance. The fungicide, azoxystrobin, was shown to induce stress-responsive genes of zebrafish at or below concentrations detected in Swiss rivers within a few days [57]. Carbendazim affected locomotor behavior of zebrafish larvae after a 5 d exposure to $0.16 \mu \mathrm{~g} / \mathrm{L}$ [58]. Exposed larvae moved more slowly than the control group, suggesting that their feeding success and predator avoidance capability might be impacted. Such non-apical sublethal effects may compromise individual fitness and ultimately lead to a decline of fish abundance, for example, through disruption of olfactory and reproductive functions, as well as impairment of their swimming and feeding ability [19].


Figure 3. Comparison of $\mathrm{LC}_{50^{\prime}} \mathrm{s}$, no observed effect concentrations for apical sublethal endpoints (NOEC) and selected non-apical sublethal effect concentrations (sublethal) in red, with maximum environmental concentrations (MEC; in orange) measured in Switzerland in 2015 ( $0.5-24$ day TP composite samples, [5]), 2017 ( 3.5 day TP composite samples, [6,7]) and 2018 (14 day TP composite sample, [7]). Values < LOQ were substituted with zero; 2017 MEC for chlorpyrifos from [7]. References are provided in SI, Table S7.

### 3.4. Vulnerability of Resident Fish Species

Small- and medium-sized streams are important spawning and rearing habitats for fish. While the data on indigenous fish species and their distribution in Switzerland are still incomplete, 47 are currently known to be resident in Switzerland [28]. Of those, approximately 25 species inhabit small- and medium-sized streams either permanently, periodically or sporadically [35] and are, thus, potentially exposed to pesticides. This includes species that are potentially threatened (e.g., brown trout (Salmo trutta)) or endangered (e.g., lake trout (Salmo trutta)) (SI, Table S9). Compilation of toxicity data for several data-rich pesticides identified as fish-relevant in our study showed that several model species commonly used in standardized tests (Pimephales promelas, Oryzias latipes, Danio rerio) tend to have moderate to low sensitivity, while salmonids and non-model species tend to be more sensitive (up to $>100$-fold) to such compounds (SI, Figure S2).

Taxonomy, life history and life stage can modify fish responses to environmental stressors, and specialist species are less likely to show tolerance stressors than generalists [59-62]. Species that reproduce once a year, and whose early life stages (ELS) hatch and rear during a period when highest pesticide risk occurs (April-May and August-September, Figures 1 and 2) may be most vulnerable, because the population cannot compensate by reproducing during times when risk from pesticides is low [63-65]. Fish ELS are often most sensitive to chemicals [66]. In addition, larval and juvenile fish tend to be more specialized with regard to their prey $[67,68]$, which puts them at greater risk of indirect effects in case their invertebrate prey species are depleted due to pesticides [27,69]. Among such species are brown trout who spawn from November to January, and embryos develop slowly to hatch in spring [70,71], and spring spawners with short developmental periods, such as common nase (Chondrostoma nasus), the salmonid, grayling (Thymallus thymallus), as well as two endangered species, stroemer (Leuciscus souffia) and spirlin (Alburnoides bipunctatus). The results of our vulnerability analysis are, however, based on few data and must be considered descriptive.

### 3.5. Uncertainties and Limitations of Our Risk Assessment

Sampling designs, analytes and analytical methods, as well as availability of toxicological data are important variables influencing the outcome of the risk assessment. The AF applied to effect data when deriving risk thresholds is intended to account for uncertainties when extrapolating data from laboratory to field and from model to non-model species. However, the general lack of toxicological effect data for non-model species often makes it necessary to choose the AF without an appropriate knowledge base. For this study, we applied an AF of 10 based on existing guidelines; however, the assembled toxicity data show that inter-species sensitivity can vary by a factor of $>100$ (SI Figure S2, Table S8). Additional uncertainty arises from the exposure assessment. Analytical constraints, such as limits of quantification that are above effect concentrations, and the selection of analytes, which should be based on usage information and include toxic metabolites, as well as the sampling method and period are important factors to take into consideration. Recent studies by $[5,41]$ showed that the temporal resolution of sampling can significantly influence the results of a risk assessment. MEC detected in grab and composite samples collected over several days or weeks can greatly underestimate short-term peak concentrations of environmental pollutants, especially in small- to medium-sized streams where runoff and flow dynamics tend to be more extreme. This means that acute toxicity is more likely to occur than previously considered. Moreover, resident fish are commonly exposed to multiple stressors including elevated water temperature, changes in the food web and pathogens, which may modulate chemical toxicity $[26,27,46,59,72]$. The reliance of standard risk assessment approaches on analytical data from grab or composite samples, and on toxicity data derived from laboratory tests with model species and apical endpoints is, therefore, likely to result in an underestimation of the environmental risk of chemicals in Switzerland and elsewhere.

## 4. Conclusions

Our fish-specific risk analysis of pesticide data from monitoring studies conducted in Switzerland showed that pesticides, both as single substance and in mixture, posed a risk for fish at multiple sampling sites. Among hundreds of pesticides analyzed, only three pyrethroid insecticides and two fungicides exceeded fish-specific risk thresholds individually $\left(R_{i} \geq 1\right)$. Another eighteen pesticides were detected at lower risk concentrations $\left(R Q_{i} \geq 0.1\right)$. All but five of these relevant pesticides contributed at least $30 \%$ to acute or chronic mixture risk in the same water sample and were therefore categorized as highly relevant. Based on available information on sublethal toxicity of highly relevant pesticides, we conclude that fish health may be impaired at environmental concentrations. Sensitive fish species, whose early life stages are present when highest pesticide concentrations occur, are likely to be most vulnerable. However, significant knowledge gaps still exist with regard to the sensitivity of non-model indigenous species, the consequences of non-apical sublethal effects for ecological fitness, the effects of long-term, low-level exposure in combination with intermittent acutely toxic peak pesticide concentrations and the combined effects of multiple stressors on fish health. Current data suggest that such uncertainties are likely to result in an underestimation of environmental risk of pesticides on fish.

Comprehensive long-term monitoring programs together with focused laboratory studies are essential for a better understanding of the ecological risk and impact of chemical pollution. To achieve this, the selection of analytes for characterizing exposure must be checked and revised on a regular basis, as pesticides applied in agriculture change annually depending on weather conditions, crop and regulatory actions. Substances such as the highly relevant pyrethroid insecticides, may have been overlooked by many monitoring programs due to analytical constraints and inadequate limits of quantification. The increased use of bioassays to monitor biological effects rather than individual chemicals could circumvent some of these challenges. However, methods, both analytical and ecotoxicological, to detect chemicals and their effects must continue to be improved utilizing technological advances. Furthermore, methods to extrapolate genomic, ecological and
mechanistic information across species may strengthen our understanding of the intrinsic sensitivity of resident species. Eventually, this will lead to better prospective and retrospective risk assessment as well as management to reduce the impact of chemicals in the environment.

Supplementary Materials: The following are available online at https:/ / www.mdpi.com/article/ 10.3390 /toxics 9040079 /s1, Table S1: Sources of chemical-analytical data from monitoring studies conducted in Switzerland 2012-2018, Table S2: Acute effect values used in the fish-specific risk assessment (Excel-file: SI_T2), Table S3: Chronic effect values used in the fish-specific risk assessment (Excel-file: SI_T3), Table S4: Acute and chronic single and mixture risk quotients of pesticides detected in water samples of Swiss monitoring studies 2012-2018 (Excel-file: SI_T4 RQmix), Table S5: Effect values, MECs and mode of action of highly relevant substances, Table S6. Sublethal effect data for data-rich relevant pesticides, Table S7: Toxic effect data for data-rich relevant pesticides shown in Figure 3, Table S8: Acute toxicity data ( 96 h LC50) for data-rich relevant pesticides shown in Figure S2, Table S9: Fish species monitored in small- to medium-sized Swiss streams and their current status, Figure S1: Maximal cumulative ratios (MCR) as a function of the mixture risk quotient of pesticides detected in water samples from monitoring campaigns 2012-2018, Figure S2: Species sensitivity distributions (SSDs) of acute toxicity concentrations for data-rich relevant pesticides. Data for this study is available at https: / / doi.org /10.25678/00038Y.
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## References

1. Norman, J.E.; Mahler, B.J.; Nowell, L.H.; Van Metre, P.C.; Sandstrom, M.W.; Corbin, M.A.; Qian, Y.; Pankow, J.F.; Luo, W.; Fitzgerald, N.B.; et al. Daily stream samples reveal highly complex pesticide occurrence and potential toxicity to aquatic life. Sci. Total Environ. 2020, 715, 136795. [CrossRef]
2. Stehle, S.; Buba, S.; Schulz, R. Compilation and analysis of global surface water concentrations for individual insecticide compounds. Sci. Total Environ. 2018, 639, 516-525. [CrossRef] [PubMed]
3. Budd, R.; Wang, D.; Ensminger, M.; Phillips, B. An evaluation of temporal and spatial trends of pyrethroid concentrations in California surface waters. Sci. Total Environ. 2020, 718, 137402. [CrossRef] [PubMed]
4. Moschet, C.; Wittmer, I.; Simovic, J.; Junghans, M.; Piazzoli, A.; Singer, H.; Stamm, C.; Leu, C.; Hollender, J. How a Complete Pesticide Screening Changes the Assessment of Surface Water Quality. Environ. Sci. Technol. 2014, 48, 5423-5432. [CrossRef]
5. Spycher, S.; Mangold, S.; Doppler, T.; Junghans, M.; Wittmer, I.; Stamm, C.; Singer, H. Pesticide risks in small streams: How to get as close as possible to the stress imposed on aquatic organisms. Environ. Sci. Technol. 2018, 52, 4526-4535. [CrossRef]
6. Spycher, S.; Teichler, R.; Vonwyl, E.; Longrée, P.; Stamm, C.; Singer, H.; Daouk, S.; Doppler, T.; Junghans, M.; Kunz, M. Anhaltend hohe PSM-Belastung in Bächen. Aqua Gas 2019, 4, 14-25.
7. Rösch, A.; Beck, B.; Hollender, J.; Singer, H. Picogram per liter quantification of pyrethroid and organophosphate insecticides in surface waters: A result of large enrichment with liquid-liquid extraction and gas chromatography coupled to mass spectromtry using atmospheric pressure chemical Ionization. Anal. Bioanal. Chem. 2019, 411, 3151-3164. [CrossRef] [PubMed]
8. Rösch, A.; Beck, B.; Hollender, J.; Stamm, C.; Singer, H.; Doppler, T.; Junghans, M. Geringe Konzentrationen mit grosser Wirkung: Nachweis von Pyrethroid-und Organophosphatinsektiziden in Schweizer Bächen im pg/l Bereich. Aqua Gas 2019, 11, 54-66.
9. Langer, M.; Junghans, M.; Spycher, S.; Koster, M.; Baumgartner, C.; Vermeirssen, E.; Werner, I. Hohe ökotoxikologische Risiken in Bächen-NAWA SPEZ untersucht Bäche in Gebieten landwirtschaftlicher Nutzung. Aqua Gas 2017, 4, 58-68.
10. Junghans, M.; Langer, M.; Baumgartner, C.; Vermeirssen, E.; Werner, I. Ökotoxikologische Risiken in Bächen durch Effekte aus Organismen bestätigt. Aqua Gas 2019, 4, 26-34.
11. Moermond, C.T.; Kase, R.; Korkaric, M.; Ågerstrand, M. CRED: Criteria for reporting and evaluating ecotoxicity data. Environ. Toxicol. Chem. 2016, 35, 1297-1309. [CrossRef]
12. EFSA. Conclusion on the peer review of epoxiconazole. EFSA Sci. Rep. 2008, 138, 1-80.
13. EFSA. Conclusion on the peer review of the pesticide risk assessment of the active substance tebuconazole. EFSA J. 2014, 12, 3485.
14. Sabra, F.S.; Mehana, E.S.E.D. Pesticides toxicity in fish with particular reference to insecticides. Asian J. Agric. Food Sci. 2015, 3, 40-60.
15. Ullah, S.; Zorriehzahra, M.J. Ecotoxicology: A review of pesticides induced toxicity in fish. Adv. Anim. Vet. Sci. 2015, 3, $40-57$. [CrossRef]
16. Perez-Parada, A.; Goyenola, G.; Texeira de Mello, F.; Heinzen, H. Recent advances and open questions around pesticide dynamics and effects on freshwater fishes. Curr. Opin. Environ. Sci. Health 2018, 4, 38-44. [CrossRef]
17. Bojarski, B.; Witeska, M. Blood biomarkers of herbicide, insecticide, and fungicide toxicity to fish-A review. Environ. Sci. Pollut. Res. 2020, 27, 19236-19250. [CrossRef] [PubMed]
18. Werner, I.; Young, T. Pyrethroid insecticides-Exposure and impacts in the aquatic environment. In The Encyclopedia of the Anthropocene; DellaSala, D.A., Goldstein, M.I., Eds.; Elsevier: Oxford, UK, 2018; Volume 5, pp. 119-126. [CrossRef]
19. Baldwin, D.H.; Spromberg, J.A.; Collier, T.K.; Scholz, N.L. A fish of many scales: Extrapolating sublethal pesticide exposures to the productivity of wild salmon populations. Ecol. Appl. 2009, 19, 2004-2015. [CrossRef] [PubMed]
20. Sih, A.; Bell, A.V.; Kerby, J.L. Two stressors are deadlier than one. Trends Ecol. Evol. 2004, 19, 274-276. [CrossRef]
21. Laetz, C.A.; Baldwin, D.H.; Collier, T.K.; Hebert, V.; Stark, J.D.; Scholz, N.L. The synergistic toxicity of pesticide mixtures: Implications for risk assessment and the conservation of endangered Pacific salmon. Environ. Health Persp. 2009, 117, 348-353. [CrossRef]
22. Bacchetta, C.; Rossi, A.; Ale, A.; Campana, M.; Parma, M.J.; Cazenave, J. Combined toxicological effects of pesticides: A fish multi-biomarlker approach. Ecol. Indic. 2014, 36, 532-538. [CrossRef]
23. Relyea, R.; Hoverman, J. Assessing the ecology in ecotoxicology: A review and synthesis in freshwater systems. Ecol. Lett. 2006, 9, 1157-1171. [CrossRef] [PubMed]
24. Yamamuro, M.; Komuro, T.; Kamiya, H.; Kato, T.; Hasegawa, H.; Kameda, Y. Neonicotinoids disrupt aquatic food webs and decrease fishery yields. Science 2019, 366, 620-623. [CrossRef]
25. Blackwell, B.R.; Ankley, G.T.; Bradley, P.M.; Houck, K.A.; Makarov, S.S.; Medvedev, A.V.; Swintek, J.; Villeneuve, D.L. Potential toxicity of complex mixtures in surface waters from a nationwide survey of US streams: Identifying in Vitro Bioactivities and Causative Chemicals. Environ. Sci. Technol. 2019, 53, 973-983. [CrossRef]
26. Burkhardt-Holm, P.; Giger, W.; Güttinger, H.; Ochsenbein, U.; Peter, A.; Scheurer, K.; Segner, H.; Staub, E.; Suter, M.J.-F. Where have all the fish gone? Environ. Sci. Technol. 2005, 39, 441A-447A. [CrossRef] [PubMed]
27. Brooks, M.L.; Fleishman, E.; Brown, L.; Lehman, P.; Werner, I.; Johnson, M.L.; Scholz, N.; Mitchelmore, C.; Parker, A.; Stoms, D.; et al. Life histories, salinity zones, and sublethal contributions of contaminants to pelagic fish declines illustrated with a case study of San Francisco Estuary, California, USA. Estuaries Coasts 2011, 35, 603-621. [CrossRef]
28. Kirchhofer, A.; Breitenstein, M.; Zaugg, B. Rote Liste der Fische und Rundmauler der Schweiz; Umwelt-Vollzug Nr. 0734; Bundesamt für Umwelt, Bern, und Schweizer Zentrum für die Kartographie der Fauna: Neuenburg, Switzerland, 2007; p. 64.
29. EU Commission. European Union Guideline for the Derivation of Environmental Quality Standards; Guidance Document No. 27; European Commission: Brussels, Belgium, 2011. [CrossRef]
30. Ashauer, R.; Kuhl, R.; Zimmer, E.; Junghans, M. Effect modeling quantifies the difference between the toxicity of average pesticide concentrations and time-variable exposures from water quality monitoring. Environ. Toxicol. Chem. 2020, 39, 2158-2168. [CrossRef]
31. Price, P.; Dhein, E.; Hamer, M.; Han, X.; Heneweer, M.; Junghans, M.; Kunz, P.; Magyar, C.; Penning, H.; Rodriguez, C. A decision tree for assessing effects from exposures to multiple substances. Environ. Sci. Eur. 2012, 24, 26. [CrossRef]
32. Price, P.; Han, X.; Junghans, M.; Kunz, P.; Watts, C.; Leveret, D. An application of a decision tree for assessing effects from exposures to multiple substances to the assessment of human and ecological effects from combined exposures to chemicals observed in surface waters and wastewater effluents. Environ. Sci. Eur. 2012, 24, 34-47. [CrossRef]
33. Van Straalen, N.M. Biodiversity of ecotoxicological responses in animals. Neth. J. Zool. 1994, 44, 112-129. [CrossRef]
34. Ibrahim, L.; Preuss, T.G.; Ratte, H.T.; Hommen, U. A list of fish species that are potentially exposed to pesticides in edge-of-field water bodies in the European Union-A first step towards identifying vulnerable representatives for risk assessment. Environ. Sci. Poll. Res. 2013, 20, 2679-2687. [CrossRef]
35. Spalinger, L.; Dönni, W.; Guthruf, J. NAWA Trend Biologie: 2. Kampagne (2015) Fachbericht Fische Report; Federal Office for Environment: Bern, Switzerland, 2017; p. 75.
36. EFSA. Guidance on tiered risk assessment for plant protection products for aquatic organisms in edge-of-field surface waters. EFSA J. 2013, 11, 3290.
37. R Core Team. R: A Language and Environment for Statistical Computing; R Foundation for Statistical Computing: Vienna, Austria, 2020. Available online: https:/ / R-project.org / (accessed on 3 February 2021).
38. Brady, J.A.; Wallender, W.W.; Werner, I.; Fard, B.M.; Zalom, F.G.; Oliver, M.N.; Wilson, B.W.; Mata, M.M.; Henderson, J.D.; Deanovic, L.A.; et al. Pesticide runoff from orchard floors: A comparative analysis of diazinon and esfenvalerate. Agric. Ecosyst. Environ. 2006, 115, 56-68. [CrossRef]
39. Jorgenson, B.C.; Wissel-Tyson, C.; Young, T.M. Factors contributing to the off-target transport of pyrethroid insecticides from urban surfaces. J. Agric. Food Chem. 2012, 60, 7333-7340. [CrossRef] [PubMed]
40. Jorgenson, B.; Brown, L.; Macneale, K.; Schlenk, D.; Scholz, N.L.; Spromberg, J.; Werner, I.; Weston, D.; Young, T.; Zhang, M.; et al. Predicted transport of pyrethroid insecticides from an urban landscape to surface water. Environ. Toxicol. Chem. 2013, 32, 2469-2477. [CrossRef]
41. Dax, A.; Stravs, M.; Stamm, C.; Ort, C.; La Cecilia, D.; Singer, H. MS²Field: Mikroverunreinigungen mobil messen. Zeitlich hochaufgelöste Messungen zeigen realistisches Ausmass akuter Gewässerbelastungen. Aqua Gas 2020, 12, 14.
42. Federal Office for Agriculture. Zurückgezogene Wirkstoffe Aus Anhang 1 der PSMV. 2017. Available online: https:/ /www.blw. admin.ch/blw/de/home/nachhaltige-produktion/pflanzenschutz/pflanzenschutzmittel/zugelassene-pflanzenschutzmittel. html (accessed on 28 January 2021).
43. Jeffries, K.M.; Hinch, S.G.; Gale, M.K.; Clark, T.D.; Lotto, A.G.; Casselman, M.T.; Li, S.; Rechisky, E.L.; Porter, A.D.; Welch, D.W.; et al. Immune response genes and pathogen presence predict migration survival in wild salmon smolts. Mol. Ecol. 2014, 23, 5803-5815. [CrossRef]
44. Hayes, T.B.; Hansen, M. From silent spring to silent night: Agrochemicals and the anthropocene. Elem. Sci. Anthr. 2017, 5, 57. [CrossRef]
45. Scholz, N.L.; Fleishman, E.; Brown, L.; Brooks, M.L.; Mitchelmore, C.; Werner, I.; Johnson, M.L.; Schlenk, D. Pesticides and the decline of pelagic fishes in western North America's largest estuarine ecosystem. BioScience 2012, 62, 428-434. [CrossRef]
46. Fong, S.; Louie, S.; Werner, I.; Davis, J.; Connon, R.E. Contaminant Effects on California Bay-Delta Species and Human Health. San Fr. Estuary Watershed Sci. 2016, 14. Available online: http:/ /escholarship.org/uc/item/52m780xj (accessed on 28 January 2021).
47. Brander, S.M.; Gabler, M.K.; Fowler, N.L.; Connon, R.E.; Schlenk, D. Pyrethroid Pesticides as endocrine disruptors: Molecular mechanisms in vertebrates with a focus on fishes. Environ. Sci. Technol. 2016, 50, 8977-8992. [CrossRef] [PubMed]
48. Vieira, C.E.D.; Dos Reis Martinez, C.B. The pyrethroid $\lambda$-cyhalothrin induces biochemical, genotoxic, and physiological alterations in the teleost Prochilodus lineatus. Chemosphere 2018, 210, 958-967. [CrossRef]
49. Clifford, M.A.; Eder, K.J.; Werner, I.; Hedrick, R.P. Synergistic effects of esfenvalerate and infectious hematopoietic necrosis virus on juvenile Chinook salmon mortality. Environ. Toxicol. Chem. 2005, 24, 1766-1772. [CrossRef]
50. Muranli, F.D.G.; Güner, U. Induction of micronuclei and nuclear abnormalities in erythrocytes of mosquito fish (Gambusia affinis) following exposure to the pyrethroid insecticide lambda-cyhalothrin. Mutat. Res. Genet. Toxicol. Environ. Mutagenesis 2011, 726, 104-108. [CrossRef]
51. Tierney, K.B.; Ross, P.S.; Kennedy, C.J. Linuron and carbaryl differentially impair baseline amino acid and bile salt olfactory responses in three salmonids. Toxicology 2007, 231, 175-187. [CrossRef] [PubMed]
52. Sandahl, J.F.; Baldwin, D.H.; Jenkins, J.J.; Scholz, N.L. Odor-evoked field potentials as indicators of sublethal neurotoxicity in juvenile coho salmon (Oncorhynchus kisutch) exposed to copper, chlorpyrifos, or esfenvalerate. Can. J. Fish. Aquat. Sci. 2004, 61, 404-413. [CrossRef]
53. Sandahl, J.F.; Baldwin, D.H.; Jenkins, J.J.; Scholz, N.L. Comparative thresholds for acetylcholinesterase inhibition and behavioral impairment in coho salmon exposed to chlorpyrifos. Environ. Toxicol. Chem. 2005, 24, 136-145. [CrossRef]
54. Moore, A.; Waring, C.P. Sublethal effects of the pesticide diazinon on olfactory function in mature male Atlantic salmon parr. J. Fish Biol. 1996, 48, 758-775. [CrossRef]
55. Uren Webster, T.M.; Perry, M.H.; Santos, E.M. The herbicide linuron inhibits cholesterol biosynthesis and induces cellular stress responses in brown trout. Environ. Sci. Technol. 2015, 49, 3110-3118. [CrossRef]
56. Saglio, P.; Trijasse, S. Behavioral responses to atrazine and diuron in goldfish. Arch. Environ. Contam. Toxicol. 1998, 35, 484-491. [CrossRef]
57. Jiang, J.; Shi, Y.; Yu, R.; Chen, L.; Zhao, X. Biological response of zebrafish after short-term exposure to azoxystrobin. Chemosphere 2018, 202, 56-64. [CrossRef] [PubMed]
58. Andrade, T.S.; Henriques, J.; Almeida, A.; Machado, A.L.; Koba, O.; Giang, P.T.; Soares, A.M.V.M.; Domingues, I. Carbendazim exposure induces developmental, biochemical and behavioural disturbance in zebrafish embryos. Aquat. Toxicol. 2016, 170, 390-399. [CrossRef] [PubMed]
59. Lange, K.; Bruder, A.; Matthaei, C.D.; Brodersen, J.; Paterson, R.A. Multiple-stressor effects on freshwater fish: Importance of taxonomy and life stage. Fish Fish. 2018, 19, 974-983. [CrossRef]
60. Baird, D.J.; Van den Brink, P.J. Using biological traits to predict species sensitivity to toxic substances. Ecotox. Environ. Saf. 2007, 67, 296-302. [CrossRef]
61. Segner, H. Moving beyond a descriptive aquatic toxicology: The value of biological process and trait information. Aquat. Toxicol. 2011, 105, 50-55. [CrossRef]
62. Spurgeon, D.; Lahve, E.; Robinson, L.; Short, S.; Kille, P. Species sensitivity to toxic substances: Evolution, ecology and applications. Front. Environ. Sci. 2020, 8, 588380. [CrossRef]
63. Schäfers, C.; Nagel, R. Fish toxicity and population dynamics: Effects of 3,4-dichloroaniline and the problems of extrapolation. In Sublethal and Chronic Effects of Pollutants on Frehswater Fish; Müller, R., Lloyd, R., Eds.; Fishing News Books for FAOP; Blackwell Scientific Publisher: London, UK, 1994; Chapter 20; pp. 229-238.
64. Spromberg, J.A.; Birge, W.S. Modelling the effect of chronic toxicity on fish populations: The influence of life history strategy. Enviro. Toxicol. Chem. 2000, 24, 1532-1540. [CrossRef]
65. Segner, H. Reproductive and developmental toxicity in fishes. In Reproductive and Developmental Toxicology; Gupta, R.C., Ed.; Elsevier: Amsterdam, The Netherlands, 2011; pp. 1145-1166.
66. Hutchinson, T.H.; Solbe, J.; Kloepper-Sams, P.J. Analysis of the ecetoc aquatic toxicity (EAT) database III-Comparative toxicity of chemical substances to different life stages of aquatic organisms. Chemosphere 1998, 36, 129-142. [CrossRef]
67. Mayer, C.M.; Wahl, D.H. The relationship between prey selectivity and growth and survival in a larval fish. Can. J. Fish. Aquat. Sci. 1997, 54. [CrossRef]
68. Sullivan, L.J.; Ignoffo, T.R.; Baskerville-Bridges, B.; Ostrach, D.J.; Kimmerer, W.J. Prey selection of larval and juvenile planktivorous fish: Impacts of introduced prey. Environ. Biol. Fishes 2016, 99, 633-646. [CrossRef]
69. Boyle, T.P.; Fairchild, J.F.; Haverland, P.S.; Lebo, J.A.; Robinson-Wilson, E. Ecological restructuring in experimental aquatic mesocosms due to the application of diflubenzuron. Environ. Toxicol. Chem. 1996, 15, 1806-1814. [CrossRef]
70. Kottelat, M.; Freyhof, J. Handbook of European Freshwater Fishes; Kottelat, Cornol and Freyhof: Berlin, Germany, 2007; 646p, ISBN 978-2-8399-0298-4.
71. Riedl, C.; Peter, A. Timing of brown trout spawning in Alpine rivers with special consideration of egg burial depth. Ecol. Freshw. Fish 2013, 22, 384-397. [CrossRef]
72. Birk, S.; Chapman, D.; Carvalho, L.; Spears, B.M.; Andersen, H.E.; Argillier, C.; Auer, S.; Baattrup-Pedersen, A.; Banin, L.; Beklioğlu, M.; et al. Impacts of multiple stressors on freshwater biota across spatial scales and ecosystems. Nat. Ecol. Evol. 2020, 4, 1060-1068. [CrossRef] [PubMed]
