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Differential diagnosis of chorea (guidelines of the German Neurological Society)

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Abstract

Introduction Choreaform movement disorders are characterized by involuntary, rapid, irregular, and unpredictable movements of the limbs, face, neck, and trunk. These movements often initially go unnoticed by the affected individuals and may blend together with seemingly intended, random motions. Choreaform movements can occur both at rest and during voluntary movements. They typically increase in intensity with stress and physical activity and essentially cease during deep sleep stages. In particularly in advanced stages of Huntington disease (HD), choreiform hyperkinesia occurs alongside with dystonic postures of the limbs or trunk before they typically decrease in intensity.

Summary or definition of the topic The differential diagnosis of HD can be complex. Here, the authors aim to provide guidance for the diagnostic process. This guidance was prepared for the German Neurological Society (DGN) for German-speaking countries.

Recommendations Hereditary (inherited) and non-hereditary (non-inherited) forms of chorea can be distinguished. Therefore, the family history is crucial. However, even in conditions with autosomal-dominant transmission such as HD, unremarkable family histories do not necessarily rule out a hereditary form (e.g., in cases of early deceased or unknown parents, uncertainties in familial relationships, as well as in offspring of parents with CAG repeats in the expandable range (27–35 CAG repeats) which may display expansions into the pathogenic range).

Conclusions The differential diagnosis of chorea can be challenging. This guidance prepared for the German Neurological Society (DGN) reflects the state of the art as of 2023.

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Introduction

This guideline is an abridged and translated short version of the guidance on chorea and Huntington's Disease prepared for the German Neurological Society, covering diagnostic aspects as well as symptomatic treatment options. A complete version of this guideline (in German) can be found on the website of the Deutsche Gesellschaft für Neurologie (www.dgn.org/leitlinien) and the AWMF (Arbeitsgemeinschaft wissenschaftlicher Medizinischer Gesellschaften).

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This guideline has been approved by the German Neurological Society (DGN) and the German Association for Psychiatry, Psychotherapy, and Psychosomatics; German Society of Human Genetics; Swiss Neurological Society; Austrian Society of Neurology, and was reviewed by the German HD Association (Deutsche Huntington-Hilfe e.V.)

What's new?

The differential diagnosis of chorea can be complex. Taking a family history can be helpful in many cases. A variety of new mutations causing chorea as a symptom has been described. For sporadic cases, it can be helpful to differentiate based on: course, age at disease onset, symmetry, additional neurological, clinical and paraclinical findings such as concurrent ataxia, polyneuropathy, cutaneous or scleral telangiectasias, liver pathology, laboratory abnormalities or specific MRI findings.

Guidelines in detail

The recommendations of this guideline were established through a Delphi process (strength of consensus > 75–95% for all recommendations), achieved over two rounds of voting. All statements for which consensus strength is not specified were met with a consensus of > 95%.

Hereditary disease entities presenting with chorea or featuring choreiform movements

- Huntington's disease [14, 95]
- C9orf72 expansion mutation (frontotemporal dementia, motor neuron disease and movement disorders, probably frequent phenocopy of HD [43])
- Spinocerebellar ataxia 17 (corresponds to Huntington's-disease-like-4; HDL4 [95])
- Spinocerebellar ataxia types 3, 2, 1 and 7 [78]
- Spinocerebellar ataxia type 8 [95]
- Spinocerebellar ataxia type 12 (mainly India [48, 95])

- Spinocerebellar ataxia type 48/SCAR 16 due to mutations in the STUB1 gene [61, 91]
- Dentato-rubro-pallido-Luysian atrophy (mainly Japan, DRPLA; [78])
- Ataxia telangiectasia and ataxia telangiectasia like disease (serum alpha-fetoprotein ↑ [14])
- Ataxia with oculomotor apraxia (AOA1 (serum albumin ↓) and AOA2 (serum alpha-fetoprotein↑; now also called SCAN2 [2, 14, 57])
- McLeod syndrome (CK↑, ± acanthocytes in blood smear↑, myocardial abnormalities, striatal atrophy, analysis of the Kell/Kx blood group phenotype or mutations in XK gene [25])
- Chorea-acanthocytosis (CK↑, ± acanthocytes in blood smear↑, striatal atrophy, ChoreinWestern blot (LMU, Munich), detection of mutation in the CHAC gene (VPS13A [25, 95]))
- Huntington's disease-like 2 (HDL2; predominantly in patients of African origin [14, 95])
- Benign hereditary chorea (including thyroid transcription factor 1 gene, TITF1/NKX2-1; L-Dopa or methylphenidate therapy might be helpful [32, 106])
- ADCY5 mutation [13, 14]

Other rare inherited disease entities

- Huntington's disease-like 1 and 3, only described in individual families [14]
- HDL1 with prion protein (PrP) gene mutations (PRNP) and rapid progression [14, 95]
- HDL3, a family [47]
- RNF216 mutation (autosomal recessive, leukoencephalopathic lesions and possibly Serum gonadotropin ↓ [93])
- ANO3 mutations [52]
- FRRS1L mutations (Saudi Arabia; also epilepsy [95])
- Primary Familial Brain Calcification (formerly "Fahr's disease", cMRI/CCT helpful (SLC20A2-, PDGFB, PDGFRB or XPR1 gene [95]))
- POLG gene mutations (dystonia, myoclonus, discrete chorea [101])
- Leigh's disease [63]
- SETX mutation (with motor neuron disease [94])
- Laurence-Moon-Biedl-Bardet syndrome [65]
- Friedreich ataxia [41]
- NBIA "neurodegeneration with brain iron accumulation" (umbrella term for e.g. Pantothenate kinase-associated neurodegeneration (PKAN 2), neuroferritinopathies (FTL), Aceruloplasminemia (CP), phospholipase-associated neurodegeneration (PLAN), Betapropeller protein-associated neurodegeneration (WDR45), infantile neuroaxonal

- dystrophy (PLA2G6, C19orf12, C2orf37, FA2H, ATP13A2, COASY and DCAF17 mutations—more likely no chorea)) with iron deposits in the basal ganglia as a typical MRI finding [3, 14, 82, 95, 103, 113])
- Wilson's disease [14, 95]
 - TAR DNA binding protein variation (TARDBP; with frontotemporal dementia [51])
 - Lesch-Nyhan syndrome; X-linked [1, 14]
 - Niemann-Pick type C [46]
 - Cereoid lipofuscinosis [72]
 - Lipidoses, aminoacidosis and carbohydrate metabolism disorders [69, 96]
 - Phenylketonuria [44]
 - Paroxysmal kinesiogenic dyskinesia (PKD; paroxysmal kinesiogenic choreoathetosis; dystonia 10 [14, 50], weekly parenteral doses of vitamins and minerals might be helpful [10])
 - Paroxysmal non-kinesiogenic dyskinesia (PNKD); dystonia type 8 [14, 50]
 - Paroxysmal choreoathetosis with infantile febrile convulsions; ICCA [14]
 - Tuberous brain sclerosis [116]
 - FUS-related ALS [34]
 - Mutations in iron-responsive element-binding protein 2; IREB2 [20]
 - 18p deletions syndrome [22]
 - X-linked Dystonia-Parkinsonism; Lubag; Dyt3 [31]
 - FXTAS [56]
 - Dopamine D2 receptor variant (also dystonia [107])
 - GLRB mutations (GlyR β-subunit; with hyperekplexia [29])
 - CAMK4 variant (with dystonia, autism, developmental delay, later chorea [120])
 - Eukaryotic translation elongation factor (EEF1A2) mutations (also associated with epilepsy, autism, intellectual impairment, sudden onset of chorea [55])
 - ERCC4 gene mutations in xeroderma pigmentosa (with ataxia [15])
 - Replication factor complex subunit 1 (RFC1) mutations (cerebellar ataxia, neuropathy, vestibular areflexia syndrome (CANVAS) and leading cause of late-onset ataxia, 11% Chorea [105])
 - Polynucleotide kinase phosphatase (PNKP) mutation (rather benign course, early onset, with microcephaly, epilepsy, developmental delay, ataxia with oculomotor apraxia (AOA type 4) and polyneuropathy [12])
 - “Benign chorea type 2” (with onset of the disease around the age of 40; Japan [97])

Hereditary chorea primarily presenting in children and adolescents

- NXX2-1, ADCY-5, FOXG1, GNAO1, GPR88, SLC2A1, SQSTM1, ATP8A2 or SYT-1 Mutations [5]
- Hereditary disorders of glycosylation (CDG; in children [71])
- ELAC2 gene mutations (rare mitochondrial disease with cardiomyopathy, children with developmental delay, possibly acanthocyttes [76])
- SCN2A mutation (neonatal, early childhood epilepsy, developmental disorders, possibly autism and episodic ataxia [115])
- PDE10A mutations (MRI with bilateral striatal lesions [68])
- KCNQ2 mutations (associated with fever [27])
- ATP1A3 mutations (alternating hemiplegia of childhood (AHC), rapid-onset dystonia, parkinsonism, CAPOS (cerebellar ataxia, areflexia, pes cavus, optic atrophy, sensorineural hearing loss [100]))
- ATP1A2 mutations (regression, hemiplegia, epilepsy [11])
- SUCLA2 mutation in mitochondrial DNA (hypotonia, Dyston/Leigh-like syndrome, deafness, but also myopathy, ataxia [36])
- Glutaric aciduria (AR [44])
- ARX loss (intellectual impairment [87])

Autoimmune and paraneoplastic choreiform syndromes

Sydenham's chorea (chorea minor, post-streptococcal infection disease): anti-streptolysin O (ASL), anti-DNAse B (anti-streptodornase B, ASNB). The relevance of anti-basal ganglia antibodies (immunohistochemical analysis [38] and dopamine D2 receptor Ab (cell-based testing is unclear; so far laboratory analysis established in one centre; specificity not clearly proven [24]).

Systemic lupus erythematosus (SLE); antiphospholipid antibody syndrome (APS); Chorea gravidarum (often SLE, APS or other autoimmune diseases are discussed as the underlying cause for chorea gravidarum [14, 86]).

Multiple sclerosis [37, 104]

Behçet's disease [40].

Chorea in synaptic (idiopathic and paraneoplastic) autoimmune encephalitis is possible, but rarely isolated. GAD65-AK; CASPR2-Ab; NMDA-R-AK; CRMP-5 IgG [108], N-type or P/Q-type calcium channel antibodies [17, 70]; Anti-SOX1-Ab [117], LGI-1-Ab (30–60 high-frequency, short daily brachiofacial dystonic seizures per day preceding or paralleling limbic encephalitis), and probably also other autoimmune, partly post-infectious

or -vaccine encephalitis, e.g. B. IgLON5-Ab (usually with sleep disorders and stridor); [39, 60, 74, 88, 110].

Very rare: paraneoplastic chorea with antibodies against onconeural (intracellular) antigens, most of these cases are progressive and multisystemic syndromes with subacute onset (anti-CV2/CRMP-5, Anti-Hu, Anti-Yo, Anti-Ma2 [14, 21, 30]).

Phosphodiesterase 10A antibody [6]; Takayasu vasculitis [62]; Rasmussen syndrome [35]; celiac disease with anti-gliadin antibodies [112]; steroid responsive Encephalopathy in Autoimmune Thyroiditis (SREAT [102]); Microscopic polyangiitis (MPA [45]).

Infectious causes

HIV encephalopathy [84]; viral encephalitis (mumps, measles, varicella-zoster virus, herpes simplex virus, ECHO group viruses); new variant of Creutzfeldt-Jakob disease; diphtheria; bacterial endocarditis; neurobrucellosis; neurosyphilis; neuroborreliosis; other bacterial encephalitides; cerebral toxoplasmosis; CNS cryptococcosis; neurocysticercosis [14]; Whipple's disease (including ataxia, vertical gaze palsy, oculomasticatory myorhythmias, cognitive impairment [79]); subacute sclerosing panencephalitis (SSPE [98]); Influenza A encephalopathy [81]; SARS-CoV-2 encephalitis [42].

Structural lesions of the basal ganglia

Ischemic or hemorrhagic infarcts; neoplasms; abscessing lesions (incl. toxoplasmosis abscesses and tuberculomas); demyelinating lesions; central pontine/extrapontine myelinolysis; neurosarcoidosis [4, 14]; cavernoma [83]; structural lesions, may also cause hemichorea. Vascular encephalopathy, lacunar infarcts may also cause intermittent chorea [92].

Metabolic, endocrine and toxic causes

Nonketotic hyperglycemia in diabetes mellitus (T1-weighted MRI sequences often show localized hyperintensity, especially in the putamen [18]).

Hypoglycemia; hypo/hypernatremia; hypocalcemia; hypoparathyroidism (sometimes presenting as hemichorea and with calcification of the basal ganglia [26]; hyperthyroidism; acute intermittent porphyria; liver failure including chronic hepatocerebral degeneration; kidney failure; carbon monoxide; manganese; mercury; thallium; organophosphates, 3-NP [14]; vitamin B12 deficiency [28]; Wernicke encephalopathy [89]; 3-Hydroxy-sobutyryl-CoAHydrolase (HIBCH) deficiency (children, sometimes only paroxysmal chorea, e.g. during stress [99]), Tay-Sachs disease [59].

Medication and drug-induced chorea

Dopamine receptor antagonists (e.g. phenothiazine, butyrophenone, benzamide) including antiemetics (metoclopramide); medications for the treatment of Parkinson's disease (such as L-dopa, dopamine agonists, anticholinergics); antiepileptic drugs (e.g. phenytoin, carbamazepine, valproic acid, gabapentin, lamotrigine, pregabalin [85], levetiracetam [118]; calcium channel blockers (cinnarizine, flunarizine, verapamil); lithium; tricyclic antidepressants; SSRI [54]; anti-malarial drugs; steroids; oral contraceptives; antihistamines (H1 and H2); psychostimulants (methylphenidate, amphetamines, pemoline, cocaine); baclofen, digoxin, cyclosporine, theophylline [14]; buprenorphine, hydromorphone and other opiates [66]; ceftriaxone [119]; memantine [9].

Other causes

Polycythaemia vera [67]; essential thrombocythaemia [109]; post-pump chorea after cardiac surgery [90]; superficial siderosis [58]; Moyamoya disease [16]; chorea gravidarum (idiopathic or secondary, see under: "Autoimmune and paraneoplastic choreiform syndromes" [49]; Covid-19 vaccination [7]; intoxications with wood or plant protection products, e.g. propiconazole [77].

Important differential diagnoses for the syndrome chorea include

1. Focal epilepsy [33]. Chorea can sometimes be mistaken for focal epilepsy.
2. Tic Disorders: Unlike chorea, tic disorders are characterized by typical premonitory urges and the ability to suppress tics for a short time [8].
3. Akathisia: It can be challenging to differentiate akathisia from chorea, especially since many anti-dopaminergic drugs including presynaptic dopamine depleters, such as tetrabenazine, can induce akathisia. If akathisia is suspected, a reduction in antihyperkinetic medication may be indicated. Propranolol, anticholinergics, benzodiazepines, and postsynaptic serotonin-5-HT2a receptor antagonists like ritanserin, cyproheptadine, trazodone, mianserin, or mirtazapine can be helpful in managing drug-induced akathisia [80].
4. Myoclonus-dystonia disorders: Conditions like epsilon-sarcoglycan (SGCE) DYT11 gene mutations (often alcohol-sensitive) or VPS16 gene mutations can present as myoclonus-dystonia disorders [75]. Additionally, many other conditions discussed above may exhibit a myoclonus-dystonia phenotype (e.g., benign chorea or PKAN2).

Table 1 Overview of key recommendations for the differential diagnosis of chorea (modified from: Cardoso et al. [14], Hermann and Walker [44], Nguyen et al. [73], Schneider and Bird [95])

Pattern of inheritance	Autosomal-dominant	Huntington's disease (most common inherited chorea, generally with positive family history and typical clinic, molecular genetic testing can be carried out as a next step; but ~8% without positive family history [111]) C9orf72 mutations Spinocerebellar ataxia type 3, 2, 1, 7, 8, 12, 17, 48 DRPLA (especially Japan) HDL2 (especially of African origin) Neuroferritinopathy (NBIA) NKX2-1 (benign course of the disease)
	Autosomal-recessive	Wilson's disease Neuroacanthocytosis Syndroms, VPS13A- and XK-disease / McLeod (CK, blood smear, chorein western blot) PLAN, PKAN, aceruloplasminemia (NBIA) Friedreich's Ataxia Niemann-Pick type C disease AOA1, AOA2 (now SCAN2), AT (AFP elevantion, Albumin reduced) Bilateral striatal necrosis, glutaric aciduria and similiar diseases in childhood
	X-linked	McLeod-Syndrome (CK, blood smear, Kx and Kell blood group phenotype) FXTAS Lesch-Nyhan-Syndrome RETT Syndrome Metabolic diseases in childhood
According to course	Acute	Stroke/ICB
	Subacute	Metabolic Paraneoplastic Drug side effects Malignancies Prion diseases
	Chronic progressive	Neurodegenerative
	Not progressive	Drug side effects Benign Chorea (NKX2-1)
	Episodic	Paroxysmal dyskinesias (PED, SLC2A1, Dyt 18)

Table 1 (continued)

Presenting predominantly in childhood (a selection)	Benign hereditary chorea (including thyroid-transcription-factor-1-gene, TITF1/NKX2-1 mutations; L-Dopa or methylphenidate treatment potentially helpful [32, 106]) ADCY5 mutation [13, 14] Paroxysmal dyskinesias (PED, SLC2A1, Dyt 18) NBIA Lesch-Nyhan-Syndrome, X-linked [44] RETT-Syndrome, X-linked [44] Mitochondriopathies [44] Polynucleotide kinase phosphatase (PNKP) mutation (rather benign course, early onset, with microcephaly, epilepsy, developmental delay, ataxia with oculomotor apraxia (AOA type 4) and polyneuropathy [12]) ELAC2 gene mutations, rare mitochondrial disease with cardiomyopathy, children with developmental delay, possibly acanthocytes [76] FOXP1, GNAO1-, GPR88-, SLC2A1-, SQSTM1-, ATP8A2- oder SYT-1-mutation [5] Hereditary disorders of glycosylation (CDG; in children [71]) SCN2A mutation (neonatal, early childhood epilepsy, developmental disorders, possibly autism and episodic ataxia [115]) PDE10A mutations, MRI with bilateral striatal lesions [68] KCNQ2 mutations, associated with fever [27] ATP1A3 mutations, alternating hemiplegia of childhood (AHC), rapid-onset dystonia, parkinsonism, CAPS (cerebellar ataxia, areflexia, pes cavus, optic atrophy, sensorineural hearing loss [100]) ATP1A2 mutations with regression, hemiplegia, epilepsy [11] SUCLA2 mutations, mitochondrial DNA, hypotonia, Dyston/Leigh-like syndrome, deafness, but also myopathy, ataxia [36] Glutaric aciduria; AR [44] Non-inherited: Sydenham chorea
Symmetry	Asymmetric in diseases with structural lesion or metabolic cause (also generalized possible)
Signs on	Asymmetric Subcortical dementia/Frontal lobe syndrome Ataxia Loss of reflexes/CK Seizures
MRI findings	Iron deposits Calcium depositis (formerly "M. Fahr") Leukenzephalopathy Atrophy pattern Structural lesion

Table 1 (continued)

Recommended laboratory tests	Especially in sporadic cases	Routine lab, including liver parameters, CK (neuroacanthocytosis, but also after a fall, possibly blood smear asking for acanthocytes), vitamin B12, methyl-maleonate, ferritin, AFP (increased in AT and AOA II), albumin (decreased in AOA I) antistreptolysin (AST), Anti-DNAse B, Ceruloplasmin, Copper in serum and 24-h urine collection, ANA, ENA, antidiouble-strand DNA (dsDNA), ANCA, RF, anti-gliadin Ab, paraneoplastic or antineuronal antibodies: e.g.: Anti-HU, -Yo, -Ma, -CRMP-5/ CV2, anti-NMDA-Rec-Ab, anti-GAD-, anti-IgG5, anti-LGI-1-, phospholipid-Ab, cardiolipin-Ab, TSH (basal), anti-thyroid peroxidase (MAK) Ab, TSH receptor auto Ab (TRAK), parathyroid hormone, erythropoietin and hematocrit (Polycythemia vera), Treponema pallidum screening test, borrelia IgG/IgM Ab, HIV, possibly pregnancy test
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Suggested work-up in a patient with chorea

Please refer to Table 1. Medical history, particularly family history, medication history, presence of other relevant medical conditions (as mentioned above).

Neurological assessment to determine if chorea is the only presenting movement disorder and if additional neurological or systemic signs are present. In general, chorea is often more noticeable during conversations, especially when emotionally charged, such as during discussions of stressful topics or during neuropsychological testing. However, during the actual neurological examination, chorea may be less pronounced [8].

Cerebral imaging studies (MRI, if contraindicated CT scan, asking for focal lesions or caudate and/or cortical atrophy. Note that for the determination of caudate atrophy, coronal sections should be available. Signal changes in T2-weighted imaging? Contrast-enhancing lesions? Hypointensities suggesting iron deposition on iron-sensitive magnetic resonance images in order to rule out symptomatic causes or provide evidence for pathognomonic alterations (e.g. "Eye of the tiger" sign for PKAN2; hypointensities in the basal ganglia, particularly in bilateral globus pallidus and substantia nigra for NBIA; "Face of the giant panda" sign for Wilson's disease; cerebellar or pontine atrophy for hereditary ataxia).

Depending on the findings and circumstances: Comprehensive laboratory testing, including cerebrospinal fluid analysis to explore the differential diagnoses mentioned above. FDG-PET-CT or FDG-PET-MRI for tumor screening in suspected paraneoplastic etiology. Heavy Metal Assessment (mercury, manganese, thallium), "drug test" in serum and/or urine.

Additional tests in selected cases: Positron Emission Tomography (e.g., FDG-PET) to detect reduced glucose utilization/hypometabolism in basal ganglia (e.g., in HD and other neurodegenerative disorders causing chorea) or increased glucose utilization/hypermetabolism in

Sydenham's chorea or autoimmune encephalitis including SLE with cerebral involvement, associated with hypometabolism in the prefrontal and premotor cortex [53, 114], hypermetabolism in the hippocampus and orbitofrontal cortex [64], and potentially parieto-occipital hypometabolism in patients with neuropsychiatric disorders [23], contralateral striatal hypoperfusion in patients with non-ketotic hyperglycemia.

If Huntington's disease is suspect

Molecular genetic testing (determination of CAG repeats in the Huntingtin gene, Chromosome 4p) after informed consent following the Genetic Diagnosis Act (GenDG).

Unified Huntington's Disease Rating Scale (UHDRS'99) total motor score.

Neuropsychological or Behavioral Neurological Assessment (psychomotor slowing, frontal-executive dysfunction, memory impairment, decreased speech fluency, spatial-visual disturbances, formal cognitive testing according to UHDRS).

Psychiatric Examination (personality changes, changes in motivation, irritability, aggression, depression, suicidal ideation, delusions, hallucinations, obsessive-compulsive disorders, and sexual disorders; it is recommended to use the "Problem Behaviour Assessment" scale (PBA-s), which is also used in the ENROLL-HD observational study).

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Methodological approach

The project was managed by the coordinator Carsten Saft. The topics were worked on by all other authors based on the current data situation and then coordinated in two Delphi rounds by the guideline group. This S2k level guideline (AWMF-registry number 030/028) is based on a systematic pubmed search. The core statements were evaluated according to the guidelines of the Oxford Centre for Evidence-based Medicine—Levels of Evidence and from this a strength of recommendation was derived. A strong recommendation corresponds in the formulation to a "should", a recommendation to a "should" and an open recommendation to a "can". In a second Delphi round, all recommendations were finally agreed upon by the neurological guidelines group. Based on this expert consensus, the formulation of the core statements was evaluated as strong agreement in the case of > 95% of all experts, as agreement in the case of 75–95%, as majority agreement in the case of > 50–75%, and as no agreement in the case of < 50%. In this abbreviated guideline we only refer to agreements of 90–100%. The final recommendations of this guideline were established through a Delphi process (strength of consensus > 75–95% for all recommendations), achieved over two rounds of voting. All statements for which consensus strength is not specified separately were met with a consensus of > 95%. The guideline was reviewed by the Guideline Committee of the German Neurological Society and approved by the German Neurological Society. Interdisciplinarity was established. As a Patient organization the German HD Association (Deutsche Huntington-Hilfe e.V.) was involved.

Author contributions

CS: leading author of the S2k-Guideline of the German Neurological Society (DNG) for German-Speaking Countries, conception of guideline development process, literature research, interpretation of literature, discussion, approval of recommendations, preparation of the manuscript. GBL: discussion, approval of recommendation, interpretation of literature, editing of manuscript. JMB, MD, HHJ, RK, JP, HPN, KR, RR, KS: interpretation of literature, discussion, approval of recommendations, editing of manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Given by all authors and the German Neurological society.

Competing interests

All participants in the guideline have submitted their declarations of interest (AWMF form for the declaration of interests in the context of guideline projects) to the coordinator or the Editorial Office for Guidelines of the DGN in time and completely filled out. The evaluation of the declarations of interest with regard to thematic relevance to the guideline was carried out by Carsten Saft. The external evaluation of the interests in the overall view was also carried out by NN AWMF. No conflicts of interest were found, so no consequences, e.g. abstentions, were taken. For reasons of transparency, the interests of the participants and the consequences drawn from them are listed on the respective AWMF guideline website and are also shown in the appendix of the short version. No competing interests with regard to the contents (see attached Col-satement A detailed listing is available at <https://dgn.org/leitlinien/>.

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References

1. Adler, C. H., & Wrabetz, L. (1996). Lesch-Nyhan variant: Dystonia, ataxia, near-normal intelligence, and no self-mutilation. *Movement Disorders*, 11(5), 583–584. <https://doi.org/10.1002/mds.870110519>
2. Anheim, M., Monga, B., Fleury, M., Charles, P., Barbot, C., Salih, M., & Koenig, M. (2009). Ataxia with oculomotor apraxia type 2: Clinical, biological and genotype/phenotype correlation study of a cohort of 90 patients. *Brain*, 132(Pt 10), 2688–2698. <https://doi.org/10.1093/brain/awp211>
3. Arber, C. E., Li, A., Houlden, H., & Wray, S. (2016). Review: Insights into molecular mechanisms of disease in neurodegeneration with brain iron accumulation: Unifying theories. *Neuropathology and Applied Neurobiology*, 42(3), 220–241. <https://doi.org/10.1111/nan.12242>
4. Ataya, A., & Harman, E. (2015). Images in clinical medicine. Hemichorea-hemiballismus in neurosarcoidosis. *The New England Journal of Medicine*, 372(21), e27. <https://doi.org/10.1056/NEJMcm1407763>
5. Baizabal-Carvallo, J. F., & Cardoso, F. (2020). Chorea in children: Etiology, diagnostic approach and management. *Journal of Neural Transmission (Vienna)*, 127(10), 1323–1342. <https://doi.org/10.1007/s00702-020-02238-3>
6. Balint, B., & Bhatia, K. P. (2021). Autoimmune movement disorders with neuronal antibodies—An update. *Current Opinion in Neurology*, 34(4), 565–571. <https://doi.org/10.1097/WCO.0000000000000956>
7. Batot, C., Chea, M., Zeidan, S., Mongin, M., Pop, G., Mazoyer, J., & Degos, B. (2022). Clinical and radiological follow-up of a pfizer-BioNTech COVID-19 vaccine-induced hemichorea-hemiballismus. *Tremor and Other Hyperkinetic Movements*, 12, 1–6. <https://doi.org/10.5334/tohm.688>
8. Bonomo, R., Latorre, A., Balint, B., Smilowska, K., Rocchi, L., Rothwell, J. C., & Bhatia, K. P. (2020). Voluntary inhibitory control of chorea: A case series. *Movement Disorders Clinical Practice*, 7(3), 308–312. <https://doi.org/10.1002/mdc3.12907>
9. Borges, L. G., & Bonakdarpour, B. (2017). Memantine-induced chorea and dystonia. *Practical Neurology*, 17(2), 133–134. <https://doi.org/10.1136/practneurol-2016-001470>
10. Bruton, A., & Fuller, L. (2019). Paroxysmal kinesigenic dyskinesia symptoms markedly reduced with parenteral vitamins and minerals: A case report. *Perm Journal*. <https://doi.org/10.7812/TPP/19.036>
11. Calame, D. G., Houck, K., Lotze, T., Emrick, L., & Parnes, M. (2021). A novel ATP1A2 variant associated with severe stepwise regression, hemiplegia, epilepsy and movement disorders in two unrelated patients. *European Journal of Paediatric Neurology*, 31, 21–26. <https://doi.org/10.1016/j.ejpn.2021.01.004>
12. Caputi, C., Tolve, M., Galosi, S., Inghilleri, M., Carducci, C., Angeloni, A., & Leuzzi, V. (2019). PNKP deficiency mimicking a benign hereditary chorea: The misleading presentation of a neurodegenerative disorder. *Parkinsonism & Related Disorders*, 64, 342–345. <https://doi.org/10.1016/j.parkreldis.2019.03.012>
13. Carapito, R., Paul, N., Untrau, M., Le Gentil, M., Ott, L., Alsahel, G., & Bahrami, S. (2015). A de novo ADCY5 mutation causes early-onset autosomal dominant chorea and dystonia. *Movement Disorders*, 30(3), 423–427. <https://doi.org/10.1002/mds.26115>
14. Cardoso, F., Seppi, K., Mair, K. J., Wenning, G. K., & Poewe, W. (2006). Seminar on choreas. *Lancet Neurology*, 5(7), 589–602.
15. Carre, G., Marelli, C., Anheim, M., Geny, C., Renaud, M., Rezvani, H. R., & Tranchant, C. (2017). Xeroderma pigmentosum complementation group F: A rare cause of cerebellar ataxia with chorea. *Journal of the Neurological Sciences*, 376, 198–201. <https://doi.org/10.1016/j.jns.2017.03.021>
16. Cavallieri, F., Zedde, M., Assenza, F., & Valzania, F. (2021). Steroid-responsive acute left-arm chorea as a presenting symptom of moyamoya disease. *Canadian Journal of Neurological Sciences*, 48(2), 287–289. <https://doi.org/10.1017/cjn.2020.155>
17. Chang, K., Lwanga, A., Kaur, T., & Helgason, C. (2018). P/Q and N-type voltage-gated calcium channel binding antibodies associated with paraneoplastic chorea and mixed invasive ductal and lobular carcinoma of the breasts in an elderly patient. *Cureus*, 10(8), e3097. <https://doi.org/10.7759/cureus.3097>
18. Chang, K. H., Tsou, J. C., Chen, S. T., Ro, L. S., Lyu, R. K., Chang, H. S., & Chen, C. J. (2010). Temporal features of magnetic resonance imaging and spectroscopy in non-ketotic hyperglycemic chorea-ballism

- patients. *European Journal of Neurology*, 17(4), 589–593. <https://doi.org/10.1111/j.1468-1331.2009.02867.x>
- 19. Cincotta, M., & Walker, R. H. (2022). One side of the story; clues to etiology in patients with asymmetric chorea. *Tremor Other Hyperkinet Movement (NY)*, 12, 3. <https://doi.org/10.5334/tohm.675>
 - 20. Costain, G., Ghosh, M. C., Maio, N., Carnevale, A., Si, Y. C., Rouault, T. A., & Yoon, G. (2019). Absence of iron-responsive element-binding protein 2 causes a novel neurodegenerative syndrome. *Brain*, 142(5), 1195–1202. <https://doi.org/10.1093/brain/awz072>
 - 21. Crespo-Burillo, J. A., Hernando-Quintana, N., Ruiz-Palomino, P., & Martin-Martinez, J. (2015). Chorea secondary to striatal encephalitis due to anti-CV2/CRMP5 antibodies. Case description and review of the literature. *Neurologia*, 30(7), 451–453. <https://doi.org/10.1016/j.nrl.2013.10.007>
 - 22. Crosiers, D., Blaumeiser, B., & Van Goethem, G. (2019). Spectrum of movement disorders in 18p deletion syndrome. *Movement Disorders Clinical Practice*, 6(1), 70–73. <https://doi.org/10.1002/mdc3.12707>
 - 23. Curiel, R., Akin, E. A., Beaulieu, G., DePalma, L., & Hashefi, M. (2011). PET/CT imaging in systemic lupus erythematosus. *Annals of the New York Academy of Sciences*, 1228, 71–80. <https://doi.org/10.1111/j.1749-6632.2011.06076.x>
 - 24. Dale, R. C., Merheb, V., Pillai, S., Wang, D., Cantrell, L., Murphy, T. K., & Brilot, F. (2012). Antibodies to surface dopamine-2 receptor in autoimmune movement and psychiatric disorders. *Brain*, 135(Pt 11), 3453–3468. <https://doi.org/10.1093/brain/aws256>
 - 25. Danek, A. (2002). Progress in molecular chorea diagnosis. McLeod syndrome and chorea acanthocytosis. *Der Nervenarzt*, 73(6), 564–569. <https://doi.org/10.1007/pl00020831>
 - 26. Desai, K., Walzade, P., Ravat, S. H., & Agarwal, P. A. (2019). Adult-onset isolated hemichorea revealing iatrogenic hypoparathyroidism and bilateral basal ganglia calcification. *Annals of Indian Academy of Neurology*, 22(4), 496–499. https://doi.org/10.4103/aian.AIAN_123_18
 - 27. Dhamija, R., Goodkin, H. P., Bailey, R., Chambers, C., & Brenton, J. N. (2017). A case of KCNQ2-associated movement disorder triggered by fever. *Journal of Child Neurology*, 32(14), 1123–1124. <https://doi.org/10.1177/0883073817736702>
 - 28. Edvardsson, B., & Persson, S. (2011). Chorea associated with vitamin B12 deficiency. *European Journal of Neurology*, 18(10), e138–139. <https://doi.org/10.1111/j.1468-1331.2011.03478.x>
 - 29. Estevez-Fraga, C., Magrinelli, F., Latorre, A., Cordivari, C., Houlden, H., Tinazzi, M., & Bhatia, K. P. (2020). A new family with GLRB-related hyperkplexia showing chorea in homo- and heterozygous variant carriers. *Parkinsonism & Related Disorders*, 79, 97–99. <https://doi.org/10.1016/j.parkreldis.2020.08.016>
 - 30. Etemadifar, M., Salari, M., Badiee, H., & Mirmosayeb, O. (2017). Anti-ma2 receptor encephalitis mimicking Huntington chorea. *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, 22, 31. <https://doi.org/10.4103/1735-1995.202148>
 - 31. Evidente, V. G. H. (1993). X-linked dystonia-parkinsonism. In M. P. Adam, G. M. Mirzaa, R. A. Pagon, S. E. Wallace, L. J. H. Bean, K. W. Gripp, & A. Amemiya (Eds.), *GeneReviews(R)*. Seattle (WA).
 - 32. Farrenburg, M., & Gupta, H. V. (2020). Levodopa-responsive chorea: A review. *Annals of Indian Academy of Neurology*, 23(2), 211–214. https://doi.org/10.4103/aian.AIAN_221_19
 - 33. Fasano, A., Di Bonaventura, C., Bove, F., Espay, A. J., Morgante, F., Fabbrini, G., & Berardelli, A. (2019). Movement disorders phenomenology in focal motor seizures. *Parkinsonism & Related Disorders*, 61, 161–165. <https://doi.org/10.1016/j.parkreldis.2018.10.021>
 - 34. Flies, C. M., & Veldink, J. H. (2020). Chorea is a pleiotropic clinical feature of mutated fused-in-sarcoma in amyotrophic lateral sclerosis. *Amyotroph Lateral Scler Frontotemporal Degener*, 21(3–4), 309–311. <https://doi.org/10.1080/21678421.2020.1733021>
 - 35. Gambardella, A., Andermann, F., Shorvon, S., Le Piane, E., & Aguglia, U. (2008). Limited chronic focal encephalitis: Another variant of Rasmussen syndrome? *Neurology*, 70(5), 374–377. <https://doi.org/10.1212/01.wnl.0000298723.96653.99>
 - 36. Garone, C., Gurgel-Giannetti, J., Sanna-Cherchi, S., Krishna, S., Naini, A., Quinzii, C. M., & Hirano, M. (2017). A novel SUCLA2 mutation presenting as a complex childhood movement disorder. *Journal of Child Neurology*, 32(2), 246–250. <https://doi.org/10.1177/0883073816666221>
 - 37. Ghosh, R., Roy, D., Dubey, S., Das, S., & Benito-Leon, J. (2022). Movement disorders in multiple sclerosis: An update. *Tremor Other Hyperkinet Movement (NY)*, 12, 14. <https://doi.org/10.5334/tohm.671>
 - 38. Gupta, H. V., Barnes, H., Radhi, F. A., & Jassam, Y. (2020). Chorea and Parkinsonism with elevated striatal antibody. *Annals of Indian Academy of Neurology*, 23(2), 223–224. https://doi.org/10.4103/aian.AIAN_364_19
 - 39. Hacohen, Y., Wright, S., Waters, P., Agrawal, S., Carr, L., Cross, H., & Lim, M. J. (2013). Paediatric autoimmune encephalopathies: Clinical features, laboratory investigations and outcomes in patients with or without antibodies to known central nervous system autoantigens. *Journal of Neurology, Neurosurgery and Psychiatry*, 84(7), 748–755. <https://doi.org/10.1136/jnnp-2012-303807>
 - 40. Hamid, M., Adan, K., Satte, A., & Bourazza, A. (2021). Chorea in neuro-Behcet's disease. *Cureus*, 13(10), e19039. <https://doi.org/10.7759/cureus.19039>
 - 41. Hanna, M. G., Davis, M. B., Sweeney, M. G., Noursadeghi, M., Ellis, C. J., Elliott, P., & Marsden, C. D. (1998). Generalized chorea in two patients harboring the Friedreich's ataxia gene trinucleotide repeat expansion. *Movement Disorders*, 13(2), 339–340. https://doi.org/10.1002/mds.87013_0223
 - 42. Hassan, M., Syed, F., Ali, L., Rajput, H. M., Faisal, F., Shahzad, W., & Badshah, M. (2021). Chorea as a presentation of SARS-CoV-2 encephalitis: A clinical case report. *Journal of Movement Disorders*. <https://doi.org/10.14802/jmd.20098>
 - 43. Hensman Moss, D. J., Poulter, M., Beck, J., Hehir, J., Polke, J. M., Campbell, T., & Tabrizi, S. J. (2014). C9orf72 expansions are the most common genetic cause of Huntington disease phenocopies. *Neurology*, 82(4), 292–299. <https://doi.org/10.1212/WNL.0000000000000061>
 - 44. Hermann, A., & Walker, R. H. (2015). Diagnosis and treatment of chorea syndromes. *Current Neurology and Neuroscience Reports*, 15(2), 514. <https://doi.org/10.1007/s11910-014-0514-0>
 - 45. Iftikhar, S. (2018). Chorea and calcifications: Atypical presentation of microscopic polyangiitis. *Mayo Clinic Proceedings*, 93(7), 961–962. <https://doi.org/10.1016/j.mayocp.2018.05.023>
 - 46. Josephs, K. A., Van Gerpen, M. W., & Van Gerpen, J. A. (2003). Adult onset Niemann-Pick disease type C presenting with psychosis. *Journal of Neurology, Neurosurgery and Psychiatry*, 74(4), 528–529. <https://doi.org/10.1136/jnnp.74.4.528>
 - 47. Kambouris, M., Bohlega, S., Al-Tahan, A., & Meyer, B. F. (2000). Localization of the gene for a novel autosomal recessive neurodegenerative Huntington-like disorder to 4p15.3. *American Journal of Human Genetics*, 66(2), 445–452. <https://doi.org/10.1086/302744>
 - 48. Kaur, J., Parveen, S., Shamim, U., Sharma, P., Suroliya, V., Sonkar, A. K., & Faruq, M. (2020). Investigations of Huntington's disease and Huntington's disease-like syndromes in Indian choreatic patients. *Journal of Huntington's Disease*, 9(3), 283–289. <https://doi.org/10.3233/JHD-200398>
 - 49. Kim, A., Choi, C. H., Han, C. H., & Shin, J. C. (2009). Consecutive pregnancy with chorea gravidarum associated with moyamoya disease. *Journal of Perinatology*, 29(4), 317–319. <https://doi.org/10.1038/jp.2008.183>
 - 50. Klein, C. (2014). Genetics in dystonia. *Parkinsonism & Related Disorders*, 20(Suppl 1), S137–142. [https://doi.org/10.1016/S1353-8020\(13\)70033-6](https://doi.org/10.1016/S1353-8020(13)70033-6)
 - 51. Kovacs, G. G., Murrell, J. R., Horvath, S., Haraszti, L., Majtenyi, K., Molnar, M. J., & Spina, S. (2009). TARDBP variation associated with frontotemporal dementia, supranuclear gaze palsy, and chorea. *Movement Disorders*, 24(12), 1843–1847. <https://doi.org/10.1002/mds.22697>
 - 52. Koya Kutty, S., Mulroy, E., Magrinelli, F., Di Lazzaro, G., Latorre, A., & Bhatia, K. P. (2021). Huntington disease-like phenotype in a patient with ANO3 mutation. *Parkinsonism & Related Disorders*, 90, 120–122. <https://doi.org/10.1016/j.parkreldis.2021.02.022>
 - 53. Krakauer, M., & Law, I. (2009). FDG PET brain imaging in neuropsychiatric systemic lupus erythematosus with choreic symptoms. *Clinical Nuclear Medicine*, 34(2), 122–123. <https://doi.org/10.1097/RNU.0b013e318192c4d2>
 - 54. Laban, T., Larroche, C., Comparon, C., Dhote, R., & Degos, B. (2021). Fluoxetine-induced chorea. *Revue Neurologique (Paris)*, 177(8), 1010–1011. <https://doi.org/10.1016/j.neurol.2020.12.007>
 - 55. Lance, E. I., Kronenburger, M., Cohen, J. S., Furmanski, O., Singer, H. S., & Fatemi, A. (2018). Successful treatment of choreo-athetotic movements in a patient with an EEF1A2 gene variant. *SAGE Open Medical Case*

- Reports, 6, 2050313X18807622. <https://doi.org/10.1177/2050313X18807622>
56. Lapostolle, A., Delion, T., Arnaud, S., Manceau, P., & Degos, B. (2021). FXTAS patient presenting as Huntington-like generalized chorea. *Revue Neurologique (Paris)*, 177(4), 445–446. <https://doi.org/10.1016/j.neurol.2020.08.008>
57. Le Ber, I., Moreira, M. C., Rivaud-Pechoux, S., Chamayou, C., Ochsner, F., Kuntzer, T., & Durr, A. (2003). Cerebellar ataxia with oculomotor apraxia type 1: Clinical and genetic studies. *Brain*, 126(Pt 12), 2761–2772. <https://doi.org/10.1093/brain/awg283>
58. Le Rhun, E., Soto Ares, G., Pecheux, N., Destee, A., & Defebvre, L. (2008). Superficial hematosiderosis of the central nervous system improved by corticosteroids. *Revue Neurologique (Paris)*, 164(3), 264–270. <https://doi.org/10.1016/j.neurol.2007.08.010>
59. Lefter, S., O'Mahony, O., Sweeney, B., & Ryan, A. M. (2021). Late-onset Tay-Sachs disease in an Irish family. *Movement Disorders Clinical Practice*, 8(1), 106–110. <https://doi.org/10.1002/mdc3.13096>
60. Leyboldt, F., Armanagé, T., & Dalmau, J. (2015). Autoimmune encephalopathies. *Annals of the New York Academy of Sciences*, 1338(1), 94–114. <https://doi.org/10.1111/nyas.12553>
61. Lieto, M., Riso, V., Galatolo, D., De Michele, G., Rossi, S., Bargigiani, M., & Silvestri, G. (2020). The complex phenotype of spinocerebellar ataxia type 48 in eight unrelated Italian families. *European Journal of Neurology*, 27(3), 498–505. <https://doi.org/10.1111/ene.14094>
62. Lopes, A. S., Clemente, G., Len, C. A., Masruha, M. R., & Terrier, M. T. (2015). Chorea: A rare manifestation of Takayasu's arteritis. *Revista Brasileira de Reumatologia*, 55(4), 384–386. <https://doi.org/10.1016/j.rbr.2013.09.003>
63. Macaya, A., Munell, F., Burke, R. E., & De Vivo, D. C. (1993). Disorders of movement in Leigh syndrome. *Neuropediatrics*, 24(2), 60–67. <https://doi.org/10.1055/s-2008-1071515>
64. Mackay, M., Tang, C. C., Volpe, B. T., Aranow, C., Mattis, P. J., Korff, R. A., & Edelberg, D. (2015). Brain metabolism and autoantibody titres predict functional impairment in systemic lupus erythematosus. *Lupus Science and Medicine*, 2(1), e000074. <https://doi.org/10.1136/lupus-2014-000074>
65. Malik, G. M., Mubarik, M., Khan, M. D., Lone, B. A., Kadla, S. A., & Bhat, F. A. (1995). Laurence-Moon-Biedl-Bardet syndrome with chorea. *Journal of the Association of Physicians of India*, 43(4), 295–296.
66. Martin, E. J., Vaughan, C. L., Atayee, R., Hirst, J. M., O'Donnell, K., & Edmonds, K. P. (2018). Hydromorphone-induced chorea as an atypical presentation of opioid neurotoxicity: A case report and review of the literature. *Palliative Medicine*, 32(9), 1529–1532. <https://doi.org/10.1177/0269216318786861>
67. Marvi, M. M., & Lew, M. F. (2011). Polycythemia and chorea. *Handbook of Clinical Neurology*, 100, 271–276. <https://doi.org/10.1016/B978-0-444-52014-2.00019-7>
68. Mencacci, N. E., Kamsteeg, E. J., Nakashima, K., R'Bilo, L., Lynch, D. S., Balint, B., & Bhatia, K. P. (2016). De novo mutations in PDE10A cause childhood-onset chorea with bilateral striatal lesions. *American Journal of Human Genetics*, 98(4), 763–771. <https://doi.org/10.1016/j.ajhg.2016.02.015>
69. Mizuguchi, M., & Kamoshita, S. (1993). Movement disorders in miscellaneous disorders—Inherited metabolic diseases. *Nihon Rinsho*, 51(11), 2919–2923.
70. Morgan, T. T., Armitage, A., Stone, B., & Benge, J. (2019). Non paraneoplastic immune-mediated calcium channel chorea. *Proceedings (Baylor University Medical Center)*, 32(2), 281–282. <https://doi.org/10.1080/0898280.2019.1581318>
71. Mostile, G., Barone, R., Nicoletti, A., Rizzo, R., Martinelli, D., Sturiale, L., & Zappia, M. (2019). Hyperkinetic movement disorders in congenital disorders of glycosylation. *European Journal of Neurology*, 26(9), 1226–1234. <https://doi.org/10.1111/ene.14007>
72. Nass, R., Petito, C., Stoner, E., & New, M. (1986). Neuronal ceroid lipofuscinosis with hypergonadotropic hypogonadism. *Journal of Child Neurology*, 1(2), 142–144. <https://doi.org/10.1177/088307388600100209>
73. Nguyen, Q. T. R., Ortigoza Escobar, J. D., Burgunder, J. M., Mariotti, C., Saft, C., Hjermind, L. E., & Bachoud-Levi, A. C. (2022). Corrigendum: Combining literature review with a ground truth approach for diagnosing huntington's disease phenocopy. *Frontiers in Neurology*, 13, 891800. <https://doi.org/10.3389/fneur.2022.891800>
74. O'Toole, O., Lennon, V. A., Ahlskog, J. E., Matsumoto, J. Y., Pittcock, S. J., Bower, J., & McKeon, A. (2013). Autoimmune chorea in adults. *Neurology*, 80(12), 1133–1144. <https://doi.org/10.1212/WNL.0b013e3182886991>
75. Park, J., Reilaender, A., Petry-Schmelzer, J. N., Stobe, P., Cordts, I., Harmuth, F., & Haack, T. B. (2022). Transcript-specific loss-of-function variants in VPS16 are enriched in patients with dystonia. *Neuro Genetics*, 8(1), e644. <https://doi.org/10.1212/NXG.0000000000000644>
76. Paukar, M., Pajak, A., Freyer, C., Bergendal, A., Dory, M., Laffita-Mesa, J. M., & Svenssonsson, P. (2018). Chorea, psychosis, acanthocytosis, and prolonged survival associated with ELAC2 mutations. *Neurology*, 91(15), 710–712. <https://doi.org/10.1212/WNL.0000000000006320>
77. Paul, B. S., Paul, G., Kaur, J., & Singh, G. (2017). Chorea as unusual complication of fungicide poisoning. *Journal of Postgraduate Medicine*, 63(1), 53–54. <https://doi.org/10.4103/0022-3859.198156>
78. Pedroso, J. L., de Freitas, M. E., Albuquerque, M. V., Saraiva-Pereira, M. L., Jardim, L. B., & Barsottini, O. G. (2014). Should spinocerebellar ataxias be included in the differential diagnosis for Huntington's disease-like syndromes? *Journal of the Neurological Sciences*, 347(1–2), 356–358. <https://doi.org/10.1016/j.jns.2014.09.050>
79. Peregrin, J., & Malikova, H. (2015). Primary whipple disease of the brain: Case report with long-term clinical and MRI follow-up. *Neuropsychiatric Disease and Treatment*, 11, 2461–2469. <https://doi.org/10.2147/Ndt.S92066>
80. Poyurovsky, M., & Weizman, A. (2020). Treatment of antipsychotic-induced akathisia: Role of serotonin 5-HT(2a) receptor antagonists. *Drugs*, 80(9), 871–882. <https://doi.org/10.1007/s40265-020-01312-0>
81. Prasuhn, J., Royl, G., Wandinger, K. P., Bruggemann, N., Neumann, A., & Munte, T. F. (2018). Transient generalized chorea in influenza A encephalopathy. *Tremor Other Hyperkinet Movement (N Y)*, 8, 591. <https://doi.org/10.7916/D8F495TP>
82. Prohaska, R., Sibon, O. C., Rudnicki, D. D., Danek, A., Hayflick, S. J., Verhaag, E. M., & Walker, R. H. (2012). Brain, blood, and iron: Perspectives on the roles of erythrocytes and iron in neurodegeneration. *Neurobiology of Diseases*, 46(3), 607–624. <https://doi.org/10.1016/j.nbd.2012.03.006>
83. Qiu, J., Cui, Y., Sun, L., Guo, Y., & Zhu, Z. (2018). Hemichorea associated with cavernous angioma and a small hemorrhage: A case report and literature review. *Medicine (Baltimore)*, 97(43), e12889. <https://doi.org/10.1097/MD.00000000000012889>
84. Rajakaruna, G. K., Italiano, C. M., John, M., & Nolan, D. (2020). Chorea associated with persistent low-level viremia in a patient living with HIV: A case report. *Journal of Virus Eradication*, 6(1), 27–29.
85. Rissardo, J. P., & Caprara, A. L. F. (2020). Pregabalin-associated movement disorders: A literature review. *Brain Circulation*, 6(2), 96–106. https://doi.org/10.4103/bcb.bcb_57_19
86. Robottom, B. J., & Weiner, W. J. (2011). Chorea gravidarum. *Handbook of Clinical Neurology*, 100, 231–235. <https://doi.org/10.1016/B978-0-444-52014-2.00015-X>
87. Rodgers, J., Calvert, S., Shoubridge, C., & McGaughran, J. (2021). A novel ARX loss of function variant in female monozygotic twins is associated with chorea. *European Journal of Medical Genetics*, 64(11), 104315. <https://doi.org/10.1016/j.ejmg.2021.104315>
88. Sabater, L., Gaig, C., Gelpi, E., Battaller, L., Lewerenz, J., Torres-Vega, E., & Graus, F. (2014). A novel non-rapid-eye movement and rapid-eye-movement parasomnia with sleep breathing disorder associated with antibodies to IgLON5: A case series, characterisation of the antigen, and post-mortem study. *Lancet Neurology*, 13(6), 575–586. [https://doi.org/10.1016/S1474-4422\(14\)70051-1](https://doi.org/10.1016/S1474-4422(14)70051-1)
89. Sabatini, J. S., Schutz-Pereira, G. L., Feltrin, F., Teive, H. A. G., & Camargo, C. H. F. (2016). Wernicke's encephalopathy with chorea: Neuroimaging findings. *Dement Neuropsychol*, 10(4), 370–372. <https://doi.org/10.1590/s1980-5764-2016dn1004020>
90. Saft, C., Reber, D., Streuer, M., & Andrich, J. (2011). Post pump chorea in a 77-year-old male. *Neurological Sciences*, 32(4), 699–701. <https://doi.org/10.1007/s10072-011-0583-7>
91. Saft, C., Skodda, S., Nguyen, H. P., Park, J., & Haack, T. B. (2021). Teaching video neuroimage: New STUB1 variant causes chorea, tremor, dystonia, myoclonus, ataxia, depression, cognitive impairment, epilepsy, and superficial siderosis. *Neurology*, 97(17), E1749–E1750. <https://doi.org/10.1212/WNL.00000000000012264>
92. Salgado, P., Taipa, R., Domingos, J., Dias, D., Pires, M. M., & Magalhaes, M. (2017). Vascular pathology causing late onset generalized chorea: A

- clinico-pathological case report. *Movement Disorders Clinical Practice*, 4(6), 819–823. <https://doi.org/10.1002/mdc3.12528>
93. Santens, P., Van Damme, T., Steyaert, W., Willaert, A., Sablonniere, B., De Paep, A., & Dermaut, B. (2015). RNF216 mutations as a novel cause of autosomal recessive Huntington-like disorder. *Neurology*, 84(17), 1760–1766. <https://doi.org/10.1212/WNL.0000000000001521>
 94. Saracchi, E., Castelli, M., Bassi, M. T., Brighina, E., Cereda, D., Marzorati, L., & Brighina, L. (2014). A novel heterozygous SETX mutation in a patient presenting with chorea and motor neuron disease. *Amyotroph Lateral Scler Frontotemporal Degener*, 15(1–2), 138–140. <https://doi.org/10.3109/21678421.2013.865751>
 95. Schneider, S. A., & Bird, T. (2016). Huntington's disease, Huntington's disease look-alikes, and benign hereditary chorea: What's new? *Movement Disorders Clinical Practice*, 3(4), 342–354. <https://doi.org/10.1002/mdc3.12312>
 96. Shah, P. A., & Kuchhai, F. A. (2009). Galactosemia with chorea—An unusual presentation. *Indian Journal of Pediatrics*, 76(1), 97–98. <https://doi.org/10.1007/s12098-009-0037-x>
 97. Shimohata, T., Hara, K., Sanpei, K., Nunomura, J., Maeda, T., Kawachi, I., & Honma, Y. (2007). Novel locus for benign hereditary chorea with adult onset maps to chromosome 8q21.3 q23.3. *Brain*, 130(Pt 9), 2302–2309. <https://doi.org/10.1093/brain/awm036>
 98. Singh, P., Saini, A. G., Sankhyan, N., Gupta, P., & Vyas, S. (2015). Blindness, dancing extremities, and corpus callosum and brain stem involvement: An unusual presentation of fulminant subacute sclerosing panencephalitis. *Journal of Child Neurology*, 30(1), 87–90. <https://doi.org/10.1177/0883073813520498>
 99. Spitz, M. A., Lenaers, G., Charif, M., Wirth, T., Chelly, J., Abi-Warde, M. T., & Roubertie, A. (2021). Paroxysmal dyskinesias revealing 3-hydroxy-isobutyryl-CoA hydrolase (HIBCH) deficiency. *Neuropediatrics*, 52(5), 410–414. <https://doi.org/10.1055/s-0040-1722678>
 100. Sweeney, M. T., Newcomb, T. M., & Swoboda, K. J. (2015). The expanding spectrum of neurological phenotypes in children with ATP1A3 mutations, alternating hemiplegia of childhood, rapid-onset dystonia-parkinsonism, CAPOS and beyond. *Pediatric Neurology*, 52(1), 56–64. <https://doi.org/10.1016/j.pediatrneurol.2014.09.015>
 101. Synofzik, M., Schule, R., Schulte, C., Kruger, R., Lindig, T., Schols, L., & Asmus, F. (2010). Complex hyperkinetic movement disorders associated with POLG mutations. *Movement Disorders*, 25(14), 2472–2475. <https://doi.org/10.1002/mds.23307>
 102. Taurin, G., Golfer, V., Pinel, J. F., Deburghgraeve, V., Poirier, J. Y., Edan, G., & Verin, M. (2002). Chorea syndrome due to Hashimoto's encephalopathy. *Movement Disorders*, 17(5), 1091–1092. <https://doi.org/10.1002/mds.10230>
 103. Tonekaboni, S. H., & Mollamohammadi, M. (2014). Neurodegeneration with brain iron accumulation: An overview. *Iranian Journal of Child Neurology*, 8(4), 1–8.
 104. Tranchant, C., Bhatia, K. P., & Marsden, C. D. (1995). Movement disorders in multiple sclerosis. *Movement Disorders*, 10(4), 418–423. <https://doi.org/10.1002/mds.870100403>
 105. Traschutz, A., Cortese, A., Reich, S., Dominik, N., Faber, J., Jacobi, H., & Group, R. F. C. S. (2021). Natural history, phenotypic spectrum, and discriminative features of multisystemic RFC1 disease. *Neurology*, 96(9), e1369–e1382. <https://doi.org/10.1212/WNL.00000000000011528>
 106. Tubing, J., Bohnenpoll, J., Spiegler, J., Gillessen-Kaesbach, G., Baumer, T., Max, C., & Munchau, A. (2018). Methylphenidate can improve chorea in NKX2.1 and ADCY5 mutation-positive patients—a report of two children. *Mov Disorders Clinical Practice*, 5(3), 343–345. <https://doi.org/10.1002/mdc3.12608>
 107. van der Weijden, M. C. M., Rodriguez-Contreras, D., Delnooz, C. C. S., Robinson, B. G., Condon, A. F., Kielhold, M. L., & Verbeek, D. S. (2021). A gain-of-function variant in dopamine D2 receptor and progressive chorea and dystonia phenotype. *Movement Disorders*, 36(3), 729–739. <https://doi.org/10.1002/mds.28385>
 108. Vaswani, P. A., Kimchi, E. Y., & Hung, A. Y. (2020). CRMP-5-IgG associated paraneoplastic chorea. *Movement Disorders Clinical Practice*, 7(6), 713–715. <https://doi.org/10.1002/mdc3.13019>
 109. Venkatesan, E. P., Ramadoss, K., Balakrishnan, R., & Prakash, B. (2014). Essential thrombocythemia: Rare cause of chorea. *Annals of Indian Academy of Neurology*, 17(1), 106–107. <https://doi.org/10.4103/0972-128569>
 110. Vynogradova, I., Savitski, V., & Heckmann, J. G. (2014). Hemichorea associated with CASPR2 antibody. *Tremor Other Hyperkinet Movements (NY)*, 4, 239. <https://doi.org/10.7916/D8VM49C5>
 111. Walker, F. O. (2007). Huntington's disease. *Lancet*, 369(9557), 218–228.
 112. Walker, R. H. (2011). Further evidence for celiac disease-associated chorea. *Tremor Other Hyperkinet Movements (NY)*. <https://doi.org/10.7916/D82806BC>
 113. Wang, Z. B., Liu, J. Y., Xu, X. J., Mao, X. Y., Zhang, W., Zhou, H. H., & Liu, Z. Q. (2019). Neurodegeneration with brain iron accumulation: Insights into the mitochondria dysregulation. *Biomedicine & Pharmacotherapy*, 118, 109068. <https://doi.org/10.1016/j.biopha.2019.109068>
 114. Weiner, S. M., Otte, A., Schumacher, M., Brink, I., Juengling, F. D., Sobanksi, T., & Peter, H. H. (2000). Alterations of cerebral glucose metabolism indicate progress to severe morphological brain lesions in neuropsychiatric systemic lupus erythematosus. *Lupus*, 9(5), 386–389. <https://doi.org/10.1191/096120300678828370>
 115. Wolff, M., Brunklaus, A., & Zuberi, S. M. (2019). Phenotypic spectrum and genetics of SCN2A-related disorders, treatment options, and outcomes in epilepsy and beyond. *Epilepsia*, 60(Suppl 3), S59–S67. <https://doi.org/10.1111/epi.14935>
 116. Wright, R. A., Pollock, M., & Donaldson, I. M. (1992). Chorea and tuberous sclerosis. *Movement Disorders*, 7(1), 87–89. <https://doi.org/10.1002/mds.870070119>
 117. Yamagishi, T., Inoue, K., Ouchi, H., Shibano, K., & Hara, K. (2020). A case of anti-SRY-related HMG-box gene 1 (SOX1) antibody-positive chorea. *Rinsho Shinkeigaku*, 60(12), 852–856. <https://doi.org/10.5692/clinicalneuro.cn-001454>
 118. Yim, S. H., Choi, Y. H., Heo, K., & Cho, K. H. (2019). A case of dyskinesia after levetiracetam administration. *BMC Neurology*, 19(1), 292. <https://doi.org/10.1186/s12883-019-1519-8>
 119. Yokoyama, Y., Hosokawa, N., Kudo, T., Goda, H., Ito, K., Suzuki, M., & Funakoshi, R. (2020). Chorea-like symptoms and high blood concentration of ceftriaxone in a patient undergoing hemodialysis: A case report. *Journal of Infection and Chemotherapy*, 26(3), 285–288. <https://doi.org/10.1016/j.jiac.2019.10.005>
 120. Zech, M., Bardakjian, T. M., Stoklosa, M., Ploski, R., Jech, R., Gonzalez-Alegre, P., & Winkelmann, J. (2021). A neurodevelopmental disorder with dystonia and chorea resulting from clustering CAMK4 variants. *Movement Disorders*, 36(2), 520–521. <https://doi.org/10.1002/mds.28398>

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