

OP-163

Sulfated hydrogels as primary intervertebral disc cell culture systems

Paola Bermudez Lekerika¹, Exarchos Kanelis², Andreas Shaun Croft¹, Katherine Briana Crump¹, Karin Wuertz Kozak³, Christine Le Maitre⁴, Leonidas Alexopoulos⁵, Dominic Rütsche⁶, Benjamin Gantenbein¹

¹Tissue Engineering for Orthopaedics and Mechanobiology, Bone & Joint Program, Department for BioMedical Research (DBMR), University of Bern, Switzerland; Inselspital, University of Bern and Department of Orthopedic Surgery & Traumatology, Bern, Switzerland,

²ProtATonce Ltd., Athens, Greece

³Department of Biomedical Engineering, Rochester Institute of Technology, Rochester, United States

⁴Oncology and Metabolism Department Medical School, The University of Sheffield, Sheffield, United Kingdom

⁵Department of Mechanical Engineering, National Technical University of Athens, Zografou, Greece

⁶Tissue Engineering + Biofabrication Laboratory, Department of Health Sciences and Technology, ETH, Zürich, Switzerland

INTRODUCTION: Intervertebral disc (IVD) degeneration is a key contributor for low back pain, a leading cause of disability worldwide¹. During degeneration, IVD aging is accelerated, leading to progressive structural changes, including blood vessel and nerve ingrowth that promote discogenic pain². In vitro studies require novel biomaterials that mimic the IVD extracellular matrix (ECM) to provide mechanical support and a reservoir of cytokines and growth factors. As proteoglycans with their attached sulfated glycosaminoglycans (GAGs) are one of the major components of the ECM, the ECM's sulfation state could be a key factor for IVD cell-fate³. Thus, we aim to explore human NP cell fate using a novel sulfated alginate model with varying degrees of sulfation (DS).

METHODS: Primary human NP cells were expanded, mixed with solutions of i) 2.5% of standard alginate, ii) 0.1 DS, and iii) 0.2 DS alginate (4 x 10⁶ cells/ml) and casted in 27 µl cylindrical-shaped carriers (4 mm diameter, 2 mm height). Carriers were cultured for two weeks for phenotype recovery and were collected with the culture media on day 0, 7 and 14.

RESULTS: A significant decrease of cell density ($p < 0.05$) was observed in 0.2 DS alginate after 7 and 14 days of culture. Similarly, cell viability was significantly reduced ($p < 0.05$) in 0.2 DS alginate after 7 days of culture ($N = 4$). In addition, cell metabolic activity tended to be decreased in 0.2 DS alginate compared to standard alginate after 14 days of culture. Surprisingly, ECM remodeling factors such as MMP2 and TIMP1 were slightly upregulated in the 0.1 DS group ($N = 1$), whereas catabolic cytokines were downregulated in the 0.1% DS group.

DISCUSSION & CONCLUSIONS: We demonstrate significant cellular differences between 0.2 DS alginate vs standard alginate and 0.1 DS alginate. Particularly, a significant decrease in cell density, metabolic activity and viability were observed in the 0.2 DS alginate after 7 days of culture. According to the secretome, the sulfated alginate group seems to possess increased catabolic ECM remodeling with lower secretion of catabolic factors, suggesting less responsive NP cells to ECM structural changes. Overall, standard alginate seems to be the best option for NP cell 3D culture models.

ACKNOWLEDGEMENTS: This project was supported by the Marie Skłodowska Curie International Training Network "disc4all" under the grant agreement #955735.

REFERENCES: 1FY. Wang et al (2020) JOR Spine 5:1186.

2P. Bermudez-Lekerika et al (2022) Front Cell Dev Biol 29(10):924692.

3E. Lazarus et al (2021) Cells 10(12):3568.

Keywords: Hydrogels and injectable systems, In vitro microenvironments