Clinical utility of sFlt-1 and PIGF in screening, prediction, diagnosis and monitoring of preeclampsia and fetal growth restriction

H. Stepan¹, A. Galindo², M. Hund³, D. Schlembach⁴, J. Sillman³, D. Surbek⁵, M. Vatish⁶

¹University Hospital Leipzig, Leipzig, Germany; ²Hospital Universitario 12 de Octubre, Madrid, Spain; ³Roche Diagnostics International Ltd, Rotkreuz, Switzerland; ⁴Clinicum Vivantes Neukoelln, Berlin, Germany; ⁵University Hospital, University of Bern, Bern, Switzerland; ⁶Nuffield Department of Women's & Reproductive Health, University of Oxford, Oxford, UK

Correspondence to: Dr Holger Stepan, Liebigstrasse 20a, Department of Obstetrics, University Hospital Leipzig, Leipzig, Germany; Holger.Stepan@medizin.uni-leipzig.de

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/uog.26032

Abstract

Preeclampsia (PE) is characterized by placental and maternal endothelial dysfunction, and associated with fetal growth restriction (FGR), placental abruption, preterm delivery and stillbirth. The angiogenic factors soluble fms-like tyrosine kinase 1 (sFlt-1) and placental growth factor (PIGF) are altered in pregnancies complicated by placental-related disorders. In this review, we summarize existing literature examining the performance of maternal PIGF, sFlt-1 and sFlt-1/PIGF ratio for a) screening and diagnosing PE, b) predicting PE development in the short term, c) monitoring established PE and d) predicting other placental-related disorders. We also discuss the performance of PIGF and the sFIt-1/PIGF ratio for predicting PE in twin pregnancies. For first trimester screening, a more accurate way of identifying high-risk women than current practices is to combine PIGF levels with clinical risk factors and ultrasound markers. To support diagnosis of PE later in pregnancy, the sFlt-1/PIGF ratio has advantages over PIGF because it has a higher pooled sensitivity and specificity for diagnosing and monitoring PE. The sFlt-1/PIGF ratio has clinical value because it can rule out the development of PE in the subsequent 1–4 weeks after the test. Once diagnosis of PE is established, repeated measurement of sFlt-1 and PIGF can help monitor progression of the condition and may inform clinical decision-making around optimal time for delivery. The sFlt-1/PIGF ratio is useful for predicting FGR and preterm delivery, but the association between stillbirth and the angiogenic factors remains unclear. The sFlt-1/PIGF ratio can also be used to predict PE in twin pregnancies, although different sFlt-1/PIGF ratio cut-offs to those of singleton pregnancies should be applied for optimal performance. In summary, PIGF, sFlt-1 and the sFlt-1/PIGF ratio are useful for screening, diagnosing, predicting, and monitoring placental-related disorders in singleton and twin pregnancies; we propose further integration of these angiogenic factor tests in clinical practice.

INTRODUCTION

Preeclampsia (PE) is a complex disorder characterized by poor placental function and maternal endothelial dysfunction (Figure 1).^{1, 2} PE can deteriorate into eclampsia (a severe complication of PE, characterized by maternal seizures) or HELLP syndrome (maternal hemolysis, elevated liver enzymes and low platelet count), which are leading causes of maternal morbidity and mortality.³⁻⁶ PE is also associated with adverse fetal outcomes, including fetal growth restriction (FGR), preterm delivery and stillbirth.^{7, 8} Globally, PE is estimated to affect 2–8% of pregnancies and is responsible for >70,000 maternal deaths and >500,000 fetal deaths every year.^{9, 10}

FGR is defined as the failure of the fetus to reach its genetically determined growth potential.¹¹ This condition is diagnosed using ultrasound, and is defined as an estimated fetal weight of <10th percentile for gestational age.^{12, 13} FGR increases the risk of fetal morbidity and mortality and long-term ill health in adulthood.^{12, 14-18} The main treatment for PE and/or FGR is delivery of the fetus and the pathological placenta, thus PE and FGR are among the leading causes of preterm delivery.¹⁹

PE rarely develops before 20 weeks' gestation, and is often classified as early-onset (<34 weeks' gestation) or late-onset (≥34 weeks' gestation).²⁰⁻²² Early-onset PE is associated with a higher risk of adverse maternal and fetal outcomes than late-onset PE.²³ PE symptoms include *de novo* hypertension, proteinuria and/or evidence of maternal acute kidney injury, pulmonary edema, liver dysfunction, neurological features, hemolysis, thrombocytopenia, or FGR.²² Although there is no cure for PE, being able to effectively screen, diagnose, predict and monitor PE development is important as it allows for the implementation of preventative clinical management strategies.²⁴ For example, several studies have demonstrated that treating women who have a high risk of developing PE with low-dose aspirin starting early in pregnancy can reduce the risk of developing the preterm forms of the disease and associated adverse outcomes.²⁵⁻²⁸

The American College of Obstetricians and Gynecologists (ACOG), the National Institute for Health and Care Excellence (NICE) and the International Society for the Study of Hypertension in Pregnancy (ISSHP) recommend screening pregnant women for PE in the first trimester based on the following maternal risk factors: diagnosed with PE or hypertension in a previous pregnancy; having chronic hypertension or kidney disease; having pre-pregnancy diabetes mellitus or autoimmune disease; primiparity or multiple pregnancy or pregnancy being conceived by assisted reproductive technology; mother aged >35 years or having a pre-pregnancy body mass index of >30 kg/m²; pregnancy interval of >10 years; or a family history of PE (Table 1).^{10, 23, 29-31} These organizations further advise that high-risk women are offered low-dose aspirin to try to prevent the onset of PE (Table 1).^{10, 23, 31} It is recommended that women with a diagnosis of PE are monitored closely for the development of severe PE, eclampsia and HELLP syndrome and their fetuses should be monitored for the development of FGR.²²

Certain circulating maternal blood pro-angiogenic and anti-angiogenic factors, such as placental growth factor (PIGF) and soluble fms-like tyrosine kinase 1 (sFlt-1), respectively, are altered in pregnancies complicated by PE, FGR, preterm delivery and stillbirth.^{11, 32-38} Altered levels of sFlt-1 and PIGF are observed when abnormal placentation occurs, leading to insufficient remodeling of maternal spiral arteries and placental ischemia.^{11, 32} The hypoxic placenta releases elevated concentrations of anti-angiogenic factors, such as sFlt-1, to promote peripheral vasoconstriction and raise maternal blood pressure, increasing the flow of oxygenated maternal blood through the intervillous space.³⁹ Meanwhile, concentrations of pro-angiogenic factors, such as PIGF, are substantially reduced due to inhibition by sFlt-1.³⁹ The imbalance of sFlt-1 and PIGF contributes to the clinical manifestation of PE, and alterations in these angiogenic factors can often be detected before the onset of clinical symptoms.^{11, 32-38} According to the 2021 ISSHP guidelines, angiogenic imbalance (i.e. an increased maternal sFlt-1/PIGF ratio or decreased maternal PIGF) is a diagnostic criterion for *de novo* PE, where maternal and fetal outcomes differ considerably to preeclampsia

superimposed on chronic hypertension.⁴⁰ Therefore, gestational hypertension combined with uteroplacental dysfunction, e.g. angiogenic imbalance, is defined as *de novo* PE.¹⁰ The NICE 2016 guidelines, the Danish Society for Obstetrics and Gynecology 2018 guidelines, the European Society of Cardiology 2018 guidelines, the German Society of Obstetrics and Gynecology, the Austrian Society of Obstetrics and Gynecology, the Swiss Society for Obstetrics and Gynecology 2019 guidelines and the Spanish Society of Gynecology and Obstetrics 2020 guidelines recommend measuring the maternal sFlt-1/PIGF ratio to help diagnose and/or predict PE development.⁴¹⁻⁴⁵

The purpose of this review is to summarize the existing literature examining the performance of maternal sFlt-1 and PIGF levels and the sFlt-1/PIGF ratio in PE.

Angiogenic imbalance underlying PE and placental-related disorders

sFlt-1 is an anti-angiogenic factor that is important for the regulation of angiogenic homeostasis during pregnancy.¹ sFlt-1 concentration steadily increases during the third trimester but increases prematurely in women who go on to develop PE and in women with pregnancies complicated by FGR.⁴⁶ Circulating maternal sFlt-1 has been reported to increase approximately 5 weeks before the onset of symptoms.⁴⁶ sFlt-1 binds to PIGF, and a recent study suggests that lower circulating levels of PIGF are largely mediated by excess circulating sFlt-1.⁴⁷ PIGF is a pro-angiogenic factor that is expressed in the placenta and enhances the actions of vascular endothelial growth factor-A (VEGF-A), which is essential for placental vascular development.¹ Maternal PIGF concentration increases initially, peaking in mid-gestation and then gradually decreases toward term.^{46, 48} This decrease in PIGF concentration is known to occur prematurely in women who go on to develop PE and is often detectable before the onset of symptoms.³⁷ Increasing levels of sFlt-1 and decreasing levels of PIGF result in an increasing sFlt-1/PIGF ratio, thus the sFlt-1/PIGF ratio can be a useful tool for predicting and/or diagnosing placental-related disorders, including PE, FGR, stillbirth and preterm birth.⁴⁹ Other anti-angiogenic factors, such as soluble endoglin, have previously been investigated as biomarkers for diagnosis and prediction of PE, but have not been implemented in clinical practice.^{46, 50}

Using PIGF in the first trimester to screen for PE

A key objective of first trimester screening is to identify women at high risk of developing PE later in pregnancy so that suitable preventative strategies can be put in place. At present, many centers do not use a combined first-trimester screening approach.²⁰ Identification of women at high risk of developing PE is often based on assessment of clinical risk factors only, as recommended by the ACOG 2018 and NICE 2019 guidelines.^{29, 31} Combining clinical risk factors, maternal blood pressure (including mean arterial pressure [MAP]), mean uterine artery pulsatility index (MUTPI) measurements, and maternal angiogenic biomarkers into an algorithm may be a more accurate way of identifying high-risk women.^{20, 51}

One of the most robust PE screening algorithms has been developed by the Fetal Medicine Foundation (FMF) and uses a combination of clinical risk factors, maternal age, MAP and MUTPI measurements and maternal PIGF to identify high-risk women in the first trimester (Figure 2).^{20, 52-54} The FMF algorithm was initially developed in almost 36,000 pregnant women attending a UK hospital and has since been validated in two large multicenter trials.^{20, 28, 53} The FMF algorithm for screening for PE can also be adapted for use later in pregnancy to assess the risk of developing the disease based on a combination of clinical risk factors, maternal age, MAP, MUTPI, and maternal PIGF measurements.⁵² The International Federation of Gynecology and Obstetrics recommends that all pregnant women should be screened for PE early in pregnancy by assessing both clinical risk factors and maternal biomarkers.⁷

sFlt-1 is not useful for screening for PE during the first trimester, as levels of sFlt-1 only begin to increase at 21–24 weeks of gestation in women who go on to develop PE.⁴⁶

Using PIGF to diagnose and predict PE in the second and third trimester

The normal decrease in maternal PIGF levels toward the end of pregnancy often occurs prematurely in women who develop PE; therefore, regular measurement of maternal circulating PIGF levels may help to diagnose PE promptly and before clinical onset of the disease.^{37, 55} In support of this, a metaanalysis of 40 studies (3189 cases of PE *vs* 89,498 pregnant controls at <14 weeks, \geq 14 weeks or \geq 19 weeks to term) reported that low maternal PIGF levels of 80–120 pg/mL were able to diagnose PE with a pooled sensitivity of 78%, a pooled specificity of 88%, a pooled positive likelihood ratio of 6.3 and a pooled negative likelihood ratio of 0.26 (Table 2).³⁷

In addition, the prospective, multicenter PETRA trial reported that low PIGF levels ($\leq 100 \text{ pg/mL}$) in women presenting at <35 weeks' gestation with suspected PE had a sensitivity of 76%, a specificity of 69% and a negative predictive value (NPV) of 53% for predicting a final diagnosis of PE at any time.³⁵ A secondary analysis of the PETRA trial showed that low PIGF levels ($\leq 100 \text{ pg/mL}$) were significantly associated with increased risk for a composite maternal adverse outcome compared with normal PIGF levels (6.2% vs 1.9%), and had a sensitivity and specificity of 86.8% and 34.3%, respectively, for predicting a composite maternal outcome.⁵⁶

Low maternal PIGF levels can also predict the development of PE in women with suspected PE. The prospective, multicenter PELICAN study showed that, for 287 women presenting at <35 weeks' gestation with suspected PE, a PIGF concentration of <5th percentile for gestation had a sensitivity of 96%, a specificity of 55% and an NPV of 98% for predicting the development of PE requiring delivery within the next 14 days (Table 2).⁴⁸ In the same cohort of women, a PIGF concentration of <100 pg/mL predicted PE requiring delivery within 14 days with a sensitivity of 96%, a specificity of 56% and an NPV of 98% (Table 2).⁴⁸

To assess the diagnostic accuracy of PIGF measurements in a real-world setting, the PARROT trial examined whether knowledge of the woman's PIGF concentration decreased the time taken by clinicians to diagnose PE in women with suspected PE.⁵⁵ In this study, 11 maternity units were assigned to blocks that represented an intervention initiation time that occurred at 6-week intervals throughout the trial; the units were assigned either to the intervention group or to receive usual

Accepted Article

care with additional concealed testing depending on their allocated block.⁵⁵ The study found that the availability of the PIGF test result significantly reduced the median time for clinical confirmation of PE from 4.1 to 1.9 days (P=0.027) and significantly lowered the incidence of severe maternal adverse outcomes (as defined by the fullPIERS consensus, P=0.043) but did not significantly alter adverse fetal outcomes.⁵⁵ In addition, no differences between groups in gestational age at delivery were observed in this study (mean difference = 0.52 weeks).

Using the sFlt-1/PIGF ratio to diagnose and predict PE in the second and third trimester

Several cut-off thresholds for the sFlt-1/PIGF ratio have been established using receiver operating characteristic (ROC) analysis (Table 2). Verlohren *et al.* validated a set of gestational dependent cut-offs for the diagnosis of PE.⁵⁷ For early-onset PE (<34 weeks' gestation), an sFlt-1/PIGF ratio cut-off of \leq 33 ruled out PE at the time of the test with a sensitivity of 95.0% and a specificity of 94.0%, and a cut-off of \geq 85 diagnosed PE with a sensitivity of 88.0% and a specificity of 99.5%. For late-onset PE (\geq 34 weeks' gestation), an sFlt-1/PIGF ratio cut-off of \leq 33 ruled out PE at the time of the test with a sensitivity of s33 ruled out PE at the time of the test with a sensitivity of 88.0% and a specificity of 99.5%. For late-onset PE (\geq 34 weeks' gestation), an sFlt-1/PIGF ratio cut-off of \leq 33 ruled out PE at the time of the test with a sensitivity of 89.6% and a specificity of 73.1%, and a cut-off of \geq 110 diagnosed PE with a sensitivity of 58.2% and a specificity of 95.5%. Later, Herraiz *et al.* reported that using an sFlt-1/PIGF ratio cut-off of >95th percentile at 24–28 weeks' gestation identified 100% of the women at high risk of PE who went on to develop the early-onset form of the disease in a prospective, observational study of 5601 pregnant women (Table 2).¹³

Employing the sFIt-1/PIGF ratio can also predict the development of PE and/or adverse fetal outcomes in the short term in women with suspected PE. PROGNOSIS was a prospective study conducted at 30 sites across 14 countries and recruited women with suspected PE at 24+0–36+6 weeks' gestation.⁵⁸ The study validated an sFIt-1/PIGF ratio cut-off of ≤38 for ruling out the development of PE within 1 week of the test with a sensitivity of 80.0%, a specificity of 78.3% and an NPV of 99.3% (Table 2). The study also showed an improvement in prediction, compared with clinical variables, of an sFIt-1/PIGF ratio cut-off of >38 for ruling in the development of PE within 4 weeks of

the test with a sensitivity of 66.2%, a specificity of 83.1% and a positive predictive value (PPV) of 36.7% (Table 2). These sFIt-1/PIGF ratio cut-offs were then validated in 764 pregnant Asian women in the PROGNOSIS Asia study (Table 2).⁵⁹ Zeisler *et al.* later carried out exploratory post-hoc analysis of the data collected during the PROGNOSIS study and reported that applying an sFIt-1/PIGF ratio cut-off of \leq 38 ruled out the onset of PE for up to 4 weeks with a high NPV of 94.3% (Table 2).⁶⁰ Moreover, the INSPIRE trial reported that clinical examination combined with an sFIt-1/PIGF ratio of >38 was able to identify 100% of the women who developed PE in the subsequent week and posthoc analysis found that an sFIt-1/PIGF ratio of \geq 85 was able to rule in PE developing within the next 4 weeks with a PPV of 71.4% (Table 2).^{61, 62}

In a prospective pilot study that included 50 pregnant women at risk for developing PE, Soundararajan *et al.* found that women with a high-risk sFlt-1/PIGF ratio (>85) were more likely to have PE with severe features (90.9% *vs* 8.00%, *P*<0.001), a higher composite maternal adverse outcome rate (18.2% *vs* 0%, *P*=0.04), and to deliver at an earlier gestational age (32.6 *vs* 37.4 weeks, *P*=0.001) compared with women with a low-risk sFlt-1/PIGF ratio (<33).⁶³ Similarly, Leaños-Miranda *et al.* reported that pregnant women with PE who had a severe imbalance of angiogenic factors (sFlt-1/PIGF ratio ≥85) had significantly higher rates of preterm delivery, delivery within 14 days of the test, and infants that were small for gestational age (SGA) compared with women with PE who had no imbalance (sFlt-1/PIGF ratio ≤38) or a mild imbalance (sFlt-1/PIGF ratio >38–<85) of angiogenic factors (*P*<0.001).⁶⁴ Indeed, Tan *et al.* reported that screening for delivery due to PE within 4 weeks of assessment at 31–34 weeks' gestation by combining maternal risk factors with sFlt-1 and PIGF values performed similarly to using the sFlt-1/PIGF ratio alone.⁶⁵

The sFlt-1/PIGF ratio has been shown to have a similar sensitivity to PIGF for prediction and diagnosis of PE, but a higher specificity (Table 2).^{37, 58-62, 66-68} Consequently, many of the current guidelines recommend using the sFlt-1/PIGF ratio to aid in the diagnosis of PE.^{10, 41-45}

Using PIGF and the sFIt-1/PIGF ratio to monitor disease progression in established PE

Following a diagnosis of PE, monitoring maternal sFlt-1 and PIGF levels may help to predict the interval between diagnosis of PE and delivery, which would allow for the implementation of clinical management strategies. In a secondary analysis of an observational cohort study, Zeisler *et al.* investigated the correlation between the sFlt-1/PIGF ratio and the time to delivery and reported that women with suspected PE at 24+0–36+6 weeks' gestation had a 2.9 fold greater likelihood of imminent delivery (i.e., delivery on the day of the test) if their sFlt-1/PIGF ratio was >38, when compared with pregnant women with lower sFlt-1/PIGF ratios.⁶⁹ The sFlt-1/PIGF ratios determined in the primary analysis were not available to investigators or patients until study completion; as such, the results did not influence clinical decision-making.⁵⁸

In addition, several studies have taken serial measurements of sFIt-1/PIGF ratios once PE has been diagnosed to examine the time between diagnosis and delivery. Baltajian *et al.* conducted an observational study in which sFIt-1 and PIGF concentrations were recorded weekly from admission to delivery in pregnant women admitted to hospital with suspected PE; the mean number of days from admission to delivery was six (range: 0–35) for women with an sFIt/PIGF ratio \geq 85 at admission and 14 (range: 0–39) for women with an sFIt/PIGF ratio <85 at admission (*P*<0.001).⁷⁰ In addition to shorter time to delivery, the rate of increase in anti-angiogenic state was more pronounced in women with adverse outcomes compared with those without adverse outcomes.⁷⁰ Similarly, Schaarschmidt *et al.* took serial measurements of sFIt-1 and PIGF from women with confirmed early-onset PE and confirmed late-onset PE from admission until delivery.⁷¹ Compared with those who had late-onset PE, women with early-onset PE had greater daily increases in sFIt-1 levels (11% *vs* 3% per day, respectively, *P*<0.05), greater daily decreases in PIGF levels (21% *vs* 10% per day, respectively, *P*<0.05). Likewise, Peguero *et al.* measured sFIt-1 and PIGF revels in women with confirmed early-onset PE at admission and just before delivery and reported that longitudinal changes in maternal

sFlt-1 levels were more pronounced in pregnancies with early-onset severe PE vs uncomplicated pregnancies (median increase: 1047 vs 342 pg/mL/day, respectively; *P*=0.04) and the median time from admission to delivery was shorter (4 days vs 16 days, respectively).⁷² Daily increments in the sFlt-1 and sFlt-1/PIGF ratio values measured following PE diagnosis were associated with shorter time to delivery, and women with steeper increases in sFlt-1 had a significantly shorter time to delivery (*P*<0.001), with earlier gestational age at delivery.⁷² In a cohort of 84 women diagnosed with PE before 37 weeks' gestation, Meler *et al.* reported very low PIGF levels (<12 pg/mL) in 87.5% of women diagnosed before 28 weeks' gestation, 78.4% of women diagnosed between 28 and 32 weeks' gestation, and 41% of women diagnosed after 32 weeks' gestation, with a sensitivity of 76.9% and NPV of 76.9% for predicting maternal complications.⁷³ Importantly, in some cases of severe or early-onset PE, PIGF concentrations may be lower than the detection limit of many commercially available PIGF assays (1.9–9 pg/mL); therefore, measurement of PIGF alone may not be a useful tool for prognosis of early-onset PE.^{41, 73} However, these findings support the characterization of PE as a progressive disorder.⁷¹

PE is frequently accompanied by adverse maternal and fetal outcomes. The fullPIERS model can help to determine which women with confirmed PE will have adverse maternal outcomes and is designed to be used at any stage of pregnancy.²⁹ The model is based on a combination of maternal risk factors and clinical findings including gestational age, presence of chest pain or dyspnea, oxygen saturation, platelets, creatinine and aspartate transaminase/alanine aminotransferase ratio.⁷⁴ The fullPIERS model has been validated in several populations and is currently recommended by NICE guidelines.²⁹ Ukah *et al.* assessed whether adding maternal PIGF concentration improved the performance of the fullPIERS model; the study reported an area under the curve (AUC) of 0.67 (95% confidence interval [CI], 0.58–0.76), lower than that previously reported for the fullPIERS model alone (AUC >0.75).⁷⁵ Median gestational age at delivery was lower in the extension cohort, for which maternal PIGF concentrations were available, compared with the original fullPIERS cohort (33.9 vs 36.9 weeks).⁷⁵

Nevertheless, a Spanish multicenter study demonstrated that using an algorithm based on maternal PIGF levels to determine the optimal delivery time for women with late-onset preterm PE resulted in a lower rate of progression to severe PE (adjusted relative risk [ARR] = 0.5; 95% CI, 0.33–0.76; P=0.001) without an increase in neonatal morbidity (ARR = 0.77; 95% CI, 0.39–1.53; P=0.45).⁷⁶ In this study, median gestational age at delivery was the same in the revealed cohort, in which deliveries were planned based on maternal PIGF concentrations, compared with the concealed cohort, in which deliveries were managed under standard of care (37 vs 37 weeks).⁷⁶

Using PIGF and the sFlt-1/PIGF ratio to predict FGR

Placental dysfunction is associated with FGR and often occurs alongside PE.¹³ Pregnancies that result in FGR have a similar angiogenic factor profile to pregnancies complicated by PE.³⁸ They are characterized by low levels of PIGF throughout pregnancy, particularly in the first trimester, and higher median sFlt-1/PIGF ratios than gestation-matched pregnancies; therefore, knowledge of maternal sFlt-1 and PIGF levels may help predict which pregnancies will be complicated by FGR.^{11, 33, ^{38, 77} Importantly, the sFlt-1/PIGF ratio reflects the degree of placental dysfunction alone and cannot differentiate fully between different clinical presentations of placental dysfunction, such as PE and FGR. Additional tools, such as ultrasound, biophysical methods, and additional biomarkers, are required to make a differential diagnosis.}

In a study that examined 47 biomarkers, maternal PIGF levels were the best predictor of delivery of an SGA neonate (defined in this study as both constitutionally small and pathologically small [FGR] neonates) in women presenting at 20+0–34+6 weeks' gestation with suspected PE.⁷⁸ Low maternal PIGF concentrations (<100 pg/mL) had a sensitivity of 93.2% and an NPV of 89.7% for predicting an SGA fetus, outperforming ultrasound prediction, which yielded a sensitivity of 71.2% (95% CI, 57.9– 82.2%) and an NPV of 78.5% (95% CI, 67.8–86.9%). An elevated sFlt-1/PIGF ratio has been reported in pregnancies complicated by FGR and are particularly elevated in cases of early-onset or severe FGR.^{11, 79} The sFlt-1/PIGF ratio was shown to perform similarly to PIGF levels for predicting adverse outcomes, including FGR, in a systematic review of 33 studies.⁸⁰ Moreover, an observational study of pregnancies complicated by early-onset FGR reported that an elevated sFlt-1/PIGF ratio (>38) could be measured from 4 weeks before delivery in most (73%) of the cases of FGR included in the study; extreme sFlt-1/PIGF ratio values (≥655) could be measured in the 48 hours before delivery in 65% of the pregnancies complicated by FGR and PE, but in only 8% of pregnancies complicated by FGR only.³⁶

Combining the sFIt-1/PIGF ratio with other clinical investigations can also help predict FGR. Gaccioli *et al.* reported that combining ultrasound measurements of the fetus with the sFIt-1/PIGF ratio at 28 weeks' gestation (using a cut-off of >5.78) provided a positive likelihood ratio for premature delivery of an SGA fetus of 41.1, with a sensitivity of 38.5% and a specificity of 99.1% (Table 2).⁸¹ Combined ultrasonic estimated fetal weight and sFIt-1/PIGF ratio screening provided a higher positive likelihood ratio (41.1 vs 5.9) and higher specificity (99.1% vs 92.2%) for predicting premature delivery of an SGA fetus compared with ultrasonic estimated fetal weight alone, but lower sensitivity (38.5% vs 46.2%).⁸¹ Furthermore, in a real-world data analysis of women with symptoms of PE, an AUC of 88.7% was obtained for prediction of maternal and/or fetal adverse outcomes, where the sFIt-1/PIGF ratio was combined with all available clinical information.⁸² Notably, Ciobanu *et al.* reported that the addition of PIGF, sFIt-1, MUTPI, umbilical artery pulsatility index and fetal middle cerebral artery pulsatility index, to maternal factors and fetal biometry only marginally improved the predictive performance for the delivery of an SGA neonate compared with maternal factors and fetal biometry alone.⁸³

Using PIGF and the sFlt-1/PIGF ratio to predict preterm delivery

Both PIGF and the sFIt-1/PIGF ratio have been examined for their association with spontaneous and iatrogenic preterm delivery. In women presenting with suspected PE at 20+0–35+0 weeks' gestation,

low maternal PIGF levels strongly correlated with early delivery.³⁵ Specifically, a maternal PIGF concentration of $\leq 100 \text{ pg/mL}$ predicted preterm delivery with a sensitivity of 81.0% and a specificity of 85.3% (Table 2). A PIGF level within normal range (>100 pg/mL) was associated with pregnancy progressing closer to term, even if the women went on to develop PE. Similarly, Salahuddin et al. assessed the association between maternal sFlt-1 and PIGF levels and adverse maternal and fetal outcomes, including premature delivery, in 412 pregnant women with suspected PE and 434 pregnant women without PE.⁸⁴ The study reported that women who had an adverse outcome had a significantly higher sFlt-1, a significantly lower PIGF and consequently, a significantly higher sFlt-1/PIGF ratio than women who did not. The sFlt-1/PIGF ratio also negatively correlated with the timing of delivery. Consistent with the two aforementioned studies, Rana et al. reported that an sFlt-1/PIGF ratio cut-off of ≥85 yielded a sensitivity of 72.9% and a specificity of 94.0% for the development of a range of adverse pregnancy outcomes, including preterm delivery, in women at <34 weeks' gestation.⁸⁵ During the study, delivery occurred within 2 weeks of admission in 86.0% of women with an sFlt-1/PIGF ratio of ≥85 compared with only 15.8% of women with an sFlt-1/PIGF ratio <85 (hazard ratio = 15.2; 95% CI, 8.0–28.7). Similarly, in a population of pregnant women with chronic hypertension, Heimberger et al. reported that women with an sFlt-1/PIGF ratio ≥85 had a higher prevalence of preterm delivery compared with those with an sFlt-1/PIGF ratio <85.86

Using PIGF and the sFlt-1/PIGF ratio to predict stillbirth

The association between maternal angiogenic factors and the risk of stillbirth has been examined in several systematic reviews but remains unclear. Heazell *et al.* performed a systematic review of 21 studies including 100,687 pregnancies involving 740 stillbirths and reported that, of the four biochemical tests examined (PIGF, human placental lactogen, estriol and uric acid), PIGF was the most accurate at predicting stillbirth, with a diagnostic odds ratio (OR) of 49.2 (95% CI, 12.7–191).⁸⁷ Conversely, a systematic review performed by Townsend *et al.* reported that maternal age, parity and prior adverse pregnancy outcome were better predictors of stillbirth than any of the biomarker

tests examined, including PIGF.⁸⁸ However, in a systematic review of 12 studies including 71,668 women, Jacobs *et al.* reported no clear evidence for a consensus between sFlt-1 levels in the first trimester and adverse pregnancy outcomes.⁸⁹

Using PIGF and the sFlt-1/PIGF ratio to predict complications in twin pregnancies

Women with twin pregnancies are more likely to develop PE than women with singleton pregnancies (OR 4.07; 95% CI, 3.65–4.54).⁹⁰ sFlt-1, PIGF and the sFlt-1/PIGF ratio have demonstrated utility in predicting PE or predicting delivery due to PE in twin pregnancies.^{91, 92} In a European multicenter study of 49 twin pregnancies and 292 gestation-matched singleton pregnancies, it was observed that women with twin pregnancies complicated by PE had a similar angiogenic factor profile to that of women with singleton pregnancies complicated by PE; however, ROC analysis provided an optimal sFlt-1/PIGF ratio cut-off of 53 for diagnosing PE in twin pregnancies.⁹² Applying this cut-off resulted in a sensitivity of 94.4% and specificity of 74.2% for diagnosing PE in twin pregnancies, whereas applying the singleton cut-off values of 33 and 85 gave sensitivities of 100% and 83.3% and specificities of 67.7% and 80.6%, respectively. Conversely, in a retrospective study of 164 twin pregnancies with suspected PE, an sFlt-1/PIGF ratio cut-off of <38 was used to rule out delivery due to PE within 1 week with an NPV of 98.8% and within 2 weeks with an NPV of 96.4%.⁹¹ Furthermore, De La Calle et al. analyzed data from three prospective studies (PROGNOSIS, STEPS, and a multicenter case-control study) and reported that reference ranges for the sFlt-1/PIGF ratio were comparable in twin and singleton pregnancies until 29 weeks' gestation, but were then higher in twin pregnancies until birth.57, 58, 68, 93-95

Hayes-Ryan *et al.* reported that women with twin pregnancies who developed PE have significantly lower PIGF levels compared with controls across all gestational intervals from 12+0 to 36+6 weeks' gestation.⁹⁶ Moreover, this difference can be observed several weeks before the onset of clinical symptoms of PE.⁹⁶ Dröge *et al.* also reported significantly lower PIGF levels in women with twin pregnancies who developed PE compared with controls (*P*≤0.001), in addition to significantly lower Accepted Article

PIGF levels in women with twin pregnancies who developed PE compared with singleton pregnancies who developed PE ($P \le 0.001$).⁹² Therefore, measurement of PIGF alone may have potential as an aid in predicting PE in women with twin pregnancies.

sFlt-1, PIGF and the sFlt-1/PIGF ratio have also been examined for their association with adverse maternal and fetal outcomes in twin pregnancies. Rana *et al.* reported that the angiogenic factor profile was altered in women with twin pregnancies who had subsequent adverse outcomes when compared with those who did not; the median [25th-75th percentile] sFlt-1 level was elevated (11,461.5 pg/mL [8794.0–14,847.5] *vs* 7495.0 pg/mL [3498.0–10,482.0], respectively, *P*=0.0004), the median PIGF level was reduced (162.5 pg/mL [98.0–226.5] *vs* 224.0 pg/mL [156.0–449.0], respectively, *P*=0.005) and the median sFlt-1/PIGF ratio was elevated (74.2 [43.5–110.5] *vs* 36.2 [7.1–71.3], respectively, *P*=0.0005).⁹⁷

CONCLUSIONS

Maternal PIGF, sFIt-1 and the sFIt-1/PIGF ratio show good performance for screening and diagnosing PE, for predicting PE development in the short term, for monitoring established PE and for predicting other placental-related disorders. In particular, the sFIt-1/PIGF ratio is clinically useful in guiding management of pregnant women with unclear symptoms of PE, as it has a high NPV and can therefore rule out PE in the subsequent 1–4 weeks. The sFIt-1/PIGF ratio also shows good performance for predicting PE in twin pregnancies. The sFIt-1/PIGF ratio may be preferred over PIGF alone because PIGF decreases with disease severity and in cases of severe or early-onset disease, PIGF concentrations may be lower than the detection limit of commercially available PIGF assays. In cases of early-onset PE, measurement of PIGF alone is not a useful tool and may limit monitoring of disease progression. There remains a need for further integration of tests for these angiogenic factors in clinical practice.

Acknowledgements

Third-party medical writing support for the development of this manuscript, under the direction of the authors, was provided by Anna King, PhD, and Chloe Fletcher, MSc, of Ashfield MedComms (Macclesfield, UK), an Ashfield Health company, and was funded by Roche Diagnostics International Ltd (Rotkreuz, Switzerland). ELECSYS is a trademark of Roche. All other product names and trademarks are the property of their respective owners.

Author conflicts of interest

HS and AG received speaker and consultant fees from Roche Diagnostics. MH and JS are employees of Roche Diagnostics International Ltd (Rotkreuz, Switzerland). DSc received lecture fees from Roche and ThermoFisher Scientific; received advisory board fees from Roche; received support for studies on the sFlt-1/PIGF ratio from Roche; and received support for studies on PIGF from Perkin Elmer. DSu received speaker and advisory board fees from Roche Diagnostics. MV received speaker fees and support for studies on sFlt1/PIGF from Roche Diagnostics.

Author contributions

All authors proposed the concept and/or content of this review, provided references for inclusion, contributed to the writing, editing and/or critical review, and approved the final version for submission.

REFERENCES

1. Stepan H, Hund M, Andraczek T. Combining biomarkers to predict pregnancy complications and redefine preeclampsia: the angiogenic-placental syndrome. *Hypertension* 2020; **75**: 918–926.

2. Wang A, Rana S, Karumanchi SA. Preeclampsia: the role of angiogenic factors in its pathogenesis. *Physiology* 2009; **24**: 147–158.

3. Magley M, Hinson MR. Eclampsia. In *StatPearls*.StatPearls Publishing, Copyright © 2021, StatPearls Publishing LLC: Treasure Island (FL), 2021.

4. Weinstein L. Syndrome of hemolysis, elevated liver enzymes, and low platelet count: a severe consequence of hypertension in pregnancy. *Am J Obstet Gynecol* 1982; **142**: 159–167.

5. Trottmann F, Baumann M, Amylidi-Mohr S, Surbek D, Risch L, Mosimann B, Raio L. Angiogenic profiling in HELLP syndrome cases with or without hypertension and proteinuria. *Eur J Obstet Gynecol Reprod Biol* 2019; **243**: 93–96.

6. Abalos E, Cuesta C, Grosso AL, Chou D, Say L. Global and regional estimates of preeclampsia and eclampsia: a systematic review. *Eur J Obstet Gynecol Reprod Biol* 2013; **170**: 1–7.

Poon LC, Shennan A, Hyett JA, Kapur A, Hadar E, Divakar H, McAuliffe F, da Silva Costa F, von Dadelszen P, McIntyre HD, Kihara AB, Di Renzo GC, Romero R, D'Alton M, Berghella V, Nicolaides KH, Hod M. The International Federation of Gynecology and Obstetrics (FIGO) initiative on pre-eclampsia: a pragmatic guide for first-trimester screening and prevention. *Int J Gynaecol Obstet* 2019; **145 Suppl 1**: 1–33.

Harmon QE, Huang L, Umbach DM, Klungsøyr K, Engel SM, Magnus P, Skjærven R, Zhang J,
 Wilcox AJ. Risk of fetal death with preeclampsia. *Obstet Gynecol* 2015; **125**: 628–635.

Accepted Article

Duley L. The global impact of pre-eclampsia and eclampsia. *Semin Perinatol* 2009; **33**: 130–137.

10. Magee LA, Brown MA, Hall DR, Gupte S, Hennessy A, Karumanchi SA, Kenny LC, McCarthy F, Myers J, Poon LC, Rana S, Saito S, Staff AC, Tsigas E, von Dadelszen P. The 2021 International Society for the Study of Hypertension in Pregnancy classification, diagnosis & management recommendations for international practice. *Pregnancy Hypertension* 2022; **27**: 148-169.

11. Gaccioli F, Aye ILMH, Sovio U, Charnock-Jones DS, Smith GCS. Screening for fetal growth restriction using fetal biometry combined with maternal biomarkers. *Am J Obstet Gynecol* 2018; **218**: s725–s737.

12. Savchev S, Figueras F, Sanz-Cortes M, Cruz-Lemini M, Triunfo S, Botet F, Gratacos E. Evaluation of an optimal gestational age cut-off for the definition of early- and late-onset fetal growth restriction. *Fetal Diagn Ther* 2014; **36**: 99–105.

13. Herraiz I, Simón E, Gómez-Arriaga PI, Quezada MS, García-Burguillo A, López-Jiménez EA, Galindo A. Clinical implementation of the sFlt-1/PIGF ratio to identify preeclampsia and fetal growth restriction: a prospective cohort study. *Pregnancy Hypertens* 2018; **13**: 279–285.

14. Barker DJP, Osmond C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. *Lancet* 1986; **1**: 1077–1081.

15. Hales CN, Barker DJ. Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. *Diabetologia* 1992; **35**: 595–601.

16. Cooper C, Fall C, Egger P, Hobbs R, Eastell R, Barker D. Growth in infancy and bone mass in later life. *Ann Rheum Dis* 1997; **56**: 17–21.

Accepted Article

Barker DJP. Adult consequences of fetal growth restriction. *Clin Obstet Gynecol* 2006; **49**:
 270–283.

Jarvis S, Glinianaia SV, Torrioli MG, Platt MJ, Miceli M, Jouk PS, Johnson A, Hutton J,
 Hemming K, Hagberg G, Dolk H, Chalmers J. Cerebral palsy and intrauterine growth in single births:
 European collaborative study. *Lancet* 2003; **362**: 1106–1111.

19. Amaral LM, Wallace K, Owens M, LaMarca B. Pathophysiology and current clinical management of preeclampsia. *Curr Hypertens Rep* 2017; **19**: 61.

20. Poon LC, Galindo A, Surbek D, Chantraine F, Stepan H, Hyett J, Tan KH, Verlohren S. From first-trimester screening to risk stratification of evolving pre-eclampsia in second and third trimesters of pregnancy: comprehensive approach. *Ultrasound Obstet Gynecol* 2020; **55**: 5–12.

21. Tranquilli AL, Brown MA, Zeeman GG, Dekker G, Sibai BM. The definition of severe and earlyonset preeclampsia. Statements from the International Society for the Study of Hypertension in Pregnancy (ISSHP). *Pregnancy Hypertens* 2013; **3**: 44–47.

22. Brown MA, Magee LA, Kenny LC, Karumanchi SA, McCarthy FP, Saito S, Hall DR, Warren CE, Adoyi G, Ishaku S. The hypertensive disorders of pregnancy: ISSHP classification, diagnosis & management recommendations for international practice. *Pregnancy Hypertens* 2018; **13**: 291–310.

23. Chaemsaithong P, Sahota DS, Poon LC. First trimester preeclampsia screening and prediction. *American Journal of Obstetrics and Gynecology* 2020. DOI:

https://doi.org/10.1016/j.ajog.2020.07.020.

24. Mol BWJ, Roberts CT, Thangaratinam S, Magee LA, de Groot CJM, Hofmeyr GJ. Preeclampsia. *Lancet* 2016; **387**: 999–1011. 25. Bujold E, Roberge S, Lacasse Y, Bureau M, Audibert F, Marcoux S, Forest JC, Giguère Y. Prevention of preeclampsia and intrauterine growth restriction with aspirin started in early pregnancy: a meta-analysis. *Obstet Gynecol* 2010; **116**: 402–414.

26. Roberge S, Giguère Y, Villa P, Nicolaides K, Vainio M, Forest JC, von Dadelszen P, Vaiman D, Tapp S, Bujold E. Early administration of low-dose aspirin for the prevention of severe and mild preeclampsia: a systematic review and meta-analysis. *Am J Perinatol* 2012; **29**: 551–556.

27. Roberge S, Nicolaides KH, Demers S, Villa P, Bujold E. Prevention of perinatal death and adverse perinatal outcome using low-dose aspirin: a meta-analysis. *Ultrasound Obstet Gynecol* 2013;
41: 491–499.

28. Rolnik DL, Wright D, Poon LCY, Syngelaki A, O'Gorman N, de Paco Matallana C, Akolekar R, Cicero S, Janga D, Singh M, Molina FS, Persico N, Jani JC, Plasencia W, Papaioannou G, Tenenbaum-Gavish K, Nicolaides KH. ASPRE trial: performance of screening for preterm pre-eclampsia. *Ultrasound Obstet Gynecol* 2017; **50**: 492–495.

29. NICE. Hypertension in pregnancy: diagnosis and management. https://www.nice.org.uk/guidance/ng133 [20 July 2021].

30. National Collaborating Centre for Women's and Children's Health (UK). Hypertension in pregnancy: the management of hypertensive disorders during pregnancy. RCOG Press: London, 2010.

31. Practice CoO, Medicine SfM-F. ACOG Committee Opinion No. 743: low-dose aspirin use during pregnancy. *Obstet Gynecol* 2018; **132**: e44-52.

32. Allen RE, Rogozinska E, Cleverly K, Aquilina J, Thangaratinam S. Abnormal blood biomarkers in early pregnancy are associated with preeclampsia: a meta-analysis. *Eur J Obstet Gynecol Reprod Biol* 2014; **182**: 194–201.

33. Herraiz I, Dröge LA, Gómez-Montes E, Henrich W, Galindo A, Verlohren S. Characterization of the soluble fms-like tyrosine kinase-1 to placental growth factor ratio in pregnancies complicated by fetal growth restriction. *Obstet Gynecol* 2014; **124**: 265–273.

34. Sovio U, Gaccioli F, Cook E, Charnock-Jones DS, Smith GCS. Slowing of fetal growth and elevated maternal serum sFLT1:PIGF are associated with early term spontaneous labor. *Am J Obstet Gynecol* 2021; **225**: 1–10.

35. Barton JR, Woelkers DA, Newman RB, Combs CA, How HY, Boggess KA, Martin JN, Jr., Kupfer K, Sibai BM. Placental growth factor predicts time to delivery in women with signs or symptoms of early preterm preeclampsia: a prospective multicenter study. *Am J Obstet Gynecol* 2020; **222**: 1–11.

36. Herraiz I, Quezada MS, Rodriguez-Calvo J, Gómez-Montes E, Villalaín C, Galindo A. Longitudinal change of sFlt-1/PIGF ratio in singleton pregnancy with early-onset fetal growth restriction. *Ultrasound Obstet Gynecol* 2018; **52**: 631–638.

37. Agrawal S, Shinar S, Cerdeira AS, Redman C, Vatish M. Predictive performance of PIGF (placental growth factor) for screening preeclampsia in asymptomatic women: a systematic review and meta-analysis. *Hypertension* 2019; **74**: 1124–1135.

Alahakoon TI, Zhang W, Trudinger BJ, Lee VW. Discordant clinical presentations of
 preeclampsia and intrauterine fetal growth restriction with similar pro- and anti-angiogenic profiles.
 J Matern Fetal Neonatal Med 2014; 27: 1854–1859.

39. Herraiz I, Llurba E, Verlohren S, Galindo A. Update on the diagnosis and prognosis of preeclampsia with the aid of the sFlt-1/PIGF ratio in singleton pregnancies. *Fetal diagnosis and therapy* 2018; **43**: 81-89.

40. Boneh HR, Pariente G, Baumfeld Y, Yohay D, Rotem R, Weintraub AY. Superimposed versus de novo pre-eclampsia: Is there a difference? *International Journal of Gynecology & Obstetrics* 2022; **n/a**.

41. NICE. PIGF-based testing to help diagnose suspected preeclampsia (Triage PIGF test, Elecsys immunoassay sFlt-1/PIGF ratio, DELFIA Xpress PIGF 1-2-3 test, and BRAHMS sFlt-1 Kryptor/BRAHMS PIGF plus Kryptor PE ratio). <u>https://www.nice.org.uk/guidance/dg23/</u> [20 July 2021].

42. Regitz-Zagrosek V, Roos-Hesselink JW, Bauersachs J, Blomström-Lundqvist C, Cífková R, De Bonis M, lung B, Johnson MR, Kintscher U, Kranke P, Lang IM, Morais J, Pieper PG, Presbitero P, Price S, Rosano GMC, Seeland U, Simoncini T, Swan L, Warnes CA. 2018 ESC Guidelines for the management of cardiovascular diseases during pregnancy. *Eur Heart J* 2018; **39**: 3165–3241.

43. German Society of Obstetrics and Gynecology (DGGG), Austrian Society of Obstetrics and Gynecology (OeGGG), and Swiss Society for Obstetric and Gynecology (SGGG) Guidelines for Hypertensive Disorders in Pregnancy: Diagnosis and therapy.

https://www.awmf.org/leitlinien/detail/ll/015-018.html [28 June 2019].

44. Spanish Society of Gynaecology and Obstetrics (SEGO): Practical assistance guide: hypertensive disorders in pregnancy. *Prog en Obstet y Ginecol* 2020; **63**: 244–272.

45. Danish Society for Obstetrics and Gynecology: Danish 2018 national clinical guideline for hypertensive disorders in pregnancy and preeclampsia.

https://static1.squarespace.com/static/5467abcce4b056d72594db79/t/5bac84e7652dea0a1b5fb48 9/1538032877105/180924+PE-guideline-final+sandbjerg.pdf [3 November 2021].

46. Levine RJ, Maynard SE, Qian C, Lim KH, England LJ, Yu KF, Schisterman EF, Thadhani R, Sachs BP, Epstein FH, Sibai BM, Sukhatme VP, Karumanchi SA. Circulating angiogenic factors and the risk of preeclampsia. *N Engl J Med* 2004; **350**: 672–683.

47. Lecarpentier E, Zsengellér ZK, Salahuddin S, Covarrubias AE, Lo A, Haddad B, Thadhani RI, Karumanchi SA. Total versus free placental growth factor levels in the pathogenesis of preeclampsia. *Hypertension* 2020; **76**: 875–883.

48. Chappell LC, Duckworth S, Seed PT, Griffin M, Myers J, Mackillop L, Simpson N, Waugh J, Anumba D, Kenny LC, Redman CWG, Shennan AH. Diagnostic accuracy of placental growth factor in women with suspected preeclampsia: a prospective multicenter study. *Circulation* 2013; **128**: 2121– 2131.

49. Stepan H, Herraiz I, Schlembach D, Verlohren S, Brennecke S, Chantraine F, Klein E, Lapaire O, Llurba E, Ramoni A, Vatish M, Wertaschnigg D, Galindo A. Implementation of the sFlt-1/PIGF ratio for prediction and diagnosis of pre-eclampsia in singleton pregnancy: implications for clinical practice. *Ultrasound Obstet Gynecol* 2015; **45**: 241–246.

50. Levine RJ, Lam C, Qian C, Yu KF, Maynard SE, Sachs BP, Sibai BM, Epstein FH, Romero R, Thadhani R, Karumanchi SA. Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *N Engl J Med* 2006; **355**: 992–1005.

51. Tan MY, Wright D, Syngelaki A, Akolekar R, Cicero S, Janga D, Singh M, Greco E, Wright A, Maclagan K, Poon LC, Nicolaides KH. Comparison of diagnostic accuracy of early screening for preeclampsia by NICE guidelines and a method combining maternal factors and biomarkers: results of SPREE. *Ultrasound Obstet Gynecol* 2018; **51**: 743–750.

52. The Fetal Medicine Foundation. Risk assessment: risk for preeclampsia. https://fetalmedicine.org/research/assess/preeclampsia/first-trimester [10 June 2021].

53. Poon LCY, Kametas NA, Maiz N, Akolekar R, Nicolaides KH. First-trimester prediction of hypertensive disorders in pregnancy. *Hypertension* 2009; **53**: 812–818.

54. Mosimann B, Amylidi-Mohr SK, Surbek D, Raio L. First trimester screening for preeclampsia - a systematic review. *Hypertens Pregnancy* 2020; **39**: 1–11.

55. Duhig KE, Myers J, Seed PT, Sparkes J, Lowe J, Hunter RM, Shennan AH, Chappell LC. Placental growth factor testing to assess women with suspected pre-eclampsia: a multicentre, pragmatic, stepped-wedge cluster-randomised controlled trial. *Lancet* 2019; **393**: 1807–1818.

56. Parchem JG, Brock CO, Chen H-Y, Kalluri R, Barton JR, Sibai BM. Placental growth factor and the risk of adverse neonatal and maternal outcomes. *Obstet Gynecol* 2020; **135**: 665–673.

57. Verlohren S, Herraiz I, Lapaire O, Schlembach D, Zeisler H, Calda P, Sabria J, Markfeld-Erol F, Galindo A, Schoofs K, Denk B, Stepan H. New gestational phase-specific cutoff values for the use of the soluble fms-like tyrosine kinase-1/placental growth factor ratio as a diagnostic test for preeclampsia. *Hypertension* 2014; **63**: 346–352.

58. Zeisler H, Llurba E, Chantraine F, Vatish M, Staff AC, Sennström M, Olovsson M, Brennecke SP, Stepan H, Allegranza D, Dilba P, Schoedl M, Hund M, Verlohren S. Predictive value of the sFlt-1:PIGF ratio in women with suspected preeclampsia. *N Engl J Med* 2016; **374**: 13–22.

59. Bian X, Biswas A, Huang X, Lee KJ, Li TK, Masuyama H, Ohkuchi A, Park JS, Saito S, Tan KH, Yamamoto T, Dietl A, Grill S, Verhagen-Kamerbeek WDJ, Shim JY, Hund M. Short-term prediction of adverse outcomes using the sFIt-1 (soluble fms-like tyrosine kinase 1)/PIGF (placental growth factor) ratio in Asian women with suspected preeclampsia. *Hypertension* 2019; **74**: 164–172.

60. Zeisler H, Llurba E, Chantraine FJ, Vatish M, Staff AC, Sennström M, Olovsson M, Brennecke SP, Stepan H, Allegranza D, Schoedl M, Grill S, Hund M, Verlohren S. Soluble fms-like tyrosine kinase-1 to placental growth factor ratio: ruling out pre-eclampsia for up to 4 weeks and value of retesting. *Ultrasound Obstet Gynecol* 2019; **53**: 367–375. 61. Cerdeira AS, O'Sullivan J, Ohuma EO, Harrington D, Szafranski P, Black R, Mackillop L, Impey L, Greenwood C, James T, Smith I, Papageorghiou AT, Knight M, Vatish M. Randomized interventional study on prediction of preeclampsia/eclampsia in women with suspected preeclampsia: INSPIRE. *Hypertension* 2019; **74**: 983–990.

62. Cerdeira AS, O'Sullivan J, Ohuma EO, James T, Papageorghiou AT, Knight M, Vatish M.
 Performance of soluble fms-like tyrosine kinase-1-to-placental growth factor ratio of ≥85 for ruling in preeclampsia within 4 weeks. *Am J Obstet Gynecol* 2021; **224**: 322–323.

63. Soundararajan R, Suresh SC, Mueller A, Heimberger S, Avula S, Sathyanarayana C, Mahesh S, Madhuprakash S, Rana S. Real life outpatient biomarker use in management of hypertensive pregnancies in third trimester in a low resource SeTting: ROBUST study. *Pregnancy Hypertension* 2021; **23**: 97–103.

64. Leaños-Miranda A, Graciela Nolasco-Leaños A, Ismael Carrillo-Juárez R, José Molina-Pérez C, Janet Sillas-Pardo L, Manuel Jiménez-Trejo L, Isordia-Salas I, Leticia Ramírez-Valenzuela K. Usefulness of the sFlt-1/PIGF (Soluble fms-Like Tyrosine Kinase-1/Placental Growth Factor) Ratio in Diagnosis or Misdiagnosis in Women With Clinical Diagnosis of Preeclampsia. *Hypertension* 2020; **76**: 892–900.

65. Tan MY, Wright D, Koutoulas L, Akolekar R, Nicolaides KH. Comparison of screening for preeclampsia at 31-34 weeks' gestation by sFlt-1/PIGF ratio and a method combining maternal factors with sFlt-1 and PIGF. *Ultrasound Obstet Gynecol* 2017; **49**: 201–208.

66. Stepan H, Hund M, Gencay M, Denk B, Dinkel C, Kaminski WE, Wieloch P, Semus B, Meloth T, Dröge LA, Verlohren S. A comparison of the diagnostic utility of the sFlt-1/PIGF ratio versus PIGF alone for the detection of preeclampsia/HELLP syndrome. *Hypertens Pregnancy* 2016; **35**: 295–305.

67. Agrawal S, Cerdeira AS, Redman C, Vatish M. Meta-analysis and systematic review to assess the role of soluble FMS-like tyrosine kinase-1 and placenta growth factor ratio in prediction of preeclampsia: the SaPPPhirE study. *Hypertension* 2018; **71**: 306–316.

68. Verlohren S, Galindo A, Schlembach D, Zeisler H, Herraiz I, Moertl MG, Pape J, Dudenhausen JW, Denk B, Stepan H. An automated method for the determination of the sFlt-1/PIGF ratio in the assessment of preeclampsia. *Am J Obstet Gynecol* 2010; **202**: 1–11.

69. Zeisler H, Llurba E, Chantraine F, Vatish M, Staff AC, Sennström M, Olovsson M, Brennecke SP, Stepan H, Allegranza D, Dinkel C, Schoedl M, Dilba P, Hund M, Verlohren S. Soluble fms-Like tyrosine kinase-1-to-placental growth factor ratio and time to delivery in women with suspected preeclampsia. *Obstet Gynecol* 2016; **128**: 261–269.

70. Baltajian K, Bajracharya S, Salahuddin S, Berg AH, Geahchan C, Wenger JB, Thadhani R, Karumanchi SA, Rana S. Sequential plasma angiogenic factors levels in women with suspected preeclampsia. *Am J Obstet Gynecol* 2016; **215**: 1–10.

71. Schaarschmidt W, Rana S, Stepan H. The course of angiogenic factors in early- vs. late-onset preeclampsia and HELLP syndrome. *J Perinat Med* 2013; **41**: 511–516.

72. Peguero A, Fernandez-Blanco L, Mazarico E, Benitez L, Gonzalez A, Youssef L, Crispi F, Hernandez S, Figueras F. Added prognostic value of longitudinal changes of angiogenic factors in early-onset severe pre-eclampsia: a prospective cohort study. *BJOG* 2021; **128**: 158–165.

73. Meler E, Scazzocchio E, Peguero A, Triunfo S, Gratacos E, Figueras F. Role of maternal plasma levels of placental growth factor for the prediction of maternal complications in preeclampsia according to the gestational age at onset. *Prenat Diagn* 2014; **34**: 706–710.

74. NICE. Evidence review for prediction of complications in pre-eclampsia: Hypertension in pregnancy: diagnosis and management: Evidence review C.

https://www.ncbi.nlm.nih.gov/books/NBK577929/ [17 May 2022].

75. Ukah UV, Payne BA, Hutcheon JA, Chappell LC, Seed PT, Conti-Ramsden FI, Ansermino JM, Magee LA, von Dadelszen P. Placental growth factor for the prognosis of women with preeclampsia (fullPIERS model extension): context matters. *BMC Pregnancy Childbirth* 2020; **20**: 668.

76. Peguero A, Herraiz I, Perales A, Melchor JC, Melchor I, Marcos B, Villalain C, Martinez-Portilla R, Mazarico E, Meler E, Hernandez S, Matas I, del Rio M, Galindo A, Figueras F. Placental growth factor testing in the management of late preterm preeclampsia without severe features: a multicenter, randomized, controlled trial. *Am J Obstet Gynecol* 2021; **225**: 1–14.

77. Taylor RN, Grimwood J, Taylor RS, McMaster MT, Fisher SJ, North RA. Longitudinal serum concentrations of placental growth factor: evidence for abnormal placental angiogenesis in pathologic pregnancies. *Am J Obstet Gynecol* 2003; **188**: 177–182.

78. Griffin M, Seed PT, Duckworth S, North R, Myers J, Mackillop L, Simpson N, Waugh J, Anumba D, Kenny LC, Redman CWG, Shennan AH, Chappell LC. Predicting delivery of a small-forgestational-age infant and adverse perinatal outcome in women with suspected pre-eclampsia. *Ultrasound Obstet Gynecol* 2018; **51**: 387–395.

79. Garcia-Manau P, Mendoza M, Bonacina E, Garrido-Gimenez C, Fernandez-Oliva A, Zanini J, Catalan M, Tur H, Serrano B, Carreras E. Soluble fms-like tyrosine kinase to placental growth factor ratio in different stages of early-onset fetal growth restriction and small for gestational age. *Acta Obstet Gynecol Scand* 2021; **100**: 119–128.

80. Lim S, Li W, Kemper J, Nguyen A, Mol BW, Reddy M. Biomarkers and the prediction of adverse outcomes in preeclampsia: a systematic review and meta-analysis. *Obstet Gynecol* 2021;
137: 72–81.

81. Gaccioli F, Sovio U, Cook E, Hund M, Charnock-Jones DS, Smith GCS. Screening for fetal growth restriction using ultrasound and the sFLT1/PIGF ratio in nulliparous women: a prospective cohort study. *Lancet Child Adolesc Health* 2018; **2**: 569–581.

82. Dröge LA, Perschel FH, Stütz N, Gafron A, Frank L, Busjahn A, Henrich W, Verlohren S. Prediction of preeclampsia-related adverse outcomes with the sFlt-1 (soluble fms-like tyrosine kinase 1)/PIGF (placental growth factor)-ratio in the clinical routine: a real-world study. *Hypertension* 2021; **77**: 461–471.

83. Ciobanu A, Rouvali A, Syngelaki A, Akolekar R, Nicolaides KH. Prediction of small for gestational age neonates: screening by maternal factors, fetal biometry, and biomarkers at 35-37 weeks' gestation. *Am J Obstet Gynecol* 2019; **220**: 1–11.

84. Salahuddin S, Wenger JB, Zhang D, Thadhani R, Karumanchi SA, Rana S. KRYPTOR-automated angiogenic factor assays and risk of preeclampsia-related adverse outcomes. *Hypertens Pregnancy* 2016; **35**: 330–345.

85. Rana S, Powe CE, Salahuddin S, Verlohren S, Perschel FH, Levine RJ, Lim KH, Wenger JB, Thadhani R, Karumanchi SA. Angiogenic factors and the risk of adverse outcomes in women with suspected preeclampsia. *Circulation* 2012; **125**: 911–919.

86. Heimberger S, Mueller A, Ratnaparkhi R, Perdigao JL, Rana S. Angiogenic factor abnormalities and risk of peripartum complications and prematurity among urban predominantly obese parturients with chronic hypertension. *Pregnancy Hypertens* 2020; **20**: 124–130. **Accepted Article**

87. Heazell AE, Hayes DJ, Whitworth M, Takwoingi Y, Bayliss SE, Davenport C. Biochemical tests of placental function versus ultrasound assessment of fetal size for stillbirth and small-for-gestational-age infants. *Cochrane Database Syst Rev* 2019; **5**: CD012245.

88. Townsend R, Sileo FG, Allotey J, Dodds J, Heazell A, Jorgensen L, Kim VB, Magee L, Mol B, Sandall J, Smith G, Thilaganathan B, von Dadelszen P, Thangaratinam S, Khalil A. Prediction of stillbirth: an umbrella review of evaluation of prognostic variables. *BJOG* 2021; **128**: 238–250.

89. Jacobs M, Nassar N, Roberts CL, Hadfield R, Morris JM, Ashton AW. Levels of soluble fms-like tyrosine kinase one in first trimester and outcomes of pregnancy: a systematic review. *Reproductive Biology and Endocrinology* 2011; **9**: 77.

90. Laine K, Murzakanova G, Sole KB, Pay AD, Heradstveit S, Räisänen S. Prevalence and risk of pre-eclampsia and gestational hypertension in twin pregnancies: a population-based register study. *BMJ Open* 2019; **9**: e029908.

91. Binder J, Palmrich P, Pateisky P, Kalafat E, Kuessel L, Zeisler H, Munkhbaatar M, Windsperger K, Thilaganathan B, Khalil A. The prognostic value of angiogenic markers in twin pregnancies to predict delivery due to maternal complications of preeclampsia. *Hypertension* 2020; **76**: 176–183.

92. Dröge L, Herraiz I, Zeisler H, Schlembach D, Stepan H, Küssel L, Henrich W, Galindo A, Verlohren S. Maternal serum sFlt-1/PIGF ratio in twin pregnancies with and without pre-eclampsia in comparison with singleton pregnancies. *Ultrasound Obstet Gynecol* 2015; **45**: 286–293.

93. Verlohren S, Herraiz I, Lapaire O, Schlembach D, Moertl M, Zeisler H, Calda P, Holzgreve W,
Galindo A, Engels T, Denk B, Stepan H. The sFlt-1/PIGF ratio in different types of hypertensive
pregnancy disorders and its prognostic potential in preeclamptic patients. *Am J Obstet Gynecol* 2012;
206: 1-8.

94. De La Calle M, Delgado JL, Verlohren S, Escudero AI, Bartha JL, Campillos JM, Aguarón De La Cruz A, Chantraine F, García Hernández JÁ, Herraiz I, Llurba E, Kurka H, Guo G, Sillman J, Hund M, Perales Marín A. Gestational age-specific reference ranges for the sFlt-1/PlGF immunoassay ratio in twin pregnancies. *Fetal Diagn Ther* 2021; **48**: 288–296.

95. Perales A, Delgado JL, de la Calle M, García-Hernández JA, Escudero AI, Campillos JM, Sarabia MD, Laíz B, Duque M, Navarro M, Calmarza P, Hund M, Álvarez FV. sFlt-1/PIGF for prediction of early-onset pre-eclampsia: STEPS (Study of Early Pre-eclampsia in Spain). *Ultrasound Obstet Gynecol* 2017; **50**: 373–382.

96. Hayes-Ryan D, Meaney S, Fitzgerald AP, O'Mahony E, Normile C, Kenny LC, O'Donoghue K. A prospective study of placental growth factor in twin pregnancy and development of a dichorionic twin pregnancy specific reference range. *BJOG* 2021; **128**: 411–419.

97. Rana S, Hacker MR, Modest AM, Salahuddin S, Lim KH, Verlohren S, Perschel FH, Karumanchi SA. Circulating angiogenic factors and risk of adverse maternal and perinatal outcomes in twin pregnancies with suspected preeclampsia. *Hypertension* 2012; **60**: 451–458.

Figure 1 Pathophysiology and features of PE^{1, 2}

PE is associated with placental dysfunction and altered maternal sFlt-1 and PIGF concentrations. PE can result in a range of adverse maternal outcomes including maternal hypertension, proteinuria, cerebral edema and liver dysfunction, and a range of adverse fetal outcomes including fetal growth restriction, prematurity and stillbirth.

AT1-AAs, agonistic angiotensin II type 1 receptor autoantibodies; NK, natural killer cells; PE, preeclampsia; PIGF, placental growth factor; sFlt-1, soluble fms-like tyrosine kinase 1. This figure was adapted from Stepan et al. 2020, and originally printed by Wang et al. 2009.

Figure 2 Proposed model for screening, prediction, and monitoring PE in pregnant women²⁰

*FMF combined algorithm for the early identification of women with a high risk of developing PE. The algorithm uses a combination of maternal factors, uterine artery PI, mean arterial pressure and angiogenic factors to create an individualized risk score. BP, blood pressure; FMF, Fetal Medicine Foundation; PAPP-A, pregnancy-associated plasma protein-A; PI, pulsatility index; PE, preeclampsia; PIGF, placental growth factor; sFlt-1, soluble fms-like tyrosine kinase-1. This figure was adapted from Poon et al. 2020.

TABLES

Table 1 Summary of maternal risk factors and recommendations for prevention of PE outlined by the ACOG, NICE and ISSHP guidelines

Guideline	ACOG 2018 ³¹	NICE 2019 ²⁹	ISSHP 2021 ¹⁰	
Maternal risk factors for	PE			
High risk factors	 Diagnosis of PE in a previous pregnancy Multi-fetal gestation Renal disease Autoimmune disease Diabetes mellitus (type 1 or type 2) Chronic hypertension 	 Diagnosis of hypertensive disease during a previous pregnancy Chronic kidney disease Autoimmune disease (e.g., antiphospholipid syndrome) Diabetes mellitus (type 1 or type 2) Chronic hypertension 	 Diagnosis of PE in a previous pregnancy BMI >30 kg/m² Chronic hypertension Pregestational diabetes mellitus Chronic kidney disease (including kidney transplanted women) Systemic lupus erythematosus/ antiphospholipid syndrome Receipt of assisted reproductive treatments 	
Moderate risk factors	 First pregnancy Maternal age ≥35 years BMI >30 kg/m² Family history of PE Sociodemographic characteristics Personal history factors 	 First pregnancy Maternal age ≥40 years Pregnancy interval of >10 years BMI ≥35 kg/m² at first visit Multi-fetal pregnancy 	 First pregnancy Maternal age >40 years Multi-fetal pregnancy Prior placental abruption Prior stillbirth Prior fetal growth restriction 	

Recommendations for prevention of PE

For women at high risk of developing PE, low-dose aspirin (81 mg/day) should be initiated between 12–28 weeks of gestation (optimally before 16 weeks) and should be continued daily until delivery.

Low-dose aspirin should be considered for women with >1 moderate risk factor for PE. Advise pregnant women at high risk of developing PE, or with >1 moderate risk factor for PE, to take 75–150 mg/day aspirin from 12 weeks of gestation until delivery. For women at increased risk of developing PE, low-dose aspirin is recommended, to be taken at bedtime, preferably before 16 weeks of gestation and discontinued by 36 weeks.

After multivariable screening, aspirin should be given at a dose of 150 mg/night.

After screening with clinical risk factors and blood pressure, aspirin should be given at a dose of 100–162 mg/day.

ACOG, American College of Obstetricians and Gynecologists; BMI, body mass index; ISSHP, International Society for the Study of Hypertension in Pregnancy;

NICE, National Institute for Health and Care Excellence; PE, preeclampsia

Diagnose/ predict/rule out PE	PIGF cut-off	Sensitivity (95% CI), %	Specificity (95% CI), %	PPV (95% CI), %	NPV (95% CI), %	Positive likelihood ratio	Negative likelihood ratio	Reference (study)
Diagnose PE in	80–120 pg/mL	78	88	-		(95% CI) 6.3	(95% CI) 0.26	37
asymptomatic women		(67–86)	(75–95)			(2.7–14.7)	(0.16–0.42)	
(Meta-analysis subset, 6 studies, g=various)								
Diagnose PE in asymptomatic women	50–150 pg/mL	74 (64–82)	85 (78–90)	-	-	4.8 (3.1–7.6)	0.31 (0.21–0.45)	37
(Meta-analysis subset, 12 studies, g=various)								
Diagnose PE requiring delivery within 14 days in women with suspected PE (<i>n</i> =287,	<5 th percentile for gestation	96 (89–99)	55 (48–61)	43 (36–51)	98 (93–99.5)	2.1 (1.8–2.5)	0.07 (0.02–0.22)	48 (PELICAN)
g=20+0–<35+0 weeks)								
Diagnose PE requiring	<100 pg/mL	96 (89–99)	56 (49–63)	44 (36–52)	98 (93–99.5)	2.2 (1.9–2.6)	0.07 (0.02–0.22)	48 (PELICAN)

Table 2 PIGF and the sFIt-1/PIGF ratio cut-offs and accuracy for diagnosing and predicting PE and associated adverse outcomes

delivery within 14 days in women with suspected PE (<i>n</i> =287, g=20+0-<35+0 weeks)								
Predict adverse outcome	PIGF cut-off	Sensitivity (95% CI), %	Specificity (95% Cl), %	PPV (95% CI), %	NPV (95% CI), %	Positive likelihood ratio (95% Cl)	Negative likelihood ratio (95% CI)	Reference (study)
Predict delivery of an SGA fetus in women with suspected PE (<i>n</i> =274, g=20+0-34+6 weeks)	<100 pg/mL	93.2 (83.5–98.1)	52.2 (39.7–64.6)	63.2 (52.2–73.3)	89.7 (75.8–97.1)	2.0 (1.5–2.5)	0.13 (0.05–0.34)	⁷⁸ (PELICAN, further analysis)
Predict preterm delivery in women with suspected PE (<i>n</i> =753, g=20+0-35+0 weeks)	≤100 pg/mL	81.7 (NR)	85.3 (NR)	93.5 (NR)	64.5 (NR)	-	-	35
Diagnose/ predict/rule out PE	sFlt-1/PlGF ratio cut-off	Sensitivity (95% CI), %	Specificity (95% CI), %	PPV (95% CI), %	NPV (95% CI), %	Positive likelihood ratio (95% Cl)	Negative likelihood ratio (95% Cl)	Reference (study)

	various	80	92			10.5	0.22	67
Diagnose PE in high and low risk women (Meta- analysis, 15 studies, n=20, 121,	vanous	80 (68–88)*	92 (87–96)*	-	-	10.5 (6.2–18.0)*	0.22 (0.13–0.35)*	(SaPPPhirE)
g=various)								
Rule out early- onset PE (<i>n</i> =1149, g=20+0–33+6 weeks)	≤33⁺	95.0 (89.8–100)	94.0 (90.5–100)	-	-	15.8 (9.13–27.5)	0.05 (0.02–0.13)	57
Rule in early- onset PE (<i>n</i> =1149, g=20+0–33+6 weeks)	≥85⁺	88.0 (81.3–100)	99.5 (97.7–100)	-	-	176 (24.88–1245)	0.12 (0.07–0.21)	57
Rule out late- onset PE (n=1149, g>34+0 weeks)	≤33⁺	89.6 (84.2–100)	73.1 (68.3–100)	-	-	3.33 (2.71–4.10)	0.14 (0.09–0.24)	57
Rule in late- onset PE (<i>n</i> =1149, g>34+0 weeks)	≥110 ⁺	58.2 (50.7–100)	95.5 (92.9–100)	-	-	13 (7.34–23.0)	0.44 (0.36–0.54)	57
Rule out early- onset PE/FGR in women with suspected PE	>95 th percentile	100 (78.5–100)	80.6 (75.0–85.2)	24.1 (15.0–36.5)	100 (97.9–100)	5.2 (4.0–6.7)	-	13

(<i>n</i> =14, g=24+0–28+0 weeks)								
Rule out late- onset PE/FGR in women with suspected PE (<i>n</i> =37, g=24+0–28+0 weeks)	>95 th percentile	40.5 (26.3–56.5)	92.9 (87.7–96.0)	57.7 (38.9–74.5)	86.7 (80.7–91.1)	5.7 (2.9–11.4)	-	13
Rule out PE developing within 1 week of test in women with suspected PE (<i>n</i> =550, g=24+0-36+6 weeks)	≤38	80.0 (51.9–95.7)	78.3 (74.6–81.7)	-	99.3 (97.9–99.9)	-	-	58 (PROGNOSIS)
Rule in PE developing within 4 weeks of test in women with suspected PE (<i>n</i> =550, g=24+0-36+6 weeks)	>38	66.2 (54.0–77.0)	83.1 (79.4–86.3)	36.7 (28.4–45.7)	-	-	-	⁵⁸ (PROGNOSIS)
Rule out PE developing within 1 week	≤38	76.5 (58.8–89.3)	82.1 (79.0–85.0)	17.9 (12.1–25.2)	98.6 (97.2–99.4)	4.28 (3.34–5.48)	0.29 (0.16–0.53)	⁵⁹ (PROGNOSIS Asia)

Accepted Article

of test in women with suspected PE (<i>n</i> =700, g=20+0–36+6 weeks) [‡]								
Rule in PE developing within 4 weeks of test in women with suspected PE (<i>n</i> =700, g=20+0–36+6 weeks) [‡]	>38	62.0 (49.7–73.2)	83.9 (80.8–86.7)	30.3 (23.0–38.5)	95.1 (93.0–96.8)	3.86 (2.99–4.98)	0.45 (0.34–0.61)	⁵⁹ (PROGNOSIS Asia)
Rule out PE developing within 2 weeks of test in women with suspected PE (<i>n</i> =550, g=24+0-36+6 weeks)	≤38	78.0 (62.4–89.4)	81.1 (77.5–84.4)	25.0 (17.8–33.4)	97.9 (96.0–99.0)	4.14 (3.25–5.27)	0.27 (0.15–0.48)	60 (PROGNOSIS post-hoc analysis)
Rule out PE developing within 3 weeks of test in women with suspected PE	≤38	70.0 (56.8–81.2)	82.4 (78.8–85.7)	32.8 (24.8–41.7)	95.7 (93.3–97.5)	3.99 (3.1–5.14)	0.36 (0.25–0.54)	⁶⁰ (PROGNOSIS post-hoc analysis)

(<i>n</i> =550, g=24+0–36+6 weeks)								
Rule out PE developing within 4 weeks of test in women with suspected PE (<i>n</i> =550, g=24+0–36+6 weeks)	≤38	66.2 (54.0–77.0)	83.1 (79.4–86.3)	36.7 (28.4–45.7)	94.3 (91.7–96.3)	3.91 (3.02–5.07)	0.41 (0.29–0.56)	⁶⁰ (PROGNOSIS post-hoc analysis)
Rule in PE developing within 1 week of test in women with suspected PE (<i>n</i> =370, g=24+0-37+0 weeks)	>38	100 (85.8–100)	77.8 (70.6–83.9)	40.0 (27.6–53.5)	100 (97.1–100)	-	-	61 (INSPIRE)
Rule in PE developing within 4 weeks of test in women with suspected PE (<i>n</i> =186, g=24+0/7– 36+6/7 weeks)	≥85	57.1 (39.4–73.7)	94.7 (89.8–97.7)	71.4 (51.3–86.8)	-	-	-	62 (INSPIRE post- hoc analysis)

Predict/rule out adverse outcome	sFlt-1/PlGF ratio cut-off	Sensitivity (95% CI), %	Specificity (95% CI), %	PPV (95% CI), %	NPV (95% CI), %	Positive likelihood ratio (95% Cl)	Negative likelihood ratio (95% Cl)	Reference (study)
Rule out adverse fetal outcomes within 1 week of test in women with suspected of PE (<i>n</i> =690, g=20+0-36+6 weeks) [‡]	≤38	80.0 (61.4–92.3)	81.8 (78.7–84.7)	16.7 (11.0–23.8)	98.9 (97.6–99.6)	4.40 (3.46–5.60)	0.24 (0.12–0.50)	⁵⁹ (PROGNOSIS Asia)
Rule in adverse fetal outcomes within 4 weeks of test in women with suspected of PE (<i>n</i> =690, g=20+0-36+6	>38	61.6 (52.5–70.2)	88.1 (85.2–90.7)	53.5 (45.0–61.8)	91.2 (88.5–93.4)	5.19 (3.99–6.76)	0.44 (0.35–0.55)	⁵⁹ (PROGNOSIS Asia)
weeks) [‡] Predict preterm delivery of an SGA fetus (<i>n</i> =3981, g=28 weeks)	>5.78 ^{§,} **	38.5 (21.1–59.3)	99.1 (98.7–99.3)	21.3 (11.6–35.8).	99.6 (99.3–99.8)	41.1 (23.0–73.6)	0.62 (0.46–0.84)	81

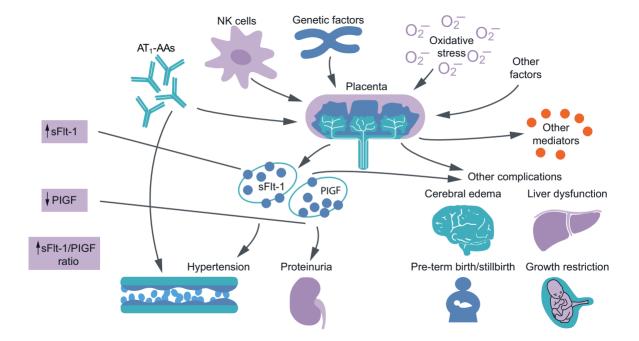
Predict delivery of an SGA fetus associated with PE or perinatal morbidity or mortality (<i>n</i> =3747, g=36 weeks)	>38**	37.9 (26.1–51.4)	97.8 (97.3–98.3)	21.6 (14.5–30.8)	99.0 (98.6–99.3)	17.5 (11.8–25.9)	0.63 (0.52–0.78)	81
Predict adverse outcome in women with PE (<i>n</i> =176, g<34+0 weeks)	≥85	72.9 (NR)	94.0 (NR)	-	87.3 (NR)	-	0.29 (NR)	85

*Pooled sensitivity and specificity values are reported; ⁺The sFlt-1/PIGF ratio cut-off values were derived and validated for the Elecsys sFlt-1/PIGF assay only;

[‡]Gestational week 18+0 in Japan; [§]an sFlt-1/PIGF ratio cut-off of >38 was not used as it would represent an extremely elevated ratio at 28 weeks' gestation; **reported values are for an elevated sFlt-1/PIGF ratio combined with a suspicion of SGA (<10th percentile for gestational age) determined by ultrasound scan measurements.

CI, confidence interval; g, gestational age at time of test; n, number of participants in study; NPV; negative predictive value; NR, not reported; PE,

preeclampsia; PIGF, placental growth factor; PPV, positive predictive value; sFlt-1, soluble fms-like tyrosine kinase 1; SGA, small for gestational age.



Week of pregnancy

0–10 11 12 13	14 15 16 17	18 19 20 21 22 23 2	24 25 26 27 28 29	9 30 31 32 33 34 3	35 36 37 38 39 40		
1 st trimester			2 nd /3 rd trimester				
1 st trimester screening (11–13 weeks)		2 nd trimester risk assessment in all asymptomatic women	Action				
			Following	g 2 nd trimester FMF combined	l algorithm*:		
All pregnant women: • Maternal risk factors • Mean arterial	High risk Aspirin	All patients receive scan and uterine artery Doppler, either as part of FMF combined algorithm* at 20–22 weeks or independently at 20–22 weeks	Low risk: Standard surveillance and reassessment at 35–37 weeks	Intermediate risk: Reassessment of risk at 30–34 weeks	High risk: High level of surveillance with weekly clinic assessment of BP and proteinuria and/or home BP monitoring at 24–31 weeks		
pressure Uterine artery PI 		High-risk patients remain as high risk following 2 nd trimester assessment, which guides frequency	Following sFlt-1 and PIGF measured at 24–28 weeks:				
• PIGF • PAPP-A (if available from			sFlt-1/PIGF ratio ≤38: Standard surveillance	sFlt-1/PIGF ratio >38–85: Increased surveillance	sFlt-1/PIGF ratio ≥85: High level of surveillance (as detailed above)		
aneuploidy screening) in		of monitoring	Low risk based on 1 st and 2 nd trimester screening:				
absence of PIGF • Risk calculation by FMF combined algorithm*	FMF combined Low risk	Low-risk patients with high risk by FMF 2 nd trimester combined algorithm or abnormal uterine artery Doppler	Risk assessment at 36 weeks by either 35–37 week FMF combined algorithm or sFIt-1 and PIGF				
		(PI >95 th percentile) are reclassified as	Clinical suspicion of preeclampsia after 24 weeks				
		high risk	sFlt-1/PIGF ≤38: progression within 1 week unlikely sFlt-1/PIGF >38: progression within 4 weeks likely				