


REVIEW

Impact of estrogens on resting energy expenditure: A systematic review

Susanna Weidlinger¹  | Katja Winterberger¹ | Janna Pape¹ |
Magdalena Weidlinger² | Heidrun Janka³ | Michael von Wolff¹ | Petra Stute¹

¹Department of Obstetrics and Gynecology, University Hospital of Bern, Bern, Switzerland

²Department of Obstetrics and Gynecology, Hospital Thun, Thun, Switzerland

³Medical Library, University Library Bern, University of Bern, Bern, Switzerland

Correspondence

Susanna Weidlinger, Department of Obstetrics and Gynecology, University Hospital of Bern, 3010 Bern, Switzerland.

Email: susanna.weidlinger@insel.ch

Summary

The fear of weight gain is one of the main reasons for women not to initiate or to early discontinue hormonal contraception or menopausal hormone therapy. Resting energy expenditure is by far the largest component and the most important determinant of total energy expenditure. Given that low resting energy expenditure is a confirmed predictive factor for weight gain and consecutively for the development of obesity, research into the influence of sex steroids on resting energy expenditure is a particularly exciting area. The objective of this systematic review was to evaluate the effects of medication with natural and synthetic estrogens on resting energy expenditure in healthy normal weight and overweight women. Through complex systematic literature searches, a total of 10 studies were identified that investigated the effects of medication with estrogens on resting energy expenditure. Our results demonstrate that estrogen administration increases resting energy expenditure by up to +208 kcal per day in the context of contraception and by up to +222 kcal per day in the context of menopausal hormone therapy, suggesting a preventive effect of circulating estrogen levels and estrogen administration on weight gain and obesity development.

KEYWORDS

estrogen, hormonal contraception, menopausal hormone therapy, resting energy expenditure

1 | INTRODUCTION

1.1 | Obesity, basic facts, and considerations

Obesity shows a concerning high and, most importantly, continuously increasing prevalence. The proportion of overweight (BMI \geq 25 kg/m²) people aged 18 years and older worldwide has almost doubled in the last 40 years: In 1975, 21.5% of the adult population was

estimated to be overweight; by 2016, this number had risen to 38.9%. The proportion of obese (BMI \geq 30 kg/m²) adults had even tripled, from an estimation of 4.7% in 1975 to 13.1% in 2016.¹ Obesity is a commonly known major risk factor for morbidity and mortality,^{2–5} which may even lead to a stagnation or decline in life expectancy.⁶

The fear of unwanted weight gain is one of the main reasons for women not to initiate or to early discontinue hormonal contraception

Abbreviations: AMPK, AMP-activated protein kinase; ATP, adenosine triphosphate; BAT, brown adipose tissue; CEE, conjugated equine estrogens; CNS, central nervous system; DEE, dietary energy expenditure; E2, estradiol; EE, ethinyl estradiol; EI, energy intake; ER, endoplasmic reticulum; ER α , estrogen receptor α ; HC, hormonal contraception; MHT, menopausal hormone therapy; OC, oral contraceptive; PAEE, physical activity energy expenditure; REE, resting energy expenditure; SNS, sympathetic nervous system; TEE, total energy expenditure; UCP1, uncoupling protein 1; VMH, ventromedial hypothalamus; WAT, white adipose tissue; β 3 AR, β 3 adrenergic receptor.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. *Obesity Reviews* published by John Wiley & Sons Ltd on behalf of World Obesity Federation.

(HC) or menopausal hormone therapy (MHT),⁷⁻¹¹ According to current data, however, this concern seems to be unfounded. Neither a sex hormone-related significant weight-increasing nor weight-reducing effect has been clearly confirmed or refuted. Cochrane analyses on the topic of the influence of combination contraceptives,¹² progestin-only contraceptives,¹³ and MHT¹⁴ on body weight showed no association between sex hormone use and weight changes. However, the duration of hormone administration in the majority of the studies included in these Cochrane analyses was probably too short to detect significant effects of sex hormone medication on body weight. Overall, the majority of studies lack clear evidence to definitively rule out the possibility of sex hormone-induced weight change.

Obesity is the result of a chronic disturbance in energy homeostasis caused by an imbalance between energy intake (EI) and total energy expenditure (TEE). The term “energy intake” is almost self-explanatory and refers to the intake of energy through macronutrients (carbohydrates, proteins, fats) and alcohol, whereas the term “total energy expenditure” is a more complex one and consists of three main components: first, resting energy expenditure (REE); second, dietary energy expenditure (DEE, also called “thermic effect of food” or “diet-induced thermogenesis”); and third, physical activity energy expenditure (PAEE).¹⁵⁻¹⁷ REE represents the largest component of TEE (60%–80%¹⁸⁻²⁰) and is defined as the metabolic rate required to maintain vital physiological functions from the cellular to the systemic level of an organism that is awake, in mental and physical rest, fasting, and in a thermoneutral environment.¹⁵⁻¹⁷ DEE describes the energy required for the digestion, absorption, and storage of ingested nutrients. For individuals consuming an average mixed diet, DEE amounts to approximately 10% of TEE.^{15,16,18,21} PAEE is the most variable component of TEE and can be significantly increased by intentional physical exercise. Given the typically sedentary lifestyle of Western societies, it averages 25%–35%²² of TEE ranging from 15% in very inactive individuals to up to 50% in highly active individuals.¹⁸

Both a reduction in EI and an increase in PAEE are necessary for sustainable weight loss but are difficult to implement in the long term. For this reason, research has recently focused increasingly on the possibility of achieving a negative energy balance by increasing REE.

1.2 | Resting energy expenditure and basal proton leak

As mentioned above, REE is defined as the energy required by an organism to maintain homeostasis.¹⁵⁻¹⁷ The majority of the energy required for REE is generated through the breakdown of organic substrates in the course of cellular respiration in the mitochondria. The goal of cellular respiration is to produce adenosine triphosphate (ATP), the universal energy source for cellular functions. The main part of ATP production takes place in the respiratory chain, the final step of cellular respiration. However, the coupling of oxidative processes to ATP production has limited efficiency. Part of the energy of the proton motive force is “lost” through a basal proton leak at the inner mitochondrial membrane and is thus uncoupled from ATP production

and directly converted into heat. Hence, REE is the sum of the energy required for ATP production (about 70%–80% of REE) on the one hand, and the energy converted into heat through the basal proton leak (20%–30% of REE), on the other.²²⁻²⁸ The latter enables the organism to maintain body temperature in a thermoneutral situation without the need to actively initiate other homeostatic thermoregulatory mechanisms.^{29,30} The basal proton leak accounts for a significant proportion of this so-called obligatory thermogenesis.^{27,31,32} Still, the exact mechanism behind this process remains controversial.³³ However, reinforcing the process of “wasting” energy through proton leaks represents a possibility to modulate the energy balance toward the negative and is therefore considered a potential and promising target for the treatment of obesity. Whereas the basal mitochondrial proton leak cannot be intentionally modulated, there is an inducible/adaptive/facultative proton leak exclusively in brown adipose tissue (BAT) mitochondria, which, in contrast to the basal proton leak, can directly be regulated due to its dependence on the activity of the tissue-specific so-called uncoupling protein 1 (UCP1).^{29,30}

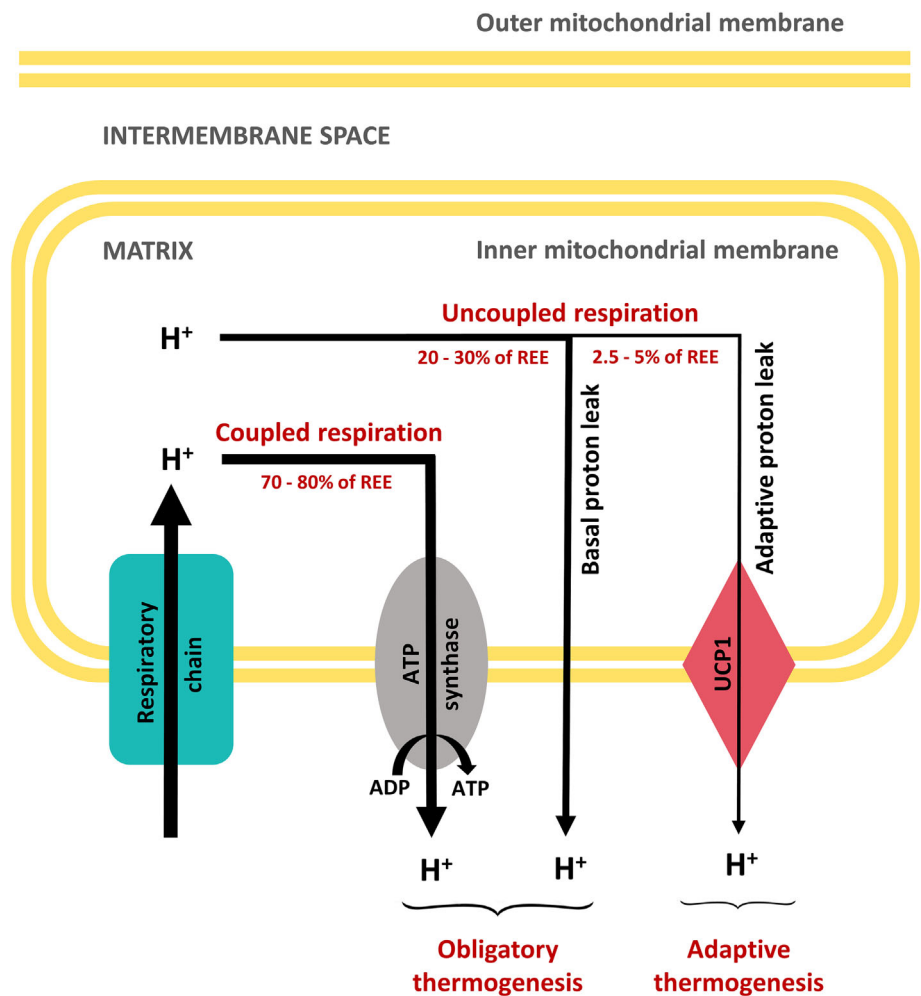
1.3 | Brown adipose tissue and adaptive proton leak

Mammals have two types of adipose tissue that play opposite roles in the body's energy metabolism: white lipid-storing adipose tissue (WAT) and brown lipid-burning adipose tissue. WAT cells contain a single large lipid vacuole and a few mitochondria, whereas BAT cells are characterized by multilocular small lipid droplets and a large number of well-developed mitochondria. WAT not only has the ability to store excess energy in the form of triglycerides and release this energy as needed in the form of free fatty acids but, in recent years, has also gained an expanded role as a metabolically and endocrinologically highly active organ. Through the release of various biochemically active substances, such as adipokines, it is involved in communication between cells, tissues, and organs.³⁴

In contrast to energy storage, the main function of BAT is to generate heat on demand through the oxidation of free fatty acids, thus consuming energy. This process is part of the so-called adaptive thermogenesis (synonyms: facultative or nonshivering thermogenesis).^{29,30,35,36}

It is the mass of mitochondria in the BAT cells that enables this form of energy release and gives this particular fat tissue its characteristic brown color. In contrast to the regular mitochondria, the BAT mitochondria are characterized by the ability to actively uncouple cellular respiration from the synthesis of ATP and consequently to convert the energy released during fatty acid oxidation directly into heat, circumventing ATP production. This biochemical short circuit is made possible by the unique expression of UCP1, an inducible ion channel at the inner mitochondrial membrane that allows the re-entry of protons, from the intermembrane space into the mitochondrial matrix, bypassing the membrane-bound ATP synthase and thus dissipating the energy of the proton gradient directly into heat, as described previously (Figure 1).^{25,29,35}

FIGURE 1 Obligatory versus adaptive thermogenesis. The goal of cellular respiration is the production of adenosine triphosphate (ATP), the universal energy source for cellular functions. Only 70%–80% of the energy of the proton motive force is used for ATP production. 20%–30% of this energy is “lost” through a basal proton leak at the inner mitochondrial membrane and is converted directly to heat, thus uncoupling it from ATP production. These two processes together are referred to as obligatory thermogenesis. Although the basal mitochondrial proton leak cannot be intentionally modulated, there is also an adaptive proton leak that, unlike the basal proton leak, can be directly regulated because of its dependence on the activity of the so-called uncoupling protein 1 (UCP1) at the inner mitochondrial membrane. Thus, cellular respiration can be actively uncoupled from ATP synthesis when needed, and the energy released during cellular respiration can be directly converted to heat bypassing ATP production. This process is called adaptive thermogenesis.



The concept of additional stimulation of this energetically wasteful tissue with the resulting increase in energy expenditure and its potential influence to counteract the development of obesity and related diseases has kept scientists busy for more than 40 years.³⁷ In fact, in animals (rodents), the enhancement of BAT thermogenesis to maintain healthy body weight is now well established.³⁸⁻⁴³ In humans, however, it has long been assumed that only infants possess a physiologically relevant amount of BAT.^{29,44} Following the recent rediscovery of metabolically active BAT also in adult humans,⁴⁵⁻⁴⁹ its experimental use has once again attracted increased interest in clinical and non-clinical studies.^{38,50-54}

BAT thermogenesis is controlled by the sympathetic nervous system (SNS). The ventromedial nucleus of the hypothalamus (also referred to as the ventromedial hypothalamus, VMH) is the central key center responsible for the regulation of BAT thermogenesis and thus is superior to the SNS.^{29,55-58} The physiological stimulus to activate the VMH-SNS-BAT axis is cold.^{45,46,49,59-62} Subsequent release of norepinephrine from the SNS leads to activation of BAT via binding to its β_3 adrenergic receptors (β_3 AR). This is followed by an increase in the activity of intracellular lipolytic enzymes and the breakdown of cytosolic triglycerides to free fatty acids, which are directed into the mitochondria, where they both allosterically disinhibit UCP1 present in the inner mitochondrial membrane and themselves serve as fuel for fatty acid oxidation. Simultaneously, glucose

and fatty acid uptake from the periphery into BAT cells is increased to replenish intracellular lipid stores.^{29,30,35,49} Hence, the BAT is the only tissue that can literally “burn fat.”

However, stimulation of the BAT by the SNS leads not only to an increase in its metabolic activity but also to an expansion of its volume as well as to the so-called “browning,” which refers to the recruitment of brown adipocytes by differentiation from brown adipocyte precursors/preadipocytes in WAT or by transdifferentiation of existing white adipocytes. These stimulation-induced thermogenically competent cells in WAT are referred to as brite (brown-in-white) or beige adipocytes. As the name implies, they exhibit phenotypic and functional features common to white and brown adipocytes. In summary, BAT is a flexible tissue that can be recruited by specific triggers and atrophies in their absence.^{35,38,53,56,58,63,64}

There is growing evidence that BAT volume and activity in humans are associated with increased energy expenditure,^{46,51,65-67} decreased BMI^{45-47,68-71} as well as improved glucose,⁷²⁻⁷⁵ and lipid metabolism.^{73,75} Thus, people with a higher volume of metabolically active BAT also appear to be more likely to be protected from developing obesity and its related diseases.

Because more or less intensive, intermittent as well as continuous cold stimuli are not tolerable for humans in the long run to provide continuous activation of the BAT, other substances have been

investigated. Hormonal substances include adrenal steroids,⁷⁶ thyroid hormones,^{77–80} insulin,^{81,82} and estradiol.^{83–85}

1.4 | Estrogens and energy metabolism

The function of estrogens as important regulators of energy balance is well established.^{86–88} Using the example of ovarian insufficiency, for example, at menopause, as a consequence of ovariectomy or ovarian function suppression, it has been sufficiently demonstrated that low estrogen levels are associated with increased EI⁸⁷ and decreased PAEE,⁸⁹ hence leading to weight gain and promoting the development of obesity in the long term,^{87,90–98} with MHT counteracting this development.^{99–101} Furthermore, several studies in premenopausal women show an increase in REE during the luteal phase, when sex steroid levels (estradiol and progesterone) are elevated, compared with the follicular phase,^{102–108} when sex steroid levels are low, suggesting a role for sex steroids, particularly estrogens,¹⁰⁹ in the regulation of REE. In contrast, other studies have found no difference in REE at different phases of the menstrual cycle.^{110–113} Consequently, if physiological variations of estrogens affect REE, it is reasonable to assume that exogenous administration of these hormones also has an impact on REE. However, data on this are scarce.

1.5 | Estradiol and its central and peripheral effects on brown adipose tissue thermogenesis

Estrogen receptor α (ER α), expressed by both the central nervous system (CNS) and adipose tissue, is considered the key mediator of estrogen action on energy homeostasis.^{86,87,114} With respect to energy expenditure, estrogens primarily exert both central and peripheral stimulatory effects on BAT thermogenesis.^{84–86,115} Consistent with this is the observation of a sexual dimorphism in the BAT.¹¹⁶ As far as is currently known, women exhibit greater BAT mass and are more responsive to BAT-activating stimuli than men.^{47,117–120} Moreover, these sex differences disappear after menopause.^{121,122}

In the CNS, estrogens exert their multiple functions in energy balance by binding to ER α , which is expressed at a variety of sites, predominantly in the hypothalamus.^{123–125} The most important actions of estrogens on energy expenditure, in particular on BAT thermogenesis, take place nucleus-specifically in the VMH. Any electrical, pharmacological, and hormonal stimulation of this nucleus increases sympathetic outflow to the BAT via intermediate interconnections in brainstem areas such as the nucleus raphe pallidus and the inferior olive, thereby increasing BAT thermogenesis.^{55,58,83,126–129}

Among the major molecular mechanisms mediating the action of estrogens in the hypothalamus is the inhibition of hypothalamic AMP-activated protein kinase (AMPK).⁸³ AMPK is an enzyme that plays a key role in regulating cellular processes in situations of cellular energy deficiency. Central activation of AMPK leads to an increase in energy production, that is, synthesis of ATP, and a decrease in energy wasting, ensuring the maintenance of cellular energy homeostasis.^{130–134} Thus, centrally, estrogens lead to increased sympathetic tone via central inhibition of AMPK, with subsequent activation of BAT thermogenesis and

energy “wasting.”⁸³ Other substances known for their effects on metabolic homeostasis and whose common pathway is to increase BAT-specific energy expenditure via AMPK inhibition in the VMH include leptin, thyroid hormones, nicotine, bone morphogenetic protein 8b, and glucagon-like peptide 1 and its analogs.^{78,126,135–137} Finally, recent data have shown that not only BAT activation but also the browning of WAT is regulated via AMPK inhibition in the VMH.¹¹⁵

Another key mechanism by which estrogens lead to an increase in sympathetic tone and thus activation of BAT and energy wasting is through modulation of the lipotoxic effects of reactive lipid species in the VMH. The deposition of lipophilic metabolites such as ceramides in the VMH associated with overeating, also known as hypothalamic lipotoxicity, leads to the impairment of various cellular functions such as the triggering of stress in the endoplasmic reticulum (ER). The result is reduced sympathetic tone, which leads to reduced BAT thermogenesis and food-independent weight gain.¹³⁸ It has been demonstrated that centrally administered estradiol (E2) can counteract the central dysregulation of the energy metabolism triggered by the accumulation of lipid metabolites in the hypothalamus. The binding of E2 to ER α results in a selective reduction of ceramide-induced lipotoxicity and consequently of ER stress in neuronal cells of the VMH, which in turn leads to a markedly increased BAT activity, weight loss, and improvement of glucose metabolism.^{139,140}

Peripherally estrogen modulation of BAT occurs by a direct action on ER α in adipocytes. As in the CNS, the effect of estrogens in adipocytes is mediated by AMPK. In contrast to central regulation, in the periphery, AMPK is activated by the binding of estrogen to its receptor. As in BAT triggered by the binding of norepinephrine to the β 3-AR, activation of ER α in WAT leads to intracellular lipolysis with release of free fatty acids followed by disinhibition of UCP1. Furthermore, estrogen activity in white adipocytes leads to the expression of BAT-specific genes (e.g., of β 3 AR and other markers for mitochondrial biogenesis) and consequently to browning (Figure 2).¹⁴¹

The mechanisms underlying estrogen-induced BAT thermogenesis described herein suggest promising clinical effects on REE, body weight development, and glucose and lipid metabolism. To date, results on this topic have almost exclusively come from animal studies, whereas findings in humans are still sparse and inconclusive.

Thus, the aim of this systematic review was to evaluate the effects of medication with natural and synthetic estrogens on REE in healthy normal weight and overweight (BMI 18.5–29.9 kg/m²) women.

2 | METHODS

2.1 | Information sources and search strategy

The systematic review was registered in PROSPERO, an international database of prospectively registered systematic reviews. To identify all potentially relevant documents on the topic, complex literature searches were designed and executed by a medical information specialist (HJ) according to the guidelines of PRISMA 2020¹⁴² and PRISMA-Search¹⁴³ in the following information sources: Medline

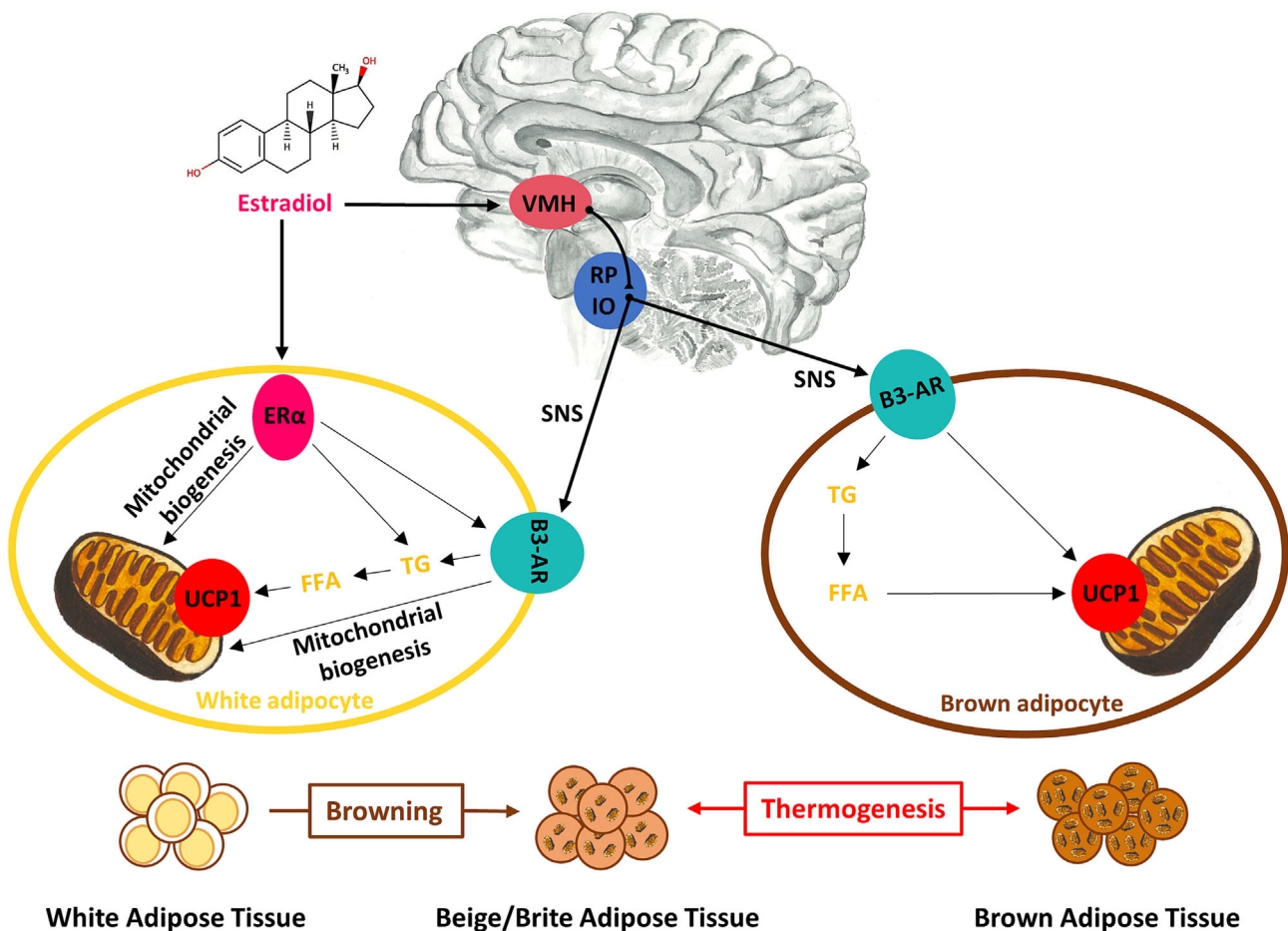


FIGURE 2 Estradiol and its central and peripheral effects on brown adipose tissue thermogenesis. Binding of estradiol (E2) to estrogen receptor α (ER α), expressed by both the central nervous system (CNS) and the adipose tissue, exerts stimulatory effects on brown adipose tissue (BAT) thermogenesis. Centrally, estradiol leads to increased sympathetic nervous system (SNS) activity by binding to its receptor in the ventromedial hypothalamus (VMH) and via intermediate connections in brainstem areas such as the nucleus raphe pallidus (RP) and the inferior olive (IO). Ultimately, triggered by the binding of noradrenergic to the β 3 adrenergic receptor (β 3 AR), intracellular lipolysis occurs in the BAT and white adipose tissue (WAT) with release of free fatty acids (FFA) from triglycerides (TG), followed by disinhibition of UCP1. However, stimulation of the BAT by the SNS not only leads to an increase in its metabolic activity but also to an expansion of its volume as well as to the so-called “browning” of WAT. In the periphery, estradiol leads directly to metabolic activation and browning of WAT via binding to the ER α .

(1946–March 16, 2021), Embase (1974–March 16, 2021), CINAHL (1937–March 16, 2021), Cochrane Library (1996–March 16, 2021), Web of Science (1900–March 16, 2021), and [ClinicalTrials.gov](https://www.clinicaltrials.gov) (NLM).

The searches were conducted on March 17 and 18, 2021. The search concepts used were (1) “women” (of all ages), (2) “resting energy expenditure”/“basal metabolic rate,” and (3) “gonadal steroids” (including androgens, estrogens, and progestogens as well as natural and synthetic hormones). In addition to electronic database searching, reference lists and bibliographies from relevant publications were checked for relevant studies. The final detailed search strategies are presented in the [Supporting Information](#).

2.2 | Study selection process

The search strategy yielded a total of 973 articles published between 1929 and 2021. After the removal of duplicates by HJ 429, unique

articles were identified. Titles, abstracts, and full texts were double-screened independently by the authors SW and KW and tested against the inclusion criteria. Disagreements between individual judgments were resolved in a bilateral discussion. All English-, German-, or French-language clinical trials that investigated the effects of medication with natural and synthetic estrogens on REE in healthy normal-weight and overweight (BMI 18.5–29.9 kg/m²) women were included (Figure 3).

2.3 | Data extraction

Relevant information from the included studies was collected for data extraction according to a predesigned protocol established by the two reviewers (SW and KW). For all studies, predefined study data (see Tables 1 and 2) were collected independently by the two reviewers (SW and KW), tabulated according to the PICO elements,

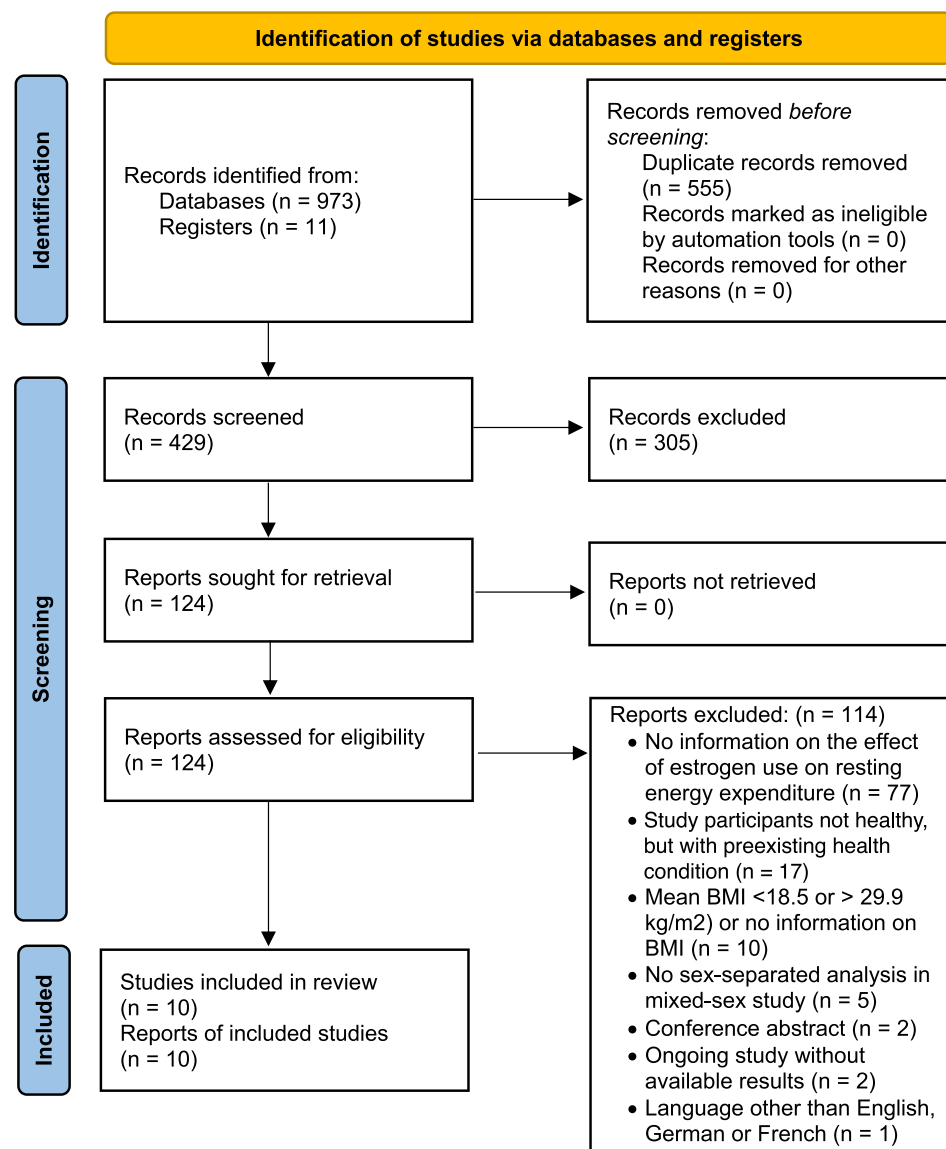


FIGURE 3 PRISMA 2020 flow diagram.

among others, and then cross-checked against each other to ensure correct data collection. The main result of interest, the change in REE as a result of estrogen medication, was expressed variously in the different studies, either in absolute numbers, in *p*-values to indicate the significance of differences, or in purely literal textual terms.

2.4 | Data analysis and risk of bias assessment

The risk of bias was assessed for all included randomized controlled trials (RCTs) using the Cochrane Risk of Bias 2 (RoB 2) tool¹⁴⁴ and for all other included non-RCTs using the Thomas H. Quality assessment tool for quantitative studies,¹⁴⁵ which is independent of study design. All included studies were reviewed by two of the review authors (SW and KW) to independently assess the risk of bias (assessment of methodological quality). Disagreements were resolved by bilateral consensus. With the exception of the study by Eck et al.¹⁴⁶ in which the methodological quality was rated as strong, corresponding to a

low risk of bias, the methodological quality of all other studies^{147–154} was only rated as moderate (Tables 3 and 4).

3 | RESULTS

3.1 | Characteristics of included articles

Our systematic review identified a total of 10 studies investigating the effects of medication with natural and synthetic estrogens on REE and encompassing 461 healthy normal-weight and overweight women (Tables 1 and 2).^{146–155} The 10 included studies consisted of six studies in which estrogens were used in the context of HC^{146,149–152,154} and four studies in which estrogens were used in the context of MHT.^{147,148,153,155} Study designs were either RCTs,^{150,151,153} prospective cohort studies,¹⁴⁷ or cross-sectional studies.^{146,148,149,152,154,155} The sample size of each cohort with the population of interest ranged from 6¹⁵⁵ to 99 subjects.¹⁵² The type of

TABLE 1 Overview over the basic characteristics of the included studies.

Author, year of publication, journal	Country	Title	Study design	Primary objective	Population of interest	Groups of interest	Number of participants	Age of participants, years (mean \pm SD)	BMI, kg/m ² (mean \pm SD)
Duhita et al., 2019, Nutrients	Switzerland, New Zealand	Assessment of the Dose-Response Relationship between Meal Protein Content and Postprandial Thermogenesis: Effect of Sex and the Oral Contraceptive Pill	RCT	Effect of sex and combined monophasic OC use on the dose-response relationship between meal protein content and dietary-induced thermogenesis, respiratory quotient, hunger, and satiety	Healthy, premenopausal women	OC users OC non-users	7 7	22.6 \pm 0.6 24.4 \pm 0.9	21.0 \pm 0.6 22.8 \pm 0.7
Duhita et al., 2017, Obesity	Switzerland	Oral Contraceptive Pill Alters Acute Dietary Protein-Induced Thermogenesis in Young Women	RCT	Effect of menstrual cycle phase and combined monophasic OC use on the dose-response relationship between meal protein content and dietary-induced thermogenesis	Healthy, premenopausal women	OC users OC non-users	8 8	23 \pm 1 24 \pm 1	21.9 \pm 1.2 22.0 \pm 0.6
Melanson et al., 2015, J Appl Physiol	USA	Regulation of energy expenditure by estradiol in premenopausal women	RCT	Effects of sex hormone suppression with GnRH-Ag with estradiol vs. placebo add-back therapy for 5 months on total daily energy expenditure and its components	Healthy, premenopausal women	GnRH-Ag + MHT GnRH-Ag + placebo	21 24	33 \pm 9 37 \pm 8	26.9 \pm 6.2 27.3 \pm 6.2

(Continues)

TABLE 1 (Continued)

Author, year of publication, journal	Country	Title	Study design	Primary objective	Population of interest	Groups of interest	Number of participants	Age of participants, years (mean \pm SD)	BMI, kg/m ² (mean \pm SD)
Aubertin-Leheudre et al., 2008, J Aging Phys Act	Canada	Enhanced Rate of Resting Energy Expenditure in Women Using Hormone-Replacement Therapy: Preliminary Results	Cross-sectional study	Effect of long-term MHT treatment (>1 year) on total daily energy expenditure and its components as well as, body composition in actual MHT-users vs. MHT never-users	Healthy, postmenopausal women	MHT users MHT never-users	13 13	59.2 \pm 3.1 59.6 \pm 2.7	24.1 \pm 2.6 23.7 \pm 2.9
Reimer et al., 2005, Physiol Behav	Canada	Dietary and metabolic differences in premenopausal women taking or not taking hormone replacement therapy	Cross-sectional study	Dietary intake analysis in premenopausal women during the luteal and follicular phases as well as in postmenopausal MHT users and never-users	Healthy, postmenopausal women	MHT users MHT never-users	6 7	54.5 \pm 1.0 53.4 \pm 1.7	26.3 \pm 2.0 22.9 \pm 1.2
Anderson et al., 2001, Metabolism	USA	Body Composition and Energy Balance: Lack of Short-Term Hormone Replacement in Postmenopausal Women	Prospective cohort study	Effect of physiologic MHT on body weight, body composition, and energy balance	Healthy, postmenopausal women	45–55-year-old postmenopausal women before and after initiation of MHT 70–80-year-old postmenopausal women before and after initiation of MHT	18 15	52.4 \pm 0.6 72.3 \pm 0.6	25.0 \pm 0.8 26.2 \pm 1.0
Kimm et al., 2001, Am J Epidemiol	USA	Effects of race, cigarette smoking, and use of contraceptive medications on resting energy expenditure in young women	Cross-sectional study	Assessment of racial differences in the effects of cigarette smoking, use of contraceptive medication use, and menstrual cycle on REE in a biracial cohort	Healthy, premenopausal women	Contraception users Contraception non-users	48 99	19.4	24.6 \pm 5.0 (white women); 27.8 \pm 7.18 (black women)

TABLE 1 (Continued)

Author, year of publication, journal	Country	Title	Study design	Primary objective	Population of interest	Groups of interest	Number of participants	Age of participants, years (mean \pm SD)	BMI, kg/m ² (mean \pm SD)
Piers et al., 1997, Eur J Clin Nutr	Australia	The validity of predicting the basal metabolic rate of young Australian men and women	Cross-sectional study	Assessment of the accuracy of different equations for the prediction of the basal metabolic rate of young Australians	Healthy premenopausal women	OC users OC non-users	42 47	24 \pm 3 23 \pm 2	21.7 \pm 2.0 22.2 \pm 2.6
Eck et al., 1997, Eur J Clin Nutr	USA	Differences in macronutrient selections in users and nonusers of an oral contraceptive	Cross-sectional study	Effect of menstrual cycle phase and OC use on energy balance and its components	Healthy premenopausal women	OC users OC non-users	17 15	21.7 \pm 3.2 19.9 \pm 3.1	22.2 \pm 2.3 20.8 \pm 2.8
Diffey et al., 1997, Br J Nutr	Australia	The effect of oral contraceptive agents on the basal metabolic rate of young women	Cross-sectional study	Effect of OC use on REE in young women	Healthy premenopausal women	OC users OC never-users	24 22	26 \pm 3 25 \pm 3	21.8 \pm 2.4 22.3 \pm 2.2

Abbreviations: GnRH-Ag, gonadotropin-releasing hormone agonist; MHT, menopausal hormone therapy; OC, oral contraceptive; RCT, randomized controlled trial; REE, resting energy expenditure.

TABLE 2 Overview of the results of interest of the included studies.

Author, year of publication, Journal	Groups of interest	Interventions/ medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
Duhita et al., 2019, Nutrients	OC users	OCs used: comb. monophasic OC formulations differed in terms of EE concentration (mean: $2.6 \pm 3 \mu\text{g/d}$) and progestin type and dose	IC. Calc. of baseline REE from the av. of 3 meas's. of REE on 3 different d in the week following withdrawal with an interval of at least 2 d between each meas.	0.89 ± 0.03 kcal/min	-	$p = 0.74$ (between groups)	-	Δ OC users vs. OC non-users: -14 kcal/ $d \triangleq -1.1\%$; $p = 0.74$
	OC non-users	-	IC. Calc. of baseline REE from the av. of 3 meas's. of REE on 3 different d in follicular phase with an interval of at least 2 d between each meas.	0.90 ± 0.03 kcal/min	-	-	-	
Duhita et al., 2017, Obesity	OC users	OCs used: comb. monophasic OC formulations differed in terms of EE concentration (EE $20 \mu\text{g/d}$; $n = 4$, EE $30\text{--}35 \mu\text{g/d}$; $n = 4$); no information on type and dose of progestin	IC. Calc. of baseline REE from the av. of 2 meas's. of REE on cycle d 5–12 and 2 meas's. on cycle d 19–26 with an interval of at least 2 d between each meas.	-	-	$p = 0.6$ (between groups)	-	No effect of OC use on REE.
	OC non-users	-	IC. Calc. of baseline REE from the av. of 2 meas's. of REE in follicular phase (cycle d 5–12) and 2 meas's. in luteal phase (cycle d 19–26) with an interval of at least 2 d between each meas.	-	-	-	-	

TABLE 2 (Continued)

Author, year of publication, Journal	Groups of interest	Interventions/medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
Melanson et al., 2015, J Appl Physiol	GnRH-Ag + E2 (+ MPA)	Leuprolide acetate 3.75 mg i.m. monthly, E2 75 μ g/d-transdermal patch + MPA 5 mg/d orally for 12 d every other month (end of month 2 and 4)	IC. Meas. of pre-treatment REE in follicular phase (cycle d 2–6) in most women. Meas. of post-treatment REE after 5 months of sex hormone suppression with E2 add-back therapy.	1456.1 \pm 167.9 kcal/d	1462.6	\pm 169.8 kcal/d	n.s. (within-group) $p < 0.05$ (between groups)	$p < 0.05$ (between-groups), adj. for changes in FM and FFM
GnRH-Ag + placebo	Δ within GnRH-Ag + E2: +6 kcal/d; n.s. Δ within GnRH-Ag + placebo: -54 kcal/d Δ $\hat{=}$ -3.7%; $p \leq 0.05$ Δ GnRH-Ag + placebo vs. GnRH-Ag + E2: -60 kcal/d Δ $\hat{=}$ -4.1%; $p \leq 0.05$	Leuprolide acetate 3.75 mg i.m. monthly + placebo patch for 5 months	IC. Meas. of pre-treatment REE in follicular phase (cycle d 2–6) in most women. Meas. of post-treatment REE after 5 months of sex hormone suppression.	1455.4 \pm 181.4 kcal/d	1401.1	\pm 167.9 kcal/d	$p < 0.05$ (within group) $p < 0.05$ (between groups)	
Aubertin-Leheudre et al., 2008, J Aging Phys Act	MHT users	Actual MHT use for at least 1 year; oral: $n = 11$, transdermal: $n = 2$; CEE 0.625 mg/d; $n = 2$, CEE 0.625 mg/d + MPA 2.5 mg/d; $n = 11$; no information on the regime; av. treatment time of MHT 8.8 years (range 4–12 years)	IC. No further details on the calc. of baseline REE.	1392 \pm 228 kcal/d	-	$p = 0.038$ (between groups)	$p = 0.05$ (between groups), adj. for total daily energy expenditure and time since menopause	Δ MHT users vs. MHT never-users: +222 kcal/d; $p = 0.038$

(Continues)

TABLE 2 (Continued)

Author, year of publication, Journal	Groups of interest	Interventions/medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
Reimer et al., 2005, Physiol Behav	MHT never-users	-	IC. No further details on the calc. of baseline REE.	1170 \pm 283 kcal/d	-	n.s.	-	No effect of MHT on REE.
	MHT users	Actual MHT use for at least 1 year; estrogen and progestin; no information on type and dose of estrogen and progestin, regime, and type of application; av. 7 years since menopause	IC. No further details on the calc. of baseline REE.	-	-	n.s.	-	No effect of MHT on REE.
Anderson et al., 2001, Metabolism	MHT never-users	Av. 4 years since menopause	IC. No further details on the calc. of baseline REE.	-	-	-	-	-
	Postmenop. women of 45–55 years, before and after initiation of MHT	First month of MHT: E2 50 μ g/d transdermal patch; second month of MHT: E2 50 μ g/d transdermal patch + vaginal progesterone 200 mg/d for the final 7 d	IC. No further details on the calc. of REE pre-treatment, after 1 and 2 months of treatment.	6424 \pm 170 kJ/d	E2: 6881 \pm 232 kJ/d E2 + P: 6238 \pm 173 kJ/d	n.s. (within group)	n.s. (within group), adj. for FFM	No effect of E2 or E2 + P on REE.
Postmenop. women of 70–80 years, before and after initiation of MHT	Postmenop. women of 70–80 years, before and after initiation of MHT	First month of MHT: E2 50 μ g/d transdermal patch; second month of MHT: E2 50 μ g/d transdermal patch + vaginal progesterone 200 mg/d for the final 7 d	IC. No further details on the calc. of REE pre-treatment, after 1 and 2 months of treatment.	6054 \pm 197 kJ/d	E2: 5994 \pm 266 kJ/d E2 + P: 6023 \pm 166 kJ/d	n.s. (within group)	n.s. (within group), adj. for FFM	No effect of E2 or E2 + P on REE.

TABLE 2 (Continued)

Author, year of publication, journal	Groups of interest	Interventions/ medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
Kimm et al., 2001,	Am J Epidemiol	Hormonal contraception users	Hormonal contraception use in general, incl. all active pharmaceutical substances, dosages, regimens, and forms of administration, incl. progestin-only contraceptives	IC. Calc. of baseline REE from the av. of 2 meas's. of REE with an interval of 10–14 d between each meas.	-	-	$p = 0.07$ (between groups)	-
Hormonal	Δ hormonal contraception users vs. hormonal contraception non-users: +46 kcal/d; $p = 0.07$	contraception non-users	-	IC. Calc. of baseline REE from the av. of 2 meas's. of REE with an interval of 10–14 d between each meas.	-	-	-	-
Piers et al., 1997, Eur J Clin Nutr	OC users	OCs used: either phasic or constant dose comb. OCs. The most common phasic dose pill used was a comb. of EE (30–50 μ g/d) + LNG (50–125 μ g/d). The most common constant dose pill was EE (50 μ g/d) + LNG (125 μ g/d). Other comb. included EE (30 μ g/d) + DSG (150 μ g/d); EE (30 μ g/d) + LNG (150 μ g/d); and EE (35 μ g/d) + NET (500 μ g/d). OC use for ≥ 6 months.	IC. No further details on the calc. of baseline REE.	5725 \pm 485 kJ/d; 5748 kJ/d (adj. for body weight); 5735 kJ/d (adj. for FFM and FM)	-	-	$p = 0.014$ (between groups), adj. for body weight $p = 0.024$ (between groups), adj. for FFM and FM	Δ OC users vs. OC non-users, adj. for body weight: +32 kcal/d \triangleq +2.4%; $p = 0.014$ Δ OC users vs. OC non-users, adj. for FM and FFM: +37 kcal/d \triangleq +2.8%; $p = 0.024$

(Continues)

TABLE 2 (Continued)

Author, year of publication, journal	Groups of interest	Interventions/medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
	OC non-users	-	IC. No further details on the calc. of baseline REE.	5590 \pm 530 kJ/d 5569 kJ/d (adj. for body weight) 5580 kJ/d (adj. for FFM and FM)	-			
Eck et al., 1997, Eur J Clin Nutr	OC users	OCs used: comb. triphasic OCs; 28 d regimen: constant EE 35 μ g/d for 21 d; trillevel adjustment of progestin (NET 0.5 mg d1–d7, 0.75 mg d8–d14, 1.0 mg d15–d21); placebo pills d22–d28; OC use for \geq 6 months	IC. Meas. of baseline REE 4 times within one OC cycle: during menstruation (d2, 3 or 4), in the first half (d 7, 8 or 9), in the middle (d 14, 15 or 16) and in the second half (d 22, 23 or 24) of the OC cycle.	Menstruation: 6.0 \pm 0.7 MJ/d Follicular phase: 6.0 \pm 0.9 MJ/d Ovulation: 5.8 \pm 0.7 MJ/d Luteal phase: 6.1 \pm 0.7 MJ/d	-	n.s. (within group and between groups)	n.s. (adj. for age)	Δ OC users vs. OC non-users, unadj. during: - menstruation: +24 kcal/ d Δ +1.7%; n.s. - follicular phase: +96 kcal/ d Δ +7.1%; n.s. - ovulation: -48 kcal/ d Δ -3.3%; n.s. - luteal phase: -24 kcal/ d Δ -1.6%; n.s.
	OC non-users	-	IC. Meas. of baseline REE 4 times within one menstrual cycle: during menstruation (d 2, 3 or 4), during follicular phase (d 7, 8 or 9), around ovulation (within 48 h after a positive ovulation test), and during luteal phase (d 22, 23, or 24).	Menstruation: 5.9 \pm 0.7 MJ/d Follicular phase: 5.6 \pm 0.5 MJ/d Ovulation: 6.0 \pm 0.5 MJ/d Luteal phase: 6.2 \pm 0.9 MJ/d	-	n.s. (within group and between groups)	n.s. (adj. for age)	
Diffey et al., 1997, Br J Nutr	OC users	OCs used: comb. OCs: The most used phasic-dose OC was a comb. of EE (30–50 μ g/d) + LNG (50–125 μ g/d). The most used constant dose OC was EE (50 μ g/d) + LNG	IC. No further details on the calc. of baseline REE.	5841 \pm 471 kJ/d	-	n.s.	p = 0.001 (between groups), adj. for body weight p = 0.002 (between groups), adj. for FM and FFM	Δ OC users vs. OC never-users, unadj.: +208 kcal/ d Δ +3.7%, n.s. Δ OC users vs. OC never-users, adj. for body weight: +4.9%; p = 0.001 Δ OC users vs. OC never-users, adj. for

TABLE 2 (Continued)

Author, year of publication, journal	Groups of interest	Interventions/medication details	Method, timing, and way of REE measurements	REE baseline/pre-treatment (mean \pm SD)	REE post-treatment (mean \pm SD)	p-value (Δ REE), unadjusted	p-value (Δ REE), adjusted	Outcome of interest: Δ REE
	OC never-users	(125 μ g/d). Other comb. included EE (30 μ g/d) + DSG (150 μ g/d); EE (30 μ g/d) + LNG (150 μ g/d); and EE (35 μ g/d) + NET (500 μ g/d). OC use for ≥ 6 months.		5633 \pm 614 kJ/d	-			FM and FFM: +4.8%; $p = 0.002$

Abbreviations: av., average; calc., calculation; CEE, conjugated equine estrogens; d, day/days; comb., combined/combinations; DSG, desogestrel; E2, estradiol; EE, ethinyl estradiol; FFM, fat-free mass; FM, fat mass; GnRH-Ag, gonadotropin-releasing hormone agonist; IC, indirect calorimetry; i.m., intramuscular; LNG, levonorgestrel; meas., measurement; MHT, menopausal hormone therapy; MPA, medroxyprogesteroneacetate; NET, norethisterone; OC, oral contraceptive; REE, resting energy expenditure.

estrogen used was either estradiol (E2) or conjugated equine estrogens (CEE), in the context of MHT,^{147,148,153,155} or ethinyl estradiol (EE), in the context of contraception.^{146,149-152,154} E2 dose and type of application in MHT varied from 50 to 75 μ g/d transdermally^{147,153} to no further specification.¹⁵⁵ CEE dose and mode of application in MHT varied from 0.625 mg/d orally¹⁴⁸ to no further information.¹⁵⁵ EE dose and form of application in contraception varied from 20 to 50 mcg/d orally^{146,149-151,154} to a generalized statement of inclusion of all HC methods, and thus also of progestin-only methods.¹⁵² The type of progestin used in MHT varied between medroxyprogesterone acetate (MPA),^{148,153} progesterone,¹⁴⁷ and no further details regarding the used progestin.¹⁵⁵ Two of the four MHT studies did not specifically distinguish between sequential and continuous-combined use of the progestin,^{148,155} whereas the other two studies reported sequential use.^{147,153} The progestins in the context of HC were levonorgestrel,^{149,154} desogestrel,^{149,154} and norethisterone,^{146,149,154} or there was no more specific information about this detail.¹⁵⁰⁻¹⁵² The mean age of the subjects enrolled in the study groups of interest in the six contraceptive trials ranged between 19¹⁵² and 26¹⁴⁹ years, and that of the subjects in the four MHT studies ranged between 33¹⁵³ and 72¹⁴⁷ years. The subjects in the contraceptive studies had an average BMI of 20.8¹⁴⁶ to 27.8¹⁵² kg/m², and those in the MHT studies had a BMI of 22.9¹⁵⁵ to 27.3¹⁵³ kg/m². To measure REE, the indirect calorimetry method was consistently used in all included studies.¹⁴⁶⁻¹⁵⁵ In all six contraceptive studies as well as in two of the MHT studies, between-group differences in REE were reported (HC users versus non-users or MHT users versus non-users). Within-group differences in REE before and after starting HC or MHT were not investigated.^{146,148-152,154,155} Only the MHT studies by Melanson et al. and Anderson et al. also reported within-group differences at pre-MHT^{147,153} and 1,¹⁴⁷ 2,¹⁴⁷ and 5¹⁵³ months after MHT initiation.

3.2 | Impact of estrogens on resting energy expenditure

Regarding the six studies on contraceptives,^{146,149-152,154} the picture that clearly emerges is that the various HC methods exert a neutral influence on REE or may even slightly increase it. Thus, in the cross-sectional study by Piers et al., the measured REE of women on combined oral contraceptives (OCs) adjusted for body weight was significantly higher than that of women not on combined OCs by 2.4% and +32 kcal/d ($p = 0.014$), respectively. REE adjusted for fat mass (FM) and fat-free mass (FFM), two major confounders of REE, was also significantly higher in women taking combined OCs as compared with women not taking combined OCs, by 2.8% corresponding to +37 kcal/d ($p = 0.024$).¹⁵⁴ In the study by Diffey et al., the REE-enhancing effect of combined OCs was even more pronounced. Although the raw (unadjusted) results showed a 3.7% increase in REE, equivalent to +208 kcal/d in combined OC users compared with combined OC never-users, the difference was not significant. However, after applying an analysis of covariance with either body weight or a combination of FM and FFM as covariates, the differences between

TABLE 3 Risk of bias assessment of non-randomized trials according to the quality assessment tool by Thomas H.

Study name	A	B	C	D	E	F	Global judgment
Aubertin (2008)	Strong	Moderate	Weak	Moderate	Strong	Strong	<i>Moderate</i>
Reimer (2005)	Moderate	Weak	Moderate	Moderate	Strong	Strong	<i>Moderate</i>
Anderson (2001)	Moderate	Moderate	Moderate	Moderate	Strong	Weak	<i>Moderate</i>
Kimm (2001)	Strong	Weak	Moderate	Moderate	Strong	Strong	<i>Moderate</i>
Piers (1997)	Moderate	Weak	Moderate	Moderate	Strong	Strong	<i>Moderate</i>
Eck (1997)	Strong	Moderate	Moderate	Moderate	Strong	Strong	<i>Strong</i>
Diffey (1997)	Moderate	Weak	Moderate	Moderate	Strong	Strong	<i>Moderate</i>

Note: A, selection bias; B, study design; C, confounders; D, blinding; E, data collection methods; F, withdrawals and dropouts. Thomas H. Quality assessment tool for quantitative studies. Effective Public Health Practice Project. McMaster University, Toronto. Recommended by Deeks JJ, Dinnes J, D'Amico R, Sowden AJ, Sakaravitch C, Song F, Petticrew M, Altman DG; International Stroke Trial Collaborative Group; European Carotid Surgery Trial Collaborative Group. *Evaluating non-randomised intervention studies. Health Technol Assess.* 2003;7 (27):iii-x, 1-173. doi: [10.3310/hta7270](https://doi.org/10.3310/hta7270). PMID: 14499048.

TABLE 4 Risk of bias assessment of randomized controlled trials according to the Cochrane Risk of Bias 2 (RoB 2) tool.

		Risk of bias domains					Overall
		D1	D2	D3	D4	D5	
Study	Duhita (2019)						
	Duhita (2017)						
	Melanson (2015)						

Domains:
D1: Bias arising from the randomization process.
D2: Bias due to deviations from intended intervention.
D3: Bias due to missing outcome data.
D4: Bias in measurement of the outcome.
D5: Bias in selection of the reported result.

Judgement
 Some concerns
 Low

Note: McGuinness, LA, Higgins, JPT. Risk-of-bias VISualization (robvis): An R package and Shiny web app for visualizing risk-of-bias assessments. *Res Syn Meth.* 2020; 1-7. <https://doi.org/10.1002/jrsm.1411>.

groups were found significant. Accordingly, combined OC users had a significantly higher REE than combined OC never-users, 4.9% ($p = 0.001$) and 4.8% ($p = 0.002$), respectively.¹⁴⁹ In the remaining four contraceptive studies, no significant effect of HC use on REE was found. Nevertheless, the cross-sectional study by Kimm et al. found a clear trend toward an increase in REE by +46 kcal/d ($p = 0.07$) with contraceptive medication use.¹⁵² In the cross-sectional study by Eck et al., examining the effect of combined OC use on energy balance and its components across the different phases of the menstrual cycle, no significant differences in REE were found among the different phases of the menstrual cycle itself, nor between the groups of combined OC users and non-users. Combined OC users had a higher REE than non-users during menstruation (+24 kcal/d \triangleq 1.7%) and during the follicular phase (+96 kcal/d \triangleq 7.1%), when endogenous estradiol levels are physiologically low, whereas REE was lower during ovulation (-48 kcal/d \triangleq 3.3%) and the luteal phase (-24/d kcal \triangleq 1.6%), when endogenous estradiol levels are physiologically high. This observation again speaks for the influence of sex steroids on the REE. However, as

noted above, all differences showed no statistical significance.¹⁴⁶ Likewise, the two RCTs by Duhita et al. with a very small number of subjects did not describe any significant effect of the use of combined OCs on REE either.^{150,151} At least, none of the contraceptive studies showed a significant negative effect of HC use on REE.

The four MHT studies^{147,148,153,155} show a similar picture. In the RCT by Melanson et al., in which young healthy women received ovarian function suppression with GnRH analogs with or without add-back therapy, a significant decrease in REE was observed in the GnRH-Ag + placebo substitution group (3.7%, corresponding to -54 kcal/d, $p \leq 0.05$), an effect not observed in the GnRH-Ag + E2 substitution group (+6 kcal/d). Although REE did not differ significantly between groups before the intervention, it did after the intervention (difference in between-group changes: 60 kcal/d, $p < 0.05$). Pharmacological suppression of sex hormones and thus iatrogenically induced menopause reduced REE. E2 add-back therapy not only attenuated the decrease in REE but completely counteracted this effect.¹⁵³ In the cross-sectional study by Aubertin-Leheudre et al., the

REE of actual MHT users was compared with that of MHT never-users. The physiologically postmenopausal women of both groups did not differ in baseline characteristics such as age, BMI, or body composition (FM and FFM), but MHT users had a significantly higher REE than MHT non-users ($\Delta + 222$ kcal/d; $p = 0.038$). The magnitude of the increase in REE due to MHT use corresponded to a large and significant Cohen's effect size of 0.78.¹⁴⁸ A clear trend in REE pattern, although not statistically significant, was evident in the cross-sectional study with a very small number of cases by Reimer et al. comparing the REE of postmenopausal actual MHT users with that of MHT non-users. Thus, the REE was highest in the luteal phase of premenopausal women, followed by that of the follicular phase of premenopausal women, the REE of postmenopausal women using MHT, and finally that of postmenopausal women not using MHT. The REE was significantly lower ($p < 0.05$) in postmenopausal women not taking MHT compared with premenopausal women, but not in women taking MHT.¹⁵⁵ In the prospective cohort study by Anderson et al., REE was examined in younger (45–55 years) and older (70–80 years) subjects before and after initiation of MHT. No significant effect of short-term (1 and 2 months) E2 or E2 + progesterone administration on REE was found, even when adjusted for FFM.¹⁴⁷ In summary, estrogen substitution in the context of MHT resulted in a significant increase in REE in two of the four studies,^{148,153} whereas it showed a distinct, but nonsignificant trend toward an increase in REE in one¹⁵⁵ and a neutral effect in another study.¹⁴⁷ As in the studies on contraception, no single study could be identified in which estrogen substitution led to a significant decrease in REE.

4 | DISCUSSION

4.1 | Strengths and limitations

It should be noted that only two^{147,153} of all included studies comprehensively investigated estrogen effects exclusively, without any possible confounding by the concomitant use of a progestin. Thus, the RCT by Melanson et al.¹⁵³ showing a significant positive effect and the prospective cohort study by Anderson et al.¹⁴⁷ observing no significant effect of estrogen administration on REE provide the most valid evidence regarding our research question.

It is also worth mentioning that in the contraceptive studies, the timing of the REE measurement during the menstrual cycle is of great importance in the control group, in contrast to the MHT studies with the consistently postmenopausal patients. It is striking that the two studies by Piers et al.¹⁵⁴ and Diffey et al.¹⁴⁹ reported a significant increase in REE with OC use but did not mention the timing of the REE measurement during the menstrual cycle of OC non-users. Assuming that the REE measurement in these two studies took place only during the early follicular phase in OC nonusers, the effect of estrogens on REE over the entire course of the menstrual cycle would not be accurately represented by a single measurement, given the drastic physiological fluctuations of sex steroids in the menstrual cycle. REE measurements in the follicular phase may underestimate

REE over the course of a menstrual cycle, and measurements in the luteal phase may overestimate it. Here, Eck et al.¹⁴⁶ clearly showed that the REE of OC non-users was lower in the first cycle phase than that of OC users. In the second half of the menstrual cycle, however, the exact opposite was the case. The timing of the REE measurement in the menstrual cycle is therefore of great importance. The average influence of sex steroids across the cycle of HC nonusers is better represented in the work of Duhita et al.¹⁵⁰ and Kimm et al.¹⁵² by calculating the average REE from multiple measurements across the menstrual cycle, with REE measurements evenly distributed between the follicular and luteal phases. In contrast, Eck et al.¹⁴⁶ evaluated the REE measurements in the follicular phase, at the time of ovulation, in the luteal phase, and during menstruation separately and then compared them between OC users and OC non-users.

Furthermore, it should be critically noted that in exactly those two^{149,154} of the six^{146,149–152,154} HC papers that showed a significant positive influence of estrogens on REE, high-dose OCs with up to 50 μg EE/d were used. Because these are EE dosages as they are no longer standardly used nowadays, the results cannot be uncritically extrapolated to the OCs currently in use.

Another limitation of our review and a potential reason that some studies failed to demonstrate an effect of estrogen medication on REE could be the fact of underpowerment. If the included studies are subdivided according to the number of subjects of interest, a significant estrogen use-dependent increase in REE (Piers et al. with 89 subjects,¹⁵⁴ Diffey et al. with 46 subjects,¹⁴⁹ and Melanson et al. with 45 subjects¹⁵³) and a nonsignificant but undeniable tendency in the same direction (Kimm et al. with 147 subjects¹⁵²) can be observed in four^{149,152–154} of the five^{147,149,152–154} studies with the largest sample size. However, with the exception of one (Aubertin-Leheudre et al. with 26 participants¹⁴⁸) of the five studies with the smallest number of participants, there was no effect of estrogen use on REE (Eck et al. with 32 subjects, Duhita et al. with 16 subjects,¹⁵⁰ Duhita et al. with 14 subjects,¹⁵¹ and Reimer et al. with 13 subjects¹⁵⁵).

One of the greatest weaknesses of our systematic review, which limits the significance of its results, is not at least the insufficient availability of high-quality studies on our specific research question.

One of the strengths of our systematic review is the comprehensive literature search, covering not only estrogens but all sex steroids, to identify all relevant studies on this topic and to ensure that the available evidence is presented as fully as possible. We also restricted inclusion to normal-weight and overweight women to exclude confounding factors such as metabolic adjustments and hormonal alterations that typically occur when women are obese or underweight. As is typical for systematic reviews in general, our article is characterized by an objective methodology that ensures transparency and reproducibility. By summarizing the evidence from several individual studies in this article, it is possible to make a comprehensive statement about the likely effect of estrogens on REE. This should help to draw clear conclusions and provide the basis for decisions concerning a much-discussed topic in daily practice.

In summary, our results support an REE-enhancing effect of estrogen administration in the context of HC or MHT. However, overall scientific evidence on the impact of estrogen administration on REE is limited, and the quality of available evidence is low to moderate.

4.2 | Discussion of the results with regard to the influence of estrogens on BAT

Despite the positive aspects of HC and/or MHT, the fear of unwanted weight gain is one of the main reasons for women not to initiate or to early discontinue these therapies.⁷⁻¹¹ For clinicians, easing women's anxiety is difficult regarding the currently limited understanding of the effects of sex hormones on body weight regulation. Given that a low REE is a confirmed predictive factor for weight gain¹⁵⁶ and consecutively for the development of obesity, research into the influence of sex steroids on REE is a particularly exciting area. Considering the balance of evidence, our data suggest a favorable effect of circulating estrogen levels and estrogen administration in the form of HC and MHT on REE.

Studies have shown that hormonal fluctuations during the menstrual cycle influence REE.¹⁰²⁻¹⁰⁸ According to our data, suppression of these fluctuations by HC seems to have neutral or even positive effects on body weight regulation in terms of an increase in REE.

Regarding the influence of MHT on weight development in perimenopause and postmenopause, it should be pointed out that the weight gain associated with perimenopause and postmenopause is rather a consequence of the age-related tendency to reduced physical activity, the associated loss of muscle mass, and the overall reduced energy requirement.¹⁵⁷ Contrary to mainstream belief, MHT actually counteracts this age-related tendency to gain weight, with one of the mechanisms being an increase in REE.

The estrogenic effect on BAT activity may be the underlying reason for this observation. Indeed, considering the magnitude of the estrogen-associated increase in REE (ranging from no effect^{146,147,150-152,155} to +32 kcal/d,¹⁵⁴ +37 kcal/d,¹⁵⁴ +60 kcal/d,¹⁵³ +208 kcal/d,¹⁴⁹ and +222 kcal/d¹⁴⁸), this corresponds to the additional energy expenditure associated with activation of BAT thermogenesis. It is estimated that BAT-specific energy expenditure can account for up to 2.5%–8% of REE.^{36,48,158} The limited data available on this issue quantify the actual contribution of BAT thermogenesis to TEE at 7 kcal/d in thermoneutral environments to approximately 200 kcal/d in mild cold exposure.¹⁵⁸⁻¹⁶¹ This rather small contribution of BAT to TEE is mainly due to its low overall tissue mass of about 50–500 g¹⁶² rather than its metabolic activity, which is still several times higher compared with other tissues of the human body.¹⁶¹

As mentioned at the very beginning, the development of obesity is the consequence of a chronic disturbance of energy homeostasis with a persistent positive energy balance. The average weight gain underlying obesity, which usually commences in young adulthood, is 0.5–0.7 kg per year. This corresponds to a daily positive energy balance of just 12–17 kcal/day at an average energy density of 8840 kcal

per kg body weight and lies thus within the range of the BAT-specific contribution to TEE^{161,163} and estrogen-mediated activation of BAT thermogenesis, respectively. Hence, the contribution of BAT to REE may be small but not negligible. It seems that low but sustained activity of BAT thermogenesis may significantly counteract the development of obesity. However, current estimates of BAT thermogenesis capacity are at the low end of what would potentially be clinically possible through its chronic stimulation by various endogenous or exogenous factors.¹⁶¹ It has been shown that the metabolic activity of BAT can be increased many-fold as well as BAT mass can be newly formed and existing depots can be significantly enlarged even in adulthood by long-term cold exposure or cold acclimatization^{159,161,164,165} and catecholamine-secreting tumors such as pheochromocytomas and paragangliomas.^{76,166,167} According to our results, estrogen administration could have a similar effect, although to a lesser extent. The assumption that estrogen-mediated activation of BAT thermogenesis underlies the estrogen-induced increase in REE is reasonable, although the exact mechanisms of the estrogen-induced REE increase cannot be elucidated with this systematic review. Future research needs to focus more on the metabolic effects of sex steroid administration.

Finally, it should be emphasized that estrogens have an anorexic effect and thus prevent increased EI from compensating for increased REE.¹⁶⁸ Together with this fact, our results suggest that the energy balance changes in a negative direction under the influence of estrogens. Thus, the widespread fear of weight gain solely as a result of taking HC or MHT is therefore unsubstantiated. This article is intended to help clinicians educate women accordingly and alleviate their fears in this regard.

ACKNOWLEDGMENTS

The authors are grateful for the support of Dr. Norman Bitterlich, a biostatistician, Department of Biostatistics, Medicine & Service GmbH Chemnitz, 09117 Chemnitz, Germany, with statistical questions. Open access funding provided by Inselspital Universitatsspital Bern.

CONFLICT OF INTEREST STATEMENT

No conflict of interest statement.

ORCID

Susanna Weidlinger  <https://orcid.org/0000-0002-1846-1628>

REFERENCES

1. World Health Organisation. Global Health Observatory data repository. (<https://apps.who.int/gho/data/node.main.A896?lang%3Den>).
2. Aune D, Sen A, Prasad M, et al. BMI and all cause mortality: systematic review and non-linear dose-response meta-analysis of 230 cohort studies with 3.74 million deaths among 30.3 million participants. *BMJ*. 2016;353:i2156. doi:10.1136/bmj.i2156
3. Berrington de Gonzalez A, Hartge P, Cerhan JR, et al. Body-mass index and mortality among 1.46 million white adults. *N Engl J Med*. 2010;363(23):2211-2219. doi:10.1056/NEJMoa1000367
4. di Angelantonio E, Bhupathiraju SN, Wormser D, et al. Body-mass index and all-cause mortality: individual-participant-data meta-

- analysis of 239 prospective studies in four continents. *Lancet*. 2016; 388(10046):776-786. doi:10.1016/S0140-6736(16)30175-1
5. GBD 2015 Obesity Collaborators, Afshin A, Forouzanfar MH, et al. Health effects of overweight and obesity in 195 countries over 25 years. *N Engl J Med*. 2017;377(1):13-27. doi:10.1056/NEJMoa1614362
 6. Woolf SH, Schoomaker H. Life expectancy and mortality rates in the United States, 1959-2017. *Jama*. 2019;322(20):1996-2016. doi:10.1001/jama.2019.16932
 7. Venkat P, Masch R, Ng E, Cremer M, Richman S, Arslan A. Knowledge and beliefs about contraception in urban Latina women. *J Community Health*. 2008;33(5):357-362. doi:10.1007/s10900-008-9100-1
 8. Greenwald MN, Gold MA. Do they, don't they, or why haven't they?. Contraceptive use patterns among inner-city sexually active female adolescents. *J Pediatr Adolesc Gynecol*. 2000;13(2):91-92. doi:10.1016/s1083-3188(00)00022-x
 9. Clare C, Squire MB, Alvarez K, Meisler J, Fraser C. Barriers to adolescent contraception use and adherence. *Int J Adolesc Med Health*. 2016;30(4):20160098. doi:10.1515/ijamh-2016-0098
 10. Bakken K, Eggen AE, Lund E. Side-effects of hormone replacement therapy and influence on pattern of use among women aged 45-64 years. The Norwegian Women and Cancer (NOWAC) study 1997. *Acta Obstet Gynecol Scand*. 2004;83(9):850-856. doi:10.1111/j.0001-6349.2004.00560.x
 11. Oddens BJ, Visser AP, Vemer HM, Everaerd WT, Lehert P. Contraceptive use and attitudes in Great Britain. *Contraception*. 1994;49(1):73-86. doi:10.1016/0010-7824(94)90110-4
 12. Gallo MF, Lopez LM, Grimes DA, Carayon F, Schulz KF, Helmerhorst FM. Combination contraceptives: effects on weight. *Cochrane Database Syst Rev*. 2014;1:CD003987. doi:10.1002/14651858.CD003987.pub5
 13. Lopez LM, Ramesh S, Chen M, et al. Progestin-only contraceptives: effects on weight. *Cochrane Database Syst Rev*. 2016;2016(8):CD008815. doi:10.1002/14651858.CD008815.pub4
 14. Norman RJ, Flight IH, Rees MC. Oestrogen and progestogen hormone replacement therapy for peri-menopausal and post-menopausal women: weight and body fat distribution. *Cochrane Database Syst Rev*. 2000;(2):CD001018. doi:10.1002/14651858.CD001018
 15. Westerterp KR. In: Feingold KR, Anawalt B, Boyce A, et al., eds. *Control of Energy Expenditure in Humans*. Endotext; 2000.
 16. Westerterp KR. Control of energy expenditure in humans. *Eur J Clin Nutr*. 2017;71(3):340-344. doi:10.1038/ejcn.2016.237
 17. Lam YY, Ravussin E. Analysis of energy metabolism in humans: a review of methodologies. *Mol Metab*. 2016;5(11):1057-1071. doi:10.1016/j.molmet.2016.09.005
 18. Levine JA. Non-exercise activity thermogenesis (NEAT). *Nutr Rev*. 2004;62(7 Pt 2):S82-S97. doi:10.1111/j.1753-4887.2004.tb00094.x
 19. Ravussin E, Bogardus C. A brief overview of human energy metabolism and its relationship to essential obesity. *Am J Clin Nutr*. 1992; 55(1 Suppl):242S-245S. doi:10.1093/ajcn/55.1.242s
 20. Landsberg L. Core temperature: a forgotten variable in energy expenditure and obesity? *Obes Rev*. 2012;13(Suppl 2):97-104. doi:10.1111/j.1467-789X.2012.01040.x
 21. Reed GW, Hill JO. Measuring the thermic effect of food. *Am J Clin Nutr*. 1996;63(2):164-169. doi:10.1093/ajcn/63.2.164
 22. Munzberg H, Qualls-Creekmore E, Berthoud HR, Morrison CD, Yu S. Neural control of energy expenditure. *Handb Exp Pharmacol*. 2016; 233:173-194. doi:10.1007/164_2015_33
 23. Rolfe DF, Brown GC. Cellular energy utilization and molecular origin of standard metabolic rate in mammals. *Physiol Rev*. 1997;77(3):731-758. doi:10.1152/physrev.1997.77.3.731
 24. Brand MD, Brindle KM, Buckingham JA, Harper JA, Rolfe DF, Stuart JA. The significance and mechanism of mitochondrial proton conductance. *Int J Obes Relat Metab Disord*. 1999;23(Suppl 6):S4-S11. doi:10.1038/sj.ijo.0800936
 25. Ricquier D. Fundamental mechanisms of thermogenesis. *C R Biol*. 2006;329(8):578-586. doi:10.1016/j.crv.2005.10.010
 26. Porter RK. Mitochondrial proton leak: a role for uncoupling proteins 2 and 3? *Biochim Biophys Acta*. 2001;1504(1):120-127. doi:10.1016/s0005-2728(00)00246-2
 27. Rolfe DF, Newman JM, Buckingham JA, Clark MG, Brand MD. Contribution of mitochondrial proton leak to respiration rate in working skeletal muscle and liver and to SMR. *Am J Physiol*. 1999;276(3):C692-C699. doi:10.1152/ajpcell.1999.276.3.C692
 28. Rolfe DF, Brand MD. Contribution of mitochondrial proton leak to skeletal muscle respiration and to standard metabolic rate. *Am J Physiol*. 1996;271(4 Pt 1):C1380-C1389. doi:10.1152/ajpcell.1996.271.4.C1380
 29. Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. *Physiol Rev*. 2004;84(1):277-359. doi:10.1152/physrev.00015.2003
 30. Silva JE. Thermogenic mechanisms and their hormonal regulation. *Physiol Rev*. 2006;86(2):435-464. doi:10.1152/physrev.00009.2005
 31. Porter RK, Brand MD. Body mass dependence of H⁺ leak in mitochondria and its relevance to metabolic rate. *Nature*. 1993; 362(6421):628-630. doi:10.1038/362628a0
 32. Brand MD. The proton leak across the mitochondrial inner membrane. *Biochim Biophys Acta*. 1990;1018(2-3):128-133. doi:10.1016/0005-2728(90)90232-s
 33. Nicholls DG. Mitochondrial proton leaks and uncoupling proteins. *Biochim Biophys Acta Bioenerg*. 2021;1862(7):148428. doi:10.1016/j.bbabi.2021.148428
 34. Kim JE, Kim JS, Jo MJ, et al. The roles and associated mechanisms of adipokines in development of metabolic syndrome. *Molecules*. 2022; 27(2):234. doi:10.3390/molecules27020334
 35. Nedergaard J, Cannon B. Brown adipose tissue as a heat-producing thermoeffector. *Handb Clin Neurol*. 2018;156:137-152. doi:10.1016/B978-0-444-63912-7.00009-6
 36. van Marken Lichtenbelt WD, Schrauwen P. Implications of nonshivering thermogenesis for energy balance regulation in humans. *Am J Physiol Regul Integr Comp Physiol*. 2011;301(2):R285-R296. doi:10.1152/ajpregu.00652.2010
 37. Rothwell NJ, Stock MJ. A role for brown adipose tissue in diet-induced thermogenesis. *Nature*. 1979;281(5726):31-35. doi:10.1038/281031a0
 38. Boss O, Farmer SR. Recruitment of brown adipose tissue as a therapy for obesity-associated diseases. *Front Endocrinol (Lausanne)*. 2012;3:14. doi:10.3389/fendo.2012.00014
 39. Holloway BR, Howe R, Rao BS, Stribling D. ICI D7114: a novel selective adrenoceptor agonist of brown fat and thermogenesis. *Am J Clin Nutr*. 1992;55(1 Suppl):262S-264S. doi:10.1093/ajcn/55.1.262s
 40. Kopecky J, Clarke G, Enerback S, Spiegelman B, Kozak LP. Expression of the mitochondrial uncoupling protein gene from the aP2 gene promoter prevents genetic obesity. *J Clin Invest*. 1995;96(6):2914-2923. doi:10.1172/JCI118363
 41. Nagase I, Yoshida T, Kumamoto K, et al. Expression of uncoupling protein in skeletal muscle and white fat of obese mice treated with thermogenic beta 3-adrenergic agonist. *J Clin Invest*. 1996;97(12):2898-2904. doi:10.1172/JCI118748
 42. Ghorbani M, Himms-Hagen J. Appearance of brown adipocytes in white adipose tissue during CL 316,243-induced reversal of obesity and diabetes in Zucker fa/fa rats. *Int J Obes Relat Metab Disord*. 1997;21(6):465-475. doi:10.1038/sj.ijo.0800432
 43. Kim H, Pennisi PA, Gavrilova O, et al. Effect of adipocyte beta3-adrenergic receptor activation on the type 2 diabetic MKR mice. *Am J Physiol Endocrinol Metab*. 2006;290(6):E1227-E1236. doi:10.1152/ajpendo.00344.2005

44. Lean ME. Brown adipose tissue in humans. *Proc Nutr Soc.* 1989; 48(2):243-256. doi:[10.1079/pns19890036](https://doi.org/10.1079/pns19890036)
45. Saito M, Okamatsu-Ogura Y, Matsushita M, et al. High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes.* 2009;58(7):1526-1531. doi:[10.2337/db09-0530](https://doi.org/10.2337/db09-0530)
46. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, et al. Cold-activated brown adipose tissue in healthy men. *N Engl J Med.* 2009;360(15):1500-1508. doi:[10.1056/NEJMoa0808718](https://doi.org/10.1056/NEJMoa0808718)
47. Cypess AM, Lehman S, Williams G, et al. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med.* 2009; 360(15):1509-1517. doi:[10.1056/NEJMoa0810780](https://doi.org/10.1056/NEJMoa0810780)
48. Virtanen KA, Lidell ME, Orava J, et al. Functional brown adipose tissue in healthy adults. *N Engl J Med.* 2009;360(15):1518-1525. doi:[10.1056/NEJMoa0808949](https://doi.org/10.1056/NEJMoa0808949)
49. Nedergaard J, Bengtsson T, Cannon B. Unexpected evidence for active brown adipose tissue in adult humans. *Am J Physiol Endocrinol Metab.* 2007;293(2):E444-E452. doi:[10.1152/ajpendo.00691.2006](https://doi.org/10.1152/ajpendo.00691.2006)
50. Whittle AJ, Lopez M, Vidal-Puig A. Using brown adipose tissue to treat obesity—the central issue. *Trends Mol Med.* 2011;17(8):405-411. doi:[10.1016/j.molmed.2011.04.001](https://doi.org/10.1016/j.molmed.2011.04.001)
51. Yoneshiro T, Aita S, Matsushita M, et al. Recruited brown adipose tissue as an antiobesity agent in humans. *J Clin Invest.* 2013;123(8): 3404-3408. doi:[10.1172/JCI67803](https://doi.org/10.1172/JCI67803)
52. Poekes L, Lanthier N, Leclercq IA. Brown adipose tissue: a potential target in the fight against obesity and the metabolic syndrome. *Clin Sci (Lond).* 2015;129(11):933-949. doi:[10.1042/CS20150339](https://doi.org/10.1042/CS20150339)
53. Cheng L, Wang J, Dai H, et al. Brown and beige adipose tissue: a novel therapeutic strategy for obesity and type 2 diabetes mellitus. *Adipocyte.* 2021;10(1):48-65. doi:[10.1080/21623945.2020.1870060](https://doi.org/10.1080/21623945.2020.1870060)
54. Ruiz JR, Martinez-Tellez B, Sanchez-Delgado G, Osuna-Prieto FJ, Rensen PCN, Boon MR. Role of human brown fat in obesity, metabolism and cardiovascular disease: strategies to turn up the heat. *Prog Cardiovasc Dis.* 2018;61(2):232-245. doi:[10.1016/j.pcad.2018.07.002](https://doi.org/10.1016/j.pcad.2018.07.002)
55. Perkins MN, Rothwell NJ, Stock MJ, Stone TW. Activation of brown adipose tissue thermogenesis by the ventromedial hypothalamus. *Nature.* 1981;289(5796):401-402. doi:[10.1038/289401a0](https://doi.org/10.1038/289401a0)
56. Contreras C, Nogueiras R, Dieguez C, Medina-Gomez G, Lopez M. Hypothalamus and thermogenesis: heating the BAT, browning the WAT. *Mol Cell Endocrinol.* 2016;438:107-115. doi:[10.1016/j.mce.2016.08.002](https://doi.org/10.1016/j.mce.2016.08.002)
57. Lindberg D, Chen P, Li C. Conditional viral tracing reveals that steroidogenic factor 1-positive neurons of the dorsomedial subdivision of the ventromedial hypothalamus project to autonomic centers of the hypothalamus and hindbrain. *J Comp Neurol.* 2013;521(14): 3167-3190. doi:[10.1002/cne.23338](https://doi.org/10.1002/cne.23338)
58. Contreras C, Gonzalez F, Ferno J, et al. The brain and brown fat. *Ann Med.* 2015;47(2):150-168. doi:[10.3109/07853890.2014.919727](https://doi.org/10.3109/07853890.2014.919727)
59. Huttunen P, Hirvonen J, Kinnula V. The occurrence of brown adipose tissue in outdoor workers. *Eur J Appl Physiol Occup Physiol.* 1981;46(4):339-345. doi:[10.1007/BF00422121](https://doi.org/10.1007/BF00422121)
60. Cohade C, Mourtzikos KA, Wahl RL. "USA-Fat": prevalence is related to ambient outdoor temperature—evaluation with 18F-FDG PET/CT. *J Nucl Med.* 2003;44(8):1267-1270.
61. Garcia C, Bandar V, Van Nostrand D, et al. Effective reduction of brown fat FDG uptake by controlling environmental temperature prior to PET scan: an expanded case series. *Mol Imaging Biol.* 2010; 12(6):652-656. doi:[10.1007/s11307-010-0298-9](https://doi.org/10.1007/s11307-010-0298-9)
62. Christensen CR, Clark PB, Morton KA. Reversal of hypermetabolic brown adipose tissue in F-18 FDG PET imaging. *Clin Nucl Med.* 2006;31(4):193-196. doi:[10.1097/01.rlu.0000204199.33136.05](https://doi.org/10.1097/01.rlu.0000204199.33136.05)
63. Nedergaard J, Cannon B. The browning of white adipose tissue: some burning issues. *Cell Metab.* 2014;20(3):396-407. doi:[10.1016/j.cmet.2014.07.005](https://doi.org/10.1016/j.cmet.2014.07.005)
64. Jeremic N, Chaturvedi P, Tyagi SC. Browning of white fat: novel insight into factors, mechanisms, and therapeutics. *J Cell Physiol.* 2017;232(1):61-68. doi:[10.1002/jcp.25450](https://doi.org/10.1002/jcp.25450)
65. Yoneshiro T, Aita S, Matsushita M, et al. Brown adipose tissue, whole-body energy expenditure, and thermogenesis in healthy adult men. *Obesity (Silver Spring).* 2011;19(1):13-16. doi:[10.1038/oby.2010.105](https://doi.org/10.1038/oby.2010.105)
66. van der Lans AA, Hoeks J, Brans B, et al. Cold acclimation recruits human brown fat and increases nonshivering thermogenesis. *J Clin Invest.* 2013;123(8):3395-3403. doi:[10.1172/JCI68993](https://doi.org/10.1172/JCI68993)
67. Bakker LE, Boon MR, van der Linden RA, et al. Brown adipose tissue volume in healthy lean south Asian adults compared with white Caucasians: a prospective, case-controlled observational study. *Lancet Diabetes Endocrinol.* 2014;2(3):210-217. doi:[10.1016/S2213-8587\(13\)70156-6](https://doi.org/10.1016/S2213-8587(13)70156-6)
68. Gerngross C, Schretter J, Klingenspor M, Schwaiger M, Fromme T. Active brown fat during (18)F-FDG PET/CT imaging defines a patient group with characteristic traits and an increased probability of brown fat redetection. *J Nucl Med.* 2017;58(7):1104-1110. doi:[10.2967/jnumed.116.183988](https://doi.org/10.2967/jnumed.116.183988)
69. Orava J, Nuutila P, Noponen T, et al. Blunted metabolic responses to cold and insulin stimulation in brown adipose tissue of obese humans. *Obesity (Silver Spring).* 2013;21(11):2279-2287. doi:[10.1002/oby.20456](https://doi.org/10.1002/oby.20456)
70. Vijgen GH, Bouvy ND, Teule GJ, Brans B, Schrauwen P, van Marken Lichtenbelt WD. Brown adipose tissue in morbidly obese subjects. *PLoS ONE.* 2011;6(2):e17247. doi:[10.1371/journal.pone.0017247](https://doi.org/10.1371/journal.pone.0017247)
71. Yoneshiro T, Aita S, Matsushita M, et al. Age-related decrease in cold-activated brown adipose tissue and accumulation of body fat in healthy humans. *Obesity (Silver Spring).* 2011;19(9):1755-1760. doi:[10.1038/oby.2011.125](https://doi.org/10.1038/oby.2011.125)
72. Hanssen MJ, Hoeks J, Brans B, et al. Short-term cold acclimation improves insulin sensitivity in patients with type 2 diabetes mellitus. *Nat Med.* 2015;21(8):863-865. doi:[10.1038/nm.3891](https://doi.org/10.1038/nm.3891)
73. O'Mara AE, Johnson JW, Linderman JD, et al. Chronic mirabegron treatment increases human brown fat, HDL cholesterol, and insulin sensitivity. *J Clin Invest.* 2020;130(5):2209-2219. doi:[10.1172/JCI131126](https://doi.org/10.1172/JCI131126)
74. Peirce V, Vidal-Puig A. Regulation of glucose homeostasis by brown adipose tissue. *Lancet Diabetes Endocrinol.* 2013;1(4):353-360. doi:[10.1016/S2213-8587\(13\)70055-X](https://doi.org/10.1016/S2213-8587(13)70055-X)
75. Maliszewska K, Kretowski A. Brown adipose tissue and its role in insulin and glucose homeostasis. *Int J Mol Sci.* 2021;22(4):1530. doi:[10.3390/ijms22041530](https://doi.org/10.3390/ijms22041530)
76. Dong A, Wang Y, Lu J, Zuo C. Hypermetabolic mesenteric brown adipose tissue on dual-time point FDG PET/CT in a patient with benign retroperitoneal pheochromocytoma. *Clin Nucl Med.* 2014; 39(3):e229-e232. doi:[10.1097/RLU.0b013e3182816515](https://doi.org/10.1097/RLU.0b013e3182816515)
77. Alvarez-Crespo M, Csikasz RI, Martínez-Sánchez N, et al. Essential role of UCP1 modulating the central effects of thyroid hormones on energy balance. *Mol Metab.* 2016;5(4):271-282. doi:[10.1016/j.molmet.2016.01.008](https://doi.org/10.1016/j.molmet.2016.01.008)
78. Lopez M, Varela L, Vazquez MJ, et al. Hypothalamic AMPK and fatty acid metabolism mediate thyroid regulation of energy balance. *Nat Med.* 2010;16(9):1001-1008. doi:[10.1038/nm.2207](https://doi.org/10.1038/nm.2207)
79. Lopez M, Alvarez CV, Nogueiras R, Dieguez C. Energy balance regulation by thyroid hormones at central level. *Trends Mol Med.* 2013; 19(7):418-427. doi:[10.1016/j.molmed.2013.04.004](https://doi.org/10.1016/j.molmed.2013.04.004)

80. Martinez-Sanchez N, Moreno-Navarrete JM, Contreras C, et al. Thyroid hormones induce browning of white fat. *J Endocrinol*. 2017; 232(2):351-362. doi:10.1530/JOE-16-0425
81. Guerra C, Navarro P, Valverde AM, et al. Brown adipose tissue-specific insulin receptor knockout shows diabetic phenotype without insulin resistance. *J Clin Invest*. 2001;108(8):1205-1213. doi:10.1172/JCI13103
82. Storlien LH, James DE, Burleigh KM, Chisholm DJ, Kraegen EW. Fat feeding causes widespread in vivo insulin resistance, decreased energy expenditure, and obesity in rats. *Am J Physiol*. 1986; 251(5 Pt 1):E576-E583. doi:10.1152/ajpendo.1986.251.5.E576
83. Martinez de Morentin PB, Gonzalez-Garcia I, Martins L, et al. Estradiol regulates brown adipose tissue thermogenesis via hypothalamic AMPK. *Cell Metab*. 2014;20(1):41-53. doi:10.1016/j.cmet.2014.03.031
84. Lopez M, Tena-Sempere M. Estradiol and brown fat. *Best Pract Res Clin Endocrinol Metab*. 2016;30(4):527-536. doi:10.1016/j.beem.2016.08.004
85. Gonzalez-Garcia I, Tena-Sempere M, Lopez M. Estradiol regulation of brown adipose tissue thermogenesis. *Adv Exp Med Biol*. 2017; 1043:315-335. doi:10.1007/978-3-319-70178-3_15
86. Lopez M, Tena-Sempere M. Estrogens and the control of energy homeostasis: a brain perspective. *Trends Endocrinol Metab*. 2015; 26(8):411-421. doi:10.1016/j.tem.2015.06.003
87. Mauvais-Jarvis F, Clegg DJ, Hevener AL. The role of estrogens in control of energy balance and glucose homeostasis. *Endocr Rev*. 2013;34(3):309-338. doi:10.1210/er.2012-1055
88. Frank A, Brown LM, Clegg DJ. The role of hypothalamic estrogen receptors in metabolic regulation. *Front Neuroendocrinol*. 2014;35(4): 550-557. doi:10.1016/j.yfrne.2014.05.002
89. Melanson EL, Lyden K, Gibbons E, et al. Influence of estradiol status on physical activity in premenopausal women. *Med Sci Sports Exerc*. 2018;50(8):1704-1709. doi:10.1249/MSS.0000000000001598
90. Blaustein JD, Wade GN. Ovarian influences on the meal patterns of female rats. *Physiol Behav*. 1976;17(2):201-208. doi:10.1016/0031-9384(76)90064-0
91. Carr MC. The emergence of the metabolic syndrome with menopause. *J Clin Endocrinol Metab*. 2003;88(6):2404-2411. doi:10.1210/jc.2003-030242
92. Rogers NH, Perfield JW 2nd, Strissel KJ, Obin MS, Greenberg AS. Reduced energy expenditure and increased inflammation are early events in the development of ovariectomy-induced obesity. *Endocrinology*. 2009;150(5):2161-2168. doi:10.1210/en.2008-1405
93. Lynch NA, Ryan AS, Berman DM, Sorkin JD, Nicklas BJ. Comparison of VO₂max and disease risk factors between perimenopausal and postmenopausal women. *Menopause*. 2002;9(6):456-462. doi:10.1097/00042192-200211000-00012
94. Bellino FL, Wise PM. Nonhuman primate models of menopause workshop. *Biol Reprod*. 2003;68(1):10-18. doi:10.1095/biolreprod.102.005215
95. Lovejoy JC, Champagne CM, de Jonge L, Xie H, Smith SR. Increased visceral fat and decreased energy expenditure during the menopausal transition. *Int J Obes (Lond)*. 2008;32(6):949-958. doi:10.1038/ijo.2008.25
96. Pasquali R, Casimirri F, Labate AM, et al. Body weight, fat distribution and the menopausal status in women. The VMH Collaborative Group. *Int J Obes Relat Metab Disord*. 1994;18(9):614-621.
97. Revilla R, Revilla M, Villa LF, Cortes J, Arribas I, Rico H. Changes in body composition in women treated with gonadotropin-releasing hormone agonists. *Maturitas*. 1998;31(1):63-68. doi:10.1016/s0378-5122(98)00080-2
98. Gambacciani M, Ciaponi M, Cappagli B, Benussi C, De Simone L, Genazzani AR. Climacteric modifications in body weight and fat tissue distribution. *Climacteric*. 1999;2(1):37-44. doi:10.3109/13697139909025561
99. Sorensen MB. Changes in body composition at menopause—age, lifestyle or hormone deficiency? *J Br Menopause Soc*. 2002;8(4):137-140. doi:10.1258/136218002100321974
100. Jensen LB, Vestergaard P, Hermann AP, et al. Hormone replacement therapy dissociates fat mass and bone mass, and tends to reduce weight gain in early postmenopausal women: a randomized controlled 5-year clinical trial of the Danish Osteoporosis Prevention Study. *J Bone Miner Res*. 2003;18(2):333-342. doi:10.1359/jbmr.2003.18.2.333
101. Espeland MA, Stefanick ML, Kritz-Silverstein D, et al. Effect of postmenopausal hormone therapy on body weight and waist and hip girths. Postmenopausal estrogen-progestin interventions study investigators. *J Clin Endocrinol Metab*. 1997;82(5):1549-1556. doi:10.1210/jcem.82.5.3925
102. Day DS, Gozansky WS, Van Pelt RE, Schwartz RS, Kohrt WM. Sex hormone suppression reduces resting energy expenditure and [beta]-adrenergic support of resting energy expenditure. *J Clin Endocrinol Metab*. 2005;90(6):3312-3317. doi:10.1210/jc.2004-1344
103. Matsuo T, Saitoh S, Suzuki M. Effects of the menstrual cycle on excess postexercise oxygen consumption in healthy young women. *Metabolism*. 1999;48(3):275-277. doi:10.1016/s0026-0495(99)90071-9
104. Meijer GA, Westerterp KR, Saris WH, ten Hoor F. Sleeping metabolic rate in relation to body composition and the menstrual cycle. *Am J Clin Nutr*. 1992;55(3):637-640. doi:10.1093/ajcn/55.3.637
105. Solomon SJ, Kurzer MS, Calloway DH. Menstrual cycle and basal metabolic rate in women. *Am J Clin Nutr*. 1982;36(4):611-616. doi:10.1093/ajcn/36.4.611
106. Pelkman CL, Chow M, Heinbach RA, Rolls BJ. Short-term effects of a progestational contraceptive drug on food intake, resting energy expenditure, and body weight in young women. *Am J Clin Nutr*. 2001;73(1):19-26. doi:10.1093/ajcn/73.1.19
107. Melanson KJ, Saltzman E, Russell R, Roberts SB. Postabsorptive and postprandial energy expenditure and substrate oxidation do not change during the menstrual cycle in young women. *J Nutr*. 1996; 126(10):2531-2538. doi:10.1093/jn/126.10.2531
108. Lariviere F, Moussalli R, Garrel DR. Increased leucine flux and leucine oxidation during the luteal phase of the menstrual cycle in women. *Am J Physiol*. 1994;267(3 Pt 1):E422-E428. doi:10.1152/ajpendo.1994.267.3.E422
109. Leeners B, Geary N, Tobler PN, Asarian L. Ovarian hormones and obesity. *Hum Reprod Update*. 2017;23(3):300-321. doi:10.1093/humupd/dmw045
110. Paolisso G, Rizzo MR, Mazziotti G, et al. Lack of association between changes in plasma leptin concentration and in food intake during the menstrual cycle. *Eur J Clin Invest*. 1999;29(6):490-495. doi:10.1046/j.1365-2362.1999.00488.x
111. Piers LS, Diggavi SN, Rijkskamp J, van Raaij JM, Shetty PS, Hautvast JG. Resting metabolic rate and thermic effect of a meal in the follicular and luteal phases of the menstrual cycle in well-nourished Indian women. *Am J Clin Nutr*. 1995;61(2):296-302. doi:10.1093/ajcn/61.2.296
112. Tai MM, Castillo TP, Pi-Sunyer FX. Thermic effect of food during each phase of the menstrual cycle. *Am J Clin Nutr*. 1997;66(5):1110-1115. doi:10.1093/ajcn/66.5.1110
113. Li ET, Tsang LB, Lui SS. Resting metabolic rate and thermic effects of a sucrose-sweetened soft drink during the menstrual cycle in young Chinese women. *Can J Physiol Pharmacol*. 1999;77(7):544-550. doi:10.1139/y99-038
114. Ropero AB, Alonso-Magdalena P, Quesada I, Nadal A. The role of estrogen receptors in the control of energy and glucose homeostasis. *Steroids*. 2008;73(9-10):874-879. doi:10.1016/j.steroids.2007.12.018

115. Lopez M, Tena-Sempere M. Estradiol effects on hypothalamic AMPK and BAT thermogenesis: a gateway for obesity treatment? *Pharmacol Ther.* 2017;178:109-122. doi:10.1016/j.pharmthera.2017.03.014
116. Kaikaew K, Grefhorst A, Visser JA. Sex differences in brown adipose tissue function: sex hormones, glucocorticoids, and their crosstalk. *Front Endocrinol (Lausanne).* 2021;12:652444. doi:10.3389/fendo.2021.652444
117. Herz CT, Kulterer OC, Prager M, et al. Sex differences in brown adipose tissue activity and cold-induced thermogenesis. *Mol Cell Endocrinol.* 2021;534:111365. doi:10.1016/j.mce.2021.111365
118. Ouellet V, Routhier-Labadie A, Bellemare W, et al. Outdoor temperature, age, sex, body mass index, and diabetic status determine the prevalence, mass, and glucose-uptake activity of 18F-FDG-detected BAT in humans. *J Clin Endocrinol Metab.* 2011;96(1):192-199. doi:10.1210/jc.2010-0989
119. Brendle C, Werner MK, Schmadl M, et al. Correlation of brown adipose tissue with other body fat compartments and patient characteristics: a retrospective analysis in a large patient cohort using PET/CT. *Acad Radiol.* 2018;25(1):102-110. doi:10.1016/j.acra.2017.09.007
120. van den Beukel JC, Grefhorst A, Hoogduijn MJ, et al. Women have more potential to induce browning of perirenal adipose tissue than men. *Obesity (Silver Spring).* 2015;23(8):1671-1679. doi:10.1002/oby.21166
121. Bahler L, Verberne HJ, Admiraal WM, et al. Differences in sympathetic nervous stimulation of brown adipose tissue between the young and old, and the lean and obese. *J Nucl Med.* 2016;57(3):372-377. doi:10.2967/jnumed.115.165829
122. Valle A, Santandreu FM, Garcia-Palmer FJ, Roca P, Oliver J. The serum levels of 17beta-estradiol, progesterone and triiodothyronine correlate with brown adipose tissue thermogenic parameters during aging. *Cell Physiol Biochem.* 2008;22(1-4):337-346. doi:10.1159/000149812
123. Simerly RB, Chang C, Muramatsu M, Swanson LW. Distribution of androgen and estrogen receptor mRNA-containing cells in the rat brain: an in situ hybridization study. *J Comp Neurol.* 1990;294(1):76-95. doi:10.1002/cne.902940107
124. Osterlund M, Kuiper GG, Gustafsson JA, Hurd YL. Differential distribution and regulation of estrogen receptor-alpha and -beta mRNA within the female rat brain. *Brain Res Mol Brain Res.* 1998;54(1):175-180. doi:10.1016/s0169-328x(97)00351-3
125. Cornejo MP, Hentges ST, Maliqueo M, Coirini H, Becu-Villalobos D, Elias CF. Neuroendocrine regulation of metabolism. *J Neuroendocrinol.* 2016;28(7):7. doi:10.1111/jne.12395
126. Martinez de Morentin PB, Whittle AJ, Ferno J, et al. Nicotine induces negative energy balance through hypothalamic AMP-activated protein kinase. *Diabetes.* 2012;61(4):807-817. doi:10.2337/db11-1079
127. Morrison SF, Madden CJ, Tupone D. Central neural regulation of brown adipose tissue thermogenesis and energy expenditure. *Cell Metab.* 2014;19(5):741-756. doi:10.1016/j.cmet.2014.02.007
128. Cano G, Passerin AM, Schiltz JC, Card JP, Morrison SF, Sved AF. Anatomical substrates for the central control of sympathetic outflow to interscapular adipose tissue during cold exposure. *J Comp Neurol.* 2003;460(3):303-326. doi:10.1002/cne.10643
129. Uno T, Shibata M. Role of inferior olive and thoracic IML neurons in nonshivering thermogenesis in rats. *Am J Physiol Regul Integr Comp Physiol.* 2001;280(2):R536-R546. doi:10.1152/ajpregu.2001.280.2.R536
130. Carling D, Mayer FV, Sanders MJ, Gamblin SJ. AMP-activated protein kinase: nature's energy sensor. *Nat Chem Biol.* 2011;7(8):512-518. doi:10.1038/nchembio.610
131. Hardie DG. AMP-activated protein kinase: maintaining energy homeostasis at the cellular and whole-body levels. *Annu Rev Nutr.* 2014;34(1):31-55. doi:10.1146/annurev-nutr-071812-161148
132. Hardie DG. AMPK—sensing energy while talking to other signaling pathways. *Cell Metab.* 2014;20(6):939-952. doi:10.1016/j.cmet.2014.09.013
133. Lopez M, Nogueiras R, Tena-Sempere M, Dieguez C. Hypothalamic AMPK: a canonical regulator of whole-body energy balance. *Nat Rev Endocrinol.* 2016;12(7):421-432. doi:10.1038/nrendo.2016.67
134. Martinez de Morentin PB, Urisarri A, Couce ML, Lopez M. Molecular mechanisms of appetite and obesity: a role for brain AMPK. *Clin Sci (Lond).* 2016;130(19):1697-1709. doi:10.1042/CS20160048
135. Whittle AJ, Carobbio S, Martins L, et al. BMP8B increases brown adipose tissue thermogenesis through both central and peripheral actions. *Cell.* 2012;149(4):871-885. doi:10.1016/j.cell.2012.02.066
136. Beiroa D, Imbernon M, Gallego R, et al. GLP-1 agonism stimulates brown adipose tissue thermogenesis and browning through hypothalamic AMPK. *Diabetes.* 2014;63(10):3346-3358. doi:10.2337/db14-0302
137. Tanida M, Yamamoto N, Shibamoto T, Rahmouni K. Involvement of hypothalamic AMP-activated protein kinase in leptin-induced sympathetic nerve activation. *PLoS ONE.* 2013;8(2):e56660. doi:10.1371/journal.pone.0056660
138. Contreras C, Gonzalez-Garcia I, Martinez-Sanchez N, et al. Central ceramide-induced hypothalamic lipotoxicity and ER stress regulate energy balance. *Cell Rep.* 2014;9(1):366-377. doi:10.1016/j.celrep.2014.08.057
139. Gonzalez-Garcia I, Contreras C, Estevez-Salguero A, et al. Estradiol regulates energy balance by ameliorating hypothalamic ceramide-induced ER stress. *Cell Rep.* 2018;25(2):413-423 e5. doi:10.1016/j.celrep.2018.09.038
140. Meneyrol K, Estevez-Salguero A, Gonzalez-Garcia I, et al. Ovarian insufficiency impairs glucose-stimulated insulin secretion through activation of hypothalamic de novo ceramide synthesis. *Metabolism.* 2021;123:154846. doi:10.1016/j.metabol.2021.154846
141. Santos RS, Frank AP, Fatima LA, Palmer BF, Oz OK, Clegg DJ. Activation of estrogen receptor alpha induces beiging of adipocytes. *Mol Metab.* 2018;18:51-59. doi:10.1016/j.molmet.2018.09.002
142. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ.* 2021;372:n71. doi:10.1136/bmj.n71
143. Rethlefsen ML, Kirtley S, Waffenschmidt S, et al. PRISMA-S: an extension to the PRISMA statement for reporting literature searches in systematic reviews. *Syst Rev.* 2021;10(1):39. doi:10.1186/s13643-020-01542-z
144. Sterne JAC, Savovic J, Page MJ, et al. RoB 2: a revised tool for assessing risk of bias in randomised trials. *BMJ.* 2019;366:l4898. doi:10.1136/bmj.l4898
145. Deeks JJ, Dinnes J, D'Amico R, et al. Evaluating non-randomised intervention studies. *Health Technol Assess.* 2003;7(27):iii-x, 1-173. doi:10.3310/hta7270
146. Eck LH, Bennett AG, Egan BM, et al. Differences in macronutrient selections in users and nonusers of an oral contraceptive. *Am J Clin Nutr.* 1997;65(2):419-424. doi:10.1093/ajcn/65.2.419
147. Anderson EJ, Lavoie HB, Strauss CC, Hubbard JL, Sharpless JL, Hall JE. Body composition and energy balance: lack of effect of short-term hormone replacement in postmenopausal women. *Metabolism.* 2001;50(3):265-269. doi:10.1053/meta.2001.21015
148. Aubertin-Leheudre M, Goulet ED, Dionne IJ. Enhanced rate of resting energy expenditure in women using hormone-replacement therapy: preliminary results. *J Aging Phys Act.* 2008;16(1):53-60. doi:10.1123/japa.16.1.53

149. Diffey B, Piers LS, Soares MJ, O'Dea K. The effect of oral contraceptive agents on the basal metabolic rate of young women. *Br J Nutr*. 1997;77(6):853-862. doi:[10.1079/bjn19970084](https://doi.org/10.1079/bjn19970084)
150. Duhita MR, Schutz Y, Montani JP, Dulloo AG, Miles-Chan JL. Oral contraceptive pill alters acute dietary protein-induced thermogenesis in Young women. *Obesity (Silver Spring)*. 2017;25(9):1482-1485. doi:[10.1002/oby.21919](https://doi.org/10.1002/oby.21919)
151. Duhita MR, Schutz Y, Montani JP, Dulloo AG, Miles-Chan JL. Assessment of the dose-response relationship between meal protein content and postprandial thermogenesis: effect of sex and the Oral contraceptive pill. *Nutrients*. 2019;11(7):1599. doi:[10.3390/nu11071599](https://doi.org/10.3390/nu11071599)
152. Kimm SY, Glynn NW, Aston CE, Poehlman ET, Daniels SR. Effects of race, cigarette smoking, and use of contraceptive medications on resting energy expenditure in young women. *Am J Epidemiol*. 2001; 154(8):718-724. doi:[10.1093/aje/154.8.718](https://doi.org/10.1093/aje/154.8.718)
153. Melanson EL, Gavin KM, Shea KL, et al. Regulation of energy expenditure by estradiol in premenopausal women. *J Appl Physiol*. 2015; 119(9):975-981. doi:[10.1152/jappphysiol.00473.2015](https://doi.org/10.1152/jappphysiol.00473.2015)
154. Piers LS, Diffey B, Soares MJ, et al. The validity of predicting the basal metabolic rate of young Australian men and women. *Eur J Clin Nutr*. 1997;51(5):333-337. doi:[10.1038/sj.ejcn.1600407](https://doi.org/10.1038/sj.ejcn.1600407)
155. Reimer RA, Debert CT, House JL, Poulin MJ. Dietary and metabolic differences in pre- versus postmenopausal women taking or not taking hormone replacement therapy. *Physiol Behav*. 2005;84(2):303-312. doi:[10.1016/j.physbeh.2004.12.011](https://doi.org/10.1016/j.physbeh.2004.12.011)
156. Ravussin E, Lillioja S, Knowler WC, et al. Reduced rate of energy expenditure as a risk factor for body-weight gain. *N Engl J Med*. 1988;318(8):467-472. doi:[10.1056/NEJM198802253180802](https://doi.org/10.1056/NEJM198802253180802)
157. Manini TM. Energy expenditure and aging. *Ageing Res Rev*. 2010; 9(1):1-11. doi:[10.1016/j.arr.2009.08.002](https://doi.org/10.1016/j.arr.2009.08.002)
158. Cypess AM, Weiner LS, Roberts-Toler C, et al. Activation of human brown adipose tissue by a beta3-adrenergic receptor agonist. *Cell Metab*. 2015;21(1):33-38. doi:[10.1016/j.cmet.2014.12.009](https://doi.org/10.1016/j.cmet.2014.12.009)
159. U Din M, Raiko J, Saari T, et al. Human brown adipose tissue [(15)O] O2 PET imaging in the presence and absence of cold stimulus. *Eur J Nucl Med Mol Imaging*. 2016;43(10):1878-1886. doi:[10.1007/s00259-016-3364-y](https://doi.org/10.1007/s00259-016-3364-y)
160. Muzik O, Mangner TJ, Leonard WR, Kumar A, Janisse J, Granneman JG. 15O PET measurement of blood flow and oxygen consumption in cold-activated human brown fat. *J Nucl Med*. 2013; 54(4):523-531. doi:[10.2967/jnumed.112.111336](https://doi.org/10.2967/jnumed.112.111336)
161. Carpentier AC, Blondin DP, Virtanen KA, Richard D, Haman F, Turcotte EE. Brown adipose tissue energy metabolism in humans. *Front Endocrinol (Lausanne)*. 2018;9:447. doi:[10.3389/fendo.2018.00447](https://doi.org/10.3389/fendo.2018.00447)
162. Cypess AM. Reassessing human adipose tissue. *N Engl J Med*. 2022; 386(8):768-779. doi:[10.1056/NEJMra2032804](https://doi.org/10.1056/NEJMra2032804)
163. Sanghvi A, Redman LM, Martin CK, Ravussin E, Hall KD. Validation of an inexpensive and accurate mathematical method to measure long-term changes in free-living energy intake. *Am J Clin Nutr*. 2015; 102(2):353-358. doi:[10.3945/ajcn.115.111070](https://doi.org/10.3945/ajcn.115.111070)
164. Blondin DP, Labbe SM, Tingelstad HC, et al. Increased brown adipose tissue oxidative capacity in cold-acclimated humans. *J Clin Endocrinol Metab*. 2014;99(3):E438-E446. doi:[10.1210/jc.2013-3901](https://doi.org/10.1210/jc.2013-3901)
165. Blondin DP, Tingelstad HC, Noll C, et al. Dietary fatty acid metabolism of brown adipose tissue in cold-acclimated men. *Nat Commun*. 2017;8(1):14146. doi:[10.1038/ncomms14146](https://doi.org/10.1038/ncomms14146)
166. Ricquier D, Nechad M, Mory G. Ultrastructural and biochemical characterization of human brown adipose tissue in pheochromocytoma. *J Clin Endocrinol Metab*. 1982;54(4):803-807. doi:[10.1210/jcem-54-4-803](https://doi.org/10.1210/jcem-54-4-803)
167. Fukuchi K, Tatsumi M, Ishida Y, Oku N, Hatazawa J, Wahl RL. Radio-nuclide imaging metabolic activity of brown adipose tissue in a patient with pheochromocytoma. *Exp Clin Endocrinol Diabetes*. 2004; 112(10):601-603. doi:[10.1055/s-2004-830407](https://doi.org/10.1055/s-2004-830407)
168. Eckel LA. The ovarian hormone estradiol plays a crucial role in the control of food intake in females. *Physiol Behav*. 2011;104(4):517-524. doi:[10.1016/j.physbeh.2011.04.014](https://doi.org/10.1016/j.physbeh.2011.04.014)

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Weidlinger S, Winterberger K, Pape J, et al. Impact of estrogens on resting energy expenditure: A systematic review. *Obesity Reviews*. 2023;e13605. doi:[10.1111/obr.13605](https://doi.org/10.1111/obr.13605)