

MAJOR ARTICLE

Seroepidemiology of human tularemia – systematic review and metaanalysis of seroprevalence studies

Chantal Mattatia¹¹, Philipp KA Agyeman^{1*}, Nina Schöbi¹, Simon Aebi^{1,2}, Andrea Duppenthaler¹, Michael Büttcher^{3,4,5}, Christoph Aebi¹

¹Division of Pediatric Infectious Disease, Department of Pediatrics, Bern University Hospital, University of Bern, CH-3010 Bern, Switzerland; ²Risk and Resilience Team, Center for Security Studies (CSS), Eidgenössische Technische Hochschule (ETH), CH-8092 Zurich, Switzerland; ³Paediatric Infectious Diseases Unit, Department of Paediatrics, Children's Hospital Lucerne, Lucerne Cantonal Hospital, Lucerne, Switzerland; ⁴Faculty of Medicine and Health Sciences, University Lucerne, Lucerne, Switzerland; ⁵Paediatric Pharmacology and Pharmacometrics Research Center, University Children's Hospital Basel, Basel, Switzerland

Background: Seroepidemiologic studies of human tularemia have been conducted throughout the northern hemisphere. The purposes of this study are (1) to provide an overview of *F. tularensis* seroprevalence data, and (2) to generate an estimate of the proportion of study participants whose infection remained subclinical.

Methods: We conducted a systematic review of *Francisella tularensis* seroprevalence studies according to the PRISMA guidelines. We searched Pubmed®, Embase® and Web of ScienceTMcovering the period from 1951 to 2023.

Results: The weighted pooled seroprevalence among 44'486 participants recruited in 52 studies was 3.7% (95% confidence interval (CI) 2.7-5.1). Reported seroprevalences ranged between 0.2%

© The Author(s) 2023. Published by Oxford University Press on behalf of Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*}contributed equally

Corresponding author: Christoph Aebi, Division of Pediatric Infectious Disease, Department of Pediatrics, Bern University Hospital, Inselspital, University of Bern, CH-3010 Bern, Switzerland [christoph.aebi@insel.ch]

and 31.3%. Occupational activities associated with an increased likelihood of exposure (risk ratio (RR) 3.51 (3.2-3.86)) and studies from North America vs. Europe and Asia (4.53 (4.15-4.94)) were associated with significantly increased seropositive rates. Twenty-eight datasets (47%) reported clinical information on a total of 965 seropositive participants. The weighted pooled estimate for subclinical seropositivity was 84.4% (95% CI 72.9-991.7). Studies from *F. tularensis* type A areas (RR 0.37, 95% CI 0.27-0.51) and studies from sites where pulmonary tularemia prevailed (RR 0.38, 95% CI 0.28-0.51) reported lower subclinical seropositivity rates than studies from type B areas and from areas of predominance of (ulcero-) glandular or oropharyngeal tularemia, respectively.

Conclusions: Throughout the northern hemisphere, only a small proportion of study participants showed serologic evidence of exposure to F. *tularensis*. Eight outof 10 seropositive participants had no historical evidence of past clinical tularemia.

Keywords: tularemia – seroepidemiology – seroprevalence – subclinical – systematic review

Key points: The weighted seroprevalence of human antibodies against *Francisella tularensis* in endemic areas of the northern hemisphere was 3.7%. Eight out of 10 seropositive participants had no historical evidence of past clinical tularemia.

INTRODUCTION

Human tularemia is a bacterial zoonosis caused by *Francisella tularensis*, a small, gram-negative coccobacillus with the capacity to infect a wide range of mammals, arachnids, insects and other animals. There are two main subspecies. Type A (*F. tularensis subsp. tularensis*) is mainly restricted to North America, although a few strains have been isolated in Europe [1]. Type B (*F. tularensis subsp. holarctica*) is distributed throughout the Northern hemisphere and has also been isolated in Australia [2]. Modes of acquisition in humans are diverse and include arthropod bites (ticks, mosquitos), ingestion of contaminated freshwater or soil, direct contact with infected live or dead animals, and inhalation of contaminated aerosols. Accordingly, clinical manifestations vary and include (ulcero-) glandular, oropharyngeal, typhoidal, and pulmonary manifestations [3].

While both the molecular pathogenesis of tularemia [4] and the clinical manifestations in humans have been studied in detail [3, 5], important questions remain unanswered. An issue of particular interest to clinicians is the likelihood of subclinical infection among exposed individuals (i.e., a-or oligosymptomatic, medically unattended infection). Some authors have postulated that the majority of infections remain undetected [6], while others believe that most cases cause a distinct clinical syndrome [7, 8]. A comprehensive review of the available data is lacking. Such information, however, is important, as it may elucidate whether subclinical infection as opposed to clinically overt disease is the typical human response to *F. tularensis* exposure. It may assist in

the clinical interpretation of diagnostic test results and contribute to what is known on how effectively the human immune system deals with *F. tularensis*.

One means of addressing this question is to examine seroprevalence studies. Seropositive individuals without a history of clinical disease compatible with tularemia can be considered to have experienced subclinical infection. If they represent the majority of seropositive individuals, it follows that clinical disease cannot usually be explained by pathogen virulence alone, but requires a particular set of additional conditions for it to occur. This may be particularly relevant in geographic areas, where the less virulent type B circulates [9]. Clinical disease could then be considered as evidence of some sort of immune compromise around the time of infection. Evidently, alternative explanations are possible and include the mode of acquisition, the infectious dose, strain-specific virulence determinants and genetic predisposition, which may impact on the extent of clinical disease.

Thus, the purposes of this study were (1) to generate an overview of F. tularensis seroprevalence rates reported from endemic areas worldwide, and (2) to generate an estimate of the proportion of human tularemia cases identified by detectable serum antibodies that had no history of past clinical manifestations suggestive or confirmed to be clinical tularemia (subclinical cases). Because each study usually identifies only a handful of seropositive individuals, we conducted a systematic review of tularemia seroprevalence studies between the 1940s and 2023 in accordance with the PRISMA guidelines for systematic reviews [10].

METHODS

Data source and search strategy

Pubmed® (1946 to present), Embase® (1947 to present) and Web of ScienceTM (1921 to present) were searched using the following search term combinations: (Tularemia OR Francisella) AND (seroprevalence OR seroepidemiolog*); (Tularemia OR Francisella) AND antibody AND prevalence; antibod* prevalence tulare* human. Refinements were added as needed. The list of references of each retrieved article was searched for additional suitable articles. No language limitations applied. Studies written in languages other than English, German or French were full-text translated using Deeple Pro® (www.deepl.com). The generated list of articles was screened by title and abstract independently by two authors (CM and CA) who applied the predefined inclusion/exclusion criteria (s. below). Discrepancies were resolved by consensus.

Inclusion and exclusion criteria

We included studies, research letters and abstracts that reported original data on the prevalence of serum antibodies against *F. tularensis* in humans. No restriction applied regarding the population studied (e.g., geographic location, general vs. risk populations, age, gender, ethnicity) and outbreak vs. non-outbreak time periods of serum sampling. Publications were included if published before

30 April 2023. We excluded papers without original data or those duplicating previously published data, papers focusing on clinical cases, and papers without methodologic description of antibody detection tests used. Studies reporting a seroprevalence of 0 were recorded, but excluded from analyses.

Quality assessment

We next assessed all retrieved studies using a critical appraisal checklist adapted from the Joanna Briggs Institute checklist for prevalence studies [11, 12] (supplementary file 1, table S1). We included studies, which scored ≥ 5 of 7 points (>70%) in the 7-question checklist. In addition, we devised a list of criteria identifying high-specificity serologic testing for use in subanalyses (supplementary file 1, table S2).

Data retrieved from selected articles

The following data were extracted from all articles included in the analysis: study year and location, population characteristics (age, sex, risk factors for *F. tularensis* exposure), use of a study questionnaire for participants, serologic tests used, tests for cross-reactivity used, cut-offs values defining seropositivity, number and proportion of participants testing positive, number and proportion of "subclinical" participants testing positive, narrative clinical description of participants testing positive.

Definition of subclinical infection

We defined, for the purpose of this study, a "subclinical case" as an individual who was seropositive for *F. tularensis* and explicitly asymptomatic at the time of serum sampling and had a past medical history lacking episodes of known tularenia or tularenia-like illness.

Statistical analysis and software used

Pooled counts and weighted proportions of participants seropositive for *F. tularensis* were calculated for the entire set of studies and subgroups of interest. Weighted proportions were calculated using both a fixed effect model and a random effect model using restricted maximum likelihood estimation (REML) of the between-study variance [13], because substantial heterogeneity between the studies (e.g., geographic region, risk of exposure to *F. tularensis*) other than sampling errors were expected to occur. No limit for heterogeneity as expressed by the *P* statistic applied. Forest plots are presented with a 95% confidence interval (95% CI) and group size. Weighted seroprevalences rates for subgroup analyses and risk ratios (RR) were calculated with the same model when the comparative datasets were complete. When incomplete, the RR were calculated from the weighted seroprevalences directly. For linear correlation analysis, the Pearson correlation coefficient was calculated. Funnel plots were constructed by plotting the log odds against the study size as recommended by Hunter et al [14] for metaanalyses of proportion studies (figure S3). "R" software package, version 4.0.3 (Vienna, Austria; https://www.R-

project.org/) and VassarStats (<u>www.vassarstats.net</u>) were used for analysis. The map (figure S2) was created using ArcGIS®, intellectual property of Esri (<u>www.esri.com</u>).

RESULTS

Bibliographic data and study settings

We identified 52 articles fulfilling the predefined selection criteria. A flowchart detailing the selection process adapted from PRISMA [15] is provided in figure 1. Six additional seroprevalence studies extracted during the search process were excluded, because no seropositive individuals were identified [16-21]. Seven selected articles provided 2 datasets [6, 22-27], resulting in a total of 59 datasets (tables 1 and 2). The distribution of the publication years over time is shown in figure S1 (supplementary data file 1). The geographic distribution of study countries is displayed in figure S2 (supplementary data file 1). Key study settings are summarized in table 1. Details of each study are provided in the supplementary data file 2.

Pooled seroprevalence data

A total of 44'486 participants were included. Seroprevalence rates as reported in each study ranged from 0.2 to 31.3 percent (table 2). Sample size (n) and seroprevalence rate were not significantly correlated (Pearson r -0.203, 95% CI -0.436 - 0.056, p=0.123). The weighted seroprevalence rate of 59 pooled datasets calculated with the random effects model was 3.7% (95% CI 2.7-5.1) (figure 2). The I^2 statistic was 96% (96-97; p<0.01) indicating major heterogeneity. This was expected because of real between-study differences in the participant cohorts' risk of exposure to *F. tularensis*. A funnel plot designed for proportion studies [14] (figure S3, supplementary data file 1) confirmed the heterogeneity in that there was no convergence of prevalence rates as the sample sizes increased.

Male and female individuals accounted for 18'106 and 9'885 participants, respectively, in a total of 28 datasets. Sex was not specified for another 16'477 participants in 31 datasets. The serostatus could be attributed to sex in 10'994 male and 8'094 female participants (table 3). The respective weighted seroprevalence rates were 4.7% (95% CI 3.0-7.3) and 5.2% (95% CI 3.1-8.8), and the RR was 0.83 (95% CI 0.66-1.05). We did not calculate age-specific pooled seroprevalence rates because the reporting of age and age group in the original studies was very heterogeneous and did not allow consistent grouping across a majority of studies.

Table 3 also lists the pooled weighted seroprevalence rates and estimated risk ratios according to the geographic region of the study sites, occupational risk of exposure, serologic testing strategy and the availability of clinical information. The pooled seroprevalence of studies conducted in North America was greater than that from Europe and Asia combined (9.6% vs. 2.7%; RR 4.53, 95% CI 4.15-4.94). A combined total of 16'554 study participants (37%) reported an occupational risk of exposure to *F. tularensis*. Occupational risk factors included hunting and trapping (16

studies), military service (5), animal husbandry, farming and ranching (5), butchering and slaughterhouse work (5), forestry work (4), veterinary medicine (3) and landscaping (1). Included are North American native participants whose lifestyle placed them at an increased risk of exposure. The pooled weighted seroprevalence of these at-risk populations was significantly greater than in studies of populations without such risk factors (5.5% vs. 2.4%, RR 3.51 (3.20-3.86); table 3). However, only 5 studies provided a complete comparative dataset of individuals with vs. without an increased risk of exposure and the respective seropositivity rates [28-32]. The pooled seropositivity rates of these studies were 2.7% (95% CI 1.5-4.9) and 2.4% (95% CI 1.3-4.5), respectively, with a risk ratio of 1.20 (95% CI 0.94-1.53).

Grouping of the 52 studies according to the serologic testing strategy revealed that high-specificity testing according to our ad hoc definition (supplementary file 1, table S2) was associated with a lower weighted seropositivity rate than non-high specificity testing (3.5% vs. 4.6%, RR 0.49 (0.45-0.53), table 3).

Subclinical tularemia

Clinical detail provided on study participants testing seropositive varied widely and was predominantly limited to summary statements. A total of 28 datasets [6-8, 22-24, 31, 33-53] comprised a total of 13'807 individuals, for whom clinical information was available. Of these, 965 tested positive (weighted seroprevalence 5.5% (95 CI 3.8-7.8) vs. 2.6% (95% 1.6-4.2) in studies without clinical data) (table 3). Table S3 in the supplementary data file 1 lists pertinent quotations from the narrative clinical accounts given in each study.

A past medical history compatible with tularemia or a tularemia-like illness (with or without a physician-confirmed diagnosis) among these 965 seropositive individuals could be elicited in 143, leaving 819 considered as having had subclinical tularemia. Figure 3 provides a Forest plot with the weighted rates of subclinical seropositivity reported in these studies. Overall, the weighted subclinical seropositivity rate was 84.4% (95% CI 72.9-91.7). Additional subanalyses (table 4) revealed that this rate was lower in studies from areas where, according to the literature [9, 54, 55], *F. tularensis* type A was prevalent [6, 52] than in studies from type B areas. Similarly, studies from areas with a predominance of pulmonary tularemia around the time of serum sampling [6, 8] reported lower subclinical seropositivity rates than studies from areas where (ulcero-) glandular or oropharyngeal tularemia prevailed. Subanalyses according to occupational risk of exposure and testing strategy are also listed in table 4.

DISCUSSION

This systematic review and metaanalysis covers 52 studies from North America, Europe and Asia. Studies from North America dominated the first 50 years of observation between 1951 and 2002, while studies from Europe and Asia combined (mostly its westernmost region including Turkey and Iran) prevailed between 2003 and 2023 (figures S1 and S2). The latter contributed more than three quarters of all study subjects (table 3), reflect the more recent regional seroepidemiology, and mainly used advanced, commercially available methods for testing. Their weighted pooled seroprevalence was greater than four-fold smaller than the corresponding rate derived from North American studies (table 3). Possible explanations include differences in virulence of circulating *F. tularensis* subtypes and clades, different transmission paths and dynamics, the predominance of indigenous participants in North American studies [22, 24, 33, 34, 38, 41, 44, 56-58], whose lifestyle may have led to frequent to exposure, and possibly less specific serologic methods used in the earlier decades of the observation period. However, the absolute difference in seroprevalence being 6.9% only, we chose to pool study data in subsequent analyses irrespective of their geographic origin.

The weighted overall seroprevalence rate of 3.5% for *F. tularensis* antibodies (figure 1) and the equally important finding that 90% of reported rates ranged between 0.3% and 18% (table 2) emphasize that only a small minority of individuals living in endemic areas provides serologic evidence of past infection. These findings were the result of pooling data that were generated by the use of different serologic assays. Our original intention to group and compare the studies according to the test type used was not feasible as the combinations of methods and cut-off values, the choices of confirmatory tests and the reporting formats varied widely (table 2 and supplementary data file 2). Thus, we devised an ad hoc definition for "high-specificity" testing (table S2), which indeed identified a group of studies that yielded a lower pooled seroprevalence rate suggesting greater specificity than non-"high-specificity" testing (table 3). The absolute difference of the pooled seroprevalences grouped accordingly of 1.1% (table 3) again was not clinically or epidemiologically relevant.

The finding of low seroprevalence rates across most of the reviewed studies is important both epidemiologicallyand clinically. It may reflect that tularemia transmission can be highly focal and even within endemic areas the risk of acquisition is extremely heterogeneous. Also, as serum antibodies persist for decades, cross-sectional serosurveys capture exposure events dating back many years, which underscores the rarity of tularemia. Clinically, the interpretation of a positive serology result in a patient is facilitated by knowing the pretest-likelihood of seropositivity in the community. Even when such data are not available at a particular location, this figure indicates that the "background" seropositivity rate in endemic areas is predictably low irrespective of the serologic test system used. A low seroprevalence was also found for study participants reporting occupational or lifestyle activities expected to increase the risk of exposure. While their likelihood of testing positive was indeed more than threefold that of non-at-risk participants (table 3), their pooled seroprevalence rate remained low at 5.5% with 90% of studies reporting a rate below 15% (table S2 and supplementary data file 2).

The second objective of this review was to quantify the proportion of seropositive individuals who had undergone the infection subclinically, i.e., who did not report a history compatible with past tularemia. The use of seroprevalence studies to address the issue of subclinical infection is well

established and has been used previously for other pathogens, e.g., *Borrelia burgdorferi* [59, 60]. Our calculation indicates that this proportion was 84.4% (figure 3) and was not affected by reported risk factors for exposure (table 4). Interestingly, subclinical infection appears to the norm for many vector-borne zoonoses, e.g., Lyme disease [59] or tick-borne encephalitis [61]. The interest in establishing the rate of subclinical tularemia is epidemiologic, clinical and scientific. In clinical epidemiology, mandatory reporting of tularemia cases is a common public health tool used to monitor the disease activity over time. Our data suggest that this tool likely catches 10-20% of infections at best as subclinical cases remain unreported. Knowledge of the rate of subclinical infection may not be applicable in outbreak situations and when pneumonic tularemia indicates aerosol transmission [6, 8].

In clinical practice, knowing the rate of subclinical seropositivity provides additional information for the interpretation of a positive serology, even when seroconversion around the time of an acute illness appears to strongly suggest true tularemia. Considering the high rate of subclinical tularemia indicates that the positive predictive value (PPV) of a given serologic test may be lower than commonly reported, e.g., for ELISA tests [62, 63]. Definitions for "true positive" used to calculate the PPV often rely on compatible clinical illness (reviewed by Maurin [1]) and may lead to overestimation of the PPV as compatible clinical illness, such as acute lymphadenopathy or pneumonia, has multiple causes with tularemia being an infrequent one. Thus, when tularemia is suspected, care should be taken to identify the organism by culture or PCR whenever possible.

In science, an estimate of the rate of subclinical seropositivity may add a puzzle piece to what is known about host susceptibility to *F. tularensis*. Our finding suggests that the immune defenses of most individuals, who mount a detectable humoral immune response, control and eliminate *F. tularensis* in the absence of substantial clinical manifestations. The question arises why then a 10-20% minority of becomes clinically ill at all. Only isolated cases of patients with defined immunodeficiencies and severe tularemia have been described (reviewed by Bahuaud and coworkers [64]). The vast majority of patients appear immunologically healthy before and after the disease. Future research could be directed towards the identification of subtle deviations within the framework of virulence factors and immunoprotective events associated with tularemia that could explain a temporal susceptibility to symptomatic disease in otherwise immunocompetent individuals. This could entail a comprehensive "systems-level" approach [65] comparing immune functions in previously healthy individuals with severe tularemia with individuals with subclinical infection.

The results of this systematic review need to be taken with caution. In the former Sovjet Union, tularemia was extensively studied, but publications were inaccessible using the search strategy we used. Limitations also include the large time window of study dates and widely scattered locations, virulence differences of and within the two main subspecies, partly incomplete description of participant cohorts, diverse serologic test systems, and the often sketchy descriptions of how the clinical history of seropositive participants were obtained. It also needs to be kept in mind that the

fundamental problem of serosurveys for estimating subclinical infection rates is the often low disease prevalence in these settings. Consequently, the PPV of serology is low with a tendency to overestimate subclinical infection because of contamination by false-positive results. In addition, as the majority of clinical information obtained was historical in nature and reflected the participants' recollections, it can be assumed that the data are incomplete and likely overestimate the rate of subclinical infections.In conclusion, we find that in temperate and arctic zones of the northern hemispheres where human tularemia occurs, only a small proportion of the population has ever been exposed to *F. tularensis*. Eighty to 90% of exposed persons are not aware of ever having had overt tularemia or a clinical illness compatible with it.

Author contributions:CA conceived the study and wrote the study protocol. CM and CA independently conducted the literature search, study screening and critical study appraisal, agreed on the final list of studies for analysis, and aggregated the data. PKAA and SA conducted statistical analyses. NS, MB and AD commented on analysis and drafting of the manuscript, which was prepared by CM and CA. All authors critically reviewed the various versions of the manuscript and approved the content and format of its final version.

Patient consent statement: All patient data used in this systematic reviewed had previously been published. Informed consent was not required for systematic reviews according to the competent local ethics committee.

Conflict of interest statement: The authors declare no conflict of interest.

Funding sources: No external funding.

DOI: 10.1093/ofid/ofad636

Figure 1 Bibliographic search and selection flow diagram adapted from the PRISMA statement [46].



Figure 2 Forest plot of the pooled seroprevalence rates for antibodies against F. tularensis.



DOI: 10.1093/ofid/ofad636

Figure 3 Forest plot of the pooled rates of subclinical seropositivity for antibodies against *F. tularensis*.



Table 1 Main settings of human F. tularensis seroprevalence studies included in this systematic review

Characteristic	n (%)
No. of studies	52 (100)
No. of studies with 2 datasets	7 (13)
No. of datasets	59
<i>Geography</i> Study site (world region)	
North America	13 (21)
Eastern Europe	4 (7)
Northern Europe	4 (7)
Western Europe	11 (21)
Middle East	18 (34)
East Asia	2 (4)
Υ.	
Study site in <i>F. tularensis</i> type B region	50 (96)
Study site in <i>F. tularensis</i> type A region	2 (4)
Participant characteristics	
Age span specified	52 (100)
Participants below 15 years of age included	10 (19)

Participants exclusively below 18 years of age		1 (2)
Sex distribution of seropositive individuals specified	28	3 (54)
Risk factors for F. tularensis exposure identified	33	3 (64)
Clinical information on seropositive participants provided	28	3 (56)
Participant questionnaire used	39	9 (75)
Serology for F. tularensis Primary antibody test used		Â
Tube agglutination test (TAT)	10	5 (36)
Microagglutination test (MAT)	1:	2 (23)
Enzyme Linked Immuno Sorbent Assay (ELISA)	19	9 (36)
Other		5 (8)
Confirmatory secondary test(s) used	12	2 (23)
Cross-reactivity against Brucella sp. tested	20	6 (50)
High-specificity test strategy used	39	9 (75)
Non high-specificity test strategy used	1;	3 (25)

Table 2 Human F. tularensis	seroprevalence	studies included	in this	systematic	review
------------------------------------	----------------	------------------	---------	------------	--------

		All p	ipants	Sei	Seropositive participants					
Author (year)	Cou ntry	Age (y)	n	Male/fe male (n)	n	%	Clinically evaluated (n)	Subclini cal (n)	Antibod y test(s)	
	Canad	2-88	2'9	1'623/1'319	3	1	344	344	TAT ¹	
Wood (1951) [33]	а		42		4	1.				
					4	7				
Greenberg et al	Canad	3-93	79		5	7.			TAT ¹	
(1957) [56]	а		7		8	3				
Greenberg et al	Canad	0-78	1'0		1	1			TAT ¹	
(1958) [57]	а		31		3	3.				
					9	5				
Philip et al (1962)	USA	all	11		3	2			TAT	
[34]			5		3	8.				
					-	7				
	USA	18-65	79	793/0	6	7.	20	15	TAT	
			3	000/10	0	6		10		
Philip et al (1967)	USA	15-84	34	332/12	4	1	45	42	IAI	
[22]			4		5	3.				
	• •		410		-	1	74	~~~	- • - 1	
Danistrand et al	Swede	all	12			5.	/1	20	IAL	
(1971) [8]	n Na mua	40.47	01		1	9				
Haug et al (1972)	Norwa	13-17	81		1	1.			TAT (Widai)	
[23]	y No ruo	odult	Э 5 Г Г		1	5	2	0		
<i>V</i>	Norwa	adun	55		3	э. Б	3	0	TAT (Widai)	
Kackala at al (1082)	y Finlon	adult	1'0		7	0	7	6	$T\Lambda T^{1}$	
105Keid et di (1902)	d	auun	70		'	0. 7	1	0		
[June of al (1903)		adult	17		2	1			$T\Lambda T^{1}$	
[66]	034	auun	14		2	1				
[00]						т . З				
l évesque et al	Canad	40+1	16	157/8	4	2	4	4	I AT	
(1995) [24]	a	2	5		•	4				
(/L J			-							

	Canad	adult	16		1	0.			LAT
Aquilini et al (200	a 0) Italv	21-65	5 50	507/0	1	6 2.	13	13	IF
[36]	o) ((a))	2.00	7	00170	3	6			
Feldman et al	USA	adult	13	104/28	1	9.	12	8	MAT
(2003) [6]			2		2	1			
	USA	adult	31	154/156	1	0.			MAT
			0			3			
Gutiérrez et al	Spain	14-92	4'8	2'324/2'486	9	0.			MAT
(2003) [67]			25			2			
Deutz et al (2003)	Austri	adult	14	146/3	5	3.			MAT
[68]	a	40 70	9			4			
Porsch-Ozcurum	ez Germa	18-79	66		1	0.			ELISA+WB
et at (2004) [03]	5) Cormo	adult	J∠ 1'1		5 1	2			
1601	5) Germa	auun	10		4	0. 3			LLISATIVD
Gürcan et al (200	6) Turkev	all	49 26		1	3	10	. 3	МАТ
[37]		un	6		0	8	10		100 (1
Dedeoglu Kilinc e	et Turkey	6-92	1'7	1'213/569	5	0.			MAT
al (2007) [28]			82			3			
Campagna et al	Canad	>15	24	105/146	4	1	42	33	TAT
(2011) [41]	а		9		2	6.			
						9			
Lévesque et al	Canad	50	50	22/28	2	4.	2	1	TAT
(2007) [38]	а					0			
Jenzora et al (200	8) Germa	adult	28		5	1.	5	3	ELISA+WB+
[39]	ny		6	11100/11000		7			IFA
Splettstoesser	Germa	10-65	24	1/169/1/263	5	2.			ELISA+WB
(2009) [70] Bazavaka at al	ny Slovek	odult	16		0	ა ე	11	11	TAT ¹
Dazovska el al	Siuvak	adun	29			з. 7	11	11	IAI
(2010) [71] Wölfel et al (2010		adult	9 76	670/95	1	1			FLISA
[72]	lia	adun	5	010/00	3	7			and/or IF
Otkun et al (2011)) Turkev	0.5-	11	60/55	3	3	36	34	TAT
[42]	,	76	5		6	1.		• •	
[]			\mathbf{X}		-	3			
Yagzi et al (2011)	Turkey	16-77	24	134/106	5	2.	5	4	ELISA+MAT
[43]			0			1			
Sampasa-Kanying	ga Canad	>18	26	110/157	4	1	48	39	TAT
et al (2012) [44]	а		4		8	8.			
						2			
Messier et al (201	2) Canad	18-74	91		1	1			TAT
[73]	a		7		7	8.			
Marsh water at	Tendens	40.07	0.4	0.4/0	3	9	4	0	
Yesilyurt et al	Turkey	18-67	64	64/0	4	6.	4	2	MAT+ELISA
(2012) [45] Clark at al (2012)	Azorb	<u>\18</u>	70	347/440	1	3	102	100	ELISA
[74]	Azero	>10	6	347/449	2	ו 5	125	122	ELISA
נידין	aijan		0		2	5			
Tobudicetal (201	4) Austri	18-60	54	534/12	3	0.			FLISA
[75]	a	10 00	6	001/12	U	5			22.0,1
Esmaeili et al	Iran	>18	18	184/0	1	6.	12		ELISA
(2014) [47]			4		2	5			
Esmaeili et al	Iran	>18	25	206/44	3	1			ELISA
(2014) [29]			0		6	4.			
						4			
Khoshdel et al	Iran	2-18	18	89/94	1	6.	11	11	ELISA
(2014) [48]		o -	3	1-0/10	1	0	-		
Zukiewicz-Sobcza	ak Polan	35-55	21	176/40	7	3.	0		ELISA
et al (2014) [30]	a		6			2			

Baryam et al (2015)	Turkey	18-93	49 5	152/343	1	3. 6	18	15	MAT
Zákutná et al (2015)	Slovak	adult	12	77/47	5	4.			ELPAGA+W
[76]	ia		4			0			В
Rossow et al (2015)	Finlan	30-92	1'0	481/564	1	1.	16	15	ELISA+WB
[7]	d		45		6	5			
Jurke et al (2015)	Germa	18-66	72	569/153	2	4.			ELISA+WB
	ny Turkau	40.4	2	450/400	9	0	22	22	FLICA
	тикеу	49±1	32 1	100/100	2	1.	23	23	ELISA
[JU] Büyük et al (2016)	Turkov	, 15	4 20	178/23	1	7	15	15	MATTELISA
[51]	тиксу	215	20	170/23	5	5	15	15	
Rigaud et al (2016)	Franc	17-81	2'8	2916/59	1	5			TAT+FUSA
[77]	e		75	2010/00	6	7			
[]					4				
De Keukeleire et al	Belgiu	25-72	14	128/20	3	2.			ELISA+ICT
(2017) [25]	m		8			0			
	Belgiu	18-68	40	118/90	2	0.			ELISA+ICT
	m		2			5	Ċ		
Akhvlediani et al	Georgi	18-59	50	476/13	1	2.			MAT
(2018) [26]	а		0		0	0			
	Georgi	18-65	69	310/387	3	5.			MAT
	а		7		5	0			
Esmaeili S et al	Iran	30-50	14	144/0	4	2.			ELISA
(2019) [27]			4			8			
	Iran	27-53	14	145/0	7	4.	Y		ELISA
		40 -0	5	075/05		8	40	10	
Esmaeili S et al	Iran	18-78	36	275/85	1	2.	10	10	ELISA
(2019) [31]			0	19/10	0	8	0	0	MAT
Harrist et al (2019)	USA		23	13/10	3	1	3	2	IVIAI
[52]						J. □			
Takeda et al (2019)	lanan	18-90	1'1		1	1	12	12	RSA
[53]	Japan	10 50	52		2	0	12	12	NOA
Özdemir et al	Turkey	20-80	36	180/180	2	7			FUSA
(2019) [78]	runkey	20 00	0	100/100	7	5			ELION
Obaidatetal (2020)	Jordan	all	82	339/489	6	7.			ELISA
[79]			8		4	7			
Karatas Yeni et al	Cypru	>18	43		4	0.			MAT
(2022) [80]	s		0			9			
Davarci et al (2023)	Turkey	2-89	41	226/184	6	1.			MAT
[81]			0			5			

Abbreviations: TAT, Tube Agglutination Test; LAT, Latex Agglutination Test; IF, Immunofluorescence Assay; MAT,MicroAgglutinationTest;ELISA, EnzymeLinkedImmunoSorbentAssay;WB,WesternBlot;ICT,ImmunoChromatographyTest;RSA,RapidSlideAgglutination¹The term «agglutination reaction» is used in the original publication.

	Downloaded from https://academic.oup.com/ofid/advance-article/doi/10.1093/ofid/ofad636/7473454 by E-Library
	7473454 by E-Librar
	y Insel user on 04 Ja
	inuary 2024

Table 3 Pooled weighted seroprevalence of F. tularensis antibodies according to sex, geographic region, risk of exposure, testing strategy and clinical data availability in study participants of 52 studies included in this review

	Char	acteristic	prese	nt		Characteristic absent							-		
			Pro (95	portio % CI	ons)				Prop (959	portic % CI	ons)	Risk CI)	ratio	o (95%	
Characterist				Fixe						Fixe			Fixe		
ic				d	Rando					d	Rando		d	Rando	
		Seroposi	iti Ra	effec	cm		Ser	opositi	Ra	effec	cm		effec	m	
	n	ve	W	t	effect	n	ve	1	w	t	effect	Raw	t	effect	
Male sex*	1099	566	5.1	8.8	4.7	8054		394	4.9	9.1	5.2	1.05	0.99	0.83	
	4			(8.1- 9.5)	(3.0-7.3)					(8.3- 10)	(3.1-8.8)	(0.93 - 1.19)	(0.92 - 1.05)	(0.66- 1.05)	
North America	8311	967	11.	12.9	9.6	3617		929	2.6	5.0	2.7	4.53	4.53	4.53	
(vs. Europe or Asia)			6	(12.1 - 13.7)	(6.3- 14.3)	5				(4.7- 5.3)	(1.9-3.9)	(4.15 - 4.94)	(4.33 - 4.74)	(4.15- 4.94)	
Occupational	1655	1281	7.7	10.0	5.5	2793		615	2.2	5.1	2.4	3.51	3.51	3.51	
exposure	4			(9.5- 10.6)	(3.9-7.8)	2				(4.7- 5.5)	(1.5-3.8)	(3.19 - 3.86)	(3.35 - 3.69)	(3.20- 3.86)	
High-	3448	1189	3.4	7.5	3.5	1000		707	7.1	8.9	4.6	0.49	0.49	0.49	
specificity serologic testing used	6		Ó	(7.1- 8)	(2.4-5.1)	0				(8.3- 9.6)	(2.6-8.1)	(0.45 - 0.54)	(0.47 - 0.51)	(0.45- 0.53)	
Clinicial	1380	965	7.0	9.4	5.5	3067		931	3.0	6.8	2.6	2.30	2.30	2.30	
information available	7			(8.9- 10)	(3.8-7.8)	9				(6.4- 7.2)	(1.6-4.2)	(2.11 - 2.51)	(2.20 - 2.41)	(2.11- 2.51)	

* information on sex was available in a subset of studies only

Table 4 Pooled weighted subclinical seropositivity rate for *F. tularensis* according to study location, geographic predominance of type A vs. type B, predominance of different clinical syndromes in the study area, presence or absence of occupational risk of exposure, and serologic testing strategy

	Ch	ara	cteri	stic pre	esent	Charac	teri	istic a	bsent				
			Prop	portion (95%		Proportion			n (95%	Risk r	5% CI)		
			CI)			<u>c</u>	,1)					570 CI)	
Characteristic				Fixed	Random			Fixed	Random		Fixed	Random	
			Raw	reffect	effect	R	law	effect	effect	Raw	effect	effect	
North America (vs. Europe or	560	488	87.1	65.6	80.7	405331 8	31.7	63.0	86.3	1.07	1.07	1.07	
Asia)				(57.6- 72.9)	(54.5-93.5)			(54.5- 70.8)	(72.5-93.8)	(1.01- 1.13)	(1.04- 1.10)	(1.01-1.13)	
F. tularensis type A area (vs.	75	25	33.3	33.4	47.1	890794 8	9.2	72.5	87.3	0.37	0.37	0.37	
type B)				(23.1- 45.6)	(18.7-77.6)			(66.6- 77.7)	(76.7-93.5)	(0.27- 0.51)	(0.32- 0.44)	(0.27-0.51)	
Predominance of pulmonary	83	28	33.7	33.6	44.6	882791 8	89.7	74.1	86.7	0.38	0.38	0.38	
tularemia (vs. (ulcero-) glandular or oropharyngeal tularemia)				(23.9- 44.9)	(14.2-79.8)			(68.3- 79.2)	(76.1-93.1)	(0.28- 0.51)	(0.32- 0.44)	(0.28-0.51)	
Occupational risk of exposure	e 639	559	87.5	68.3	82	326260 7	9.8	57.8	88.1	1.10	1.10	1.10	
				(61.2- 74.7)	(66.2-91.3)			(48.1- 66.9)	(67.6-96.4)	(1.03- 1.17)	(1.06- 1.13)	(1.03-1.17)	
High-specificity serologic testing	g 446	321	72.0	51.0	78.3	519498 9	96.0	86.4	93.0	0.75	0.75	0.75	
				(43.7- 58.2)	(61.1-89.3)			(80.3- 90.8)	(82.3-97.4)	(0.71- 0.80)	(0.73- 0.77)	(0.71-0.80)	

REFERENCES

- 1. Maurin, M., *Francisella tularensis, Tularemia and Serological Diagnosis*. Front Cell Infect Microbiol, 2020. **10**: p. 512090.
- 2. Aravena-Roman, M., A. Merritt, and T.J. Inglis, *First case of Francisella bacteraemia in Western Australia*. New Microbes New Infect, 2015. **8**: p. 75-7.
- 3. Maurin, M. and M. Gyuranecz, *Tularaemia: clinical aspects in Europe*. Lancet Infect Dis, 2016. **16**(1): p. 113-124.
- 4. Pechous, R.D., T.R. McCarthy, and T.C. Zahrt, *Working toward the future: insights into Francisella tularensis pathogenesis and vaccine development.* Microbiol Mol Biol Rev, 2009. **73**(4): p. 684-711.
- 5. Schobi, N., et al., *Pediatric Tularemia-A Case Series From a Single Center in Switzerland*. Open Forum Infect Dis, 2022. **9**(7): p. ofac292.
- 6. Feldman, K.A., et al., *Tularemia on Martha's Vineyard: seroprevalence and occupational risk*. Emerg Infect Dis, 2003. **9**(3): p. 350-4.
- 7. Rossow, H., et al., *Incidence and seroprevalence of tularaemia in Finland*, 1995 to 2013: regional epidemics with cyclic pattern. Euro Surveill, 2015. **20**(33): p. 21209.

- 8. Dahlstrand, S., O. Ringertz, and B. Zetterberg, *Airborne tularemia in Sweden*. Scand J Infect Dis, 1971. **3**(1): p. 7-16.
- 9. Schmid, G.P., et al., *Clinically mild tularemia associated with tick-borne Francisella tularensis*. J Infect Dis, 1983. **148**(1): p. 63-7.
- 10. Page, M.J., et al., *PRISMA 2020 explanation and elaboration: updated guidance and exemplars for reporting systematic reviews.* BMJ, 2021. **372**: p. n160.
- 11. Munn, Z., et al., *The development of a critical appraisal tool for use in systematic reviews addressing questions of prevalence.* Int J Health Policy Manag, 2014. **3**(3): p. 123-8.
- Munn, Z., et al., Methodological guidance for systematic reviews of observational epidemiological studies reporting prevalence and cumulative incidence data. Int J Evid Based Healthc, 2015. 13(3): p. 147-53.
- Kulinskaya, E., S. Morgenthaler, and R.G. Staudte, *Combining the evidence using stable weights*. Res Synth Methods, 2010. 1(3-4): p. 284-96.
- 14. Hunter, J.P., et al., *In meta-analyses of proportion studies, funnel plots were found to be an inaccurate method of assessing publication bias.* J Clin Epidemiol, 2014. **67**(8): p. 897-903.
- 15. Page, M.J., et al., *The PRISMA 2020 statement: an updated guideline for reporting systematic reviews.* BMJ, 2021. **372**: p. n71.
- Juncker-Voss, M., et al., [Screening for antibodies against zoonotic agents among employees of the Zoological Garden of Vienna, Schonbrunn, Austria]. Berl Munch Tierarztl Wochenschr, 2004. 117(9-10): p. 404-9.
- 17. de Carvalho, I.L., et al., *Francisella tularensis, Portugal*. Emerg Infect Dis, 2007. **13**(4): p. 666-7.
- 18. Tokarska-Rodak, M., et al., Serological surveillance of vector-borne and zoonotic diseases among hunters in eastern Poland. J Vector Borne Dis, 2016. **53**(4): p. 355-361.
- 19. Tokarska-Rodak, M., et al., Seroprevalence of Selected Zoonotic Agents among Hunters from Eastern Poland. Pol J Microbiol, 2018. 67(2): p. 233-236.
- 20. Cetinkol, Y., et al., *Investigation of zoonotic infections in risk groups in Ordu University Hospital, Turkey*. Niger J Clin Pract, 2017. **20**(1): p. 6-11.
- 21. Miernyk, K.M., et al., *Human Seroprevalence to 11 Zoonotic Pathogens in the U.S. Arctic, Alaska.* Vector Borne Zoonotic Dis, 2019. **19**(8): p. 563-575.
- 22. Philip, R.N., E.A. Casper, and D.B. Lackman, *The skin test in an epidemiologic study of tularemia in Montana trappers*. J Infect Dis, 1967. **117**(5): p. 393-402.
- Haug, R.H. and A.D. Pearson, *Human infections with Francisella tularensis in Norway*. Development of a serological screening test. Acta Pathol Microbiol Scand B Microbiol Immunol, 1972. 80(2): p. 273-80.
- 24. Levesque, B., et al., Seroepidemiologic study of three zoonoses (leptospirosis, Q fever, and tularemia) among trappers in Quebec, Canada. Clin Diagn Lab Immunol, 1995. **2**(4): p. 496-8.
- 25. De Keukeleire, M., et al., Seroprevalence of Borrelia burgdorferi, Anaplasma phagocytophilum, and Francisella tularensis Infections in Belgium: Results of Three Population-Based Samples. Vector Borne Zoonotic Dis, 2017. 17(2): p. 108-115.
- 26. Akhvlediani, N., et al., *Tularemia transmission to humans: a multifaceted surveillance approach*. Epidemiol Infect, 2018. **146**(16): p. 2139-2145.
- 27. Esmaeili, S., et al., Seroepidemiological study of Q fever, brucellosis and tularemia in butchers and slaughterhouses workers in Lorestan, western of Iran. Comp Immunol Microbiol Infect Dis, 2019. **66**: p. 101322.

- 28. Dedeoglu Kilinc, G., et al., [Investigation of tularemia seroprevalence in the rural area of Thrace region in Turkey]. Mikrobiyol Bul, 2007. **41**(3): p. 411-8.
- 29. Esmaeili, S., et al., Seroepidemiological survey of tularemia among different groups in western *Iran*. Int J Infect Dis, 2014. **18**: p. 27-31.
- Zukiewicz-Sobczak, W., et al., Prevalence of antibodies against selected zoonotic agents in forestry workers from eastern and southern Poland. Ann Agric Environ Med, 2014. 21(4): p. 767-70.
- 31. Esmaeili, S., et al., *Epidemiological survey of tularemia in Ilam Province, west of Iran.* BMC Infect Dis, 2019. **19**(1): p. 502.
- 32. Jurke, A., et al., Serological survey of Bartonella spp., Borrelia burgdorferi, Brucella spp., Coxiella burnetii, Francisella tularensis, Leptospira spp., Echinococcus, Hanta-, TBE- and XMRvirus infection in employees of two forestry enterprises in North Rhine-Westphalia, Germany, 2011-2013. Int J Med Microbiol, 2015. **305**(7): p. 652-62.
- Wood, W.J., Tularemia; a study based on the incidence of positive agglutination tests against P. tularensis in the Indian population of Manitoba and North-Western Ontario. Manit Med Rev, 1951. 31(10): p. 641-4.
- 34. Philip, R.N., et al., Serologic and skin test evidence of tularemia infection among Alaskan Eskimos, Indians and Aleuts. J Infect Dis, 1962. **110**: p. 220-30.
- 35. Koskela, P. and E. Herva, *Immunity against Francisella tularensis in northern Finland*. Scand J Infect Dis, 1982. **14**(3): p. 195-9.
- 36. Aquilini, D., Parola, P. Salvo, E., Paladini, A., Seroepidemiology of the rickettsioses, human granulocytic ehrlichiosis, Lyme disease, Q fever, and tularemia in forestry workers in Tuscany, Italy. Journal of Spirochetal and Tick-borne Diseases, 2000. 7(Fall): p. 35-41.
- 37. Gurcan, S., et al., *Tularemia re-emerging in European part of Turkey after 60 years*. Jpn J Infect Dis, 2006. **59**(6): p. 391-3.
- 38. Levesque, B., et al., *Seroprevalence of zoonoses in a Cree community (Canada)*. Diagn Microbiol Infect Dis, 2007. **59**(3): p. 283-6.
- 39. Jenzora, A., et al., *Seroprevalence study of Francisella tularensis among hunters in Germany*. FEMS Immunol Med Microbiol, 2008. **53**(2): p. 183-9.
- 40. Bazovska, S., Vyrostekova, G.D., Jarekova, J., Bakoss, P., Machacova, E., Spalekova, M., Antibodies against the causative agents of some natural focal infections in blood donor sera from western Slovakia. Epidemiol. Mikrobiol. Imunol., 2010. **59**(4): p. 168-171.
- 41. Campagna, S., et al., Seroprevalence of 10 zoonotic infections in 2 Canadian Cree communities. Diagn Microbiol Infect Dis, 2011. **70**(2): p. 191-9.
- 42. Tatman Otkun, M., et al., *[Epidemiological evaluation of a rapidly-prevented tularemia outbreak in Canakkale province, Turkey]*. Mikrobiyol Bul, 2011. **45**(1): p. 48-57.
- 43. Yazgi, H., et al., [Tularemia seroprevalence in the risky population living in both rural and urban areas of Erzurum]. Mikrobiyol Bul, 2011. 45(1): p. 67-74.
- 44. Sampasa-Kanyinga, H., et al., *Zoonotic infections in native communities of James Bay, Canada.* Vector Borne Zoonotic Dis, 2012. **12**(6): p. 473-81.
- 45. Yesilyurt, M., et al., [*Tularemia: are hunters really a risk group?*]. Mikrobiyol Bul, 2012. **46**(1): p. 153-5.

- 46. Liberati, A., et al., *The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration.* Ann Intern Med, 2009. **151**(4): p. W65-94.
- 47. Esmaeili, S., et al., *Serological survey of tularemia among butchers and slaughterhouse workers in Iran.* Trans R Soc Trop Med Hyg, 2014. **108**(8): p. 516-8.
- 48. Khoshdel, A., Saedi Dezaki, E., Ganji, F., Habibian, R., Imani, R., Taheri, E., Nikkhah, A., *First* seroprevalence survey of children with tularemia infection in Chaharmahal va Bakhtiari Province, *Iran*. Iranian Journal of Pathology, 2014. **9**(1): p. 23-27.
- 49. Bayram, Y., et al., [Seroprevalence of tularemia in risk groups of humans and animals in Van, eastern Turkey]. Mikrobiyol Bul, 2015. **49**(4): p. 532-41.
- 50. Gazi, H., et al., Seroprevalence of West Nile virus, Crimean-Congo hemorrhagic fever virus, Francisella tularensis and Borrelia burgdorferi in rural population of Manisa, western Turkey. J Vector Borne Dis, 2016. **53**(2): p. 112-7.
- 51. Buyuk, F., et al., *The prevalence of tularemia in occupational groups that have contact with animals.* Turk J Med Sci, 2016. **46**(2): p. 451-6.
- 52. Harrist, A., et al., *Francisella tularensis Exposure Among National Park Service Employees During an Epizootic: Devils Tower National Monument, Wyoming, 2015.* Vector Borne Zoonotic Dis, 2019. **19**(5): p. 316-322.
- 53. Takeda, T., et al., *Positive rates of anti-acari-borne disease antibodies of rural inhabitants in Japan.* J Vet Med Sci, 2019. **81**(5): p. 758-763.
- 54. Kugeler, K.J., et al., *Molecular Epidemiology of Francisella tularensis in the United States*. Clin Infect Dis, 2009. **48**(7): p. 863-70.
- 55. Staples, J.E., et al., *Epidemiologic and molecular analysis of human tularemia, United States,* 1964-2004. Emerg Infect Dis, 2006. **12**(7): p. 1113-8.
- 56. Greenberg, L. and J.D. Blake, *An immunological study of the Canadian Indian*. Can Med Assoc J, 1957. **77**(3): p. 211-6.
- 57. Greenberg, L. and J.D. Blake, *An immunological study of the Canadian Eskimo*. Can Med Assoc J, 1958. **78**(1): p. 27-31.
- Messier, V., Levesque, B., Proulx, J.F., Rochette, L., Serhir, B., Couillard, M., Ward, B.J., Libman, M.D., Dewailly, E., Dery, S., Seroprevalence of seven zoonotic infections in Nunavik, Quebec (Canada). Zoonoses Public Health, 2012. 59(107-117): p. 107.
- 59. Fahrer, H., et al., *The prevalence and incidence of clinical and asymptomatic Lyme borreliosis in a population at risk.* J Infect Dis, 1991. **163**(2): p. 305-10.
- 60. Steere, A.C., et al., *Asymptomatic infection with Borrelia burgdorferi*. Clin Infect Dis, 2003. **37**(4): p. 528-32.
- 61. Bogovic, P., et al., *Comparison of laboratory and immune characteristics of the initial and second phase of tick-borne encephalitis.* Emerg Microbes Infect, 2022. **11**(1): p. 1647-1656.
- 62. Chaignat, V., et al., *Performance of seven serological assays for diagnosing tularemia.* BMC Infect Dis, 2014. **14**: p. 234.
- 63. Porsch-Ozcurumez, M., et al., *Comparison of enzyme-linked immunosorbent assay, Western blotting, microagglutination, indirect immunofluorescence assay, and flow cytometry for serological diagnosis of tularemia.* Clin Diagn Lab Immunol, 2004. **11**(6): p. 1008-15.
- 64. Bahuaud, O., C. Le Brun, and A. Lemaignen, *Host Immunity and Francisella tularensis: A Review of Tularenia in Immunocompromised Patients.* Microorganisms, 2021. **9**(12).

- 66. Liles, W.C. and R.J. Burger, *Tularemia from domestic cats*. West J Med, 1993. **158**(6): p. 619-22.
- 67. Gutierrez, M.P., et al., *Serologic evidence of human infection by Francisella tularensis in the population of Castilla y Leon (Spain) prior to 1997.* FEMS Immunol Med Microbiol, 2003. **35**(2): p. 165-9.
- 68. Deutz, A., et al., [Sero-epidemiological studies of zoonotic infections in hunters--comparative analysis with veterinarians, farmers, and abattoir workers]. Wien Klin Wochenschr, 2003. **115** Suppl 3: p. 61-7.
- 69. Schmitt, P., et al., A novel screening ELISA and a confirmatory Western blot useful for diagnosis and epidemiological studies of tularemia. Epidemiol Infect, 2005. **133**(4): p. 759-66.
- 70. Splettstoesser, W.D., et al., *Tularemia in Germany: the tip of the iceberg?* Epidemiol Infect, 2009. 137(5): p. 736-43.
- 71. Bazovska S, G.D., Vyrostekova V, Jarekova J, Bakoss P, Machacova E, Spalekova M, *Antibodies against the causative agents of some natural focal infections in blood donor sera from western Slovakia*. Epidemiologie, Mikrobiologie, Immunologie, 2010. **59**(4): p. 168-171.
- 72. Wölfel, R., Altantuul, D., Mossbrugger, I., Zorig, L., Enkhtuvshin, B., Davaadorj, R. . Seroprevalence of zoonoses in Mongolia: Surveillance and risk factor assessment. in American Society of Tropical Medicine and Hygiene. 2010. Atlanta: American Society of Tropical Medicine and Hygiene.
- 73. Messier, V., et al., Seroprevalence of seven zoonotic infections in Nunavik, Quebec (Canada). Zoonoses Public Health, 2012. **59**(2): p. 107-17.
- 74. Clark, D.V., et al., *Seroprevalence of tularemia in rural Azerbaijan*. Vector Borne Zoonotic Dis, 2012. **12**(7): p. 558-63.
- 75. Tobudic, S., et al., Seroprevalence for Coxiella burnetii, Francisella tularensis, Brucella abortus and Brucella melitensis in Austrian adults: a cross-sectional survey among military personnel and civilians. Ticks Tick Borne Dis, 2014. **5**(3): p. 315-7.
- Zakutna, L., et al., Pilot Cross-Sectional Study of Three Zoonoses (Lyme Disease, Tularaemia, Leptospirosis) among Healthy Blood Donors in Eastern Slovakia. Cent Eur J Public Health, 2015. 23(2): p. 100-6.
- 77. Rigaud, E., et al., Seroprevalence of seven pathogens transmitted by the Ixodes ricinus tick in forestry workers in France. Clin Microbiol Infect, 2016. **22**(8): p. 735 e1-9.
- Özdemir, Z.Ö., Günes, T., Oyardi, Ö., *Risk factors associated with the frequency of antibodies to Francisella tularensis in two areas from Turkey*. Istambul Journal of Pharmacology, 2019. 49(3): p. 137-141.
- 79. Obaidat, M.M., et al., Seroepidemiology, Spatial Distribution, and Risk Factors of Francisella tularensis in Jordan. Am J Trop Med Hyg, 2020. **103**(2): p. 659-664.
- Karatas Yeni, D., Ruh, E., Bostanci, A., Celebi, B., Taylan Ozkan, A., *Investigation of* seropositivity of tularemia, brucellosis and leptospirosis in humans in northern Cyprus. Fresenius Environmental Bulletin, 2022. **31**(02/2022): p. 2153-2160.
- 81. Davarci, I., et al., *Tularemia seroprevalence in humans in the region of the Hittite-Arzawa War* (*Inner Aegean Region*), where the first biological weapon was used 3300 years ago. Turk J Med Sci, 2023. **53**(1): p. 310-315.