REVIEW ARTICLE

Malignant pheochromocytomas and paragangliomas: a diagnostic challenge

Oliver Gimm · Catherine DeMicco · Aurel Perren · Francesco Giammarile · Martin K. Walz · Laurent Brunaud

Received: 18 August 2011 / Accepted: 14 November 2011 / Published online: 29 November 2011 © Springer-Verlag 2011

Abstract

Introduction Malignant pheochromocytomas (PCCs) and paragangliomas (PGLs) are rare disorders arising from the adrenal gland, from the glomera along parasympathetic nerves or from paraganglia along the sympathetic trunk. According to the WHO classification, malignancy of PCCs and PGLs is defined by the presence of metastases at non-chromaffin sites distant from that of the primary tumor and not by local invasion. The overall prognosis of metastasized PCCs/PGLs is poor. Surgery offers currently the only change of cure. Preferably, the discrimination between malignant and benign PCCs/PGLs should be made preoperatively.

Methods This review summarizes our current knowledge on how benign and malignant tumors can be distinguished. Conclusion Due to the rarity of malignant PCCs/PGLs and the obvious difficulties in distinguishing benign and malignant PCCs/PGLs, any patient with a PCC/PGL should be treated in a specialized center where a multidisciplinary setting with specialized teams consisting of radiologists, endocrinologist, oncologists, pathologists and surgeons is available. This would also facilitate future studies to address the existing diagnostic and/or therapeutic obstacles.

Keywords Pheochromocytoma · Paraganglioma · Malignancy · Diagnosis · Therapy

O. Gimm

Department of Surgery, County Council of Östergötland, Linköping, Sweden

O. Gimm

Divison of Surgery, Department of Clinical and Experimental Medicine, Faculty of Health Sciences, Linköping University, Linköping, Sweden

C. DeMicco

Laboratoire d'Anatomie et de Cytologie Pathologique, Faculté de Médecine, Marseille, France

A. Perren

Institute of Pathology, University of Bern, Bern, Switzerland

F. Giammarile

Médecine Nucléaire—Centre Hospitalier Lyon Sud (Hospices Civils de Lyon), Université Claude Bernard Lyon 1—EMR 3738 (Faculté Charles Mérieux Lyon Sud), Lyon, France M. K. Walz

Klinik für Chirurgie und Zentrum für Minimal Invasive Chirurgie, Kliniken Essen-Mitte, Essen, Germany

L. Brunaud Department of Digestive, Hepatobiliary and Endocrine Surgery CHU Nancy-Brabois (hopital adultes), University of Nancy, Nancy, France

O. Gimm (\boxtimes)

Department of Surgery, Division of Endocrine Surgery, Linköping University, 58185 Linköping, Sweden e-mail: oliver.gimm@liu.se



Introduction

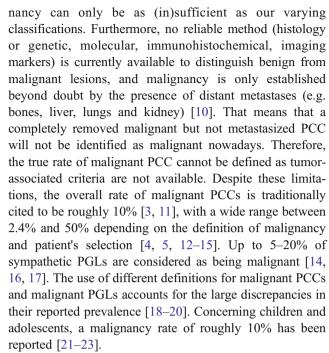
The term pheochromocytoma (PCC) refers to the color of the tumors cells when stained with chromium salts. PCCs and paragangliomas (PGLs) are tumors arising from chromaffin cells that synthesize, store, metabolize and, usually but not always, secrete catecholamines [1, 2]. According to the 2004 WHO classification, adrenal chromaffin tumors are classified as PCCs, whereas extraadrenal tumors (such as the neck, mediastinum, abdomen, pelvis and organ of Zuckerkandl) arising from the glomera along parasympathetic nerves or paraganglia along the sympathetic trunk are termed PGLs [3]. Sympathetic PGLs are typically secretory and have formerly often been termed extra-adrenal PCCs. Tumors arising in the head and neck originate almost exclusively from the parasympathetic nervous system, and approximately 95% of such tumors are non-secretory [1, 2]. Head and neck PGLs (carotid body PGLs, vagal PGLs and jugulotympanic PGLs) are not discussed in this review.

Incidence and prevalence

The epidemiology of PCCs/PGLs is not precisely known. The incidence of these tumors appears to be approximately one in 300,000/year [2]. In western countries, the estimated prevalence is between 1:6,500 and 1:2,500 [4]. The annual age-adjusted prevalence of malignant PCCs in the USA is between 0.3 and 0.7 cases per 1 million, and the incidence of malignant PCCs was 93 cases per 400 million persons in 2002 [1]. Thus, malignant PCCs are exceedingly rare.

The peak age of occurrence is in the fourth to fifth decade of life, with almost equal distribution among male and female patients, except for familial tumors occurring at an earlier age [4, 5]. A tendency towards malignancy has been reported in females by some [6].

Malignancy



Of note, propensity to malignancy is dependent on the genetic background of the tumors. In sporadic tumors, about 6–10% are malignant, and the likelihood of malignancy is even lower in patients with *RET*, *VHL* and *SDHD* gene mutations [24, 25]. However, probably up to 40% of patients with *SDHB* mutation will develop distant metastases [10].

Prognosis

The overall prognosis of metastasized catecholamine-producing tumors is poor. Five-year survival rates vary from 20% to 50% [1], with a significant heterogeneity among patients [11]. The majority of patients will succumb their disease [26], but long-term survival has been reported [27]. Survival of patients with metastatic lesions in liver and lungs tends to be shorter (<5 years) than survival of patients with bone metastases only [28]. Malignant disease is evident preoperatively in nearly 50% of patients. Metastatic disease, however, may only become evident after the primary tumor is surgically removed. Recurrence or metastases usually present within 2–5 years but may be diagnosed even after several decades in some patients [1, 14, 29–31].

Familial syndromes

Roughly 10 years ago, mainly three genes (*NF1*, *VHL* and *RET*) were associated with the occurrence of inherited PCCs, and about 10% of PCCs were considered familial. The corresponding syndromes are neurofibromatosis type 1 (NF1), von Hippel–Lindau type 2 (VHL2) and multiple



endocrine neoplasia type 2 (MEN2), respectively. In addition, various hereditary paraganglioma syndromes (PGL1-4) could be distinguished clinically and by linkage analysis, but no genes were reported. This changed in 2000, when germline mutations in the gene coding for the succinate dehydrogenase complex subunit D (SDHD) were reported for the first time in PGLs [32] and PCCs [33]. Since then, germline mutations have also been reported in other genes coding for subunits of the succinate dehydrogenase complex: SDHC [34], SDHB [35], SDHAF2/SDH5 [36] and SDHA [37]. The associated syndromes are now summarized as pheochromocytoma-paraganglioma syndromes [38]. In addition, germline mutations have recently been reported in KIF1Bbeta [39], TMEM127 [40] and MAX [41], but no syndromes have been reported yet. In addition, Carney triad [42] and Carney-Stratakis dyad are associated with PGLs [43]. While mutations in SDHB, SDHC and SDHD can lead to Carney-Stratakis dyad, no gene has been identified in Carnev triad vet.

In summary, at least ten genes have been found to carry germline mutations in familial tumors, and about 25–30% of PCCs are currently considered being familial [17, 44]. From the clinical point of view, however, the knowledge of PCCs/PGLs being inherited does not make it easier to distinguish between malignant and benign tumors. Malignancy should always be expected in *SDHB*-associated tumors, and a more stringent follow-up is probably indicated in these patients.

Genotype-phenotype correlations in familial syndromes

Von Hippel–Lindau syndrome (*VHL* mutation) is an autosomal dominant disorder. Prevalence is approximately 1:36,000 live births. The frequency of PCCs in individuals with VHL is 10–30% overall. Approximately 50% of PCCs are bilateral. About 5% of VHL-related catecholamine-secreting tumors become malignant, most commonly extra-adrenal sympathetic PGLs [2].

Multiple endocrine neoplasia type 2 is an autosomal dominant syndrome caused by mutation of the *RET* protooncogene. Prevalence is estimated at 1:30,000. Approximately 50% of individuals with MEN2A and MEN2B develop PCCs. PCCs are bilateral in 50–80% of cases but are almost always benign [2].

Neurofibromatosis type 1 is an autosomal dominant disorder caused by mutation of *NF1*. Prevalence is estimated at 1:3,000 to 1:4,000. Although PCCs are rare in NF1, their frequency is as high as 20–50% in individuals with NF1 and hypertension. Most (84%) PCCs are unilateral. Extra-adrenal sympathetic PGLs can occur. These tumors may be malignant in about 10% [2].

Hereditary pheochromocytoma-paraganglioma syndromes are inherited in an autosomal dominant manner.

SDHD (PGL1), SDHC (PGL3) and SDHB (PGL4) are the three nuclear genes responsible for the hereditary pheochromocytoma-paraganglioma syndromes. A fourth nuclear gene, SDHAF2 (PGL2), also known as SDH5, has been recently reported [2]. Mutations in SDHD (PGL1) demonstrate parent-of-origin effects and generally cause disease only when the mutation is inherited from the father. However, an individual who inherits an SDHD mutation from his/her mother has a low but not negligible risk of developing disease. Initial data suggest that mutations in SDHAF2 (PGL2) exhibit parent-of-origin effects similar to those of mutations in SDHD [2]. The pheochromocytomaparaganglioma syndrome should be considered in all individuals with PCCs and/or PGLs, particularly those with the following findings: multiple tumors including bilateral tumors, multifocal with multiple synchronous or metachronous tumors, recurrent tumors, early onset (i.e. age <40 years) tumors and family history of PCCs or PGLs [2]. Most recently, a mutation in SDHA has been found in a patient with PGL [37]. Of interest is the clinical behavior of malignant PCCs and PGLs that appears to be most aggressive in patients with germline SDHB mutation as opposed to patients having sporadic malignant PCCs and PGLs [45].

Although heterogeneous, the following correlations between the gene involved and tumor characteristics can be used to guide management: Germline mutations in SDHB are strongly associated with extra-adrenal sympathetic PGLs [13]. Chromaffin tumors in persons with germline SDHB mutations are sixfold more likely to be extra-adrenal than chromaffin tumors in general. A possible relationship between SDHB exon 1 deletions and abdominal extra-adrenal PGLs has recently been proposed [2]. PGLs in persons with a germline SDHB mutation are more likely to become malignant than sporadic PGLs or those that develop in persons with germline SDHD and SDHC mutations. SDHB mutations may also predict a shorter survival in persons with malignant PCCs and PGLs. Up to 50% of persons with malignant extra-adrenal PGLs have a germline SDHB mutation. Because extra-adrenal sympathetic PGLs have long been known to have a greater predisposition to malignancy than PCCs and head and neck PGLs, it is not clear whether this effect is the result of location, mutation status or both [2, 12]. Although less common than malignant extra-adrenal sympathetic PGLs, malignant PCCs do occur and may be more common in individuals with a germline SDHB mutation than in those with a germline SDHD or SDHC mutation or with sporadic PCCs [2]. However, persons with a germline SDHD mutation can develop malignant disease at any paraganglion site [2]. Mutations in SDHD and SDHC are more frequently associated with parasympathetic head and neck PGLs than



other tumor types. However, thoracic and abdominal localizations remain possible [2].

Carney triad is an extremely rare disorder that primarily affects young women. As initially described, the classic Carney triad included extra-adrenal sympathetic PGLs, gastric stromal sarcoma and pulmonary chondroma. PCCs were later shown to be associated with the syndrome (with adrenal cortical adenoma and esophageal leiomyoma). Carney triad may be familial, but a causative gene has yet to be identified [2, 42].

Carney–Stratakis dyad, also termed Carney–Stratakis syndrome, is the association of PGLs and GISTs and is distinct from the Carney triad [43]. PGLs and GISTs in these families appear to be inherited in an autosomal dominant manner with incomplete penetrance. PGLs occur in the head and neck, thorax and abdomen. *SDHx* mutations have been reported in individuals from six unrelated families with the Carney–Stratakis dyad, and the significance of these findings is not yet clear [2].

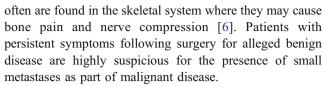
More recently, further genes have been shown to be associated with hereditary PCCs and PGLs: $KIF1B\beta$ [39, 46], TMEM127 [40, 47, 48] and MAX [41]. Concerning these genes, no specific syndrome has been reported yet. While the risk of developing bilateral PCCs appears to be high, the risk of developing PGLs is considered to be low. If the currently available data are correct, patients with MAX germline mutations have a risk of about 25% of developing malignant PCCs [41].

Preoperative diagnosis

PCCs and extra-adrenal sympathetic PGLs mainly come to medical attention in four clinical settings: signs and symptoms associated with catecholamine hypersecretion, incidentally discovered mass on CT/MRT performed for other reasons, signs and symptoms related to mass effects from the neoplasm and screening at-risk relatives [2]. Preferably, the discrimination between malignant and benign PCCs/PGLs should be made preoperatively.

Clinical diagnosis

Most patients with PCCs have hypertension, often associated with palpitations, headache and diaphoresis ("typical signs"). Functioning malignant chromaffin cell tumors have a clinical presentation similar to benign tumors, but patients may present with variable symptoms and signs, such as dyspnea, nausea, weakness, weight loss, visual disturbance, arrhythmias and mental problems [11, 49]. A lack of the "typical signs" may also raise the suspicion that one is dealing with a malignant case [50, 51]. Symptoms suggestive for malignancy may arise from metastases that



In the case of large cystic PCCs, many patients present without hypertension [52]. Patients with malignant PCCs may even lack clinical signs until the late stage [53].

Biochemical diagnosis

The diagnosis of PCCs and sympathetic PGLs is based on biochemical testing and imaging studies.

Unspecific biomarkers

Neuroendocrine cells as neurons contain vesicles that produce and secrete chromogranins and secretogranins. Both belong to a group of acidic, soluble proteins. A markedly increased preoperative chromogranin A plasma level in patients with malignant PCCs ($n=14,2,932\pm960$ ng/mL) in comparison to patients with benign pheochromcytomas (n=13, 188 ± 40.5 ng/mL) has been reported [54]. A chromogranin A level higher than 500–600 ng/mL was highly suggestive for a malignant PCC. In accordance, chromogranin A has been used to monitor patients during chemotherapy of malignant PCCs [55]. Some investigators have reported a high serum concentration of neuron-specific enolase (NSE) [56].

Hormones

Catecholamines hypersecreted by PCCs and PGLs can be any of the following: epinephrine (adrenaline), norepinephrine (noradrenaline) and dopamine. Concerning the biochemical diagnosis of PCCs, it is recommended to measure plasma or 24-h urinary excretion of fractionated metanephrines [1, 2, 4, 5]. The latter is preferred as it is more sensitive than the measurement of catecholamine concentrations [57, 58].

Concerning malignant PCCs, the same recommendations exist. Rarely, non-functioning benign and malignant PCCs are reported [59–61]. Malignant PCCs, however, may lack various enzymes. One of them, PNMT, converts norepinephrine (noradrenaline) to epinephrine (adrenaline). Lack of PNMT thus leads to dominating production of norepinephrine in malignant PCCs, and high levels of norepinephrine have even been reported to be associated with a shorter metastases-free interval [62]. False positive results may be reduced by follow-up testing for plasma chromogranin A and/or urine fractionated metanephrine levels when plasma fractionated metanephrine concentrations are less than fourfold above the reference range [11, 63].



It has also been reported that high dopamine levels, representing more premature catecholamine secretion due to decreased expression of dopamine-β-hydroxylase, are more common in malignant PCCs [64]. PCCs expressing solely dopamine but not epinephrine/norepinephrine appear to have a very high likelihood of being malignant [65], and higher levels of dopamine are associated with a shorter metastases-free interval [65]. Consequently, patients with high preoperative 24-h urinary dopamine levels (>5,000–6,000 nmol/24 h) have an increased likelihood of having malignant PCC [66]. Of note, a low ratio of plasma epinephrine to total catecholamines was reported to predict recurrence [67]. The secretion of norepinephrine with little or no epinephrine suggests an extra-adrenal PGL or a PCC associated with von Hippel–Lindau syndrome [28].

Patients with persistently elevated catecholamine levels following surgery for alleged benign disease are highly suspicious for the presence of small metastases as part of malignant disease.

Imaging

PCCs can be detected using a variety of imaging techniques, but due to their inconsistent fashion, they may mimic various tumors, both benign and malignant, including metastases [68]. Distant metastases of PCCs are most often found in bone (50%), liver (50%) and lung (30%) [49].

Diagnostic techniques include conventional radiological imaging with computed tomography (CT), magnetic resonance imaging (MRI), ultrasonography (US), contrastenhanced US (CEUS), endoscopic US (EUS) and intraoperative US (IOUS); selective angiography with hormonal sampling; and nuclear medicine imaging by 111Inoctreotide (OctreoScan), 123I-metaiodobenzylguanidine (MIBG), 99mTc-EDDA/HYNIC-Tyr3-octreotide scintigraphy or, more recently, somatostatin receptor PET with 68Ga-octreotide, [18F]DOPA, [18F]Dopamine and [11C]5hydroxytryptophan. No technique is the gold standard, and specific sequences of exams might be needed for each tumor type. A combination of two or more imaging techniques is often required for diagnosis and staging. Usually, radiological techniques (such as ultrasound, CT or MRI) are useful in the localization of the primary tumor, particularly if non-functioning, while nuclear medicine aids in the evaluation of the extent of disease, staging and therapy decision making [69–71].

Computed tomography

CT has a very high sensitivity in detecting PCCs but a relatively low specificity [57, 72–74]. PCCs may mimic both adenomas and malignant masses on both CT densitometry and washout [68].

CT densitometry has been shown as being helpful in distinguishing between benign and malignant adrenal lesions. A density of approximately 40-50 HU after injection of contrast medium is suggestive for a PCC. Inhomogeneous appearance is not uncommon and may be due to hemorrhage or necrosis. A homogeneous adrenal mass with a density of less than 10 Hounsfield units (HU) on an unenhanced CT is almost certainly a benign adrenal lesion [57]. Therefore, lesions with an unenhanced density greater than 10 HU requires further evaluation. The attenuation of PCCs on an unenhanced CT scan is significantly higher than the attenuation of adrenal cortical adenomas (44±11 Hounsfield units versus 8±18 HU). However, adrenocortical carcinomas (39±14 HU) are difficult to differentiate from PCCs based on CT findings alone without biochemical testing [1]. On contrast enhancement, the tumors are usually irregular with peripheral enhancement.

It was found that adrenocortical adenomas enhance rapidly after administration of contrast medium and also show rapid loss of contrast medium, a phenomenon called contrast washout. If the "washout" after 15 min is higher than 60%, the sensitivity is 86–88%, and the specificity is 92–96% for the lesion being an adenoma [75]. Others have proposed a "washout" greater than 50% after 10 min [76]. The washout of malignant lesions is used to be less than 40% [68].

If malignancy is suspected, an initial abdominal CT scan from the neck to the pelvis floor should be performed, and contiguous thin sections with 1.5- to 3-mm cuts must be obtained. CT scan reliably localizes most PCCs >1 cm, with accuracy approaching 100% [1, 11]. Because 90% of all sympathetic tumors are located within the adrenal gland and 98% "abdominal", a high-quality CT scan will likely identify most tumors and image the normal contralateral gland [1]. PCCs have a wide range of imaging characteristics on CT, from a well-circumscribed homogeneous mass to a heterogeneous, cystic or hemorrhagic mass. Despite these features, diagnosis of malignancy is unreliable unless distant metastases are apparent [1].

Magnetic resonance imaging

MRI sensitivity is similar to CT but is less available [72–74, 77]. It is, however, the preferred initial imaging procedure for PCCs in children, pregnant women and patients allergic to contrast medium [57]. Since PCCs are hypervascular, they characteristically have intermediate to high signal intensity on T2-weighted sequences ("light bulb" sign) [77]. However, this holds only true for about 70% of PCCs, and high signal intensities can also be found in other conditions such as hemorrhage, hematoma, etc. MRI is highly specific for PCCs if the T2-weighted image brightness is three times greater than the liver [11]. The



hyperintensity on T2-weighted images makes MRI more accurate and sensitive than CT scanning for recurrent, metastatic and extra-adrenal PGLs [1].

Chemical shift MRI relies on the different resonance frequencies of protons in fat and water molecules. Electrons surrounding the proton shield it from the applied external field. The effective magnetic field experienced by a shielded proton is less than the effective magnetic field experienced by an unshielded proton. Fat protons are more shielded than water protons and thus resonate at a lower frequency. It is the difference in resonance that is used in chemical shift imaging. Two T1-weighted acquisitions are performed: out-of-phase and in-phase acquisitions. In adrenal tumors that do not contain fat (e.g. metastases), there is no significant signal loss. Applying these techniques, the sensitivity and specificity to differentiate adenomas from metastases range from 81–100% and 94–100%, respectively [78].

Diffusion-weighted MRI (DWI) with high *b*-value has been successfully used to detect metastases from malignant PCCs and PGLs and may be superior to 123I-MIBG and even FDG-PET [79]. In particular, this technique may be advantageous in detecting lymph node and liver metastases but less sensitive in mediastinal and lung metastases.

Endosonography

Endosonography has been used successfully in localizing PCCs [80]. Hyperechoic echogeneity and echostructure was only seen in benign tumors, but no specific appearance could be found for malignant PCCs. The experience, however, is limited.

MIBG scintigraphy

Metaiodobenzylguanidine or iobenguane is the combination of the benzyl group of bretylium and the guanidine group of guanethidine. This aralkylguanidine norepinephrine analog is an adrenergic neuron blocker and the so-called false neurotransmitter [81].

Since MIBG structurally resembles noradrenaline, it enters in neuroendocrine cells by either the neuronal uptake-1 mechanism or by passive diffusion. The transfer of MIBG from the intracellular cytoplasm into catecholamine storage vesicles (neurosecretory granules or vesicles) is mediated by an ATPase-dependent proton pump. Unlike neuradrenaline, MIBG is not metabolized and is excreted unchanged. Since MIBG storage in neurosecretory granules enables a specific concentration in neuroendocrine cells in contrast to cells of other tissues, uptake of MIBG by the various organ systems reflects either rich adrenergic innervation or catecholamine excretion (or both) [82, 83].

MIBG radiolabeled with 131- or 123-iodine was developed in the early 1980s to visualize tumors of neuroendocrine origin, particularly those of the neuroectodermal (sympatho-adrenal) system (PCCs, PGLs and neuroblastomas), although other neuroendocrine neoplasms (e.g. carcinoids, medullary thyroid carcinoma, etc.) are also visualized (Table 1) [82–85].

Theoretical considerations and clinical experience indicate that the 123I-labeled agent is to be considered as the radiopharmaceutical of choice, at least in children, as it has a more favorable dosimetry and provides better image quality. Gamma emission energy of 159 KeV for 123I is more suitable for imaging (especially in tomographic

Table 1 Clinical indications to perform MIBG

Oncological indications

- 1. Detection, localization, staging and follow-up of neuroendocrine neoplasms and their metastases, in particular:
- -Pheochromocytomas
- -Neuroblastomas
- -Ganglioneuroblastomas
- -Ganglioneuromas
- -Paragangliomas
- -Carcinoid tumors
- -Medullary thyroid carcinomas
- -Merkel cell tumors
- 2. Study of the tumor uptake in order to decide and plan possible treatment with high activities of radiolabeled MIBG. In this case, the dosimetric evaluation should be individual and not based on the ICRP tables, which have only an indicative value limited to diagnostic procedures
- 3. Evaluation of tumor response to therapy by measuring the intensity of MIBG uptake and the number of focal MIBG uptake sites
- 4. Confirmation of suspected neoplasms derived from neuroendocrine tissue

Other (non-oncological) indications

1. Functional studies of the adrenal medulla (hyperplasia), sympathetic innervation of the myocardium, salivary glands and lungs



modality) than 360 KeV for 131I, and the difference in terms of radiation burden permits us to inject higher activities of 123I-MIBG. Furthermore, results with 123I-MIBG are more rapidly available. Nonetheless, 131I-MIBG is still widely employed because of its lower costs, ready availability and longer half-life and the possibility of obtaining delayed scans. Furthermore, 131I-MIBG may be preferred when estimation of the tumor's retention is required for MIBG therapy planning [86, 87]. Indeed, the high tumor affinity of MIBG for the detection of primary and secondary tumor sites have led to the use of the compound labeled with γ/β emitter 131iodine (131I-MIBG) as radiotherapeutic agents in neuroectodermally derived tumors [88].

The sensitivity and specificity of MIBG for the diagnosis of the primary tumor have been estimated to be 73% and 94%, respectively. Since there is no physiological uptake of MIBG in bone and bone marrow, the sensitivity and specificity of MIBG for detecting osteomedullary metastases are even higher (90% and 100%, respectively) [86]. A recent meta-analysis based upon the literature results reports a sensitivity of 97% [95% confidence interval (CI), 95% to 99%] for detection of neuroblastoma, while data were insufficient to estimate specificity. For PCCs, sensitivity and specificity were 94% (95% CI, 91-97%) and 92% (95% CI, 87–98%), respectively [89].

In malignant tumors, however, the expression of noradrenaline transporters decreases, and thus, the sensitivity is lower [74, 90], and it is not uncommon that both MIBGpositive and MIBG-negative lesions coexist. Also, VHLassociated PCCs are more likely to be missed, most likely due the lower expression of the norepinephrine transporter [91]. For these reasons, one unequivocal MIBG-positive lesion at a distant site is sufficient to define a metastatic disease. A single equivocal lesion on MIBG requires confirmation by another imaging modality. In particular, extra-adrenal and small adrenal PCCs are more likely to result in false negatives on MIBG. Furthermore, adrenal PCCs containing minimal solid tissue due to extensive necrosis may predict a negative MIBG result [92–95].

Fig. 1 CT (left) and 123I-MIBG SPECT-CT (right) in a right adrenal pheochromocytoma

SPECT) allows for better depiction of small focal uptake that is difficult to visualize on planar MIBG, especially in areas close to intense physiological uptake such as the liver and the bladder. The particular strengths of MIBG SPECT/ CT are detection of local recurrence, small extra-adrenal PCCs, multifocal tumors or the presence of metastatic disease (Fig. 1) [96–101]. It is important to notice that 123I-MIBG has even been used, combined with intraoperative gamma probe, to

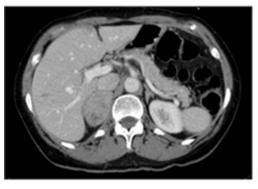
Single photon emission computed tomography (MIBG-

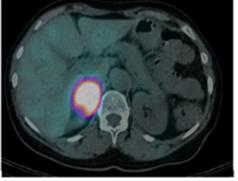
identify malignant foci [102].

Currently, radio-iodinated MIBG is prepared by an exchange radio-iodination method and, thus, is of low specific activity. For possible better targeting and to ward off pharmacological effects, especially in therapeutic purposes, its preparation at a no-carrier-added level both by solution-phase and solid-phase syntheses has been developed [103].

Somatostatin analogs

For over two decades, somatostatin receptor scintigraphy (SRS) with indium-111 DTPA D Phe octreotide (Octreoscan®) has been widely used as a diagnostic agent, particularly to image neuroendocrine neoplasms [104]. Octreotide scintigraphy can be used in addition to MIBG scintigraphy as some MIBG-negative tumors are positive with octreotide scintigraphy [58]. Primary or metastatic tumors expressing somatostatin receptors can be visualized with the help of radiolabeled somatostatin analogs. Expression of somatostatin receptors has been found in roughly 75% of PCCs [105]. While 111In-pentetreotide scintigraphy generally has a lower detection rate for malignant PCCs than 123I-MIBG scintigraphy [105–107], they can have a complementary role for the staging of malignant PCCs [105, 106, 108]. However, dedifferentiated tumors tend to have lower expression of somatostatin receptors [74, 90]. 111In-pentetreotide imaging appears to have a high sensitivity in detecting PGLs of the head and neck [109].







The expression rate of somatostatin receptor in these tumors has been reported to be higher than 90% [105].

PET images

Because of growing clinical applications of positron emission tomography (PET), several PET radiopharmaceuticals have been developed in the exploration of neuroendocrine neoplasms such as [11C]hydroxyephedrine ([11C] HED), [11C] 5-hydroxytryptophan ([11C]HTP), [18F]fluoro-2-deoxy-D-glucose ([18F]FDG), [11C/18F]fluoro-dihydroxyphenylalanine ([11C/18F] FDOPA), [18F]fluoro-dopamine ([18F]FDA) and [68Ga]somatostatinanalogs ([68Ga]DOTA-TOC and [68Ga]DOTA-NOC) [87, 110–112].

11C-HED was the first positron-emitting probe of the sympathoedrenal system used in humans, and first results in the early 1990s were very promising. Due to the short half-time (about 20 min) of 11C, on-site production is required. Since the synthesis of 11C-HED is very complex, the need for other positron-emitting compounds was evident. 18F, having a half-time of about 110min, proved to be more promising.

In general, malignant and inflammatory tissues show increased uptake 18F-FDG due to increased glucose utilization [113]. This phenomenon has been investigated, improving both imaging and therapy of malignancies [114].

Concerning PCCs, the advantages of 18F-FDG in tumors that do not accumulate MIBG have been shown early [115]. It has been repeatedly shown that 18F-FDG PET imaging may be useful in those malignant PCCs that fail to accumulate either 123I-MIBG [116] or 131I-MIBG [117, 118]. One advantage of 18F-FDG in contrast to other tracers is that its uptake appears to be unrelated to the secretory status of the tumor. In patients with *SDHB*-associated PGLs/PCCs, 18F-FDG has been shown to be superior to 131I-MIBG, 123I-MIBG, 111In-pentetreotide and even 18F-FDA in detecting metastatic lesions [119].

Finally, 18F-FDG may have a prognostic role [79, 118]. Sodium 18F has been previously used for bone imaging and can be used as a PET skeletal tracer. A pilot study analyzing administration of combined 18FNa/18F-FDG showed promising results in metastatic PGLs [120]. Since only a single PET scan is needed, health care costs could be reduced.

18F-FDA, developed as a radiopharmaceutical agent for the detection of noradrenergic pathways, is difficult to produce and has limited availability [121]. Also, normal adrenal glands may give false-positive results. In this regard, the recent recommendation to the use of standardized uptake values (SUV) may be very helpful. If the SUV is <7.3, the presence of a PCC is very unlikely, while a SUV >10.1 confirms its presence [122–124]. In addition,

18F-FDA has been reported to be superior to MIBG in the localization of metastatic PCC [125]. In the case of bone metastases, 18F-FDA-PET was superior, followed by bone-scintigraphy, CT/MRI, 18F-FDG and 123/131I-MIBG [126].

18F-DOPA, which is incorporated into the cell via the amino acid transporter system, seems to have the advantage that there is no significant uptake in normal adrenal glands [127]. Its overall sensitivity and specificity concerning PCCs are high [128–130]. However, its sensitivity in metastatic PGLs is *low* [131]. This seems to be true also for malignant PCCs, where 18F-FDG appears to be more sensitive [132].

While PET with the 11β -hydroxylase tracer 11C-metomidate has been used successfully to discriminate between adrenocortical and non-adrenocortical lesions, it could not differentiate between benign and malignant diseases [133].

Recently, the introduction of 68Ga-labeled somatostatin analogues has enabled somatostatin imaging with PET agents [134]. In a small pilot-study analyzing patients with metastatic PCCs, 68Ga-DOTATATE proved to be superior to 123I-MIBG [135].

Development of MIBG analogues labeled with positronemitting radionuclides such as 124I, 18F and 76Br has been also reported [103].

Biopsy

Biopsy of PCCs and/or PGLs is contraindicated because this invasive procedure has the risk of precipitating a hypertensive crisis, hemorrhage and tumor cell seeding [2, 136].

Summary

Most patients with malignant PCCs have the "typical signs" suggestive for PCCs: hypertension, palpitations, headache and diaphoresis. A lack of these "typical signs" or the presence of variable symptoms and signs, such as dyspnea, nausea, weakness, weight loss, visual disturbance, arrhythmias and mental problems, may raise the suspicion that one is dealing with a malignant tumor. Biochemically, high preoperative 24-h urinary dopamine levels (>5,000–6,000 nmol/24 h) and chromogranin A levels (>500–600 ng/mL) are suggestive but not proving that one might be dealing with a malignant tumor. Overall, it is considered that diagnosing patients with malignant tumors is more difficult when solely based on preoperative clinical and/or biochemical markers.

If PCCs are confined to the adrenal gland, both computed tomography (CT) and magnetic resonance imaging (MRI) have a very high sensitivity to identify the



lesion. CT of the abdomen should be performed first, followed by head and neck CT if the abdominal CT is negative [73]. In children and during pregnancy, MRI is recommended instead.

To confirm the presence of a PCC, some recommend a functional imaging technique in every patient [73]. In any case, functional imaging techniques are required when malignancy is suspected on other grounds. The most preferred method is 123I-MIBG scintigraphy, especially if imaging is correlated with SPECT-CT technique. However, MIBG may have lower performances in subsets of PCC patients, like some familial paraganglioma syndromes, malignant disease and extra-adrenal PGLs. In the case of negative MIBG scans, PET imaging with specific ligands is recommended [137]. While 18F-FDA is highly recommended [73, 138], 18F-FDG and other imaging techniques such as somatostatin receptor scintigraphy are more readily available and more commonly used. The sensitivity and specificity of various imaging techniques for PCCs and PGLs are shown in Tables 2 and 3. The estimated functional imaging performances for sporadic and various familial syndromes is shown in Table 4.

Therapy

Treatments for malignant PCCs include surgical resection, pharmacologic control of hormone-mediated symptoms, targeted methods such as external irradiation and systemic antineoplastic therapy [1, 5, 10, 11, 28]. The management of tumors in individuals with hereditary syndromes is similar to the management of sporadic tumors [58].

Surgery

Following preoperative treatment to block the effects of catecholamine excess (e.g. with α -adrenoceptor antagonists or channel blockers), surgery is the treatment of choice in almost all patients with PCCs and PGLs [1, 10, 12]. Basically, PCCs and PGLs can be divided into potentially malignant, suspicious malignant and overt malignant tumors. Overt malignant tumors are those with proven distant metastases [1, 11]. Suspicious malignant tumors are those with locoregional infiltration without distant metastases [1, 5, 11]. The remaining tumors are potentially malignant [1, 10, 11]. Surgical strategies differ between potentially malignant, suspicious malignant and overt malignant tumors [11].

For potentially malignant PCCs and PGLs, the surgical treatment follows the principle of benign diseases [1]. Local excision of the lesion usually performed by a minimally invasive approach is the accepted standard [139]. Partial adrenalectomy with complete removal of the PCCs may be indicated in bilateral and/or inherited diseases with low likelihood of malignancy, e.g. MEN2- or VHL-associated PCCs [140, 141]. In general, tumor capsular effraction should be avoided to prevent pheochromocytomatosis [142]. Regardless of tumor size, laparoscopic adrenalectomy for PCCs should be converted to open adrenalectomy for difficult dissection, invasion, adhesions or surgeon inexperience [139]. Data after minimally invasive removal of incidentally discovered malignant PCC are rare. Rabii and colleagues reported a case of a patient with a malignant PCC who underwent laparoscopic adrenalectomy followed by transperitoneal laparoscopic metastatic paraaortic lymph node resection 6 years later [143]. Walz and colleagues

Table 2 Pheochromocytomas (PCCs): sensitivity (%) and specificity (%) of various imaging techniques for detecting primary tumors and metastases

	Primary PCC				Metastases		References
	All PCC		Mal PCC				
	Sens	Spec	Sens	Spec	Sens	Spec	
СТ	85–95	29–93			90		[57, 72–74]
MRI	65–95	50-93					[72–74, 77]
123I-MIBG	83-100	95-100	91		50-80		[72, 74, 87, 106, 135, 203, 204]
131I-MIBG	58-90	90-100	67–77				[78, 87, 204, 205]
111In-octreotide	75–90				44-88		[74, 78, 106]
18F-FDG	58		77				[204]
18F-DOPA	85	100					[128]
68Ga-DOTATATE					83		[135]

All PCC benign and malignant pheochromocytomas, Mal PCC malignant pheochromocytomas, Sens sensitivity, Spec specificity



Table 3 Paragangliomas (PGLs): sensitivity (%) and specificity (%) of various imaging techniques for detecting primary tumors and metastases

	Primary PGL				Metastases		References	
	All PGL		Mal PGL					
	Sens	Spec	Sens	Spec	Sens S	Spec		
CT	90–100	50			78–96		[73, 119, 126, 206]	
MRI	100	50			78–95		[119, 126, 206]	
111In-octreotide					<80		[105]	
111In-pentetreotide					59-81		[119]	
123-MIBG	98				65-80		[119, 126, 203]	
131I-MIBG	50-82	100			30-71		[119, 126, 205, 206]	
18F-FDG					76-100		[119, 126]	
18F-FDA					70–90		[119, 126]	

All PGL benign and malignant paragangliomas, Mal PGL malignant paragangliomas, Sens sensitivity, Spec specificity

removed four malignant PCCs by the posterior retroperitoneoscopic approach. One patient had skin metastases prior to adrenalectomy, two developed liver metastases within 3 years, and one patient had an interaortocaval lymph node metastasis 3 years postoperatively. Local recurrence was not observed in any of these cases [14].

Suspicious malignant cases should be treated by complete excision [12, 144]. This may include the resection of adjacent tissue and organs (liver, kidney, vena cava, spleen and pancreas) as the only curative option [1, 5, 11, 12, 58]. Unfortunately, locoregional invasion cannot usually be defined by preoperative imaging studies. Therefore, it has been recommended that potential invasive tumors should initially be explored by laparoscopy or retroperitoneoscopy followed by conversion to open surgery in case of critical adhesions [14, 144].

In patients with overt malignant PCCs (with distant metastases), surgery is palliative and tries to reduce the hormonal effects on the cardiovascular system by reduction of tumor tissue [1, 4]. Thereby, subsequent radiotherapy or chemotherapy may be facilitated [10].

Surgical treatment of PGLs follows the same principles as that for PCCs. Complete removal has to be achieved. This usually requires meticulous dissection from major vascular structures. Minimally invasive approaches have been demonstrated to be feasible and safe but must be

categorized as advanced and challenging procedures [14, 145]. Therefore, open accesses are still the standard.

Non-surgical treatment modalities

MIBG

To date, 131I-labeled MIBG therapy is the single most valuable adjunct to surgical treatment of malignant PCC [10, 49]. As early as 1984, 131I-MIBG was administered in large doses for the treatment of malignant PCCs [11]. In a retrospective review of 33 patients with metastatic PCCs (n=22) or PGLs (n=11) treated over a 10-year period with 131I-MIBG, the median survival of patients was 4.7 years [146]. Approximately 60% of metastatic sites are 131I-MIBG avid, and in a recent meta-analysis including 166 patients, the objective response rate was 30%, and disease stabilization was achieved in an additional 43% of patients [49]. Recently, it has been reported that ultratrace iobenguane I-131 (Ultratrace 131I-MIBG) might provide improved efficacy and tolerability over carrier-added 131I-MIBG in the treatment of malignant PCCs [103, 147].

Finally, for potential use in the treatment of micrometastatic diseases, synthesis of an analogue labeled with the alpha emitter (211)At was devised [103].

Table 4 Estimated functional imaging performances (modified from [87])

	123I/131I-MIBG	Specific PET (18F-FDA/18F-DOPA)	Non-specific PET (18F-FDG)
Sporadic	+	++	++
MEN2	+	+	Insufficient data
VHL	(+)	++	Insufficient data
SDHB	+	++	++/+++
SDHC	Insufficient data	Insufficient data	Insufficient data
SDHD	+	Insufficient data	Insufficient data



Chemotherapy

Experience with cytotoxic chemotherapy is limited, and the best results were seen with a combination of cyclophosphamide, vincristine and dacarbazine (CVD) [148]. CVD showed varied success in several series, with survival benefit ranging from 1 to 4 years [11, 148]. In a 22-year follow-up study, there was no difference in overall survival between patients whose tumors objectively shrank and those with stable or progressive disease [148]. The authors conclude that CVD therapy is not indicated in every patient with metastatic PCCs/PGLs, but should be considered in the management of patients with symptoms and where tumor shrinkage might be beneficial [10, 148].

Up to now, no single-phase II or III trial investigating "new drugs" in malignant catecholamine-producing tumors has been published. More specific targeted therapy with imatinib mesylate has been tried with malignant PCCs, but no significant benefit was found [10]. The same holds true for everolimus, an inhibitor of mTOR that was not effective in four patients [10, 49]. The most promising but still immature results are reported with the multiple TKI sunitinib [10].

External radiation

External beam irradiation of bone metastases and other targeted methods (such as radiofrequency ablation of hepatic and other lesions, cryoablation, chemoembolization, cyber/gamma knife and arterial embolization) can help to alleviate local tumor complications and are treatment alternatives [10].

Recurrence

Even when seemingly complete, operative excision of malignant PCCs/PGLs does not preclude the later development of local-regional recurrence [149]. Several hypotheses for recurrent disease exist, including failure to identify and completely resect the primary tumor (especially if found in ectopic locations), tumor seeding (during needle biopsy or surgery) or the presence of metastases (nodes or distant) [11]. A recent study showed that recurrence can occur in nearly 25% of patients undergoing resection for an abdominal PGL, when followed up beyond 10 years after an apparently complete initial excision, which is a figure superior to those previously reported for PGLs in general [149]. In inherited syndromes, it is sometimes difficult to differentiate between local-regional recurrence and new ectopic locations. If the recurrence is determined resectable, reoperation remains the only option for cure but often represents a major technical challenge [11, 149]. Surgeons

should also consider metastasectomy in organs where feasible, although it has not yet been determined if this will increase survival [150]. If recurrence presents as widespread metastases, then surgical excision is likely not an option. In those cases, systemic treatments as noted earlier will be the only option [11].

Summary

Surgery offers the only potential cure for malignant PCCs and PGLs but is rarely curative. The number of other supportive treatment modalities is very limited. Current research puts major efforts into identifying genes that contribute to the development of malignant PCCs and PGLs. Those genes may be targets for future therapeutic approaches.

Pathological aspects

The pathological diagnosis of malignancy in PCCs and PGLs remains a controversial issue [20, 151]: Nuclear hyperchromasia and pleiomorphism are common in these tumors but unrelated to their evolution. At the opposite, distant metastasis may occur up to 20 years after a histological diagnosis of benign PCC [7, 18, 29, 152-156]. Thus, there is a general feeling that histology is unable to predict the evolution of PCC [157]. The rarity, fluctuating definition and long evolution of malignant PCCs have been important limiting factors to the determination of histopathological criteria of malignancy: Locally invasive and metastasizing tumors were often analyzed indistinctly [8, 12, 158]. The mean follow-up period of benign PCC in studies exceeded scarcely 8 years, whereas the mean reported delay before the occurrence of metastasis was 8 to 12 years [8, 9, 154, 155, 159-164].

After a comprehensive analysis of the literature devoted to the pathology of malignant PCCs/PGLs (MEDLINE 1980-2010), we could find only four publications evaluating the gross and histopathological features of more than ten malignant cases with comparison to benign PCC followed at least 10 years [165-168]. Three other papers analyzed large numbers of malignant PCCs with lower mean follow-up periods for the benign control groups [158, 159, 169]. A series of differences were reported, with discrepancies about their significance, but there is a general agreement that no single parameter is pertinent enough to separate benign from malignant PCC. Then, scoring systems combining several variables were elaborated thereafter. In the main course, immunohistochemistry was used for assessment of malignant potential with disputable results.



Gross features

The mean size of malignant PCCs (8–9 cm) is greater than that of benign cases (4–5 cm), but authors agree that the size is not an independently helpful criterion to the diagnosis of malignancy [139, 166, 170]. Cystic degeneration and hemorrhage are not significant either.

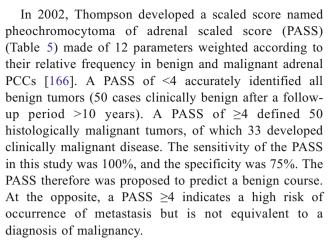
Histological parameters

The histological parameters that have been analyzed in PCC fall under five categories: (1) architecture and cell-density: "Zellballen", large nest, diffuse architecture, high cellular density, cellular monotony and spindle cells (more than 10%); (2) invasiveness: in blood, lymphatic vessels, within capsule and into peri-adrenal adipose tissue; (3) cellular morphology: pleïomorphism, nuclear hyperchromasia and cytoplasmic hyaline globule; (4) cellular proliferation: mitotic count $\geq 3/10$ HPF) and atypical form; and (5) degenerative changes: cystic changes and necrosis (focal or confluent).

Criteria concerning individual cellular morphology were poorly correlated to the malignant phenotype in most studies, whereas those concerning architecture were found significant almost constantly [8, 158, 169]. Vascular invasion exhibited borderline significance, and tumor invasiveness seems to reach significance when there is extensive invasion in the peri-adrenal tissue. Among degenerative changes, only confluent necrosis appears commonly useful, although its weight was diversely appreciated. The value of proliferative activity (mitotic count) was considered a useful criterion [166, 168] or not [158, 169], with cut-off values of 3/10 or 5/30 HPF, respectively. As discussed elsewhere, mitotic count can be difficult to interpret due to technical defects or pyknotic nuclei [151, 159]. The presence of atypical mitosis is highly significant but infrequent [166].

Multiparameter scoring systems

The model presented by Linnoila et al. in 1990 [158] used two gross features, i.e. extra-adrenal location and coarse nodularity, and two histological features, i.e. confluent necrosis and absence of cytoplasmic hyaline globules. It had an overall correct classification rate of 88% and enabled correct classification of 9 out of 12 clinically malignant PCCs and 20 out of 21 benign PCCs. In this model, the weight of the extra-adrenal location exceeded largely that of the other parameters. As the extra-adrenal location is linked to *SDHx* mutations and *SDHx* mutations are linked to malignancy [13], it can be anticipated that genetic testing will be more efficient than histopathology in predicting a malignant risk for extra-adrenal locations.



In 2005, Kimura et al. constructed a score using six clinical and histopathological features to characterize 116 adrenal PCCs and 30 extra-adrenal PGLs, of which 38 had metastasis: growth pattern, cellularity, necrosis, vascular/capsular invasion, Ki-67 immuno-reactivity and type of catecholamine [167]. Tumors were classified as well, moderately and poorly differentiated (WD, MD and PD) according to their scores. Metastases were observed in 13% of 113 WD, 63% of 27 MD and 100% of 6 PD. The respective 10-year survival was 83%, 38% and 0%. The main interest of this system is the correlation of the score with prognosis in malignant cases, but it is unable to predict a risk of malignant evolution in the WD group.

In 2006, the diagnostic performance of PASS evaluated by ROC curve analysis in 130 patients (48 malignant PCCs and 82 benign followed >10 years) was compared to a logistic regression model using 15 variables: Sex, age and

Table 5 Pheochromocytoma of adrenal gland scaled score (PASS) (according to [166])

Feature	Score if present (no. of points assigned)
Large nests or diffuse growth (> 10% of tumor volume)	2
Central (middle or large nests) or confluent tumor necrosis	2
High cellularity	2
Cellular monotony	2
Tumor cell spindling (even if focal)	2
Mitotic figures >3/10 HPF	2
Atypical mitotic figures	2
Extension into adipose tissue	2
Vascular invasion	1
Capsular invasion	1
Profound nuclear pleiomorphism	1
Nuclear hyperchromasia	1
Total	20



tumor size were added to the 12 criteria of the PASS score [168]. The Cohen' kappa value of these variables was >86%, confirming their reproducibility. The nine variables having the largest odd ratio were retained in the logistic model. The area under the ROC curve improved from 0,899 with PASS alone to 0,983 with the logistic model. This study confirmed the diagnostic interest of the PASS and showed that its performance might be improved through a different variable selection and weighting: Variables containing redundant information (diffuse growth/hypercellularity) or less reproducible criteria (nuclear pleiomorphism/hyperchromasia) might be removed.

The value of the PASS was confirmed in the study by Strong et al., where 32 of 43 benign adrenal PCCs had PASS <4 (74% specificity), whereas 5/5 malignant PCC had PASS ≥ 6 (100% sensitivity) [162]. In a study by Agarwaal et al., examining PASS in 93 patients, including 68 with a follow-up period of more than 5 years, as much as 27 of 84 (32%) benign cases showed a PASS >4, and the specificity was 68%, while five of five malignant adrenal PCCs had a PASS >4 (100%) [171]. In this study, a bladder malignant PCC had a PASS score of 2, confirming that this system should not be used for extra-adrenal tumors. The reproducibility of the PASS was addressed by a study among PCC experts that found significant interobserver and intraobserver variations in its assignment, and the experts concluded that it could not be currently recommended for clinical prognostication [163]. This study, however, included only two truly malignant cases (with distant metastasis), and 15 of the 52 benign cases had follow-up of less than 1 year. Moreover, some criteria were extracted from surgical reports instead of reviewing the slides, and four of the five pathologists attributed scores >4 to the majority of benign cases.

There is currently no agreement on the utility and reproducibility of the PASS scoring systems that still require validation in larger series of cases [20, 31, 150, 157]. Guidelines and templates provided by colleges of pathologists for reporting of adrenal tumors incorporate some of the elements of the PASS systems but not all [157]. At this timepoint, a high PASS score should not be considered as diagnostic of malignancy as some tumors with high score never metastasize. In an effort of standardization of pathological reports for prospective validation, it may be worthwhile analyzing separately all of the elements of the PASS score.

Immunohistochemistry

A large number of markers have been analyzed by immunohistochemistry in PCC, with discrepant results among investigators (reviewed in [151, 172]). In almost all of the studies, the level of correlation with malignancy,

even if statistically significant, was never sufficient to have a diagnostic utility due to overlapping between benign and metastasizing tumors. Except for few markers, none of them was superior to standard histology to predict the malignancy of PCC. CD44s and telomerase reverse transcriptase hTERT [173, 174] showed a better discriminating power, but the results need further confirmation. The disappearance of pS100 positive sustentacular cells in malignant PCCs initially demonstrated by Lloyd in 1985 [175] has been confirmed in several studies [159, 166, 176, 177] and may be useful to underline diffuse growth areas, but it has no absolute value, as some metastasizing PCC may keep sustentacular cells. Finally, the Ki-67 proliferative index, extensively analyzed in PCC, remains the only marker whose interest seems largely accepted at the present time as no benign PCC had scores >2-3\% [159, 162, 178, 179]. The counterpart of this high specificity, however, is a poor sensitivity as approximately 50% of malignant PCCs have a Ki-67 index below the 2-3% threshold value. The Ki-67 proliferation index is thus considered as a useful adjunct [162, 174].

Summary

PCCs and PGLs are rare tumors exhibiting a wide range of malignant phenotypes. Between the most common benign and obviously aggressive tumors, fatal in few years, exists a group of proliferations that follow a more progressive course characterized either by local invasiveness and relapse or by late distant metastasis: The creation of an intermediate group of borderline malignancy has been suggested for these tumors and might be clinically relevant. Actually, the histopathological diagnosis of the two first categories seems reliable, but becomes more uncertain in the borderline group. Significant histological parameters were determined, but none is diagnostic enough to be used alone to predict evolution. Among the multiparametric systems that have been developed, none is ready to be used in daily routine. If the PASS is used at the present time, it must be kept in mind that a high score will define a risk group but is not diagnostic of malignancy.

Molecular biomarkers predicting malignancy

As conventional histopathology/morphology is of little help in identifying malignant PCCs and PGLs, much effort has been invested into the identification of molecular markers that may help distinguishing benign from malignant tumors (Table 6). Analysis on the level of genes, epigenetic changes, RNA and protein expression has been performed mainly on tumor tissue.



Table 6 Genetic tumoral changes that may be helpful in differentiating malignant and benign PCCs and paragangliomas

Source	Investigated material	Results (malignant vs. benign/normal)			
Human					
PCC	DNA/protein/RNA	Up: GNAS, INSM1, DOK5, ETV1, RET, NTRK1,; down: TGIF1, DSC3, TNFRSF10B, RASSF2,	[197]		
PCC	MicroRNA	Over-expression: miR-483-5p; under-expression: miR-15a and miR-16	[196]		
PCC	RNA/protein	>Twofold difference: 19 genes up-regulated; 113 down-regulated	[186]		
PCC	Protein	Over-expression: c-erbB-2/Her2, Ki-67/MIB-1	[207]		
PCC	Protein	Over-expression: SNAIL	[208]		
PCC	Protein	Over-expression: SNAIL	[164]		
PCC	Protein	Over-expression: c-erbB-2/Her2	[209]		
PCC	RNA	>Twofold difference: 16 genes up-regulated; ca. 90 down-regulated	[187]		
PCC	Protein	Over-expression: galectin-3	[210]		
PCC	RNA	Under-expression: Secretogranin II, prohormone convertases (PC)1 and PC2, EM66	[211]		
PCC	Protein/DNA	Over-expression of Ki-67/MIB-1; gain 17q; loss 1p, 3q	[174]		
PCC	RNA	Over-expression: hTERT	[191]		
PCC	Protein	High frequency of spindle-shaped cells detected by antibodies against chromogranin B and C	[212]		
PCC	RNA/protein	Over-expression: hTERT; increased activity of telomerase	[173]		
PCC	Protein/RNA	Over-expression: VEGF	[213]		
PCC	RNA/protein	Up-regulation: HSP90, ETB, EPAS1, VEGF; down-regulation: EM66	[154]		
PCC	Protein	Over-expression: N-cadherin	[214]		
PCC	Protein/RNA	Over-expression: Cox-2	[215]		
PCC	Protein	Over-expression: Tenascin	[216]		
PCC	Protein	Expression of only one chromogranin A region in all malignant tumors	[194]		
PCC	Protein	Over-expression: Ki-67/MIB-1	[217]		
PCC	Protein	Over-expression: p53	[218]		
PCC	Protein	Over-expression: ACTH; under-expression: NSE	[219]		
PCC	Protein	Increased activity of telomerase	[190]		
PCC	RNA	Increased activity of telomerase	[192]		
PCC	Protein	Over-expression: Ki-67/MIB-1; under-expression: S-100	[179]		
PCC	RNA/protein	Threefold up-regulation of c-myc	[220]		
PCC/PGL	DNA	Malignant PCC: gain at 19q, trisomy 12; loss at 11q malignant PGL: gain at 1q	[184]		
PCC/PGL	DNA	Higher intratumoral molecular heterogeneity for LOH	[185]		
PCC/PGL	Protein	Over-expression: topoisomerase II alpha, Ki-67/MIB-1, under-expression: RB	[169]		
Mouse					
PCC	Tumor-RNA	Metastatic phenotype, up: MAMDC2, JUB, MMP14,; down: ZNF35, ASF1B, RIMS3,	[200]		

PCC pheochromocytoma, Para paraganglioma, LOH loss of heterozygosity

Tissue-based analysis of DNA

LOH analysis of familial PCC/PGL revealed a non-random association of genetic losses depending on the existing germline mutation. Losses of 1p and 3p are frequently found in both sporadic and MEN2-related PCCs, suggesting a common genetic etiology [180, 181]. In contrast, VHL-related PCCs have losses of chromosome 11 [182]. Head and neck PGLs show very few genomic copy number alterations, with a dominance of 11q deletions [183]. Using high-resolution wholegenome array comparative to genomic hybridization

(CGH), recurrent genomic alterations in benign and malignant PCCs and PGLs have been identified [184]. It is of interest that DNA gain was significantly more often identified among malignant cases. Moreover, gain at 19q, trisomy 12 and loss at 11q were positively associated with malignant PCCs, while gain at 1q was commonly observed in the malignant PGLs. These results could speak in favor of malignant PCCs and malignant PGLs following different genetic pathways, as do their non-metastasizing counterparts.

Recently, a higher intratumoral heterogeneity for loss of heterozygosity (LOH) has been reported in malignant PCC/



PGL compared to benign tumors, underlining their different pathogenesis [185].

Tissue-based RNA genome wide analysis

Using microarray analysis of total RNA, a more than twofold difference in expression between benign and malignant PCs was reported in 132 genes, with the majority of them being down-regulated in malignant tumors [186]. In a similar study, another group identified slightly more than 100 genes to be differently expressed [187]. Again, in malignant PCC/PGL, the majority of the genes were down-regulated. Comparing the gene lists of both studies, only very few (e.g. QPCT (glutaminyl-peptide cyclotransferase), MAOA (monoamine oxidase) and DKK3 (Dickkopf homolog 3)) of the described genes were found in both studies. Despite the identification of numerous differently expressed genes, not a single gene has been identified, enabling a clear differentiation between malignant and benign tumors.

Tissue-based RNA/protein analysis of candidate genes

Numerous studies have aimed at defining prognostic biomarkers for detecting malignant PCC/PGL. Using RT-PCR and immunohistochemistry, a variety of such candidate genes and/or proteins have been found to be overexpressed (e.g. p53, c-erbB2/Her2, bcl-2, c-myc, Ki-67/MIB-1, EPAS1, ETB, VEGF, HSP90, Cox-2, Tenascin, N-cadherin, ACTH and SNAIL) or under-expressed (e.g. RB, EM66, S-100, NSE, Secretogranin II, prohormone convertases (PC)1 and (PC)2) in malignant versus benign PCCs/PGLs (Table 1). However, most of these markers remain at least controversial, and none could be convincingly confirmed by independent follow-up studies: While initially a high (40%) frequency of *p53* mutations in malignant PCCs was reported [187], this could not be confirmed [188].

Over-expression of c-erbB-2/Her2 has been reported by some authors to be associated with malignant PCCs [189]. In the same study, however, over-expression has also been reported in MEN2-associated PCCs that extremely rarely exhibit a malignant phenotype.

The significance of telomerase activity remains controversial. In one study, high telomerase activity was found in all malignant PCCs [190]. However, only three malignant tumors were investigated as opposed to 16 benign cases. In the same study, the telomerase catalytic subunit (hTERT) was also suggested as helpful, while the telomerase RNA component was not. These results could be confirmed by others [9, 173, 191]. In other/further studies, however, telomerase activity was only found in a minority of the malignant PCCs [192, 193]. Over-expression of HSP90 was

described to be helpful in distinguishing between malignant (over-expression) and benign PCCs [154].

In one study, analyzing antibodies to various specific regions of the chromogranin A molecule showed immunoreactivity with antibodies to all regions examined in benign PCCs, whereas only one region was expressed in all malignant tumors [194].

Recently, the loss of SDHB expression detected by immunohistochemistry has been associated with a high risk of malignancy in a retrospective series of 60 consecutive PCC/PGL patients. This study only included sympathetic tumors, and head and neck PGLs were not analyzed [195].

Tissue-based analysis of epigenetic changes

The analysis of microRNA profiling in benign and malignant PCCs revealed an over-expression of miR-483-5p, while miR-15a and miR-16 were under-expressed [196]. Using the rat PCC cell line PC12, these mRNAs were found to regulate cell proliferation via their effect on cyclin D1 and apoptosis. Integrative epigenomic and genomic analysis has further revealed a variety of additional potential tumor suppressor genes (e.g. TGIF1, DSC3, TNFRSF10B and RASSF2) and oncogenes (e.g. GNAS, INSM1, DOK5, ETV1, RET and NTRK1) [197] involved.

Animal models

Since human PCC/PGL are rare and about 10% of these tumors are malignant, models for studying the process of malignant transformation are needed/necessary. Conditional Pten knock-out mice are used to investigate metastatic PCCs [198]. A spontaneous rat model with p27 mutations develops PCCs with high proliferation rates. These tumors share expression profiles with human tumors and might be useful to understand the mechanism of tumor progression and therapy studies [199]. By comparing microarray gene expression of parental mouse PCC cells and more aggressive cells, several genes which may be important for this metastatic process have been identified, e.g. Fyn-related kinase (FRK) and keratin 8 (KRT8) (Table 1) [200]. In this model, however, the expression of many genes was very different in human tissues compared to mouse tissue, showing the difficulties of drawing direct conclusions from animal models.

Blood

None of the potential markers identified by molecular genetics has been convincingly shown to be a useful blood marker.



Summary

In summary, we are currently in the position of having a few more or less promising candidate markers to detect malignant PCC/PGL; none of them having been proven to be reproducible in a clinical setting, to our knowledge. In order to be useful (for indicating adjuvant therapy, for example), such a marker must prove very high specificity. Most of the studies are hampered due to the low number of investigated samples. At this point, no single marker has been identified enabling differentiation between malignant and benign PCCs/PGLs. Considering the genetic data, it might well be possible that different subtypes of PCC, sympathetic PGL and head and neck PGL will exist, with different risks and mechanisms of malignant transformation. It could be useful to introduce an intermediate category in the classification for those tumors exhibiting local aggressiveness at the time of diagnosis. It has also been proposed to abandon the term "malignant PCC/PGL" and just use the term PCC/ PGL as has been done for melanomas [201]. PCCs/PGLs should instead be classified "non-invasive", "minimally invasive" or "extensively invasive" (the last two similarly to follicular thyroid carcinomas).

Concluding remarks

All of the recommendations given in this review can only be considered as level IV. Due to the rarity of malignant PCCs/PGLs and the obvious difficulties in distinguishing benign and malignant PCCs/PGLs, any patient with a PCC/PGL should be treated in a specialized center where a multidisciplinary setting with specialized teams consisting of radiologists, endocrinologist, oncologists, pathologists and surgeons is available [202]. This would also facilitate future studies to address the aforementioned diagnostic and/or therapeutic obstacles.

Conflicts of interest None.

References

- Zarnegar R, Kebebew E, Duh QY, Clark OH (2006) Malignant pheochromocytoma. Surg Oncol Clin N Am 15(3):555–571
- Klein RD, Lloyd RV, Young WF (2009) Hereditary paraganglioma-pheochromocytoma syndromes. In: Pagon RA, Dolan CR, Stephens K (eds) Gene reviews. University of Washington, Seattle
- Thompson LD, Young WF, Kawashima A, Komminoth P, Tischler AS (2004) Malignant adrenal phaeochromocytoma. In: DeLellis RA, Lloyd RV, Heitz PU, Eng C (eds) WHO classification of tumours: pathology and genetics—tumours of endocrine organs. IARC Press, pp 147–150

- Adjalle R, Plouin PF, Pacak K, Lehnert H (2009) Treatment of malignant pheochromocytoma. Horm Metab Res 41(9):687–696
- Eisenhofer G, Bornstein SR, Brouwers FM, Cheung NK, Dahia PL, de Krijger RR, Giordano TJ, Greene LA, Goldstein DS, Lehnert H, Manger WM, Maris JM, Neumann HP, Pacak K, Shulkin BL, Smith DI, Tischler AS, Young WF Jr (2004) Malignant pheochromocytoma: current status and initiatives for future progress. Endocr Relat Cancer 11(3):423–436
- Lehnert H, Mundschenk J, Hahn K (2004) Malignant pheochromocytoma. Front Horm Res 31:155–162
- Scott HW Jr, Halter SA (1984) Oncologic aspects of pheochromocytoma: the importance of follow-up. Surgery 96(6):1061– 1066
- Medeiros LJ, Wolf BC, Balogh K, Federman M (1985) Adrenal pheochromocytoma: a clinicopathologic review of 60 cases. Hum Pathol 16(6):580–589
- Elder EE, Xu D, Hoog A, Enberg U, Hou M, Pisa P, Gruber A, Larsson C, Backdahl M (2003) Ki-67 and hTERT expression can aid in the distinction between malignant and benign pheochromocytoma and paraganglioma. Mod Pathol 16(3):246–255
- Fassnacht M, Kreissl MC, Weismann D, Allolio B (2009) New targets and therapeutic approaches for endocrine malignancies. Pharmacol Ther 123(1):117–141
- Harari A, Inabnet WB 3rd (2010) Malignant pheochromocytoma: a review. Am J Surg 201:700–708
- Proye C, Vix M, Goropoulos A, Kerlo P, Lecomte-Houcke M (1992) High incidence of malignant pheochromocytoma in a surgical unit. 26 cases out of 100 patients operated from 1971 to 1991. J Endocrinol Invest 15(9):651–663
- Gimenez-Roqueplo AP, Favier J, Rustin P, Rieubland C, Crespin M, Nau V, Khau Van Kien P, Corvol P, Plouin PF, Jeunemaitre X (2003) Mutations in the SDHB gene are associated with extra-adrenal and/or malignant phaeochromocytomas. Cancer Res 63 (17):5615–5621
- 14. Walz MK, Alesina PF, Wenger FA, Koch JA, Neumann HP, Petersenn S, Schmid KW, Mann K (2006) Laparoscopic and retroperitoneoscopic treatment of pheochromocytomas and retroperitoneal paragangliomas: results of 161 tumors in 126 patients. World J Surg 30(5):899–908
- Suh I, Shibru D, Eisenhofer G, Pacak K, Duh QY, Clark OH, Kebebew E (2009) Candidate genes associated with malignant pheochromocytomas by genome-wide expression profiling. Ann Surg 250(6):983–990
- O'Riordain DS, Young WF Jr, Grant CS, Carney JA, van Heerden JA (1996) Clinical spectrum and outcome of functional extraadrenal paraganglioma. World J Surg 20(7):916–921, discussion 922
- 17. Mannelli M, Castellano M, Schiavi F, Filetti S, Giacche M, Mori L, Pignataro V, Bernini G, Giache V, Bacca A, Biondi B, Corona G, Di Trapani G, Grossrubatscher E, Reimondo G, Arnaldi G, Giacchetti G, Veglio F, Loli P, Colao A, Ambrosio MR, Terzolo M, Letizia C, Ercolino T, Opocher G (2009) Clinically guided genetic screening in a large cohort of Italian patients with pheochromocytomas and/or functional or nonfunctional paragangliomas. J Clin Endocrinol Metab 94(5):1541–1547
- 18. Goldstein RE, O'Neill JA Jr, Holcomb GW 3rd, Morgan WM 3rd, Neblett WW 3rd, Oates JA, Brown N, Nadeau J, Smith B, Page DL, Abumrad NN, Scott HW Jr (1999) Clinical experience over 48 years with pheochromocytoma. Ann Surg 229(6):755–764, discussion 764–756
- Kopetschke R, Slisko M, Kilisli A, Tuschy U, Wallaschofski H, Fassnacht M, Ventz M, Beuschlein F, Reincke M, Reisch N, Quinkler M (2009) Frequent incidental discovery of phaeochromocytoma: data from a German cohort of 201 phaeochromocytoma. Eur J Endocrinol 161(2):355–361



- McNicol AM (2010) Update on tumours of the adrenal cortex, phaeochromocytoma and extra-adrenal paraganglioma. Histopathology:(in press)
- Ciftci AO, Tanyel FC, Senocak ME, Buyukpamukcu N (2001)
 Pheochromocytoma in children. J Pediatr Surg 36(3):447–452
- Barontini M, Levin G, Sanso G (2006) Characteristics of pheochromocytoma in a 4- to 20-year-old population. Ann N Y Acad Sci 1073:30–37
- Bissada NK, Safwat AS, Seyam RM, Al Sobhi S, Hanash KA, Jackson RJ, Sakati N, Bissada MA (2008) Pheochromocytoma in children and adolescents: a clinical spectrum. J Pediatr Surg 43 (3):540–543
- Neumann HP, Berger DP, Sigmund G, Blum U, Schmidt D, Parmer RJ, Volk B, Kirste G (1993) Pheochromocytomas, multiple endocrine neoplasia type 2, and von Hippel–Lindau disease. N Engl J Med 329(21):1531–1538
- Modigliani E, Vasen HM, Raue K, Dralle H, Frilling A, Gheri RG, Brandi ML, Limbert E, Niederle B, Forgas L et al (1995) Pheochromocytoma in multiple endocrine neoplasia type 2: European study. The Euromen Study Group. J Intern Med 238 (4):363–367
- Bravo EL, Tagle R (2003) Pheochromocytoma: state-of-the-art and future prospects. Endocr Rev 24(4):539–553
- Sisson JC, Shulkin BL, Esfandiari NH (2006) Courses of malignant pheochromocytoma: implications for therapy. Ann N Y Acad Sci 1073:505–511
- 28. Pacak K, Eisenhofer G, Ahlman H, Bornstein SR, Gimenez-Roqueplo AP, Grossman AB, Kimura N, Mannelli M, McNicol AM, Tischler AS (2007) Pheochromocytoma: recommendations for clinical practice from the First International Symposium. October 2005. Nat Clin Pract Endocrinol Metab 3(2):92–102
- Tanaka S, Ito T, Tomoda J, Higashi T, Yamada G, Tsuji T (1993) Malignant pheochromocytoma with hepatic metastasis diagnosed 20 years after resection of the primary adrenal lesion. Intern Med 32(10):789–794
- Edstrom Elder E, Hjelm Skog AL, Hoog A, Hamberger B (2003)
 The management of benign and malignant pheochromocytoma and abdominal paraganglioma. Eur J Surg Oncol 29(3):278–283
- Ahlman H (2006) Malignant pheochromocytoma: state of the field with future projections. Ann N Y Acad Sci 1073:449–464
- Baysal BE, Ferrell RE, Willett-Brozick JE, Lawrence EC, Myssiorek D, Bosch A, van der Mey A, Taschner PE, Rubinstein WS, Myers EN, Richard CW 3rd, Cornelisse CJ, Devilee P, Devlin B (2000) Mutations in SDHD, a mitochondrial complex II gene, in hereditary paraganglioma. Science 287(5454):848– 851
- Gimm O, Armanios M, Dziema H, Neumann HP, Eng C (2000) Somatic and occult germ-line mutations in SDHD, a mitochondrial complex II gene, in nonfamilial pheochromocytoma. Cancer Res 60(24):6822–6825
- Niemann S, Muller U (2000) Mutations in SDHC cause autosomal dominant paraganglioma, type 3. Nat Genet 26 (3):268–270
- 35. Astuti D, Latif F, Dallol A, Dahia PL, Douglas F, George E, Skoldberg F, Husebye ES, Eng C, Maher ER (2001) Gene mutations in the succinate dehydrogenase subunit SDHB cause susceptibility to familial pheochromocytoma and to familial paraganglioma. Am J Hum Genet 69(1):49–54
- 36. Hao HX, Khalimonchuk O, Schraders M, Dephoure N, Bayley JP, Kunst H, Devilee P, Cremers CW, Schiffman JD, Bentz BG, Gygi SP, Winge DR, Kremer H, Rutter J (2009) SDH5, a gene required for flavination of succinate dehydrogenase, is mutated in paraganglioma. Science 325(5944):1139–1142
- Burnichon N, Briere JJ, Libe R, Vescovo L, Riviere J, Tissier F, Jouanno E, Jeunemaitre X, Benit P, Tzagoloff A, Rustin P, Bertherat J, Favier J, Gimenez-Roqueplo AP (2010) SDHA is a

- tumor suppressor gene causing paraganglioma. Hum Mol Genet 19(15):3011-3020
- 38. Benn DE, Gimenez-Roqueplo AP, Reilly JR, Bertherat J, Burgess J, Byth K, Croxson M, Dahia PL, Elston M, Gimm O, Henley D, Herman P, Murday V, Niccoli-Sire P, Pasieka JL, Rohmer V, Tucker K, Jeunemaitre X, Marsh DJ, Plouin PF, Robinson BG (2006) Clinical presentation and penetrance of pheochromocytoma/paraganglioma syndromes. J Clin Endocrinol Metab 91(3):827–836
- 39. Schlisio S, Kenchappa RS, Vredeveld LC, George RE, Stewart R, Greulich H, Shahriari K, Nguyen NV, Pigny P, Dahia PL, Pomeroy SL, Maris JM, Look AT, Meyerson M, Peeper DS, Carter BD, Kaelin WG Jr (2008) The kinesin KIF1Bbeta acts downstream from EglN3 to induce apoptosis and is a potential 1p36 tumor suppressor. Genes Dev 22(7):884–893
- 40. Qin Y, Yao L, King EE, Buddavarapu K, Lenci RE, Chocron ES, Lechleiter JD, Sass M, Aronin N, Schiavi F, Boaretto F, Opocher G, Toledo RA, Toledo SP, Stiles C, Aguiar RC, Dahia PL (2010) Germline mutations in TMEM127 confer susceptibility to pheochromocytoma. Nat Genet 42(3):229–233
- 41. Comino-Mendez I, Gracia-Aznarez FJ, Schiavi F, Landa I, Leandro-Garcia LJ, Leton R, Honrado E, Ramos-Medina R, Caronia D, Pita G, Gomez-Grana A, de Cubas AA, Inglada-Perez L, Maliszewska A, Taschin E, Bobisse S, Pica G, Loli P, Hernandez-Lavado R, Diaz JA, Gomez-Morales M, Gonzalez-Neira A, Roncador G, Rodriguez-Antona C, Benitez J, Mannelli M, Opocher G, Robledo M, Cascon A (2011) Exome sequencing identifies MAX mutations as a cause of hereditary pheochromocytoma. Nat Genet 43(7):663–667
- Carney JA (1999) Gastric stromal sarcoma, pulmonary chondroma, and extra-adrenal paraganglioma (Carney triad): natural history, adrenocortical component, and possible familial occurrence. Mayo Clin Proc 74(6):543–552
- Carney JA, Stratakis CA (2002) Familial paraganglioma and gastric stromal sarcoma: a new syndrome distinct from the Carney triad. Am J Med Genet 108(2):132–139
- 44. Amar L, Bertherat J, Baudin E, Ajzenberg C, Bressac-de Paillerets B, Chabre O, Chamontin B, Delemer B, Giraud S, Murat A, Niccoli-Sire P, Richard S, Rohmer V, Sadoul JL, Strompf L, Schlumberger M, Bertagna X, Plouin PF, Jeunemaitre X, Gimenez-Roqueplo AP (2005) Genetic testing in pheochromocytoma or functional paraganglioma. J Clin Oncol 23(34):8812–8818
- 45. Amar L, Baudin E, Burnichon N, Peyrard S, Silvera S, Bertherat J, Bertagna X, Schlumberger M, Jeunemaitre X, Gimenez-Roqueplo AP, Plouin PF (2007) Succinate dehydrogenase B gene mutations predict survival in patients with malignant pheochromocytomas or paragangliomas. J Clin Endocrinol Metab 92(10):3822–3828
- 46. Yeh IT, Lenci RE, Qin Y, Buddavarapu K, Ligon AH, Leteurtre E, Do Cao C, Cardot-Bauters C, Pigny P, Dahia PL (2008) A germline mutation of the KIF1B beta gene on 1p36 in a family with neural and nonneural tumors. Hum Genet 124(3):279–285
- 47. Yao L, Schiavi F, Cascon A, Qin Y, Inglada-Perez L, King EE, Toledo RA, Ercolino T, Rapizzi E, Ricketts CJ, Mori L, Giacche M, Mendola A, Taschin E, Boaretto F, Loli P, Iacobone M, Rossi GP, Biondi B, Lima-Junior JV, Kater CE, Bex M, Vikkula M, Grossman AB, Gruber SB, Barontini M, Persu A, Castellano M, Toledo SP, Maher ER, Mannelli M, Opocher G, Robledo M, Dahia PL (2011) Spectrum and prevalence of FP/TMEM127 gene mutations in pheochromocytomas and paragangliomas. JAMA 304(23):2611–2619
- 48. Neumann HP, Sullivan M, Winter A, Malinoc A, Hoffmann MM, Boedeker CC, Bertz H, Walz MK, Moeller LC, Schmid KW, Eng C (2011) Germline mutations of the TMEM127 gene in patients with paraganglioma of head and neck and



- extraadrenal abdominal sites. J Clin Endocrinol Metab 96(8): E1279–E1282
- Chrisoulidou A, Kaltsas G, Ilias I, Grossman AB (2007) The diagnosis and management of malignant phaeochromocytoma and paraganglioma. Endocr Relat Cancer 14(3):569–585
- Glodny B, Winde G, Herwig R, Meier A, Kuhle C, Cromme S, Vetter H (2001) Clinical differences between benign and malignant pheochromocytomas. Endocr J 48(2):151–159
- Birrenbach T, Stanga Z, Cottagnoud P, Stucki A (2008) Unexpected metastatic pheochromocytoma—an unusual presentation. Eur J Intern Med 19(1):60–62
- Costa SR, Cabral NM, Abhrao AT, Costa RB, Silva LM, Lupinacci RA (2008) Giant cystic malignant pheochromocytoma invading right hepatic lobe: report on two cases. Sao Paulo Med J 126(4):229–231
- Honda M, Uesugi K, Yamazaki H, Dezawa A, Mizuguchi K, Yamaji T, Ishibashi M (2000) Malignant pheochromocytoma lacking clinical features of catecholamine excess until the late stage. Intern Med 39(10):820–825
- Rao F, Keiser HR, O'Connor DT (2000) Malignant pheochromocytoma. Chromaffin granule transmitters and response to treatment. Hypertension 36(6):1045–1052
- Rao F, Keiser HR, O'Connor DT (2002) Malignant and benign pheochromocytoma: chromaffin granule transmitters and the response to medical and surgical treatment. Ann N Y Acad Sci 971:530–532
- Oishi S, Sato T (1988) Elevated serum neuron-specific enolase in patients with malignant pheochromocytoma. Cancer 61 (6):1167–1170
- 57. Grossman A, Pacak K, Sawka A, Lenders JW, Harlander D, Peaston RT, Reznek R, Sisson J, Eisenhofer G (2006) Biochemical diagnosis and localization of pheochromocytoma: can we reach a consensus? Ann N Y Acad Sci 1073:332–347
- Young WF (2008) Endocrine hypertension. In: Kronenberg HM, Melmed S, Polonsky KS, Larsen PR (eds) Texbook of endocrinology. Saunders Elsevier, Philadelphia, pp 505–537
- Yeh CN, Jeng LB, Chen MF, Hung CF (2001) Nonfunctioning malignant pheochromocytoma associated with dermatomyositis: case report and literature review. World J Urol 19(2):148–150
- Holwitt D, Neifeld J, Massey G, Lanning D (2007) Case report of an 11-year-old child with a nonfunctional malignant pheochromocytoma. J Pediatr Surg 42(11):E13–E15
- 61. Lumachi F, Borsato S, Tregnaghi A, Marino F, Fassina A, Zucchetta P, Marzola MC, Cecchin D, Bui F, Iacobone M, Favia G (2007) High risk of malignancy in patients with incidentally discovered adrenal masses: accuracy of adrenal imaging and image-guided fine-needle aspiration cytology. Tumori 93 (3):269–274
- 62. van der Harst E, de Herder WW, de Krijger RR, Bruining HA, Bonjer HJ, Lamberts SW, van den Meiracker AH, Stijnen TH, Boomsma F (2002) The value of plasma markers for the clinical behaviour of phaeochromocytomas. Eur J Endocrinol 147(1):85– 94
- 63. Algeciras-Schimnich A, Preissner CM, Young WF Jr, Singh RJ, Grebe SK (2008) Plasma chromogranin A or urine fractionated metanephrines follow-up testing improves the diagnostic accuracy of plasma fractionated metanephrines for pheochromocytoma. J Clin Endocrinol Metab 93(1):91–95
- 64. Schlumberger M, Gicquel C, Lumbroso J, Tenenbaum F, Comoy E, Bosq J, Fonseca E, Ghillani PP, Aubert B, Travagli JP et al (1992) Malignant pheochromocytoma: clinical, biological, histologic and therapeutic data in a series of 20 patients with distant metastases. J Endocrinol Invest 15(9):631–642
- 65. Proye C, Fossati P, Fontaine P, Lefebvre J, Decoulx M, Wemeau JL, Dewailly D, Rwamasirabo E, Cecat P (1986) Dopamine-secreting pheochromocytoma: an unrecognized entity? Classifi-

- cation of pheochromocytomas according to their type of secretion. Surgery 100(6):1154-1162
- John H, Ziegler WH, Hauri D, Jaeger P (1999) Pheochromocytomas: can malignant potential be predicted? Urology 53(4):679–683
- 67. Plouin PF, Chatellier G, Fofol I, Corvol P (1997) Tumor recurrence and hypertension persistence after successful pheochromocytoma operation. Hypertension 29(5):1133–1139
- Blake MA, Kalra MK, Maher MM, Sahani DV, Sweeney AT, Mueller PR, Hahn PF, Boland GW (2004) Pheochromocytoma: an imaging chameleon. Radiographics 24(Suppl 1):S87–S99
- 69. Plockinger U, Rindi G, Arnold R, Eriksson B, Krenning EP, de Herder WW, Goede A, Caplin M, Oberg K, Reubi JC, Nilsson O, Delle Fave G, Ruszniewski P, Ahlman H, Wiedenmann B (2004) Guidelines for the diagnosis and treatment of neuroendocrine gastrointestinal tumours. A consensus statement on behalf of the European Neuroendocrine Tumour Society (ENETS). Neuroendocrinology 80(6):394–424
- Hillel PG, van Beek EJ, Taylor C, Lorenz E, Bax ND, Prakash V, Tindale WB (2006) The clinical impact of a combined gamma camera/CT imaging system on somatostatin receptor imaging of neuroendocrine tumours. Clin Radiol 61(7):579–587
- Kwekkeboom DJ, Krenning EP, Lebtahi R, Komminoth P, Kos-Kudla B, de Herder WW, Plockinger U (2009) ENETS consensus guidelines for the standards of care in neuroendocrine tumors: peptide receptor radionuclide therapy with radiolabeled somatostatin analogs. Neuroendocrinology 90(2):220–226
- Lumachi F, Tregnaghi A, Zucchetta P, Cristina Marzola M, Cecchin D, Grassetto G, Bui F (2006) Sensitivity and positive predictive value of CT, MRI and 123I-MIBG scintigraphy in localizing pheochromocytomas: a prospective study. Nucl Med Commun 27(7):583–587
- Ilias I, Pacak K (2004) Anatomical and functional imaging of metastatic pheochromocytoma. Ann N Y Acad Sci 1018:495– 504
- 74. van der Harst E, de Herder WW, Bruining HA, Bonjer HJ, de Krijger RR, Lamberts SW, van de Meiracker AH, Boomsma F, Stijnen T, Krenning EP, Bosman FT, Kwekkeboom DJ (2001) [(123)I]metaiodobenzylguanidine and [(111)In]octreotide uptake in benign and malignant pheochromocytomas. J Clin Endocrinol Metab 86(2):685–693
- Blake MA, Cronin CG, Boland GW (2010) Adrenal imaging.
 AJR Am J Roentgenol 194(6):1450–1460
- Szolar DH, Korobkin M, Reittner P, Berghold A, Bauernhofer T, Trummer H, Schoellnast H, Preidler KW, Samonigg H (2005) Adrenocortical carcinomas and adrenal pheochromocytomas: mass and enhancement loss evaluation at delayed contrastenhanced CT. Radiology 234(2):479–485
- Varghese JC, Hahn PF, Papanicolaou N, Mayo-Smith WW, Gaa JA, Lee MJ (1997) MR differentiation of phaeochromocytoma from other adrenal lesions based on qualitative analysis of T2 relaxation times. Clin Radiol 52(8):603–606
- Mayo-Smith WW, Boland GW, Noto RB, Lee MJ (2001) Stateof-the-art adrenal imaging. Radiographics 21(4):995–1012
- 79. Takano A, Oriuchi N, Tsushima Y, Taketomi-Takahashi A, Nakajima T, Arisaka Y, Higuchi T, Amanuma M, Endo K (2008) Detection of metastatic lesions from malignant pheochromocytoma and paraganglioma with diffusion-weighted magnetic resonance imaging: comparison with 18F-FDG positron emission tomography and 123I-MIBG scintigraphy. Ann Nucl Med 22(5):395–401
- Kann PH, Wirkus B, Behr T, Klose KJ, Meyer S (2004) Endosonographic imaging of benign and malignant pheochromocytomas. J Clin Endocrinol Metab 89(4):1694–1697
- Hattner RS, Huberty JP, Engelstad BL, Gooding CA, Ablin AR (1984) Localization of m-iodo(131I)benzylguanidine in neuroblastoma. AJR Am J Roentgenol 143(2):373–374



- Nakajo M, Shapiro B, Copp J, Kalff V, Gross MD, Sisson JC, Beierwaltes WH (1983) The normal and abnormal distribution of the adrenomedullary imaging agent m-[I-131]iodobenzylguanidine (I-131 MIBG) in man: evaluation by scintigraphy. J Nucl Med 24(8):672–682
- Shapiro B, Copp JE, Sisson JC, Eyre PL, Wallis J, Beierwaltes WH (1985) Iodine-131 metaiodobenzylguanidine for the locating of suspected pheochromocytoma: experience in 400 cases. J Nucl Med 26(6):576–585
- 84. Wieland DM, Wu J, Brown LE, Mangner TJ, Swanson DP, Beierwaltes WH (1980) Radiolabeled adrenergic neuronblocking agents: adrenomedullary imaging with [131I]iodobenzylguanidine. J Nucl Med 21(4):349–353
- Khafagi FA, Shapiro B, Gross MD (1989) The adrenal gland. In: Maisey MN, Britton KE, Gilday DL (eds) Clinical nuclear medicine. Chapman & Hall, London, pp 271–291
- Bombardieri E, Giammarile F, Aktolun C, Baum RP, Bischof Delaloye A, Maffioli L, Moncayo R, Mortelmans L, Pepe G, Reske SN, Castellani MR, Chiti A (2010) 131I/123I-metaiodobenzylguanidine (MIBG) scintigraphy: procedure guidelines for tumour imaging. Eur J Nucl Med Mol Imaging 37(12):2436– 2446
- 87. Havekes B, Lai EW, Corssmit EP, Romijn JA, Timmers HJ, Pacak K (2008) Detection and treatment of pheochromocytomas and paragangliomas: current standing of MIBG scintigraphy and future role of PET imaging. Q J Nucl Med Mol Imaging 52 (4):419–429
- Lumbroso JD, Guermazi F, Hartmann O, Coornaert S, Rabarison Y, Leclere JG, Couanet D, Bayle C, Caillaud JM, Lemerle J et al (1988) Meta-iodobenzylguanidine (MIBG) scans in neuroblastoma: sensitivity and specificity, a review of 115 scans. Prog Clin Biol Res 271:689–705
- Jacobson AF, Deng H, Lombard J, Lessig HJ, Black RR (2010)
 123I-meta-iodobenzylguanidine scintigraphy for the detection of neuroblastoma and pheochromocytoma: results of a metaanalysis. J Clin Endocrinol Metab 95(6):2596–2606
- Kaltsas G, Korbonits M, Heintz E, Mukherjee JJ, Jenkins PJ, Chew SL, Reznek R, Monson JP, Besser GM, Foley R, Britton KE, Grossman AB (2001) Comparison of somatostatin analog and meta-iodobenzylguanidine radionuclides in the diagnosis and localization of advanced neuroendocrine tumors. J Clin Endocrinol Metab 86(2):895–902
- 91. Kaji P, Carrasquillo JA, Linehan WM, Chen CC, Eisenhofer G, Pinto PA, Lai EW, Pacak K (2007) The role of 6-[18F] fluorodopamine positron emission tomography in the localization of adrenal pheochromocytoma associated with von Hippel-Lindau syndrome. Eur J Endocrinol 156(4):483–487
- Sisson JC, Shulkin BL (1999) Nuclear medicine imaging of pheochromocytoma and neuroblastoma. Q J Nucl Med 43 (3):217–223
- 93. Bhatia KS, Ismail MM, Sahdev A, Rockall AG, Hogarth K, Canizales A, Avril N, Monson JP, Grossman AB, Reznek RH (2008) 123I-metaiodobenzylguanidine (MIBG) scintigraphy for the detection of adrenal and extra-adrenal phaeochromocytomas: CT and MRI correlation. Clin Endocrinol (Oxf) 69(2):181–188
- 94. Wiseman GA, Pacak K, O'Dorisio MS, Neumann DR, Waxman AD, Mankoff DA, Heiba SI, Serafini AN, Tumeh SS, Khutoryansky N, Jacobson AF (2009) Usefulness of 123I-MIBG scintigraphy in the evaluation of patients with known or suspected primary or metastatic pheochromocytoma or paraganglioma: results from a prospective multicenter trial. J Nucl Med 50(9):1448–1454
- Donckier JE, Michel L (2010) Phaeochromocytoma: state-of-theart. Acta Chir Belg 110(2):140–148
- Tsuchimochi S, Nakajo M, Nakabeppu Y, Tani A (1997)
 Metastatic pulmonary pheochromocytomas: positive I-123

- MIBG SPECT with negative I-131 MIBG and equivocal I-123 MIBG planar imaging. Clin Nucl Med 22(10):687–690
- Fujita A, Hyodoh H, Kawamura Y, Kanegae K, Furuse M, Kanazawa K (2000) Use of fusion images of I-131 metaiodobenzylguanidine, SPECT, and magnetic resonance studies to identify a malignant pheochromocytoma. Clin Nucl Med 25 (6):440–442
- Rozovsky K, Koplewitz BZ, Krausz Y, Revel-Vilk S, Weintraub M, Chisin R, Klein M (2008) Added value of SPECT/CT for correlation of MIBG scintigraphy and diagnostic CT in neuroblastoma and pheochromocytoma. AJR Am J Roentgenol 190 (4):1085–1090
- Rufini V, Shulkin B (2008) The evolution in the use of MIBG in more than 25 years of experimental and clinical applications. Q J Nucl Med Mol Imaging 52(4):341–350
- 100. Meyer-Rochow GY, Schembri GP, Benn DE, Sywak MS, Delbridge LW, Robinson BG, Roach PJ, Sidhu SB (2010) The utility of metaiodobenzylguanidine single photon emission computed tomography/computed tomography (MIBG SPECT/ CT) for the diagnosis of pheochromocytoma. Ann Surg Oncol 17 (2):392–400
- 101. Fukuoka M, Taki J, Mochizuki T, Kinuya S (2011) Comparison of diagnostic value of I-123 MIBG and high-dose I-131 MIBG scintigraphy including incremental value of SPECT/CT over planar image in patients with malignant pheochromocytoma/ paraganglioma and neuroblastoma. Clin Nucl Med 36(1):1–7
- 102. Buhl T, Mortensen J, Kjaer A (2002) I-123 MIBG imaging and intraoperative localization of metastatic pheochromocytoma: a case report. Clin Nucl Med 27(3):183–185
- Vaidyanathan G (2008) Meta-iodobenzylguanidine and analogues: chemistry and biology. Q J Nucl Med Mol Imaging 52 (4):351–368
- 104. Krenning EP, Bakker WH, Breeman WA, Koper JW, Kooij PP, Ausema L, Lameris JS, Reubi JC, Lamberts SW (1989) Localisation of endocrine-related tumours with radioiodinated analogue of somatostatin. Lancet 1(8632):242–244
- 105. Tenenbaum F, Lumbroso J, Schlumberger M, Mure A, Plouin PF, Caillou B, Parmentier C (1995) Comparison of radiolabeled octreotide and meta-iodobenzylguanidine (MIBG) scintigraphy in malignant pheochromocytoma. J Nucl Med 36(1):1–6
- 106. Kopf D, Bockisch A, Steinert H, Hahn K, Beyer J, Neumann HP, Hensen J, Lehnert H (1997) Octreotide scintigraphy and catecholamine response to an octreotide challenge in malignant phaeochromocytoma. Clin Endocrinol (Oxf) 46(1):39–44
- 107. Limouris GS, Giannakopoulos V, Stavraka A, Toubanakis N, Vlahos L (1997) Comparison of In-111 pentetreotide, Tc-99m (V)DMSA and I-123 mlBG scintimaging in neural crest tumors. Anticancer Res 17(3B):1589–1592
- 108. de Herder WW, Kwekkeboom DJ, Valkema R, Feelders RA, van Aken MO, Lamberts SW, van der Lely AJ, Krenning EP (2005) Neuroendocrine tumors and somatostatin: imaging techniques. J Endocrinol Invest 28(11 Suppl International):132–136
- 109. Duet M, Sauvaget E, Petelle B, Rizzo N, Guichard JP, Wassef M, Le Cloirec J, Herman P, Tran Ba Huy P (2003) Clinical impact of somatostatin receptor scintigraphy in the management of paragangliomas of the head and neck. J Nucl Med 44(11):1767–1774
- Goldsmith SJ (2009) Update on nuclear medicine imaging of neuroendocrine tumors. Future Oncol 5(1):75–84
- Hodolic M, Fettich J, Banti E, Chondrogiannis S, Al-Nahhas A, Rubello D (2010) Diagnostics of neuroendocrine tumours. In Vivo 24(5):771–774
- Carrasquillo JA, Chen CC (2010) Molecular imaging of neuroendocrine tumors. Semin Oncol 37(6):662–679
- 113. Vander Heiden MG, Cantley LC, Thompson CB (2009) Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science 324(5930):1029–1033



- 114. Shanmugam M, McBrayer SK, Rosen ST (2009) Targeting the Warburg effect in hematological malignancies: from PET to therapy. Curr Opin Oncol 21(6):531–536
- 115. Shulkin BL, Koeppe RA, Francis IR, Deeb GM, Lloyd RV, Thompson NW (1993) Pheochromocytomas that do not accumulate metaiodobenzylguanidine: localization with PET and administration of FDG. Radiology 186(3):711–715
- 116. Homma K, Hayashi K, Wakino S, Irie R, Mukai M, Kumagai H, Shibata H, Saruta T (2006) Primary malignant hepatic pheochromocytoma with negative adrenal scintigraphy. Hypertens Res 29(7):551–554
- Ezuddin S, Fragkaki C (2005) MIBG and FDG PET findings in a patient with malignant pheochromocytoma: a significant discrepancy. Clin Nucl Med 30(8):579–581
- 118. Kikuchi F, Imachi H, Murao K, Ohyama T, Miyai Y, Kushida Y, Haba R, Kakehi Y, Ishida T (2010) Positron emission tomography with 18F-fluorodeoxyglucose is a useful tool for the diagnosis of pheochromocytomas without distant metastasis, where malignancy is suspected on the basis of histopathologic analysis. Am J Med Sci 340(2):160–163
- 119. Timmers HJ, Kozupa A, Chen CC, Carrasquillo JA, Ling A, Eisenhofer G, Adams KT, Solis D, Lenders JW, Pacak K (2007) Superiority of fluorodeoxyglucose positron emission tomography to other functional imaging techniques in the evaluation of metastatic SDHB-associated pheochromocytoma and paraganglioma. J Clin Oncol 25(16):2262–2269
- 120. Iagaru A, Mittra E, Yaghoubi SS, Dick DW, Quon A, Goris ML, Gambhir SS (2009) Novel strategy for a cocktail 18F-fluoride and 18F-FDG PET/CT scan for evaluation of malignancy: results of the pilot-phase study. J Nucl Med 50(4):501–505
- 121. Pacak K, Eisenhofer G, Carrasquillo JA, Chen CC, Li ST, Goldstein DS (2001) 6-[18F]fluorodopamine positron emission tomographic (PET) scanning for diagnostic localization of pheochromocytoma. Hypertension 38(1):6–8
- 122. Timmers HJ, Carrasquillo JA, Whatley M, Eisenhofer G, Chen CC, Ling A, Linehan WM, Pinto PA, Adams KT, Pacak K (2007) Usefulness of standardized uptake values for distinguishing adrenal glands with pheochromocytoma from normal adrenal glands by use of 6-18F-fluorodopamine PET. J Nucl Med 48 (12):1940–1944
- 123. Timmers HJ, Eisenhofer G, Carrasquillo JA, Chen CC, Whatley M, Ling A, Adams KT, Pacak K (2009) Use of 6-[18F]-fluorodopamine positron emission tomography (PET) as first-line investigation for the diagnosis and localization of non-metastatic and metastatic phaeochromocytoma (PHEO). Clin Endocrinol (Oxf) 71(1):11–17
- 124. Timmers HJ, Chen CC, Carrasquillo JA, Whatley M, Ling A, Havekes B, Eisenhofer G, Martiniova L, Adams KT, Pacak K (2009) Comparison of 18F-fluoro-L-DOPA, 18F-fluoro-deoxyglucose, and 18F-fluorodopamine PET and 123I-MIBG scintigraphy in the localization of pheochromocytoma and paraganglioma. J Clin Endocrinol Metab 94(12):4757–4767
- 125. Ilias I, Yu J, Carrasquillo JA, Chen CC, Eisenhofer G, Whatley M, McElroy B, Pacak K (2003) Superiority of 6-[18F]-fluorodopamine positron emission tomography versus [131I]-metaiodobenzylguanidine scintigraphy in the localization of metastatic pheochromocytoma. J Clin Endocrinol Metab 88 (9):4083–4087
- 126. Zelinka T, Timmers HJ, Kozupa A, Chen CC, Carrasquillo JA, Reynolds JC, Ling A, Eisenhofer G, Lazurova I, Adams KT, Whatley MA, Widimsky J Jr, Pacak K (2008) Role of positron emission tomography and bone scintigraphy in the evaluation of bone involvement in metastatic pheochromocytoma and paraganglioma: specific implications for succinate dehydrogenase enzyme subunit B gene mutations. Endocr Relat Cancer 15 (1):311–323

- 127. Hoegerle S, Nitzsche E, Altehoefer C, Ghanem N, Manz T, Brink I, Reincke M, Moser E, Neumann HP (2002) Pheochromocytomas: detection with 18F DOPA whole body PET—initial results. Radiology 222(2):507–512
- 128. Imani F, Agopian VG, Auerbach MS, Walter MA, Imani F, Benz MR, Dumont RA, Lai CK, Czernin JG, Yeh MW (2009) 18F-FDOPA PET and PET/CT accurately localize pheochromocytomas. J Nucl Med 50(4):513–519
- 129. Fiebrich HB, Brouwers AH, Kerstens MN, Pijl ME, Kema IP, de Jong JR, Jager PL, Elsinga PH, Dierckx RA, van der Wal JE, Sluiter WJ, de Vries EG, Links TP (2009) 6-[F-18]Fluoro-L-dihydroxyphenylalanine positron emission tomography is superior to conventional imaging with (123)I-metaiodobenzylguani-dine scintigraphy, computer tomography, and magnetic resonance imaging in localizing tumors causing catecholamine excess. J Clin Endocrinol Metab 94(10):3922–3930
- 130. Fottner C, Helisch A, Anlauf M, Rossmann H, Musholt TJ, Kreft A, Schadmand-Fischer S, Bartenstein P, Lackner KJ, Kloppel G, Schreckenberger M, Weber MM (2010) 6-18F-fluoro-L-dihydroxyphenylalanine positron emission tomography is superior to 123I-metaiodobenzyl-guanidine scintigraphy in the detection of extraadrenal and hereditary pheochromocytomas and paragangliomas: correlation with vesicular monoamine transporter expression. J Clin Endocrinol Metab 95 (6):2800–2810
- 131. Timmers HJ, Hadi M, Carrasquillo JA, Chen CC, Martiniova L, Whatley M, Ling A, Eisenhofer G, Adams KT, Pacak K (2007) The effects of carbidopa on uptake of 6-18F-fluoro-L-DOPA in PET of pheochromocytoma and extraadrenal abdominal paraganglioma. J Nucl Med 48(10):1599–1606
- 132. Taieb D, Tessonnier L, Sebag F, Niccoli-Sire P, Morange I, Colavolpe C, De Micco C, Barlier A, Palazzo FF, Henry JF, Mundler O (2008) The role of 18F-FDOPA and 18F-FDG-PET in the management of malignant and multifocal phaeochromocytomas. Clin Endocrinol (Oxf) 69(4):580–586
- 133. Zettinig G, Mitterhauser M, Wadsak W, Becherer A, Pirich C, Vierhapper H, Niederle B, Dudczak R, Kletter K (2004) Positron emission tomography imaging of adrenal masses: (18)F-fluorodeoxyglucose and the 11beta-hydroxylase tracer (11)C-metomidate. Eur J Nucl Med Mol Imaging 31 (9):1224–1230
- 134. Kroiss A, Putzer D, Uprimny C, Decristoforo C, Gabriel M, Santner W, Kranewitter C, Warwitz B, Waitz D, Kendler D, Virgolini IJ (2011) Functional imaging in phaeochromocytoma and neuroblastoma with 68Ga-DOTA-Tyr 3-octreotide positron emission tomography and 123I-metaiodobenzylguanidine. Eur J Nucl Med Mol Imaging 38(5):865–873
- 135. Win Z, Al-Nahhas A, Towey D, Todd JF, Rubello D, Lewington V, Gishen P (2007) 68Ga-DOTATATE PET in neuroectodermal tumours: first experience. Nucl Med Commun 28(5):359–363
- 136. Vanderveen KA, Thompson SM, Callstrom MR, Young WF Jr, Grant CS, Farley DR, Richards ML, Thompson GB (2009) Biopsy of pheochromocytomas and paragangliomas: potential for disaster. Surgery 146(6):1158–1166
- 137. Mamede M, Carrasquillo JA, Chen CC, Del Corral P, Whatley M, Ilias I, Ayala A, Pacak K (2006) Discordant localization of 2-[18F]-fluoro-2-deoxy-D-glucose in 6-[18F]-fluorodopamine- and [(123)I]-metaiodobenzylguanidine-negative metastatic pheochromocytoma sites. Nucl Med Commun 27(1):31–36
- 138. Havekes B, King K, Lai EW, Romijn JA, Corssmit EP, Pacak K (2010) New imaging approaches to phaeochromocytomas and paragangliomas. Clin Endocrinol (Oxf) 72(2):137–145
- 139. Shen WT, Sturgeon C, Clark OH, Duh QY, Kebebew E (2004) Should pheochromocytoma size influence surgical approach? A comparison of 90 malignant and 60 benign pheochromocytomas. Surgery 136(6):1129–1137



- 140. Neumann HP, Reincke M, Bender BU, Elsner R, Janetschek G (1999) Preserved adrenocortical function after laparoscopic bilateral adrenal sparing surgery for hereditary pheochromocytoma. J Clin Endocrinol Metab 84(8):2608–2610
- 141. Walther MM, Herring J, Choyke PL, Linehan WM (2000) Laparoscopic partial adrenalectomy in patients with hereditary forms of pheochromocytoma. J Urol 164(1):14–17
- 142. Li ML, Fitzgerald PA, Price DC, Norton JA (2001) Iatrogenic pheochromocytomatosis: a previously unreported result of laparoscopic adrenalectomy. Surgery 130(6):1072–1077
- 143. Rabii R, Cicco A, Salomon L, Hoznek A, Chopin DK, Abbou CC (2001) Laparoscopic excision of para-aortic lymphatic metastasis of malignant pheochromocytoma. Ann Urol (Paris) 35(2):81–83
- 144. Henry JF, Sebag F, Iacobone M, Mirallie E (2002) Results of laparoscopic adrenalectomy for large and potentially malignant tumors. World J Surg 26(8):1043–1047
- 145. Solorzano CC, Lew JI, Wilhelm SM, Sumner W, Huang W, Wu W, Montano R, Sleeman D, Prinz RA (2007) Outcomes of pheochromocytoma management in the laparoscopic era. Ann Surg Oncol 14(10):3004–3010
- 146. Safford SD, Coleman RE, Gockerman JP, Moore J, Feldman JM, Leight GS Jr, Tyler DS, Olson JA Jr (2003) Iodine-131 metaiodobenzylguanidine is an effective treatment for malignant pheochromocytoma and paraganglioma. Surgery 134(6):956– 962, discussion 962–953
- 147. Coleman RE, Stubbs JB, Barrett JA, de la Guardia M, Lafrance N, Babich JW (2009) Radiation dosimetry, pharmacokinetics, and safety of ultratrace iobenguane I-131 in patients with malignant pheochromocytoma/paraganglioma or metastatic carcinoid. Cancer Biother Radiopharm 24(4):469–475
- 148. Huang H, Abraham J, Hung E, Averbuch S, Merino M, Steinberg SM, Pacak K, Fojo T (2008) Treatment of malignant pheochromocytoma/paraganglioma with cyclophosphamide, vincristine, and dacarbazine: recommendation from a 22-year follow-up of 18 patients. Cancer 113(8):2020–2028
- 149. Van Slycke S, Caiazzo R, Pigny P, Cardot-Bauters C, Arnalsteen L, D'Herbomez M, Leteurtre E, Rouaix-Emery N, Ernst O, Huglo D, Vantyghem MC, Wemeau JL, Carnaille B, Pattou F (2009) Local-regional recurrence of sporadic or syndromic abdominal extra-adrenal paraganglioma: incidence, characteristics, and outcome. Surgery 146(6):986–992
- Adler JT, Meyer-Rochow GY, Chen H, Benn DE, Robinson BG, Sippel RS, Sidhu SB (2008) Pheochromocytoma: current approaches and future directions. Oncologist 13(7):779–793
- Salmenkivi K, Heikkila P, Haglund C, Arola J (2004) Malignancy in pheochromocytomas. APMIS 112(9):551–559
- Belkin A, Macqueen DG, Duffin JD (1954) Malignant pheochromocytoma of adrenal. Can Med Assoc J 71(1):59–60
- Mahoney EM, Harrison JH (1977) Malignant pheochromocytoma: clinical course and treatment. J Urol 118(2):225–229
- 154. Favier J, Plouin PF, Corvol P, Gasc JM (2002) Angiogenesis and vascular architecture in pheochromocytomas: distinctive traits in malignant tumors. Am J Pathol 161(4):1235–1246
- 155. Wangberg B, Muth A, Khorram-Manesh A, Jansson S, Nilsson O, Forssell-Aronsson E, Tisell L, Ahlman H (2006) Malignant pheochromocytoma in a population-based study: survival and clinical results. Ann N Y Acad Sci 1073:512–516
- 156. Huang KH, Chung SD, Chen SC, Chueh SC, Pu YS, Lai MK, Lin WC (2007) Clinical and pathological data of 10 malignant pheochromocytomas: long-term follow up in a single institute. Int J Urol 14(3):181–185
- Tischler AS (2008) Pheochromocytoma and extra-adrenal paraganglioma: updates. Arch Pathol Lab Med 132(8):1272–1284
- 158. Linnoila RI, Keiser HR, Steinberg SM, Lack EE (1990) Histopathology of benign versus malignant sympathoadrenal

- paragangliomas: clinicopathologic study of 120 cases including unusual histologic features. Hum Pathol 21(11):1168–1180
- 159. van der Harst E, Bruining HA, Jaap Bonjer H, van der Ham F, Dinjens WN, Lamberts SW, de Herder WW, Koper JW, Stijnen T, Proye C, Lecomte-Houcke M, Bosman FT, de Krijger RR (2000) Proliferative index in phaeochromocytomas: does it predict the occurrence of metastases? J Pathol 191(2):175–180
- 160. Lam KY, Lo CY, Wat NM, Luk JM, Lam KS (2001) The clinicopathological features and importance of p53, Rb, and mdm2 expression in phaeochromocytomas and paragangliomas. J Clin Pathol 54(6):443–448
- 161. Khorram-Manesh A, Ahlman H, Nilsson O, Friberg P, Oden A, Stenstrom G, Hansson G, Stenquist O, Wangberg B, Tisell LE, Jansson S (2005) Long-term outcome of a large series of patients surgically treated for pheochromocytoma. J Intern Med 258 (1):55–66
- 162. Strong VE, Kennedy T, Al-Ahmadie H, Tang L, Coleman J, Fong Y, Brennan M, Ghossein RA (2008) Prognostic indicators of malignancy in adrenal pheochromocytomas: clinical, histopathologic, and cell cycle/apoptosis gene expression analysis. Surgery 143(6):759–768
- 163. Wu D, Tischler AS, Lloyd RV, DeLellis RA, de Krijger R, van Nederveen F, Nose V (2009) Observer variation in the application of the pheochromocytoma of the adrenal gland scaled score. Am J Surg Pathol 33(4):599–608
- 164. Hayry V, Salmenkivi K, Arola J, Heikkila P, Haglund C, Sariola H (2009) High frequency of SNAIL-expressing cells confirms and predicts metastatic potential of phaeochromocytoma. Endocr Relat Cancer 16(4):1211–1218
- 165. Nativ O, Grant CS, Sheps SG, O'Fallon JR, Farrow GM, van Heerden JA, Lieber MM (1992) Prognostic profile for patients with pheochromocytoma derived from clinical and pathological factors and DNA ploidy pattern. J Surg Oncol 50(4):258– 262
- 166. Thompson LD (2002) Pheochromocytoma of the adrenal gland scaled score (PASS) to separate benign from malignant neoplasms: a clinicopathologic and immunophenotypic study of 100 cases. Am J Surg Pathol 26(5):551–566
- 167. Kimura N, Watanabe T, Noshiro T, Shizawa S, Miura Y (2005) Histological grading of adrenal and extra-adrenal pheochromocytomas and relationship to prognosis: a clinicopathological analysis of 116 adrenal pheochromocytomas and 30 extra-adrenal sympathetic paragangliomas including 38 malignant tumors. Endocr Pathol 16(1):23–32
- 168. Gao B, Meng F, Bian W, Chen J, Zhao H, Ma G, Shi B, Zhang J, Liu Y, Xu Z (2006) Development and validation of pheochromocytoma of the adrenal gland scaled score for predicting malignant pheochromocytomas. Urology 68(2):282–286
- 169. Gupta D, Shidham V, Holden J, Layfield L (2000) Prognostic value of immunohistochemical expression of topoisomerase alpha II, MIB-1, p53, E-cadherin, retinoblastoma gene protein product, and HER-2/neu in adrenal and extra-adrenal pheochromocytomas. Appl Immunohistochem Mol Morphol 8(4):267– 274
- 170. Wilhelm SM, Prinz RA, Barbu AM, Onders RP, Solorzano CC (2006) Analysis of large versus small pheochromocytomas: operative approaches and patient outcomes. Surgery 140 (4):553–559, discussion 559–560
- 171. Agarwal A, Mehrotra PK, Jain M, Gupta SK, Mishra A, Chand G, Agarwal G, Verma AK, Mishra SK, Singh U (2010) Size of the tumor and pheochromocytoma of the adrenal gland scaled score (PASS): can they predict malignancy? World J Surg 34 (12):3022–3028
- 172. Carlsen E, Abdullah Z, Kazmi SM, Kousparos G (2009) Pheochromocytomas, PASS, and immunohistochemistry. Horm Metab Res 41(9):715–719



- 173. Boltze C, Mundschenk J, Unger N, Schneider-Stock R, Peters B, Mawrin C, Hoang-Vu C, Roessner A, Lehnert H (2003) Expression profile of the telomeric complex discriminates between benign and malignant pheochromocytoma. J Clin Endocrinol Metab 88(9):4280–4286
- 174. August C, August K, Schroeder S, Bahn H, Hinze R, Baba HA, Kersting C, Buerger H (2004) CGH and CD 44/MIB-1 immunohistochemistry are helpful to distinguish metastasized from nonmetastasized sporadic pheochromocytomas. Mod Pathol 17(9):1119–1128
- 175. Lloyd RV, Blaivas M, Wilson BS (1985) Distribution of chromogranin and S100 protein in normal and abnormal adrenal medullary tissues. Arch Pathol Lab Med 109(7):633–635
- 176. Unger P, Hoffman K, Pertsemlidis D, Thung S, Wolfe D, Kaneko M (1991) S100 protein-positive sustentacular cells in malignant and locally aggressive adrenal pheochromocytomas. Arch Pathol Lab Med 115(5):484–487
- 177. Achilles E, Padberg BC, Holl K, Kloppel G, Schroder S (1991) Immunocytochemistry of paragangliomas—value of staining for S-100 protein and glial fibrillary acid protein in diagnosis and prognosis. Histopathology 18(5):453–458
- 178. Brown HM, Komorowski RA, Wilson SD, Demeure MJ, Zhu YR (1999) Predicting metastasis of pheochromocytomas using DNA flow cytometry and immunohistochemical markers of cell proliferation: a positive correlation between MIB-1 staining and malignant tumor behavior. Cancer 86(8):1583–1589
- 179. Clarke MR, Weyant RJ, Watson CG, Carty SE (1998) Prognostic markers in pheochromocytoma. Hum Pathol 29(5):522–526
- 180. Edstrom E, Mahlamaki E, Nord B, Kjellman M, Karhu R, Hoog A, Goncharov N, Teh BT, Backdahl M, Larsson C (2000) Comparative genomic hybridization reveals frequent losses of chromosomes 1p and 3q in pheochromocytomas and abdominal paragangliomas, suggesting a common genetic etiology. Am J Pathol 156(2):651–659
- 181. Dannenberg H, Speel EJ, Zhao J, Saremaslani P, van Der Harst E, Roth J, Heitz PU, Bonjer HJ, Dinjens WN, Mooi WJ, Komminoth P, de Krijger RR (2000) Losses of chromosomes 1p and 3q are early genetic events in the development of sporadic pheochromocytomas. Am J Pathol 157(2):353–359
- 182. Lui WO, Chen J, Glasker S, Bender BU, Madura C, Khoo SK, Kort E, Larsson C, Neumann HP, Teh BT (2002) Selective loss of chromosome 11 in pheochromocytomas associated with the VHL syndrome. Oncogene 21(7):1117–1122
- 183. Dannenberg H, de Krijger RR, Zhao J, Speel EJ, Saremaslani P, Dinjens WN, Mooi WJ, Roth J, Heitz PU, Komminoth P (2001) Differential loss of chromosome 11q in familial and sporadic parasympathetic paragangliomas detected by comparative genomic hybridization. Am J Pathol 158(6):1937–1942
- 184. Sandgren J, Diaz de Stahl T, Andersson R, Menzel U, Piotrowski A, Nord H, Backdahl M, Kiss NB, Brauckhoff M, Komorowski J, Dralle H, Hessman O, Larsson C, Akerstrom G, Bruder C, Dumanski JP, Westin G (2010) Recurrent genomic alterations in benign and malignant pheochromocytomas and paragangliomas revealed by wholegenome array comparative genomic hybridization analysis. Endocr Relat Cancer 17(3):561–579
- 185. Korpershoek E, Stobbe CK, van Nederveen FH, de Krijger RR, Dinjens WN (2010) Intra-tumoral molecular heterogeneity in benign and malignant pheochromocytomas and extra-adrenal sympathetic paragangliomas. Endocr Relat Cancer 17(3):653– 662
- 186. Waldmann J, Fendrich V, Holler J, Buchholz M, Heinmoller E, Langer P, Ramaswamy A, Samans B, Walz MK, Rothmund M, Bartsch DK, Slater EP (2010) Microarray analysis reveals differential expression of benign and malignant pheochromocytoma. Endocr Relat Cancer 17(3):743–756

- 187. Thouennon E, Elkahloun AG, Guillemot J, Gimenez-Roqueplo AP, Bertherat J, Pierre A, Ghzili H, Grumolato L, Muresan M, Klein M, Lefebvre H, Ouafik L, Vaudry H, Plouin PF, Yon L, Anouar Y (2007) Identification of potential gene markers and insights into the pathophysiology of pheochromocytoma malignancy. J Clin Endocrinol Metab 92(12):4865–4872
- 188. Petri BJ, Speel EJ, Korpershoek E, Claessen SM, van Nederveen FH, Giesen V, Dannenberg H, van der Harst E, Dinjens WN, de Krijger RR (2008) Frequent loss of 17p, but no p53 mutations or protein overexpression in benign and malignant pheochromocytomas. Mod Pathol 21(4):407–413
- 189. Castilla-Guerra L, Moreno AM, Fernandez-Moreno MC, Utrilla JC, Fernandez E, Galera-Davison H (1997) Expression and prognostic value of c-erbB-2 oncogene product in human phaeochromocytomas. Histopathology 31(2):144–149
- Kubota Y, Nakada T, Sasagawa I, Yanai H, Itoh K (1998)
 Elevated levels of telomerase activity in malignant pheochromocytoma. Cancer 82(1):176–179
- 191. Isobe K, Yashiro T, Omura S, Kaneko M, Kaneko S, Kamma H, Tatsuno I, Takekoshi K, Kawakami Y, Nakai T (2004) Expression of the human telomerase reverse transcriptase in pheochromocytoma and neuroblastoma tissues. Endocr J 51 (1):47–52
- 192. Kinoshita H, Ogawa O, Mishina M, Oka H, Okumura K, Yamabe H, Terachi T, Yoshida O (1998) Telomerase activity in adrenal cortical tumors and pheochromocytomas with reference to clinicopathologic features. Urol Res 26(1):29–32
- 193. Bamberger CM, Else T, Bamberger AM, Frilling A, Beil FU, Allolio B, Schulte HM (1999) Telomerase activity in benign and malignant adrenal tumors. Exp Clin Endocrinol Diabetes 107 (4):272–275
- 194. Portel-Gomes GM, Grimelius L, Johansson H, Wilander E, Stridsberg M (2001) Chromogranin A in human neuroendocrine tumors: an immunohistochemical study with region-specific antibodies. Am J Surg Pathol 25(10):1261–1267
- 195. Blank A, Schmitt AM, Korpershoek E, van Nederveen F, Rudolph T, Weber N, Strebel RT, de Krijger R, Komminoth P, Perren A (2010) SDHB loss predicts malignancy in pheochromocytomas/sympathethic paragangliomas, but not through hypoxia signalling. Endocr Relat Cancer 17(4):919–928
- 196. Meyer-Rochow GY, Jackson NE, Conaglen JV, Whittle DE, Kunnimalaiyaan M, Chen H, Westin G, Sandgren J, Stalberg P, Khanafshar E, Shibru D, Duh QY, Clark OH, Kebebew E, Gill AJ, Clifton-Bligh R, Robinson BG, Benn DE, Sidhu SB (2010) MicroRNA profiling of benign and malignant pheochromocytomas identifies novel diagnostic and therapeutic targets. Endocr Relat Cancer 17(3):835–846
- 197. Sandgren J, Andersson R, Rada-Iglesias A, Enroth S, Akerstrom G, Dumanski JP, Komorowski J, Westin G, Wadelius C (2010) Integrative epigenomic and genomic analysis of malignant pheochromocytoma. Exp Mol Med 42(7):484–502
- 198. Korpershoek E, Loonen AJ, Corvers S, van Nederveen FH, Jonkers J, Ma X, Ziel-van der Made A, Korsten H, Trapman J, Dinjens WN, de Krijger RR (2009) Conditional Pten knock-out mice: a model for metastatic phaeochromocytoma. J Pathol 217 (4):597–604
- 199. Molatore S, Liyanarachchi S, Irmler M, Perren A, Mannelli M, Ercolino T, Beuschlein F, Jarzab B, Wloch J, Ziaja J, Zoubaa S, Neff F, Beckers J, Hofler H, Atkinson MJ, Pellegata NS (2010) Pheochromocytoma in rats with multiple endocrine neoplasia (MENX) shares gene expression patterns with human pheochromocytoma. Proc Natl Acad Sci USA 107(43):18493–18498
- 200. Martiniova L, Lai EW, Elkahloun AG, Abu-Asab M, Wickremasinghe A, Solis DC, Perera SM, Huynh TT, Lubensky IA, Tischler AS, Kvetnansky R, Alesci S, Morris JC, Pacak K (2009) Characterization of an animal model of aggressive metastatic



- pheochromocytoma linked to a specific gene signature. Clin Exp Metastasis 26(3):239-250
- 201. Tischler AS (2008) Pheochromocytoma: time to stamp out "malignancy"? Endocr Pathol 19(4):207–208
- Andersen KF, Altaf R, Krarup-Hansen A, Kromann-Andersen B, Horn T, Christensen NJ, Hendel HW (2011) Malignant pheochromocytomas and paragangliomas—the importance of a multidisciplinary approach. Cancer Treat Rev 37(2):111–119
- 203. Van Der Horst-Schrivers AN, Jager PL, Boezen HM, Schouten JP, Kema IP, Links TP (2006) Iodine-123 metaiodobenzylguani-dine scintigraphy in localising phaeochromocytomas—experience and meta-analysis. Anticancer Res 26(2B):1599–1604
- Shulkin BL, Thompson NW, Shapiro B, Francis IR, Sisson JC (1999) Pheochromocytomas: imaging with 2-[fluorine-18]fluoro-2-deoxy-D-glucose PET. Radiology 212(1):35–41
- 205. Taieb D, Sebag F, Hubbard JG, Mundler O, Henry JF, Conte-Devolx B (2004) Does iodine-131 meta-iodobenzylguanidine (MIBG) scintigraphy have an impact on the management of sporadic and familial phaeochromocytoma? Clin Endocrinol (Oxf) 61(1):102–108
- 206. Maurea S, Cuocolo A, Reynolds JC, Neumann RD, Salvatore M (1996) Diagnostic imaging in patients with paragangliomas. Computed tomography, magnetic resonance and MIBG scintig-raphy comparison. Q J Nucl Med 40(4):365–371
- 207. Tavangar SM, Shojaee A, Moradi Tabriz H, Haghpanah V, Larijani B, Heshmat R, Lashkari A, Azimi S (2010) Immunohistochemical expression of Ki67, c-erbB-2, and c-kit antigens in benign and malignant pheochromocytoma. Pathol Res Pract 206 (5):305–309
- 208. Waldmann J, Slater EP, Langer P, Buchholz M, Ramaswamy A, Walz MK, Schmid KW, Feldmann G, Bartsch DK, Fendrich V (2009) Expression of the transcription factor SNAIL and its target gene twist are associated with malignancy in pheochromocytomas. Ann Surg Oncol 16(7):1997–2005
- 209. Yuan W, Wang W, Cui B, Su T, Ge Y, Jiang L, Zhou W, Ning G (2008) Overexpression of ERBB-2 was more frequently detected in malignant than benign pheochromocytomas by multiplex ligation-dependent probe amplification and immunohistochemistry. Endocr Relat Cancer 15(1):343–350
- 210. Gimm O, Krause U, Brauckhoff M, Hoang-Vu C, Dralle H (2006) Distinct expression of galectin-3 in pheochromocytomas. Ann N Y Acad Sci 1073:571–577

- 211. Guillemot J, Barbier L, Thouennon E, Vallet-Erdtmann V, Montero-Hadjadje M, Lefebvre H, Klein M, Muresan M, Plouin PF, Seidah N, Vaudry H, Anouar Y, Yon L (2006) Expression and processing of the neuroendocrine protein secretogranin II in benign and malignant pheochromocytomas. Ann N Y Acad Sci 1073:527–532
- 212. Portela-Gomes GM, Stridsberg M, Grimelius L, Falkmer UG, Falkmer S (2004) Expression of chromogranins A, B, and C (secretogranin II) in human adrenal medulla and in benign and malignant pheochromocytomas: an immunohistochemical study with region-specific antibodies. APMIS 112(10):663–673
- 213. Salmenkivi K, Heikkila P, Liu J, Haglund C, Arola J (2003) VEGF in 105 pheochromocytomas: enhanced expression correlates with malignant outcome. APMIS 111(4):458–464
- Khorram-Manesh A, Ahlman H, Jansson S, Nilsson O (2002) Ncadherin expression in adrenal tumors: upregulation in malignant pheochromocytoma and downregulation in adrenocortical carcinoma. Endocr Pathol 13(2):99–110
- 215. Salmenkivi K, Haglund C, Ristimaki A, Arola J, Heikkila P (2001) Increased expression of cyclooxygenase-2 in malignant pheochromocytomas. J Clin Endocrinol Metab 86(11):5615– 5619
- Salmenkivi K, Haglund C, Arola J, Heikkila P (2001) Increased expression of tenascin in pheochromocytomas correlates with malignancy. Am J Surg Pathol 25(11):1419–1423
- 217. Ohji H, Sasagawa I, Iciyanagi O, Suzuki Y, Nakada T (2001) Tumour angiogenesis and Ki-67 expression in phaeochromocytoma. BJU Int 87(4):381–385
- 218. de Krijger RR, van der Harst E, van der Ham F, Stijnen T, Dinjens WN, Koper JW, Bruining HA, Lamberts SW, Bosman FT (1999) Prognostic value of p53, bcl-2, and c-erbB-2 protein expression in phaeochromocytomas. J Pathol 188(1):51–55
- 219. Moreno AM, Castilla-Guerra L, Martinez-Torres MC, Torres-Olivera F, Fernandez E, Galera-Davidson H (1999) Expression of neuropeptides and other neuroendocrine markers in human phaeochromocytomas. Neuropeptides 33 (2):159–163
- 220. Liu J, Voutilainen R, Kahri AI, Heikkila P (1997) Expression patterns of the c-myc gene in adrenocortical tumors and pheochromocytomas. J Endocrinol 152(2):175–181

