



Near-Complete Genome Sequence of Lötschberg Virus (*Mononegavirales: Filoviridae*) Identified in European Perch (*Perca fluviatilis* Linnaeus, 1758)

Microbiology[®]

Resource Announcements

Torsten Seuberlich,^a ^bJens H. Kuhn,^b Heike Schmidt-Posthaus^c

AMERICAN SOCIETY FOR

MICROBIOLOGY

^aDivision of Neurological Sciences, Vetsuisse Faculty, University of Bern, Bern, Switzerland ^bIntegrated Research Facility at Fort Detrick, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Fort Detrick, Frederick, Maryland, USA ^cInstitute for Fish and Wildlife Health, Vetsuisse Faculty, University of Bern, Bern, Switzerland

ABSTRACT We obtained the near-complete genome sequence of a novel virus, Lötschberg virus (LTBV), from a European perch metatranscriptome. Genome organization and pairwise sequence comparison indicated that LTBV represents a tentative new species and genus of the mononegaviral family *Filoviridae*.

e previously reported the assembly of near-complete genomes of three novel viruses using metatranscriptomic data, derived from pooled organ samples of farmed European perch (Perca fluviatilis Linnaeus, 1758): Fiwi virus (FIWIV), Kander virus (KNDV), and Oberland virus (OBLV) (1). In 2022, these viruses were officially classified by the International Committee on Taxonomy of Viruses (ICTV) as members of the mononegaviral family Filoviridae in novel species Thamnovirus percae, Thamnovirus kanderense, and Oblavirus percae, respectively (2). Also, we found evidence of an additional potentially new virus, represented by four contigs [573 to 3,529 nucleotides (nt) in length]. The predicted encoded protein sequences were 40 to 47% identical to those of Huángjiāo virus (HUJV), a filovirid identified in greenfin horse-faced filefish [Thamnaconus septentrionalis (Günther, 1874)] (1). In that study, we performed high-throughput sequencing (HTS) in paired-end mode (2 \times 150 nt), but reads were unpaired for sequence assembly due to limited computational memory (300 GB RAM) (1). Using upgraded computational memory (2 TB RAM), we reassembled the HTS reads in paired-end mode, which increased the assembly efficiency and resulted in a single near-complete genomic sequence of this virus, which we named Lötschberg virus (LTBV).

Clinically sick European perch were obtained from an aquaculture farm in Switzerland (1). We extracted RNA from pooled organ samples (brain, spleen, kidneys, heart, and pyloric ceca) with TRI reagent (Sigma Life Sciences). HTS libraries were prepared with the TruSeq stranded total RNA kit (Illumina). We sequenced the libraries in paired-end mode, with 150 cycles at a sequencing depth of 2×10^9 on a HiSeq 3000 system (Illumina). Reads were trimmed using fastp (v. 0.12.5; parameters -I 33 -W 4 -M 15 -5 3 -3 3) (3). Host-derived sequences were removed by aligning reads to the European perch genome (GENO_Pfluv_1.0) (4), using STAR (v. 2.7.3a) (5). Nonaligned reads were assembled with SPAdes (v. 3.12.0) (6). The resulting scaffolds were screened for homologs against the National Center for Biotechnology Information (NCBI) nonredundant protein sequence database using the blastx command in Basic Local Alignment Search Tool (BLAST) and DIAMOND (v. 2.0.9) (7), with a default *e*-value cutoff of 0.001. The sequences were manually annotated in Geneious Prime (v. 2023.0.1).

We obtained a scaffold of 13,584 nt (all bases assigned, no ambiguities) with a GC content of 47.7% and an average depth coverage of $12.8 \times$. The sequence contained seven open reading frames (ORFs) (>70 codons), organized similarly to those of filovirid oblaviruses and thamnoviruses, encoding filovirid-typical proteins: nucleoprotein (NP),

Editor Jelle Matthijnssens, Katholieke Universiteit Leuven

Copyright © 2023 Seuberlich et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Torsten Seuberlich, torsten.seuberlich@unibe.ch.

The authors declare no conflict of interest. Received 13 January 2023 Accepted 20 February 2023

	Reported genomic sequence length (nt)	Open reading frame (ORF) length (nt)						
Virus		ORF 1	ORF 2 (NP)	ORF 3 (VP35)	ORF 4	ORF 5 (GP)	ORF 6 (VP30)	ORF 7 (L)
Lötschberg virus	13,584ª	336 ^b	1,200	1,680	294	1,800	1,206	6,215 ^b
Oberland virus	14,682 ^{<i>a</i>}	466 ^b	1,200	1,968	957	1,263	1,152	6,656 ^b
Kander virus	13,849 ^a	654 ^b	1,206	1,665	279	1,791	1,374	6,373 ^b
Fiwi virus	13,764	468	1,203	1,662	276	1,860	1,077	6,435
Huángjiāo virus	14,280	666	1,215	1,686	273	1,881	1,185	6,441

TABLE 1 Comparison of the reported genomic sequence lengths and the length of the open reading frames (ORFs) of Lötschberg virus with those of related fish filovirids

^a Near-complete genome sequence.

^b Incomplete open reading frame.

polymerase cofactor (VP35), glycoprotein (GP_{1.2}), transcriptional activator (VP30), and large protein (L)—plus two new proteins of unknown function. The 3' and 5' termini and the terminal ORFs remained incomplete (Table 1). Comparison of LTBV NP, VP35, GP_{1.2}, VP30, and L amino- acid sequences with the homologous FIWIV, HUJV, KNDV, and OBLV proteins revealed 23 to 48% sequence identity. Pairwise Sequence Comparison (PASC; https://www.ncbi.nlm .nih.gov/sutils/pasc/viridty.cgi?textpage=overview) of near-complete genome sequences indicates that LTBV is most closely related to thamnoviruses (up to 40% PASC similarity), which is also supported by phylogenetic analysis (Fig. 1). Based on the current demarcation criteria for filovirid species (≥23% PASC divergence) and genera (≥55% PASC divergence) (8, 9), LTBV represents a member of a tentative new species and genus within the family Filoviridae.

Data availability. HTS raw data have been deposited in the NCBI Sequence Read Archive (SRA) under accession no. SRR12586223 (https://www.ncbi.nlm.nih.gov/sra/?term= SRR12586223). The genome sequence of LTBV is available at GenBank under accession no. OQ186623 (https://www.ncbi.nlm.nih.gov/nuccore/?term=OQ186623).



^{0.1}

FIG 1 Phylogenetic position of Lötschberg virus (LTBV) in the mononegaviral family Filoviridae. The neighbor-joining tree was built based on near-complete genome sequences after sequence alignment with Clustal Omega (v. 1.2.2, default settings) in Geneious Prime (Dotmatics, v. 2023.0.1) and inferred with the Geneious Consensus Tree Builder setup (Jukes-Cantor model; 5,000 bootstraps). GenBank accession numbers are indicated for each sequence at branch tips.

Genus

Thamnovirus

Oblavirus Striavirus Tapjovirus Marburgvirus

Dianlovirus

Ebolavirus

Cuevavirus

ACKNOWLEDGMENTS

This work was supported by the Swiss Food Safety and Veterinary Office (grant no. MON-108 to T.S.) and the Swiss Innovation Agency (Innossuisse, grant no. 25178.1 PFLS-LS to H.S.-P.). This work was also supported in part through the Laulima Government Solutions, LLC, prime contract with the U.S. National Institute of Allergy and Infectious Diseases under contract no. HHSN272201800013C. J.H.K. performed this work as an employee of Tunnell Government Services, a subcontractor of Laulima Government Solutions, LLC, under contract no. HHSN272201800013C.

The views and conclusions contained in this document are those of the authors and should not be interpreted as necessarily representing the official policies, either expressed or implied, of the U.S. Department of Health and Human Services or of the institutions and companies affiliated with the authors, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. Government.

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

We thank Anya Crane (Integrated Research Facility at Fort Detrick, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Fort Detrick, Frederick, MD, USA) for critically editing the manuscript.

REFERENCES

- Hierweger MM, Koch MC, Rupp M, Maes P, Di Paola N, Bruggmann R, Kuhn JH, Schmidt-Posthaus H, Seuberlich T. 2021. Novel filoviruses, hantavirus, and rhabdovirus in freshwater fish, Switzerland, 2017. Emerg Infect Dis 27: 3082–3091. https://doi.org/10.3201/eid2712.210491.
- 2. Kuhn JH, Adkins S, Alkhovsky SV, Avšič-Županc T, Ayllón MA, Bahl J, Balkema-Buschmann A, Ballinger MJ, Bandte M, Beer M, Bejerman N, Bergeron É, Biedenkopf N, Bigarré L, Blair CD, Blasdell KR, Bradfute SB, Briese T, Brown PA, Bruggmann R, Buchholz UJ, Buchmeier MJ, Bukreyev A, Burt F, Büttner C, Calisher CH, Candresse T, Carson J, Casas I, Chandran K, Charrel RN, Chiaki Y, Crane A, Crane M, Dacheux L, Bó ED, de la Torre JC, de Lamballerie X, de Souza WM, de Swart RL, Dheilly NM, Di Paola N, Di Serio F, Dietzgen RG, Digiaro M, Drexler JF, Duprex WP, Dürrwald R, Easton AJ, Elbeaino T, Ergünay K, Feng G, Feuvrier C, Firth AE, Fooks AR, Formenty PBH, Freitas-Astúa J, Gago-Zachert S, García ML, García-Sastre A, Garrison AR, Godwin SE, Gonzalez JJ, de Bellocq JG, Griffiths A, Groschup MH, Günther S, Hammond J, Hepojoki J, Hierweger MM, Hongō S, Horie M, Horikawa H, Hughes HR, Hume AJ, Hyndman TH, Jiāng D, Jonson GB, Junglen S, Kadono F, Karlin DG, Klempa B, Klingström J, Koch MC, Kondō H, Koonin EV, Krásová J, Krupovic M, Kubota K, Kuzmin IV, Laenen L, Lambert AJ, Lǐ J, Li JM, Lieffrig F, Lukashevich IS, Luo D, Maes P, Marklewitz M, Marshall SH, Marzano S-Y, McCauley JW, Mirazimi A, Mohr PG, Moody NJG, Morita Y, Morrison RN, Mühlberger E, Naidu R, Natsuaki T, Navarro JA, Neriya Y, Netesov SV, Neumann G, Nowotny N, Ochoa-Corona FM, Palacios G, Pallandre L, Pallás V, Papa A, Paraskevopoulou S, Parrish CR, Pauvolid-Corrêa A, Pawęska JT, Pérez DR, Pfaff F, Plemper RK, Postler TS, Pozet F, Radoshitzky SR, Ramos-González PL, Rehanek M, Resende RO, Reyes CA, Romanowski V, Rubbenstroth D, Rubino L, Rumbou A, Runstadler JA, Rupp M, Sabanadzovic S, Sasaya T, Schmidt-Posthaus H, Schwemmle M, Seuberlich T, Sharpe SR, Shi M, Sironi M, Smither S, Song J-W, Spann KM, Spengler JR, Stenglein MD, Takada A, Tesh RB, Těšíková J, Thornburg NJ, Tischler ND, Tomitaka Y, Tomonaga K, Tordo N, Tsunekawa K, Turina M, Tzanetakis IE, Vaira AM, van den Hoogen B, Vanmechelen B, Vasilakis N, Verbeek M, von Bargen S, Wada J, Wahl V, Walker PJ, Whitfield AE, Williams JV, Wolf YI, Yamasaki J, Yanagisawa H, Ye G, Zhang Y-Z, Økland AL. 2022. 2022 taxonomic update of phylum Negarnaviricota (Riboviria:

Orthornavirae), including the large orders *Bunyavirales* and *Mononegavirales*. Arch Virol 167:2857–2906. https://doi.org/10.1007/s00705-022-05546-z.

- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/ bioinformatics/bty560.
- Ozerov MY, Ahmad F, Gross R, Pukk L, Kahar S, Kisand V, Vasemägi A. 2018. Highly continuous genome assembly of Eurasian perch (*Perca fluviatilis*) using linked-read sequencing. G3 (Bethesda) 8:3737–3743. https://doi.org/ 10.1534/g3.118.200768.
- Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR. 2013. STAR: ultrafast universal RNA-seq aligner. Bioinformatics 29:15–21. https://doi.org/10.1093/bioinformatics/bts635.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. J Comput Biol 20:714–737. https://doi.org/10.1089/cmb.2013.0084.
- Buchfink B, Reuter K, Drost H-G. 2021. Sensitive protein alignments at treeof-life scale using DIAMOND. Nat Methods 18:366–368. https://doi.org/10 .1038/s41592-021-01101-x.
- Bào Y, Amarasinghe GK, Basler CF, Bavari S, Bukreyev A, Chandran K, Dolnik O, Dye JM, Ebihara H, Formenty P, Hewson R, Kobinger GP, Leroy EM, Mühlberger E, Netesov SV, Patterson JL, Paweska JT, Smither SJ, Takada A, Towner JS, Volchkov VE, Wahl-Jensen V, Kuhn JH. 2017. Implementation of objective PASC-derived taxon demarcation criteria for official classification of filoviruses. Viruses 9:106. https://doi.org/10.3390/v9050106.
- Kuhn JH, Amarasinghe GK, Basler CF, Bavari S, Bukreyev A, Chandran K, Crozier I, Dolnik O, Dye JM, Formenty PBH, Griffiths A, Hewson R, Kobinger GP, Leroy EM, Mühlberger E, Netesov SV, Palacios G, Pályi B, Pawęska JT, Smither SJ, Takada A, Towner JS, Wahl V, ICTV Report Consortium. 2019. ICTV virus taxonomy profile: *Filoviridae*. J Gen Virol 100:911–912. https://doi.org/10 .1099/jgv.0.001252.