

# HIV-1 drug resistance in people on dolutegravir-based ART:

## A collaborative cohort analysis

Tom Loosli<sup>1,2</sup>, Stefanie Hossmann<sup>3</sup>, Suzanne M. Ingle<sup>4</sup>, Hajra Okhai<sup>5</sup>, Katharina Kusejko<sup>1,2</sup>,  
Johannes Mouton<sup>6</sup>, Pantxika Bellecave<sup>7</sup>, Ard van Sighem<sup>8</sup>, Melanie Stecher<sup>9,10</sup>, Antonella d'Arminio  
Monforte<sup>11</sup>, M. John Gill<sup>12,13</sup>, Caroline A. Sabin<sup>5</sup>, Gary Maartens<sup>6</sup>, Huldrych F. Günthard<sup>1,2</sup>, Jonathan A.  
C. Sterne<sup>4</sup>, Richard Lessells<sup>14,15,\*</sup>, Matthias Egger<sup>3,4,16,\*</sup>, Roger Kouyos<sup>1,2,\*</sup>

\* Authors contributed equally

1. Department of Infectious Diseases and Hospital Epidemiology, University Hospital Zurich, Zurich, Switzerland (Tom Loosli Msc, Katharina Kusejko PhD, Prof Huldrych F. Günthard MD, Prof Roger Kouyos PhD)
2. Institute of Medical Virology, University of Zurich, Zurich, Switzerland (Tom Loosli, Katharina Kusejko, Prof Huldrych F. Günthard, Prof Roger Kouyos)
3. Institute of Social and Preventive Medicine (ISPM), University of Bern, Switzerland (Stefanie Hossmann Msc, Prof Matthias Egger MD)
4. Population Health Sciences, Bristol Medical School, University of Bristol, UK (Suzanne M. Ingle PhD, Prof Jonathan A. C. Sterne PhD, Prof Matthias Egger)
5. Institute for Global Health, University College London, UK (Hajra Okhai PhD, Prof Caroline A. Sabin PhD)
6. Department of Medicine, University of Cape Town, Cape Town, South Africa (Johannes Mouton MD, Prof Gary Maartens MD)
7. Virology laboratory, University Hospital Bordeaux, Bordeaux, France (Pantxika Bellecave PhD)
8. Stichting hiv monitoring, Amsterdam, the Netherlands (Ard van Sighem PhD)
9. German Center for Infection Research (DZIF), Partner-Site Cologne-Bonn, Cologne, Germany (Melanie Stecher PhD)
10. University of Cologne, Faculty of Medicine and University Hospital Cologne, Department I of Internal Medicine, Center for Integrated Oncology Aachen Bonn Cologne Duesseldorf (Melanie Stecher)
11. Italian Cohort Naive Antiretrovirals, (ICONA) L'Azienda Socio Sanitaria Territoriale (ASST) Santi Paolo e Carlo, Milano, Italy (Prof Antonella d'Arminio Monforte MD)
12. Southern Alberta Clinic, Calgary, AB, Canada (Prof M. John Gill MD)
13. Department of Medicine, University of Calgary, Calgary, AB, Canada (Prof M. John Gill)
14. KwaZulu-Natal Research Innovation and Sequencing Platform (KRISP), University of KwaZulu-Natal, Durban, South Africa (Prof Richard Lessells, PhD)
15. Centre for the AIDS Programme of Research in South Africa (CAPRISA), Durban, South Africa (Prof Richard Lessells)
16. Centre for Infectious Disease Epidemiology and Research, Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa (Prof Matthias Egger)

Correspondence to: Matthias Egger, [matthias.egger@unibe.ch](mailto:matthias.egger@unibe.ch)

43 **Word counts:** Abstract 334 words, research in context 425 words, main text 3637 words, 2 tables, 4  
44 figures, 33 references, online appendix with 20 pages.

## 45 **Summary**

46

47 **Background:** The widespread use of the integrase strand transfer inhibitor (INSTI) dolutegravir (DTG)  
48 in first- and second-line antiretroviral therapy (ART) may facilitate emerging resistance. The DTG  
49 RESIST study combined data from HIV cohorts to examine patterns of drug resistance mutations  
50 (DRMs) and identify risk factors for DTG resistance.

51 **Methods:** We included cohorts with INSTI resistance data from two collaborations (ART Cohort  
52 Collaboration, International epidemiology Databases to Evaluate AIDS in Southern Africa), and the UK  
53 Collaborative HIV Cohort. Eight cohorts from Canada, France, Germany, Italy, the Netherlands,  
54 Switzerland, South Africa, and the UK contributed data on individuals who were viraemic on DTG-  
55 based ART and underwent genotypic resistance testing. Individuals with unknown DTG initiation date  
56 were excluded. Resistance levels were categorised using the Stanford algorithm. We identified risk  
57 factors for resistance using mixed-effects ordinal logistic regression models.

58 **Findings:** We included 599 people with genotypic resistance testing on DTG-based ART between 2013  
59 and 2022. Most had HIV-1 subtype B (N=351, 58·6%), a third had been exposed to first-generation  
60 INSTIs (N=193, 32·2%); 70 (11·7%) were on DTG dual therapy, and 18 (3·0%) on DTG monotherapy.  
61 INSTI DRMs were detected in 86 (14·4%) individuals; 20 (3·3%) had more than one mutation. Most  
62 (N=563, 94·0%) were susceptible to DTG, 7 (1·2%) had potential-low, 6 (1·0%) low, 17 (2·8%)  
63 intermediate and 6 (1·0%) high-level DTG resistance. The risk of DTG resistance was higher on DTG  
64 monotherapy (adjusted odds ratio (aOR) 34·1, 95% CI 9·93 to 117) and DTG lamivudine dual therapy  
65 (aOR 9·21, 95% CI 2·20 to 38·6) compared to combination ART, and in the presence of potential-  
66 low/low (aOR 5·23, 95% CI 1·32 to 20·7) or intermediate/high-level (aOR 13·4, 95% CI 4·55 to 39·7)  
67 nucleoside reverse transcriptase inhibitors (NRTI) resistance.

68 **Interpretation:** Among people experiencing viraemia on DTG-based ART, INSTI DRMs and DTG  
69 resistance were rare. NRTI resistance substantially increased the risk for DTG resistance, which is of  
70 concern, notably in resource-limited settings. Monitoring is important to prevent resistance at the  
71 individual and population level and ensure the long-term sustainability of ART.

72 **Funding:** US National Institutes of Health, Swiss National Science Foundation.

## 73 **Research in context**

### 74 **Evidence before this study**

75 We searched SCOPUS on 20 March 2023 for all publications from inception using the terms  
76 “dolutegravir” or “DTG”, “resistant” or “resistance”, and “HIV”. The available evidence on resistance  
77 evolution in people living with HIV (PLHIV) with virological failure on DTG-based ART is limited. Most  
78 studies assessed the efficacy of DTG-based regimens in clinical studies. They reported drug resistance  
79 in individuals experiencing virological failure as a secondary objective or reported single or multiple  
80 cases of individuals developing resistance on DTG-based ART. Clinical trials such as the NADIA trial  
81 showed high viral suppression even in people with NRTI resistance. Consequently, previous analyses  
82 included only a few people experiencing failure on DTG; the SINGLE trial with 39 people with virologic  
83 failure on DTG was the largest. The highest number of individuals with DTG resistance was nine study  
84 participants in the NADIA trial. There is evidence that DTG resistance in PLHIV on a DTG monotherapy  
85 may be more likely, and studies suggest that HIV-1 subtype and mutations acquired during a first-  
86 generation INSTI-based regimen might affect the risk of DTG resistance.

### 87 **Added value of this study**

88 To our knowledge, DTG RESIST is the first study systematically investigating resistance in PLHIV  
89 experiencing viraemia on DTG-based ART using a multi-cohort collaboration design reflecting real-  
90 world routine care. We collected genotypic resistance tests and clinical data from eight observational  
91 HIV cohorts. This resulted in a large dataset of PLHIV experiencing viraemia on a DTG regimen (599  
92 individuals). It allowed a robust assessment of drug resistance mutations and risk factors for DTG  
93 resistance. Cross-resistance of first-generation INSTIs does not appear to explain the mutation  
94 patterns in PLHIV who experience virological failure on DTG-based ART regimens. PLHIV who received  
95 DTG monotherapy or DTG lamivudine dual therapy and those infected with non-B subtypes were more  
96 likely to develop resistance. Resistance to NRTIs was a major risk factor for DTG resistance, indicating  
97 that PLHIV receiving functional monotherapy are more likely to develop DTG resistance.

### 98 **Implications of all the available evidence**

99 HIV-1 drug resistance is a significant threat to the sustainability of current and future antiretroviral  
100 therapy for combating the ongoing HIV-1 pandemic. Our collaborative analysis shows that cases of  
101 DTG resistance are rare at present but not negligible. Given the global DTG roll-out, this might lead to  
102 increased frequencies and transmission of DTG resistance, particularly in PLHIV with resistance to  
103 NRTIs. While the evidence regarding subtype differences is tentative, it indicates that non-B subtypes,

104 which are most relevant for the global roll-out of DTG, might be associated with an increased risk of  
105 resistance.

## 106 **Introduction**

107 The integrase strand transfer inhibitor (INSTI) dolutegravir (DTG) was approved in 2013 in the United  
108 States and shortly afterwards in the European Union to treat HIV-1. In 2019, the WHO recommended  
109 DTG as the preferred drug for first-line and second-line antiretroviral therapy (ART) in all populations,  
110 including pregnant women and those of childbearing age. Since then, DTG-based ART was rolled out  
111 globally,<sup>1</sup> with about 100 countries including DTG in their treatment guidelines by mid 2020.<sup>2</sup>

112 DTG has a high genetic barrier to resistance,<sup>3,4</sup> and relatively few people living with HIV (PLHIV) are so  
113 far known to have developed resistance.<sup>5-7</sup> The mutations leading to DTG resistance may differ  
114 between HIV-1 subtypes. In PLHIV without prior exposure to INSTI-based ART, DTG resistance is mainly  
115 associated with the Arg263Lys mutation,<sup>8,9</sup> which was observed in three cases of DTG resistance in the  
116 NADIA trial.<sup>10</sup> The Asn155His mutation was present in two individuals with subtype A and C in the  
117 SAILING trial,<sup>11</sup> while the Gly118Arg mutation appears to be facilitated by a natural polymorphism in  
118 subtype C.<sup>12</sup> In a recent study in Ethiopia, the Gln148His/Lys/Arg mutation was found to be less  
119 prevalent in subtype C.<sup>13</sup> Pre-existing mutations, such as those acquired during a first-generation INSTI  
120 regimen, may directly confer resistance to DTG or facilitate the accumulation of additional  
121 mutations.<sup>14,15</sup>

122 The risk factors and the mutational patterns that confer resistance to DTG in vivo are less well  
123 established than for older antiretroviral drugs.<sup>16</sup> The widespread use of DTG in resource-limited  
124 settings, where ART regimens are highly standardised, drugs are recycled, access to adherence  
125 support, viral load and resistance testing is limited, and the risk for drug stock-outs is higher, may  
126 facilitate the emergence of resistance. In the DTG RESIST study, we combined data from European,  
127 North American, and South African cohorts to identify risk factors for DTG resistance and examine the  
128 patterns of resistance mutations across different HIV-1 subtypes.

## 129 **Methods**

### 130 **Study design and population**

131 The DTG RESIST project was discussed in two HIV cohort collaborations: the ART Cohort Collaboration  
132 (ART-CC)<sup>17</sup> and the International epidemiology Databases to Evaluate AIDS (IeDEA)<sup>18</sup> in Southern  
133 Africa. Six of the 21 ART-CC cohorts participated: The Agence Nationale de la Recherche sur le SIDA et  
134 les hépatites virales (ANRS CO3), Aquitaine Cohort, the AIDS Therapy Evaluation in the Netherlands  
135 cohort (ATHENA), the Köln/Bonn Cohort (CBC), Germany, the Italian Cohort of Antiretroviral-Naïve  
136 Patients (ICONA), the South Alberta Clinic Cohort (SAC), Canada, and the Swiss HIV Cohort Study  
137 (SHCS). The main reason for the non-participation of the other cohorts was the lack of access to

138 resistance data. The UK Collaborative HIV Cohort (UK CHIC) Study and linked UK HIV Drug Resistance  
139 Database (UKHDRD), although not formally part of the ART-CC collaboration, also joined. In leDEA  
140 Southern Africa, the South African Aid for AIDS (AfA) cohort was the only cohort with access to INSTI  
141 resistance data. The clinical data were provided by the data centres of the two cohort collaborations,  
142 ART-CC and leDEA, and the genotypic data by the cohorts. Genotypic data were the GRT consensus  
143 nucleotide sequences. There were two exceptions: AfA provided a list of mutations, and Aquitaine  
144 provided the Stanford resistance algorithm output. UK CHIC provided all data directly to the DTG  
145 RESIST study team. The appendix provides further details (p 2).

146 We included participants who underwent genotypic resistance testing from plasma HIV-1 RNA  
147 covering the integrase gene between two weeks after starting and up to two months after stopping  
148 any DTG-based regimen. The latest test was considered in the case of multiple genotypic resistance  
149 tests. Participants with unknown dates of initiation of DTG-based ART were excluded. The analysis of  
150 risk factors for DTG resistance was restricted to individuals with at least one year of follow-up,  
151 ensuring the availability of viral load data and assessment of viral load testing frequency.

152 The Human Research Ethics Committee of the University of Cape Town and the Cantonal Ethics  
153 Committee of the Canton of Bern granted permission to analyse these data.

## 154 **Procedures**

155 We determined HIV-1 subtypes from the integrase gene using COMET (COntext-based Modeling for  
156 Expeditious Typing)<sup>19</sup> and REGA.<sup>20</sup> If REGA and COMET output differed, the subtype with higher  
157 support was assigned. As nucleotide sequences were not available for AfA, we used subtype  
158 information from the cohort based on reverse transcriptase (RT) and protease. For Aquitaine,  
159 information on subtype was used where available and otherwise considered unknown. The Aquitaine  
160 subtypes were characterised locally using Blast analysis on Smartgene HIV module on at least two  
161 genes. In the analysis, we grouped HIV-1 subtypes other than the four most common subtypes in the  
162 study population (B, C, A, G) as 'other' (F, AD, AE, D, 06\_CPX, 18\_CPX, unknown) (appendix p 3).

163 Individuals prescribed raltegravir or elvitegravir before starting the DTG-based regimen were  
164 considered exposed to first-generation INSTIs. Viral load testing frequency was calculated for  
165 individuals with more than one year of follow-up before the Genotypic Resistance Test (GRT). We  
166 quantified HIV-1 viral load as the area under the curve (AUC) of the log<sub>10</sub>-transformed viral load  
167 measurements from DTG initiation to the GRT sample date. To account for differences in detection  
168 limits, we set any viral load measurement below 50 to 0 copies per mL. For individuals who initiated  
169 ART with the DTG-based regimen, we excluded viral loads at ART initiation by setting measurements

170 within the first 180 days from the first HIV-1 RNA measurement to 0. Time on DTG-based ART was  
171 calculated in years from DTG initiation to GRT. The ART regimen at GRT was the regimen an individual  
172 took 14 days before the test. If available, GRT results from earlier time points were used to assess  
173 prior NRTI resistance. We defined monotherapy as ART consisting of DTG only. DTG dual therapy was  
174 defined as DTG combined with a second antiretroviral drug. DTG-based regimens comprised of DTG  
175 and two or more antiretroviral drugs were considered triple or intensified regimens, respectively.

## 176 **Outcomes**

177 We defined two HIV drug resistance outcomes: the level of resistance to DTG and the presence of  
178 known drug resistance mutations (DRMs). The Stanford HIV Database version 9.0 and the Stanford  
179 HIVdb algorithm<sup>21</sup> were used to categorise drug resistance levels as susceptible (score below 10),  
180 potential low (10-14), low (15-29), intermediate (30-59) or high ( $\geq 60$ ). We defined INSTI-DRMs<sup>22</sup> as all  
181 mutations associated with INSTIs by the Stanford HIVdb algorithm, including major and accessory  
182 mutations. We used the same approach to assess resistance to all other antiretroviral drugs, whereby  
183 drug resistance to tenofovir alafenamide (not covered by the Stanford algorithm) was considered  
184 equal to tenofovir disoproxil fumarate resistance. Resistance to non-nucleoside reverse transcriptase  
185 inhibitors (NNRTIs) was calculated as the median of the scores for efavirenz, etravirine, nevirapine,  
186 and rilpivirine. Finally, we calculated resistance to nucleoside reverse transcriptase inhibitors (NRTIs)  
187 as the median of abacavir, zidovudine, emtricitabine/lamivudine, and tenofovir disoproxil fumarate  
188 scores. We used alternative definitions in sensitivity analyses.

## 189 **Statistical analysis**

190 We used descriptive statistics to present the characteristics of the study population and the different  
191 INSTI drug-resistance mutations. A negative binomial generalised linear model, adjusting for HIV-1  
192 subtype, exposure to first-generation INSTIs, and sex, was used to analyse the number of major and  
193 accessory INSTI drug-resistance mutations. We used ordinal logistic regression to identify risk factors  
194 for developing resistance, including cohort as a random effect. We considered variables based on  
195 availability and clinical relevance. We included sex, age at initiation and time on the DTG-based  
196 regimen, HIV-1 subtype, type of ART (combination ART based on three drugs or more, DTG lamivudine  
197 dual therapy, other DTG dual therapy, or monotherapy), exposure to first-generation INSTIs, HIV-1  
198 viral load, viral load testing frequency, and resistance to NRTIs. If the sequencing did not cover the RT,  
199 the missing data was included as a separate category. All analyses were performed in R, version 4.0.5.

200 We performed several sensitivity analyses. First, we replaced the NRTI resistance variable with the  
201 presence or absence of the Met184Val/Ile mutation (sensitivity analysis S1). Further, we performed



202 logistic regression using the same covariables as in the main risk factor analysis, using susceptible  
203 versus any DTG resistance as the outcome (S2). We also considered DTG resistance according to the  
204 WHO definition, whereby potential low is considered susceptible (S3). We repeated the risk factor  
205 analyses excluding study participants where RT was not sequenced (S4). Given the widespread use of  
206 tenofovir disoproxil fumarate-lamivudine-dolutegravir (TLD), we restricted the analysis of NRTIs to  
207 tenofovir disoproxil fumarate and lamivudine, using the higher resistance level of the two to quantify  
208 NRTI resistance (S5). In the subset of people on a DTG + 2 NRTI regimen, we calculated NRTI resistance  
209 specific to the two NRTIs used in each participant (S6). The main analysis could not assess whether  
210 NRTI and NNRTI resistance mutations pre-existed or were acquired on DTG. Sensitivity analysis S7  
211 restricted the study population to participants with available GRTs before experiencing viraemia on  
212 the DTG-containing regimen.

### 213 **Role of the funding source**

214 The funders of the study had no role in study design, data collection, data analysis, data interpretation,  
215 or writing of the report.

## 216 **Results**

217 A total of 599 people met the eligibility criteria and were included in the analysis of mutations  
218 conferring resistance to DTG; 540 (90%) had more than one year of follow-up since starting the DTG-  
219 based regimen and were included in the analysis of risk factors for DTG resistance. Table S3 and table  
220 S4 (appendix p. 4 and p. 5) show the number of participants included in the risk factor analysis by  
221 presence or absence of INSTI DRMs, and by DTG resistance levels, respectively.

222 The study participants included in the two analyses – mutations conferring DTG resistance and risk  
223 factors for DTG resistance – were similar (**Table 1**): most participants were men living with HIV-1  
224 subtype B who were on combination ART with three or more antiretroviral drugs (see appendix p 3  
225 for details on ART regimens). The median year of starting DTG was 2016. People had been on DTG for  
226 a median of 1.4 years at the time of genotypic resistance testing, and the median AUC of log<sub>10</sub> viral  
227 load (copies per mL) accumulated during this period was 3.6. The first GRT was performed on 22. May  
228 2013, and the last on 20. December 2021. About a third of participants had previously been exposed  
229 to first-generation INSTIs; most were exposed to raltegravir (142/193), followed by elvitegravir  
230 (38/193), and both elvitegravir and raltegravir (13/193). A total of 129 people did not have a CD4  
231 measurement within a year of the GRT, ten did not have any recorded HIV-1 RNA measurements  
232 before the GRT, and in 62 people sequencing did not cover RT.

233 At least one major or accessory INSTI DRM was found in 86 (14%) of the 599 study participants; 20  
234 (3%) had more than one mutation (appendix p 6-7). The proportion of study participants with any  
235 INSTI DRM was similar between first-generation INSTI exposed and unexposed individuals (28 out of  
236 86 (33%) and 165 out of 513 (32%), respectively). Most (563; 94%) study participants were fully  
237 susceptible to DTG, with potential low, low, intermediate, and high levels of DTG resistance being  
238 observed in 7 (1%), 6 (1%), 17 (2%) and 6 (1%), respectively (**Figure 1**).

239 The most common major INSTI DRM was Arg263Lys (N = 10), which only once occurred with another  
240 major INSTI DRM (appendix p 6). Other common major mutations included Gly140Lys/Arg/Ser (N = 9),  
241 Asn155His (N = 9), Gln148His/Arg (N = 6), and Glu138Lys (N = 7). The Gly118Arg, which has the  
242 strongest impact on susceptibility to DTG, was only observed three times. Among accessory DRMs,  
243 Glu157Gln (N = 23) and Thr97Ala (N = 18) were the most common. The distribution of INSTI resistance  
244 mutations was similar in people previously exposed to first-generation INSTIs and those not exposed  
245 (**Figure 2**). There was no statistically significant association of specific DRMs with first-generation INSTI  
246 experience. For HIV-1 subtype, we found a significant association after adjusting for multiple testing  
247 for the accessory INSTI DRM Thr97Ala (adjusted p-value = 0.015, see appendix p 8). This DRM occurred  
248 in 6 of 54 (11%) people with HIV-1 subtype A, 4 of 42 (10%) people with subtype G, 6 of 351 (2%)  
249 people with subtype B, and 0 of 69 people with HIV-1 subtype C.

250 The results from the negative binomial model of the number of mutations showed little evidence of a  
251 difference between HIV-1 subtypes. The total INSTI DRM count (including both accessory and major  
252 DRMs) was higher in first-generation INSTI-exposed people (adjusted RR 1.59, 95% CI 0.98 to 2.59)  
253 (**Error! Reference source not found.**). This association became stronger when considering only the  
254 number of major INSTI DRMs (adjusted RR 2.67, 95% CI 1.25 to 5.87) (see appendix p 9 for further  
255 details).

256 The prevalence of predicted resistance (low, intermediate or high) to NRTIs and NNRTIs was  
257 substantially higher in the presence of DTG resistance (Table 2). Among GRTs with coverage of the RT,  
258 the prevalence of at least low-level NRTI resistance was 7% overall (39 of 530), but 32% (7 of 22)  
259 among those with DTG resistance. The corresponding figures for NNRTI resistance were 15% (82 of  
260 530) and 50% (11 of 22).

261 The risk of DTG resistance was higher on DTG monotherapy compared to combination ART with three  
262 or more drugs (adjusted odds ratio [aOR] 34.09, 95% CI 9.93 to 117.01) and for DTG lamivudine dual  
263 regimen (aOR 9.21, 95% CI 2.20 to 38.55) (**Error! Reference source not found.**). The risk of resistance  
264 was also increased in the presence of a potential low/low level of NRTI resistance (aOR 5.23, 95% CI  
265 1.32 to 20.71) or intermediate/high level (aOR 13.44, 95% CI 4.55 to 39.68), compared to no NRTI

266 resistance. Non-B HIV-1 subtypes were also associated with increased resistance, particularly subtype  
267 A (aOR 3.12, 95% CI 0.84 to 11.61 compared to subtype B), but associations were not statistically  
268 significant (appendix p 10). Similarly, there was weak, statistically non-significant evidence for an  
269 association of viral load with DTG resistance (aOR 1.42, 95% CI 0.92 to 2.19 per standard deviation of  
270 the  $\log_{10}$  virus load area under the curve).

271 The results of the risk factor analyses were similar when replacing the NRTI resistance variable with  
272 the Met184Val/Ile mutation (sensitivity analysis S1, N=540/540, appendix p 12), when analysing  
273 susceptible versus any DTG resistance as the outcome in a logistic regression (S2, N=540/540,  
274 appendix p 13), or when considering DTG resistance levels following the WHO definition, where  
275 potential low-level resistance is considered susceptible (S3, N=540/540, appendix p 14). The exclusion  
276 of 58 individuals with missing RT sequences allowed the inclusion of both NRTI and NNRTI resistance  
277 in the model. The results for NRTI resistance were similar, and intermediate/high-level NNRTI  
278 resistance was also associated with DTG resistance (adjusted OR 2.72, 95% CI 0.94 to 7.86) (S4,  
279 N=482/540, appendix p 15). Results were similar when excluding individuals on DTG monotherapy (S5,  
280 N=474/540, appendix p 16). The analysis restricted to lamivudine and tenofovir disoproxil fumarate  
281 (S6, N=540/540, appendix p 17) confirmed that DTG resistance was associated with both potential  
282 low/low and intermediate/high-level resistance to these NRTI drugs. Similarly, when restricting the  
283 analysis to people on a DTG regimen with two NRTIs, we found similar results for the specific NRTIs  
284 (S7, N=309/540, appendix p 18). Finally, in sensitivity analysis S8, we used data on pre-existing NRTI  
285 resistance and found that DTG resistance was associated with prior intermediate/high NRTI resistance  
286 (N=356/540, appendix p 19).

## 287 **Discussion**

288 In this collaborative analysis of eight large cohort studies, we identified INSTI DRMs in 86 of 599 (14%)  
289 PLHIV with a genotypic resistance test while viraemic on DTG-based ART. Resistance to DTG according  
290 to the Stanford algorithm was present in 36 (6%) individuals. DTG resistance was associated with DTG  
291 monotherapy, lamivudine DTG dual therapy, and resistance to NRTIs. Exposure to first-generation  
292 INSTI was associated both with more resistance mutations and higher levels of DTG resistance, but  
293 the association with DTG resistance was not statistically significant. A wide range of INSTI DRMs was  
294 present. The polymorphic accessory INSTI DRM Thr97Ala was detected more frequently in subtypes A  
295 and G (compared to subtypes B and C), consistent with previously reported data.<sup>23</sup> The major INSTI  
296 DRMs Gly140Lys/Arg/Ser and Gln148His/Arg were detected in 5 out of 6 people with high-level DTG  
297 resistance.

298 DTG monotherapy, DTG lamivudine dual therapy and resistance to the NRTI backbone were most  
299 strongly associated with DTG resistance in our study. The complete sequence analysis, which allowed  
300 us to distinguish between NRTI and NNRTI resistance, suggests that the association may be mediated  
301 via NRTI resistance. It was robust when considering only 3TC and TDF resistance in a sensitivity  
302 analysis. As the main analysis was cross-sectional, it did not allow determining the timing of NRTI  
303 resistance relative to DTG resistance. However, an additional analysis in people with prior resistance  
304 tests suggests that NRTI resistance may often have predated DTG resistance. These results suggest  
305 that resistance to NRTI backbone drugs from previous regimens may have promoted the emergence  
306 of DTG resistance. However, its also possible that prior NRTI resistance reflects adherence issues,  
307 which may facilitate the emergence of DTG resistance.

308 The results from our study align with previous studies<sup>10,24,25</sup> that showed associations of DTG  
309 resistance with DTG monotherapy or NRTI resistance. By contrast, the NADIA trial found no evidence  
310 that resistance to NRTIs affects the effectiveness of DTG-based ART.<sup>10</sup> The NADIA trial does not,  
311 however, contradict the results of our study because outcomes differed: NADIA examined the risk of  
312 virological failure, whereas our study focused on the risk of DTG resistance among PLHIV tested for  
313 drug resistance while on DTG-based ART. Resistance to NRTIs may not impact treatment failure risk  
314 but still increase the chances of acquiring resistance in case of failure. Research on other drug classes<sup>26</sup>  
315 indicates that drug regimens with high and low genetic barriers can have similar failure rates but  
316 different probabilities of acquiring resistance-

317 Our study contributes important new information on DTG resistance in PLHIV receiving different DTG-  
318 based ART regimens by examining risk factors for DTG resistance in real-world cohort data from  
319 different settings. The cohort collaboration resulted in a large dataset of GRT results in people who  
320 experienced viraemia on DTG. Our results are central to informing HIV treatment and monitoring  
321 policies in the context of the continued expansion of DTG-based treatment regimens. The pooling of  
322 data from diverse routine clinical cohorts also has limitations. The participating cohorts include PLHIV  
323 in routine care but practices regarding when and for whom GRT is done will differ between cohorts.  
324 Further, the personalised approach to ART and HIV care in the European settings will not be  
325 generalisable to other settings, particularly to low- and middle-income countries. In our regression  
326 models, we accounted for this heterogeneity between cohorts by including cohort as a random effect,  
327 but confounding by cohort may still have affected our results. Furthermore, GRT before starting or  
328 switching ART may have prevented some individuals from receiving DTG, thus introducing selection  
329 bias. However, pre-treatment resistance to DTG was unlikely during the study period.<sup>27</sup>

330 A further limitation of our study is the dominance of HIV-1 subtype B, which was expected considering  
331 that our study population is comprised mainly of PLHIV from European countries, where subtype B  
332 predominates. More data from people with non-B subtypes are needed. The prospective arm of DTG  
333 RESIST is ongoing within the framework of the International epidemiology Databases to Evaluate AIDS  
334 (IeDEA):<sup>18</sup> individuals experiencing virologic failure on DTG-based ART are prospectively enrolled in  
335 around forty sites across sub-Saharan Africa, South America, and Asia. Furthermore, the WHO plans  
336 to launch sentinel surveys of acquired HIV resistance to DTG among people receiving DTG-based ART.<sup>28</sup>  
337 We could not assess adherence or drug interactions with rifampicin, which may influence the  
338 emergence of DTG resistance.<sup>29</sup> Adherence and rifampicin use were not recorded consistently and  
339 comparably in the participating cohorts. In our study population, the DTG-based regimens were too  
340 heterogeneous to investigate DTG resistance outcomes of specific regimens and treatment histories.  
341 Lastly, there is growing evidence that mutations outside integrase may confer DTG resistance<sup>30–32</sup>. Our  
342 study was based on pol sequences, which did not allow us to investigate the effects of these  
343 mutations.

344 The associations we found with DTG resistance, resistance to NRTI backbone drugs, and trends for  
345 HIV-1 subtype-and unsuppressed virus load have important implications for ensuring the long-term  
346 sustainability of ART. While overall INSTI resistance was rare in our population, and while the low risk  
347 of virological failure will further reduce the incidence of resistance among people treated with DTG,  
348 DTG resistance is still a concern. Firstly, the duration of DTG therapy and the duration of viraemia  
349 whilst receiving DTG was relatively short in our population: the median time on DTG was less than two  
350 years, and drug resistance might emerge more frequently in settings where individuals remain  
351 viraemic for a longer time on DTG regimens. This could happen in resource-limited settings where  
352 guidelines recommend not switching from DTG-based therapy unless multiple viral loads >1000 copies  
353 per mL have been documented and where delays in regimen switching are common.<sup>33</sup> Secondly, the  
354 strong association of DTG resistance with NRTI resistance suggests that the risk of resistance might be  
355 higher in people with previous failure on NNRTI-based first-line therapy, among whom the prevalence  
356 of NRTI resistance is much higher than in our study population. The WHO guidelines recommend DTG  
357 in 1st-, 2nd- and 3rd-line ART. This multiplicity of roles combined with the recycling of drugs and  
358 limited access to viral load and drug resistance testing will facilitate the emergence of DTG resistance.  
359 Finally, even a relatively low level of acquired DTG resistance in the millions of people receiving DTG-  
360 based ART could lead to rising levels of transmitted INSTI resistance, which could negatively affect  
361 both treatment and prevention.

362 In conclusion, our study underlines the importance of resistance testing, especially in treatment-  
363 experienced people. Although rare, DTG resistance can develop in people who experience viraemia

364 on a DTG-containing ART regimen. Monitoring the emergence of such resistance is important to  
365 prevent resistance at the individual and the population level and to ensure the long-term sustainability  
366 of ART.

## 367 **Figures & tables**

368 **Table 1: Demographics and clinical characteristics in the study population.** People with virological  
369 failure on DTG-based ART with available genotypic resistance tests from eight observational HIV  
370 cohorts were included in the study. Study participants where clinical data was available for at least  
371 one year were considered for analysing risk factors for DTG resistance.

372 Numbers (%) and medians [interquartile range] are shown.

373 \* Other subtypes are comprised as follows: For the analysis of resistance conferring mutations - Unknown, N=28 (4.7%); F,  
374 N=19 (3.2%); AE, N=10 (1.7%); D, N=10 (1.7%); 06\_CPX, N=6 (1%); AG, N=4 (0.7%); 18\_CPX, N=2 (0.3%); NA, N=2 (0.3%); AD,  
375 N=1 (0.2%); and H, N=1 (0.2%). For the analysis of risk factors for DTG resistance - Unknown, N=25 (4.6%); F, N=15 (2.8%); D,  
376 N=10 (1.9%); AE, N=7 (1.3%); 06\_CPX, N=5 (0.9%); AG, N=4 (0.7%); NA, N=2 (0.4%); 18\_CPX, N=1 (0.2%); AD, N=1 (0.2%); and  
377 H, N=1 (0.2%).

378 Abbreviations: ATHENA, the AIDS Therapy Evaluation in the Netherlands cohort; Aquitaine, Agence Nationale de la  
379 Recherche sur le SIDA et les hépatites virales (ANRS) CO3 Aquitaine Cohort; ICONA, Italian Cohort of Antiretroviral-Naïve  
380 Patients; CBC, Cologne/Bonn Cohort, Germany; SHCS, Swiss HIV Cohort Study; SAC, South Alberta Clinic Cohort, Canada; AfA,  
381 Aid for AIDS, South Africa; UK CHIC/UKHDRD, UK Collaborative HIV Cohort (UK CHIC) Study/ UK HIV Drug Resistance Database.

382 **Figure 1: Prevalence of DTG resistance and INSTI DRMs.** Genotypic resistance tests of 599 people  
383 with genotypic resistance testing on DTG-based ART were analysed using the Stanford resistance  
384 algorithm to determine INSTI DRMs and resistance level to DTG. Both major and accessory INSTI DRMs  
385 were considered for the number of INSTI DRMs. People with no INSTI DRMs (N = 86, 85.6%), and who  
386 are susceptible to DTG (N = 563, 94%) are not displayed.

387 **Figure 2: INSTI drug resistance mutations found in 599 people experiencing viraemia on a DTG-based**  
388 **regimen.** Drug resistance mutations were classified as major and accessory according to the Stanford  
389 resistance database<sup>22</sup>. Bars are coloured by previous history of first-generation INSTIs (raltegravir,  
390 elvitegravir).

391 **Figure 3: Rate ratio for number of INSTI DRMs.** A negative binomial generalised linear model was fit  
392 to the number of major and accessory INSTI DRMs in 599 people with viraemia on DTG-based ART.  
393 The plot shows uni- and multivariable point estimates and 95% confidence intervals of rate ratios.

394 **Table 2: Resistance levels to DTG, non-nucleoside reverse transcriptase inhibitors and nucleotide**  
395 **reverse transcriptase inhibitors.** Number and percentage of people with corresponding drug  
396 resistance levels are given for the entire study population. NRTI resistance level is based on median  
397 resistance score to ABC, AZT, XTC and TDF/TAF. NNRTI resistance level is based on median resistance  
398 score to EFV, ETR, NVP, and RPV.

399 **Figure 4: Odds ratios for DTG resistance levels with 95% confidence intervals from uni- and**  
400 **multivariable ordinal logistic models for genotypic DTG resistance.** Cohorts were included as random  
401 effect. DTG resistance levels in people with viraemia on DTG-based ART were assessed using the  
402 Stanford resistance algorithm.

## 403 **Authors' contributions**

404 Conceptualisation (HFG, JACS, RL, ME, RK), Data curation (TL, SH, SI, HO), Methodology (TL, CS, RK,  
405 ME, JACS), Formal analysis & Validation (TL, RK), Investigation (TL, SH, SI), Project administration (SH,  
406 SI), Resources (HO, KK, JM, AvS, MS, AAM, JG, CS, GM), Software (TL, KK), Supervision (HFG, JACS, RL,  
407 ME, RK), Visualisation (TL), Writing – original draft (TL, RL, ME, RK), Writing – review & editing (All  
408 authors).

409 TL and RDK had directly accessed and verified the underlying data reported in the manuscript. ME, TL  
410 and RDK had full access to the data, other authors had access to the data from their cohort, but not  
411 to the data from the other cohorts. ME had the final responsibility for the decision to submit for  
412 publication.

## 413 **Declaration of interests**

414 SMI reports grant funding from NIH NIAAA for the work of ART-CC (payment to institution). AvS  
415 reports funding from the Dutch Ministry of Health, Welfare and Sport for the maintenance of the  
416 ATHENA database, and grant funding from the European Centre for Disease Prevention and Control  
417 (ECDC) (payment to institution). MJG reports honoraria as Ad Hoc member of HIV National Advisory  
418 Board from Merck, Gilead Sciences, and ViiV, and a leadership position as Medical Director S Alberta  
419 HIV clinic. CAS has received funding from Gilead Sciences, ViiV Healthcare and Janssen-Cilag for  
420 membership of Data Safety and Monitoring Committees, Advisory Committees and for preparation of  
421 educational material. HFG has received personal fees from Merck, Gilead Sciences, ViiV, GSK, Janssen,  
422 Johnson and Johnson and Novartis, as an advisor/consultant or for DSMB membership and has  
423 received a travel grant from Gilead. JACS reports funding for research in this publication from NIH  
424 NIAAA (payment to institution), UK NIHR (payment to institution), and the University of Bern (payment  
425 to institution). RL reports support for research in this publication by the National Institute of Allergy  
426 & Infectious Diseases of the National Institutes of Health under award number R01AI152772, and  
427 support from the National Institute of Allergy & Infectious Diseases of the National Institutes of Health  
428 under award number R01AI167699 for a separate project pertaining to HIV treatment strategies. ME  
429 reports funding for research in this publication from the Swiss National Science Foundation (32FP30-  
430 18949) and the National Institutes of Health (Cooperative Agreement AI069924 and R01 AI152772-  
431 01). RK reports funding for research in this publication from the Swiss National Science Foundation  
432 and the National Institute of Allergy & Infectious Diseases of the National Institutes of Health, and  
433 reports grant funding from Gilead Sciences. All other authors declare no competing interest.

434



## 435 **Acknowledgements**

436 We would like to thank Anthony Hauser, Suraj Balakrishna, and Marius Zeeb for helpful discussions on  
437 data analysis. This study was supported by the National Institute Of Allergy And Infectious Diseases of  
438 the National Institutes of Health under Award Number R01AI152772 and the Swiss National Science  
439 Foundation (32FP30\_207285, 324730\_207957). The participating cohorts or cohort collaborations  
440 were funded by the Swiss National Science Foundation (33CS30\_201369) and the Yvonne Jacob  
441 Foundation (for the SHCS), the UK Medical Research Council (grant numbers G0000199, G0600337,  
442 G0900274, and M004236/1; for the UK Collaborative HIV Cohort), the National Agency for AIDS  
443 Research (France REcherche Nord&Sud Sida-hiv Hépatites), the French Agency for Research on AIDS  
444 and Viral Hepatitis | Emerging Infectious Diseases (ANRS | MIE) and the CHU de Bordeaux (for the ANRS  
445 CO3 Aquitaine-AquiVIH-NA cohort), the Dutch Ministry of Health (for the ATHENA cohort), the German  
446 Center for Infection Research (8018704707) (for the CBC), ICONA Foundation is supported by  
447 unrestricted grants from BMS, Gilead Sciences, Janssen, MSD and ViiV Healthcare. AFA is supported  
448 via IeDEA-SA by the U.S. National Institutes of Health's National Institute of Allergy and Infectious  
449 Diseases, the Eunice Kennedy Shriver National Institute of Child Health and Human Development,  
450 Division of Cancer Epidemiology and Genetics, National Cancer Institute, the National Institute of  
451 Mental Health, the National Institute on Drug Abuse, the National Heart, Lung, and Blood Institute,  
452 the National Institute on Alcohol Abuse and Alcoholism, the National Institute of Diabetes and  
453 Digestive and Kidney Diseases and the Fogarty International Center under Award Number  
454 U01AI069924. The ART-CC is funded by the US National Institute on Alcohol Abuse and Alcoholism  
455 (U01-AA026209). The content is solely the responsibility of the authors and does not necessarily  
456 represent the official views of the National Institutes of Health.

## 457 **Data sharing statement**

458 Study data will not be publicly available. Data can be made available to interested researchers.  
459 Deidentified participant data and a data dictionary can be made available and shared under a data  
460 transfer agreement. Requests for access to DTG RESIST data should be sent to  
461 matthias.egger@unibe.ch. Nucleotide sequences are available on GenBank for the cohorts where  
462 local regulations allowed data sharing (see table S1, appendix p. 2)

463

## 464 References

- 465 1. WHO. Update of recommendations on first- and second-line antiretroviral regimens. Geneva,  
466 Switzerland:World Health Organization; WHO. 2019. p. 3.
- 467 2. The Lancet HIV. End resistance to dolutegravir roll-out. *Lancet HIV*. 2020 Sep 1;7(9):e593.
- 468 3. Llibre JM, Pulido F, García F, García Deltoro M, Blanco JL, Delgado R. Genetic barrier to  
469 resistance for dolutegravir. *AIDS Rev*. 2015;17(1):56–64.
- 470 4. Cottrell ML, Hadzic T, Kashuba ADM. Clinical pharmacokinetic, pharmacodynamic and drug-  
471 interaction profile of the integrase inhibitor dolutegravir. *Clin Pharmacokinet*.  
472 2013;52(11):981–94.
- 473 5. Cevik M, Orkin C, Sax PE. Emergent resistance to dolutegravir among instinaive patients on  
474 first-line or second-line antiretroviral therapy: A review of published cases. *Open Forum*  
475 *Infect Dis*. 2020;7(6).
- 476 6. Pena MJ, Chueca N, D’Avolio A, Zarzalejos JM, Garcia F. Virological failure in HIV to triple  
477 therapy with dolutegravir-based firstline treatment: Rare but possible. *Open Forum Infect*  
478 *Dis*. 2019;6(1).
- 479 7. Scherrer AU, Yang W-L, Kouyos RD, Böni J, Yerly S, Klimkait T, et al. Successful Prevention of  
480 Transmission of Integrase Resistance in the Swiss HIV Cohort Study. *J Infect Dis*.  
481 2016;214(3):399–402.
- 482 8. Cahn P, Pozniak AL, Mingrone H, Shuldyakov A, Brites C, Andrade-Villanueva JF, et al.  
483 Dolutegravir versus raltegravir in antiretroviral-experienced, integrase-inhibitor-naïve adults  
484 with HIV: Week 48 results from the randomised, double-blind, non-inferiority SAILING study.  
485 *Lancet*. 2013;382(9893):700–8.
- 486 9. Lepik KJ, Harrigan PR, Yip B, Wang L, Robbins MA, Zhang WW, et al. Emergent drug resistance  
487 with integrase strand transfer inhibitor-based regimens. *AIDS*. 2017;31(10):1425–34.
- 488 10. Paton NI, Musaaazi J, Kityo C, Walimbwa S, Hoppe A, Balyegisawa A, et al. Efficacy and safety  
489 of dolutegravir or darunavir in combination with lamivudine plus either zidovudine or  
490 tenofovir for second-line treatment of HIV infection (NADIA ): week 96 results from a  
491 prospective, multicentre, open-label, factorial, randomised, no. *Lancet HIV*. 2022;1–13.
- 492 11. Han Y-S, Mesplède T, Wainberg MA. Differences among HIV-1 subtypes in drug resistance  
493 against integrase inhibitors. *Infect Genet Evol*. 2016;46:286–91.
- 494 12. Brenner BG, Thomas R, Blanco JL, Ibanescu R-I, Oliveira M, Mesplède T, et al. Development of  
495 a G118R mutation in HIV-1 integrase following a switch to dolutegravir monotherapy leading  
496 to cross-resistance to integrase inhibitors. *J Antimicrob Chemother*. 2016;71(7):1948–53.
- 497 13. Arimide DA, Szojka ZI, Zealiyas K, Gebreegziabxier A, Adugna F, Sasinovich S, et al. Pre-  
498 Treatment Integrase Inhibitor Resistance and Natural Polymorphisms among HIV-1 Subtype C  
499 Infected Patients in Ethiopia. *Viruses*. 2022;14(4).
- 500 14. Akil B, Blick G, Hagins DP, Ramgopal MN, Richmond GJ, Samuel RM, et al. Dolutegravir versus  
501 placebo in subjects harbouring HIV-1 with integrase inhibitor resistance associated  
502 substitutions: 48-week results from VIKING-4, a randomized study. *Antivir Ther*.  
503 2015;20(3):343–8.
- 504 15. Castagna A, Maggiolo F, Penco G, Wright D, Mills A, Grossberg R, et al. Dolutegravir in  
505 antiretroviral-experienced patients with raltegravir- and/or elvitegravir-resistant HIV-1: 24-

- 506 week results of the phase III VIKING-3 study. *J Infect Dis.* 2014;210(3):354–62.
- 507 16. Inzaule SC, Hamers RL, Doherty M, Shafer RW, Bertagnolio S, Rinke de Wit TF. Curbing the  
508 rise of HIV drug resistance in low-income and middle-income countries: the role of  
509 dolutegravir-containing regimens. *Lancet Infect Dis.* 2019;19(7):e246–52.
- 510 17. May MT, Ingle SM, Costagliola D, Justice AC, de Wolf F, Cavassini M, et al. Cohort profile:  
511 Antiretroviral therapy cohort collaboration (ART-CC). *Int J Epidemiol.* 2014;43(3):691–702.
- 512 18. Chammartin F, Dao Ostinelli CH, Anastos K, Jaquet A, Brazier E, Brown S, et al. International  
513 epidemiology databases to evaluate AIDS (IeDEA) in sub-Saharan Africa, 2012-2019. *BMJ*  
514 *Open.* 2020;10(5).
- 515 19. Struck D, Lawyer G, Ternes A-M, Schmit J-C, Bercoff DP. COMET: Adaptive context-based  
516 modeling for ultrafast HIV-1 subtype identification. *Nucleic Acids Res.* 2014;42(18).
- 517 20. Pineda-Peña A-C, Faria NR, Imbrechts S, Libin P, Abecasis AB, Deforche K, et al. Automated  
518 subtyping of HIV-1 genetic sequences for clinical and surveillance purposes: Performance  
519 evaluation of the new REGA version 3 and seven other tools. *Infect Genet Evol.* 2013;19:337–  
520 48.
- 521 21. Rhee S-Y, Gonzales MJ, Kantor R, Betts BJ, Ravela J, Shafer RW. Human immunodeficiency  
522 virus reverse transcriptase and protease sequence database. *Nucleic Acids Res.*  
523 2003;31(1):298–303.
- 524 22. Stanford University. HIV Drug Resistance Database [Internet]. [cited 2023 19 July]. Available  
525 from: <https://hivdb.stanford.edu/page/release-notes/#drm.classification>
- 526 23. Abram ME, Ram RR, Margot NA, Barnes TL, White KL, Callebaut C, et al. Lack of impact of pre-  
527 existing T97A HIV-1 integrase mutation on integrase strand transfer inhibitor resistance and  
528 treatment outcome. *PLoS One.* 2017;12(2).
- 529 24. Rolle C-P, Nguyen V, Hiestrosa F, DeJesus E. Virologic outcomes of switching to dolutegravir  
530 functional mono- or dual therapy with a non-cytosine nucleoside analog: a retrospective  
531 study of treatment-experienced, patients living with HIV. *AIDS Res Ther.* 2021;18(1).
- 532 25. Naeger LK, Harrington P, Komatsu T, Deming D. Effect of dolutegravir functional  
533 monotherapy on HIV-1 virological response in integrase strand transfer inhibitor resistant  
534 patients. *Antivir Ther.* 2016;21(6):481–8.
- 535 26. Von Wyl V, Yerly S, Böni J, Bürgisser P, Klimkait T, Battegay M, et al. Emergence of HIV-1 drug  
536 resistance in previously untreated patients initiating combination antiretroviral treatment: A  
537 comparison of different regimen types. *Arch Intern Med.* 2007;167(16):1782–90.
- 538 27. De Salazar A, Viñuela L, Fuentes A, Teyssou E, Charpentier C, Lambert-Niclot S, et al.  
539 Transmitted Drug Resistance to Integrase-Based First-Line Human Immunodeficiency Virus  
540 Antiretroviral Regimens in Mediterranean Europe. *Clin Infect Dis.* 2023 May;76(9):1628–35.
- 541 28. WHO. Sentinel surveys of acquired HIV resistance to dolutegravir among people receiving  
542 dolutegravir-containing antiretroviral therapy. Geneva; 2022.
- 543 29. Naidoo A, Naidoo K, Padayatchi N, Dooley KE. Use of integrase inhibitors in HIV-associated  
544 tuberculosis in high-burden settings: implementation challenges and research gaps. *Lancet*  
545 *HIV.* 2022;9(2):e130–8.
- 546 30. Malet I, Delelis O, Nguyen T, Leducq V, Abdi B, Morand-Joubert L, et al. Variability of the HIV-  
547 1 3' polypurine tract (3'PPT) region and implication in integrase inhibitor resistance. *J*  
548 *Antimicrob Chemother.* 2019;74(12):3440–4.

- 549 31. Dekker JG, Klaver B, Berkhout B, Das AT. Mutations in the HIV-1 3-Polypurine Tract Can  
550 Confer Dolutegravir Resistance. *Antimicrob Agents Chemother.* 2022;66(1).
- 551 32. Hikichi Y, Groebner JL, Wiengand A, Mellors JW, Kearney MF, Freed EO. Mutations outside  
552 integrase lead to high-level resistance to dolutegravir [CROI Abstract 103]. In: Conference on  
553 Retroviruses and Opportunistic Infections CROI 2023 Abstract eBook. 2023.
- 554 33. Haas AD, Keiser O, Balestre E, Brown S, Bissagnene E, Chimbetete C, et al. Monitoring and  
555 switching of first-line antiretroviral therapy in adult treatment cohorts in sub-Saharan Africa:  
556 Collaborative analysis. *Lancet HIV.* 2015;2(7):e271–8.
- 557

558 **Table 3: Demographics and clinical characteristics in the study population.** People with virological  
559 failure on DTG-based ART with available genotypic resistance tests from eight observational HIV  
560 cohorts were included in the study. Study participants where clinical data was available for at least  
561 one year were considered for analysing risk factors for DTG resistance.

	<b>Analysis of resistance conferring mutations (N=599)</b>	<b>Analysis of risk factors for DTG resistance (N=540)</b>
<b>Sex</b>		
Female	187 (31.2%)	175 (32.4%)
Male	412 (68.8%)	365 (67.6%)
<b>Age at DTG Initiation (years)</b>		
	44 [36 - 52]	45 [37 - 52]
<b>HIV Subtype</b>		
B	351 (58.6%)	316 (58.5%)
C	69 (11.5%)	63 (11.7%)
A	54 (9.0%)	51 (9.4%)
G	42 (7.0%)	39 (7.2%)
Other*	83 (13.9%)	71 (13.1%)
<b>ART regimen at DTG initiation</b>		
Combination therapy with ≥3 ARVs	511 (85.3%)	455 (84.3%)
Dual therapy (DTG & other)	51 (8.5%)	50 (9.3%)
Dual therapy (DTG & Lamivudine)	19 (3.2%)	17 (3.1%)
Monotherapy	18 (3.0%)	18 (3.3%)
<b>ART duration at DTG initiation (years)</b>		
	6.7 [0.95 - 14]	7.9 [2.4 - 15]
Missing	4 (0.7%)	4 (0.7%)
<b>Year of DTG initiation</b>		
	2016 [2015 - 2017]	2016 [2015 - 2017]
<b>Year of genotypic resistance test</b>		
	2018 [2017 - 2019]	2018 [2017 - 2019]
<b>Availability of additional (prior) GRTs</b>		
Yes	395 (65.9%)	356 (65.9%)
No	204 (34.1%)	184 (34.1%)
<b>DTG-regimen initiation</b>		
Switch to DTG-based ART	486 (81.1%)	470 (87.0%)
Initiation on DTG-based ART	113 (18.9%)	70 (13.0%)
<b>Duration on DTG-based ART at GRT (years)</b>		
	1.4 [0.58 - 2.7]	1.6 [0.67 - 2.8]
<b>Exposure to first generation INSTI</b>		
Yes	193 (32.2%)	184 (34.1%)
No	406 (67.8%)	356 (65.9%)
<b>CD4 count at GRT (cells per µL)</b>		
	412 [213 - 674]	433 [218 - 681]
Missing	129 (21.5%)	115 (21.3%)
<b>Viral load AUC (of log10 copies per ml during DTG based ART)</b>		
	3.6 [2.2 - 5.0]	3.6 [2.3 - 4.9]
Missing	10 (1.7%)	0 (0%)
<b>No. of HIV tests per year</b>		
	3.0 [2.0 - 4.3]	3.3 [2.3 - 4.3]
Missing	20 (3.3%)	0 (0%)
<b>Cohort</b>		
AfA	9 (1.5%)	9 (1.7%)
Aquitaine	64 (10.7%)	59 (10.9%)
ATHENA	66 (11.0%)	64 (11.9%)

CBC	89 (14.9%)	76 (14.1%)
ICONA	8 (1.3%)	5 (0.9%)
SAC	92 (15.4%)	87 (16.1%)
SHCS	118 (19.7%)	108 (20.0%)
UK CHIC/UKHDRD	153 (25.5%)	132 (24.4%)

562 Numbers (%) and medians [interquartile range] are shown.

563 \* Other subtypes are comprised as follows: For the analysis of resistance conferring mutations - Unknown, N=28 (4.7%); F,  
564 N=19 (3.2%); AE, N=10 (1.7%); D, N=10 (1.7%); 06\_CPX, N=6 (1%); AG, N=4 (0.7%); 18\_CPX, N=2 (0.3%); NA, N=2 (0.3%); AD,  
565 N=1 (0.2%); and H, N=1 (0.2%). For the analysis of risk factors for DTG resistance - Unknown, N=25 (4.6%); F, N=15 (2.8%); D,  
566 N=10 (1.9%); AE, N=7 (1.3%); 06\_CPX, N=5 (0.9%); AG, N=4 (0.7%); NA, N=2 (0.4%); 18\_CPX, N=1 (0.2%); AD, N=1 (0.2%); and  
567 H, N=1 (0.2%).

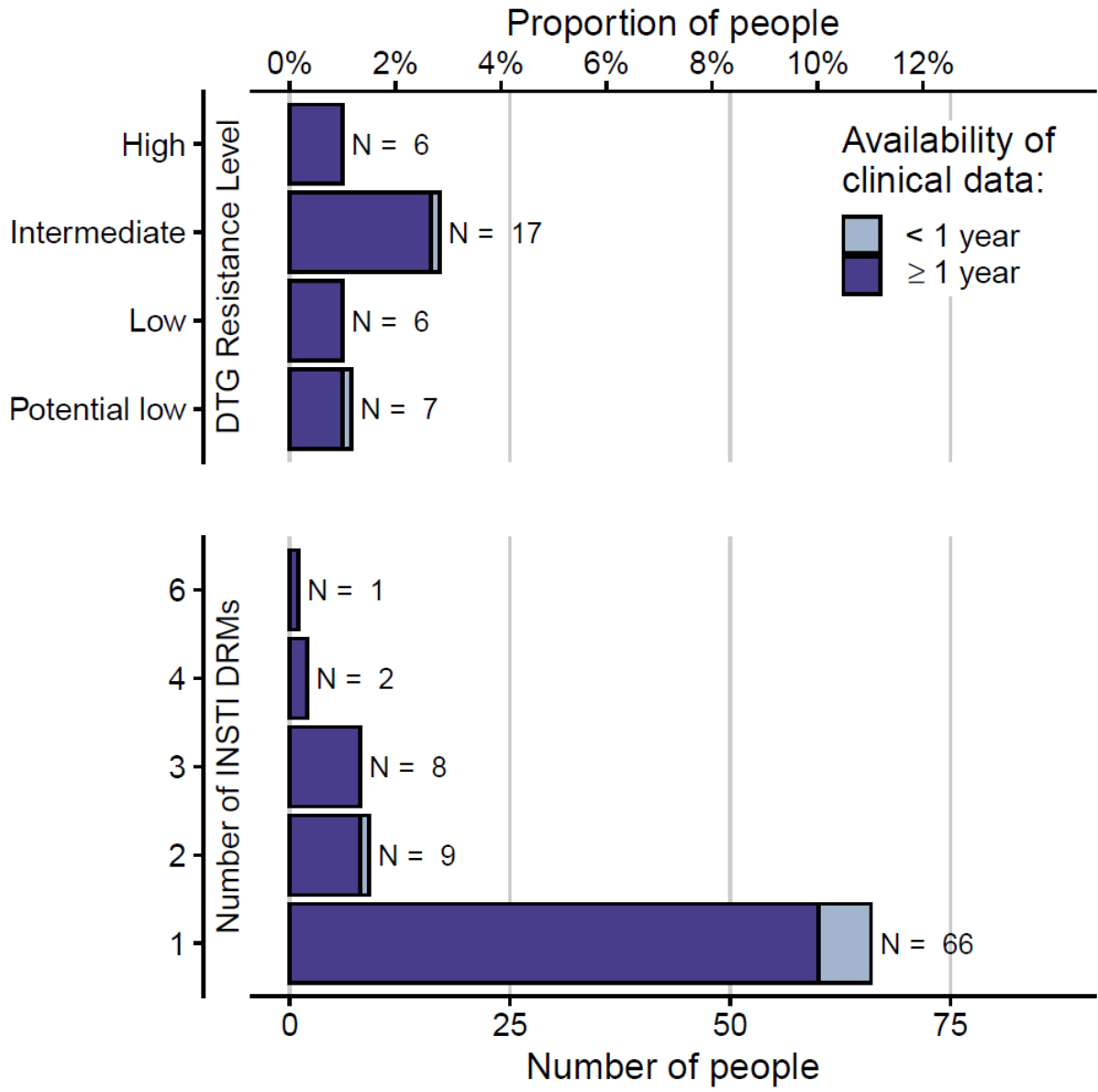
568 Abbreviations: ATHENA, the AIDS Therapy Evaluation in the Netherlands cohort; Aquitaine, Agence Nationale de la  
569 Recherche sur le SIDA et les hépatites virales (ANRS) CO3 Aquitaine Cohort; ICONA, Italian Cohort of Antiretroviral-Naïve  
570 Patients; CBC, Cologne/Bonn Cohort, Germany; SHCS, Swiss HIV Cohort Study; SAC, South Alberta Clinic Cohort, Canada; AfA,  
571 Aid for AIDS, South Africa; UK CHIC/UKHDRD, UK Collaborative HIV Cohort (UK CHIC) Study/ UK HIV Drug Resistance Database.

572

573

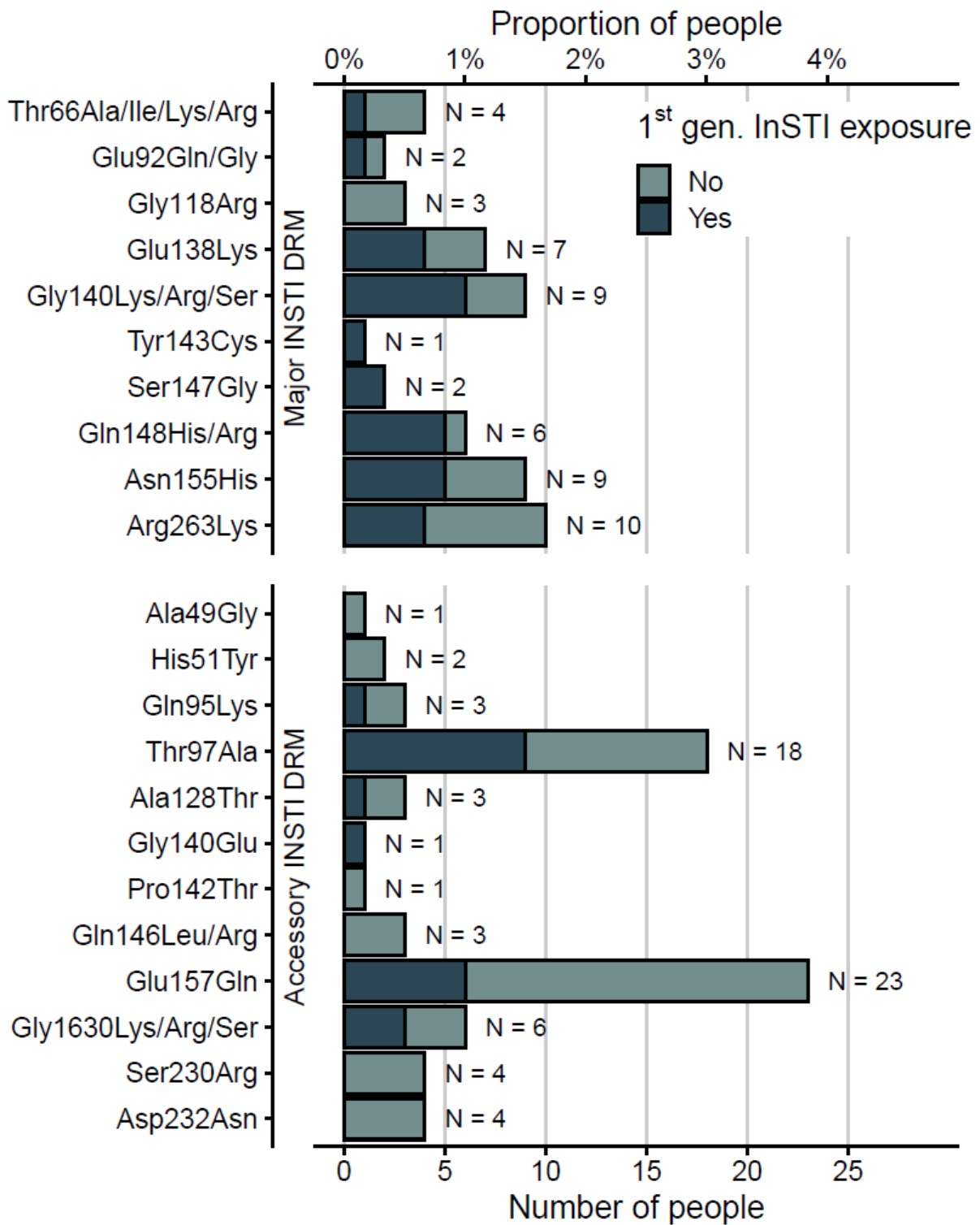
574 Figure 1

575



576

577

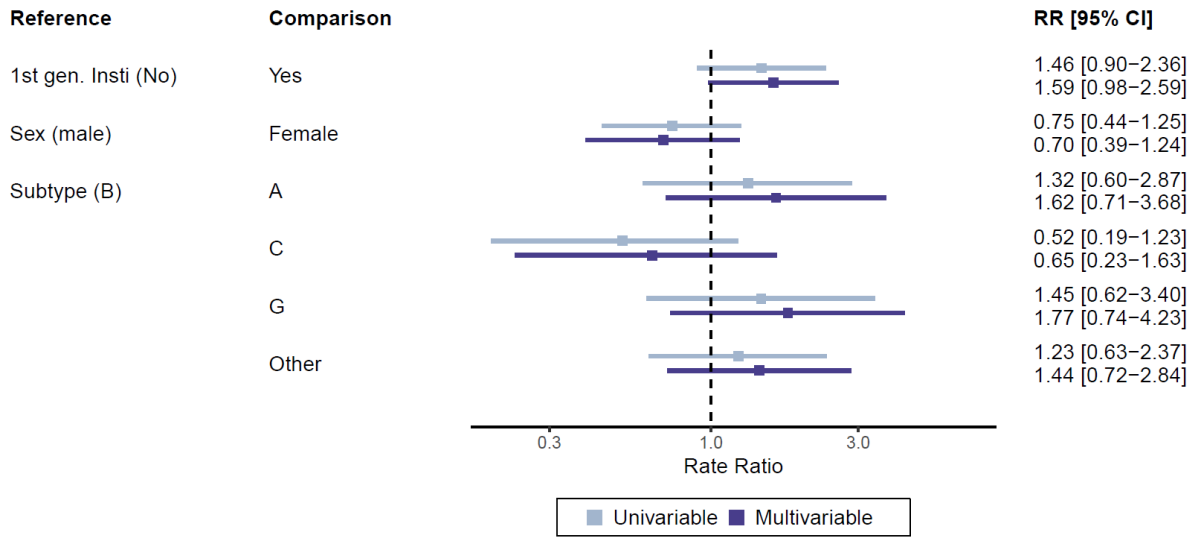




581 Figure 3

582

583



584

585

586 **Table 4: Resistance levels to DTG, non-nucleoside reverse transcriptase inhibitors and nucleotide**  
 587 **reverse transcriptase inhibitors.** Number and percentage of people with corresponding drug  
 588 resistance levels are given for the entire study population. NRTI resistance level is based on median  
 589 resistance score to ABC, AZT, XTC and TDF/TAF. NNRTI resistance level is based on median resistance  
 590 score to EFV, ETR, NVP, and RPV.

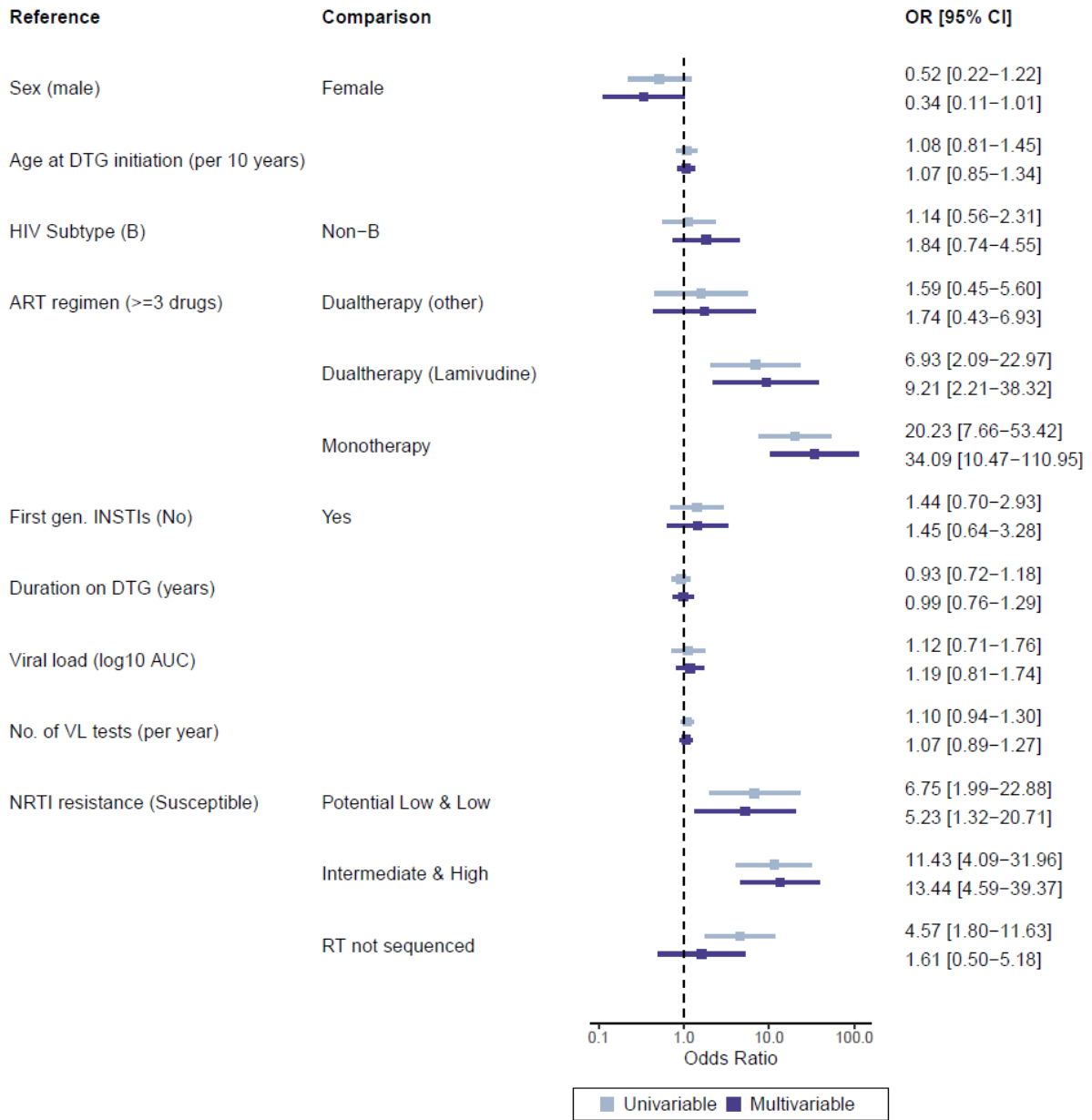
	<b>DTG resistance level</b>	
	<b>Susceptible &amp; Potential Low (N=570)</b>	<b>Low, Intermediate, High (N=29)</b>
<b>NRTI resistance level</b>		
Susceptible	467 (81.9%)	13 (44.8%)
Potential low	9 (1.6%)	2 (6.9%)
Low	10 (1.8%)	1 (3.4%)
Intermediate	9 (1.6%)	2 (6.9%)
High	13 (2.3%)	4 (13.8%)
RT not covered in GRT	62 (10.9%)	7 (24.1%)
<b>NNRTI resistance level</b>		
Susceptible	414 (72.6%)	11 (37.9%)
Potential low	23 (4.0%)	0 (0%)
Low	18 (3.2%)	0 (0%)
Intermediate	34 (6.0%)	4 (13.8%)
High	19 (3.3%)	7 (24.1%)
RT not covered in GRT	62 (10.9%)	7 (24.1%)

591

592

593 Figure 4

594



595