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Novel dimeric DOTA-coupled peptidic Y₁-receptor antagonists for targeting of neuropeptide Y receptor-expressing cancers

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Abstract

Background: Several peptide hormone receptors were identified that are specifically over-expressed on the cell surface of certain human tumors. For example, high incidence and density of the Y₁ subtype of neuropeptide Y (NPY) receptors are found in breast tumors. Recently, we demonstrated that the use of potent radiolabeled somatostatin or bombesin receptor antagonists considerably improved the sensitivity of *in vivo* imaging when compared to agonists. We report here on the first DOTA-coupled peptidic Y₁ receptor affine dimer antagonists.

Methods: Based on a Y₁ affine dimeric peptide scaffold previously reported to competitively antagonize NPY-mediated processes, we have developed new dimeric DOTA-coupled Y₁ receptor affine antagonists for scintigraphy and radiotherapy. These dimeric peptides were tested for their specific binding to Y₁ expressed in SK-N-MC cells and Y₂ expressed in SH-SY5Y as well as for their ability to mediate cAMP production in SK-N-MC cells.

Results: Introduction of two DOTA moieties at the N-termini of the dimeric NPY analogs as well as the double Asn²⁹ replacement by Dpr(DOTA) or Lys(DOTA) (**6** and **10**) moiety dramatically reduced binding affinity. However, asymmetric introduction of the DOTA moiety in one segment of the peptidic heterodimer (**8** and **11**) resulted in suitable antagonists for receptor targeting with high binding affinity for Y₁. All compounds were devoid of Y₂ binding affinity.

Conclusions: The design and the *in vitro* characterization of the first DOTA-coupled dimeric NPY receptor antagonist with high affinity and selectivity for Y₁ over Y₂ are described. This compound may be an excellent candidate for the imaging of Y₁-positive tumors and their treatment.

Keywords: neuropeptide Y receptor, tumor imaging, oncology, peptide receptor radionuclide therapy, breast cancer, antagonist

Background

Peptide hormone receptors play an increasing role in cancer medicine. This role is based primarily on the peptide receptor over-expression on tumor cells which allows a specific receptor-targeted scintigraphic tumor imaging and tumor therapy with radiolabeled peptide analogs [1]. The somatostatin receptors were the first peptide receptors identified for these purposes, and somatostatin receptor targeting has now become an integral part of the routine management of patients with gastroenteropancreatic

neuroendocrine tumors. Somatostatin receptor scintigraphy (OctreoScan[®], Covidien Ltd., St. Louis, MO, USA) detects these tumors with extremely high sensitivity and specificity [2]. Moreover, recent results from clinical studies involving somatostatin receptor radionuclide therapy of these tumors are very promising [2]. The last decade has seen the development of numerous novel somatostatin agonists suitable for tumor targeting [3,4]. Interestingly, however, it has recently been shown that potent somatostatin receptor antagonists, known to poorly internalize into tumor cells, can visualize tumors *in vivo* as well, or even better than the corresponding agonists [5]. This unexpected phenomenon was found both for sst₂- and sst₃-selective somatostatin analogs, and may be due to the

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binding of the antagonist to a larger number of sites and to its lower dissociation rate. A pilot clinical trial with radiolabeled DOTA-linked sst₂ antagonists recently confirmed the animal data [6].

Prompted by the success of somatostatin receptor targeting, the over-expression of other peptide receptor families was evaluated in tumors *in vivo* [1]. Promising new candidates for *in vivo* peptide receptor targeting of tumors are neuropeptide Y (NPY) receptors, based on their high expression in specific cancers, in particular breast carcinomas [7,8]. In humans, four NPY receptor subtypes exist, called Y₁, Y₂, Y₄, and Y₅ [9]. The natural ligands for these receptors are the peptides of the NPY family, including the neurotransmitter NPY and the two gut hormones peptide YY (PYY) and pancreatic polypeptide (PP). Via binding to the NPY receptors, these peptides regulate a wide variety of physiologic functions such as digestion, vasoconstriction, and reproduction. They also play a key role in eating behavior [10]. On this basis, Y₂ and Y₄ receptor agonists and Y₁ and Y₅ receptor antagonists have become potential drugs against overweight that are currently evaluated for this application [11]. On the other hand, since Y₁ and Y₂ receptors are highly over-expressed in breast cancer, Ewing sarcomas, neuroblastomas, and high-grade gliomas [7,12-14], the use of radiolabeled Y₁ and Y₂ receptor ligands for an NPY receptor-targeted imaging and radiotherapy of these tumors, was suggested [1,15]. It is worth mentioning that in those tumors, the presence of a significant amount of Y₄ or Y₅ receptors was not observed [8]. A daunorubicin-coupled cytotoxic NPY analog [16], a Y₂-selective, ^{99m}Tc-labeled radioactive NPY analog [17], and more recently, a ^{99m}Tc-labeled Y₁ agonist were developed [18]. Preliminary clinical data on Y₁-targeted tumor imaging with the latter compound in breast cancer patients are encouraging [18].

There is no information yet on Y₁ receptor antagonists coupled to a chelator, which are suitable for *in vivo* receptor targeting. The aim of the present study was to design and develop dimeric NPY analogs coupled to the DOTA chelator for suitable radiolabeling that could be used for imaging and for radiotherapy of Y₁-expressing tumors. We report here on the first dimeric DOTA-coupled peptidic NPY receptor antagonists with high affinity for Y₁.

Methods

Reagents

All Boc-N^α-protected amino acids with side chain protection: Arg(Tos), Asn(Xan), Cys(Acm), Tyr(2-Br-Z) were commercially available (Bachem Inc., Torrance, CA, USA). All reagents were of best grade available and were purchased from common suppliers. Silver trifluoromethanesulfonate was obtained from Sigma-Aldrich (St.

Louis, MO, USA). DOTA-NHS was from Macrocyclics (Dallas, TX, USA). The adenylate cyclase activation flash-plate assay (SMP004) was from PerkinElmer (Waltham, MA, USA).

Peptide synthesis

Each segment of the dimeric peptides was synthesized manually on a methylbenzhydrylamine (MBHA) resin using the solid phase approach and the Boc- strategy. Noteworthy, an orthogonally protected cysteine, i.e., Boc-Cys(Acm)-OH, was used to prevent dimerization of inadequate segment. Main chain assembly was mediated by diisopropylcarbodiimide (DIC) and coupling completion (45 to 60 min) was assessed by Kaiser's test. Three-fold excess of protected amino acid and DIC was used based on the original substitution of the MBHA resin (0.4 mmol.g⁻¹) and Boc removal was achieved *via* TFA-mediated deprotection (50% in dichloromethane (DCM); 10 to 15 min). An isopropyl alcohol (1% *m-cresol*) wash followed TFA treatment and then successive washes with triethylamine (TEA) solution (10% in DCM), methanol, TEA solution, methanol, and DCM completed the neutralization sequence. In order to facilitate the specific linkage of the DOTA moiety in the peptide dimers, the N-terminal amino acid of the sequence was introduced as an Fmoc derivative except when the DOTA moiety was coupled to the N-terminus. Peptide resins were then treated with anhydrous HF in the presence of anisole (5% to 10%, *v/v*) at 0°C for 1.5 h to liberate the Cys(Acm)-protected crude peptides. After elimination of HF under vacuum, the crude peptides were washed with peroxide-free diethyl ether and extracted with 0.1% TFA in 60% acetonitrile/water. After lyophilization, the orthogonally protected peptides were purified using preparative RP-HPLC and two successive solvent systems (eluent A: TEAP at pH 2.25 and 0.1% TFA, eluent B: 60% acetonitrile/A). The purified peptides were characterized by analytical RP-HPLC and MALDI-TOF-MS on a Voyager DE-STR in the reflector mode using the α -cyano-4-hydroxycinnamic acid as matrix. Conjugation of the DOTA derivative was achieved prior to the disulfide bond formation as previously reported [19]. Briefly, a solution of DOTA-NHS ester (2 eq) in DMF and N, N'-diisopropylethylamine (DIPEA) (3 eq) were added to the monomer solution in dry DMF. The mixture was stirred at room temperature and the progress of the reaction was followed by analytical RP-HPLC. After completion of the reaction, generally observed after 3 h, a preparative RP-HPLC purification was performed yielding the DOTA-conjugated analog. Homogeneity of each fraction was assessed by analytical RP-HPLC. Removal of the Acm group was achieved by silver trifluoromethanesulfonate (100 eq/Acm) treatment of each monomer (dissolved in TFA/anisole; 99:1, 1 mg/mL) at 4°C for 2 h.

The peptide silver salt was then precipitated with diethyl ether and separated by centrifugation. Dimers were obtained by the treatment of two identical monomers (homodimer) or two different monomers (heterodimer) in equimolar concentration with aqueous 1 M HCl/DMSO (1:1) overnight at room temperature resulting in the removal of the silver ions as AgCl and disulfide bond formation. Following the filtration of silver chloride, dimeric peptides were once again purified and analyzed as described above. The structure of the dimeric NPY scaffold as well as the structure of the incorporated substitutions is shown in Figure 1.

Cell lines

The neuroepithelioma cell line SK-N-MC endogenously expressing the NPY Y₁ receptor was obtained from ATCC (HTB-10; LGC Standards, Teddington, Middlesex, UK) and cultured at 37°C and 5% CO₂ in MEM with GlutaMax I and supplemented with 10% FBS, 100 U/ml

penicillin and 100 µg/ml streptomycin, 1 mM MEM sodium pyruvate, and MEM non-essential amino acids (× 1). The neuroblastoma cell line SH-SY5Y endogenously expressing the NPY Y₂ receptor, kindly provided by Dr. Paolo Paganetti (Novartis, Basel, Switzerland), was cultured at 37°C and 5% CO₂ in MEM/Ham's F12 with GlutaMax I and supplemented with 10% FBS, 100 U/ml penicillin and 100 µg/ml streptomycin, 1 mM MEM sodium pyruvate, and MEM non-essential amino acids (× 1). All culture reagents were from Gibco BRL, Life Technologies, (Grand Island, NY, USA).

Receptor autoradiography

Binding affinities of the compounds were assessed using sections of cell membrane pellets of SK-N-MC for Y₁ or SH-SY5Y for Y₂, as described before [7,13,20]. Briefly, membrane pellets were prepared and stored at -80°C, and receptor autoradiography was performed on 20-µm-thick cryostat (Microm HM 500, Walldorf, Germany)

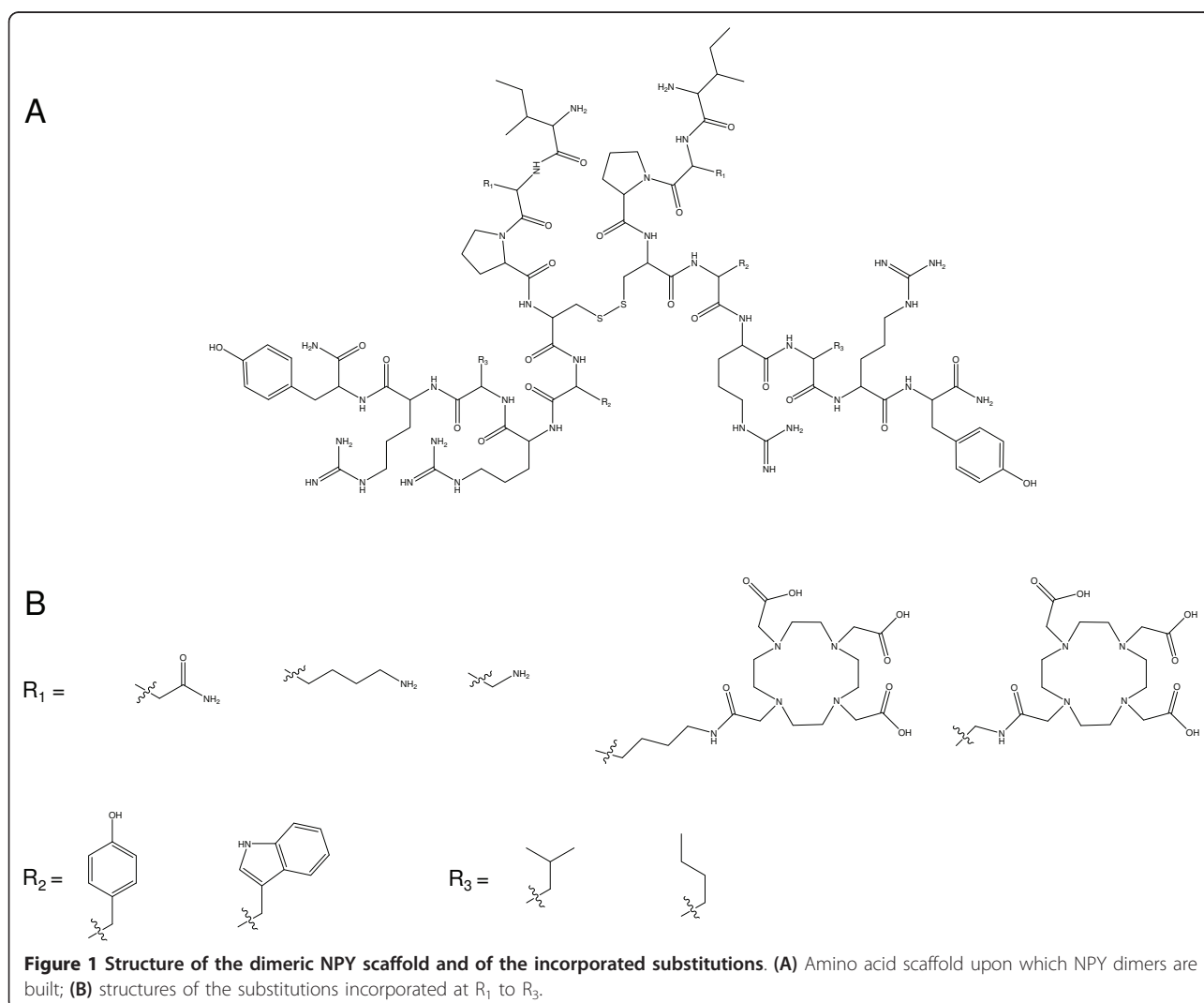


Figure 1 Structure of the dimeric NPY scaffold and of the incorporated substitutions. (A) Amino acid scaffold upon which NPY dimers are built; **(B)** structures of the substitutions incorporated at R₁ to R₃.

sections of the membrane pellets, mounted on microscope slides and stored at -20°C as previously described for other cell lines and receptors [21]. The slides were preincubated in Krebs-Ringer solution (NaCl 119 mM, KCl 3.2 mM, KH_2PO_4 1.19 mM, MgSO_4 1.19 mM, NaHCO_3 25 mM, CaCl_2 2.53 mM, D-glucose 10 mM; pH 7.4) for 60 min at room temperature. Then, they were incubated for 120 min in the incubation solution containing the Krebs-Ringer solution, 0.1% BSA, 0.05% bacitracin, and 10,000 cpm/100 μl of the ^{125}I -labeled human PYY (hPYY; 2,000 Ci/mmol; Anawa, Wangen, Switzerland). Membrane pellet sections were incubated with ^{125}I -hPYY in increasing concentrations ranging from 0.1 nM up to 1,000 nM of non-labeled hPYY, as control, or with the compounds to be tested. After the incubation, the slides were washed two times for 5 min and then rinsed four times in ice-cold preincubation solution. After drying, the slides were exposed to Kodak films Biomax MR[®] for 7 days. IC_{50} values were calculated after quantification of the data using a computer-assisted image processing system (Analysis Imaging System, Interfocus, Mering, Germany). Tissue standards (autoradiographic [^{125}I] and/or [^{14}C] microscales, GE Healthcare, Little Chalfont, UK) containing a known amount of isotope, cross-calibrated to tissue-equivalent ligand concentrations were used for quantification [22].

Adenylate cyclase activity

Adenylate cyclase activity was determined in SK-N-MC cells using the adenylate cyclase activation flashplate assay (SMP004) from PerkinElmer (Waltham, MA, USA). SK-N-MC cells were seeded in 96-well culture plates at 25,000 cells/well and cultured for 48 h at 37°C and 5% CO_2 . Culture medium was then removed from the wells and fresh medium (100 μL) containing 0.5 mM 3-isobutyl-1-methylxanthine (IBMX) was added to each well. Cells were incubated for 30 min at 37°C . Medium was then removed and replaced with fresh medium containing 0.5 mM IBMX, with or without 10 μM forskolin and various concentrations of the peptides to be analyzed. Cells were incubated for 30 min at 37°C . After removal of the medium, cells were lysed and cAMP accumulation was determined using the SMP004 kit from PerkinElmer according to the instructions of the manufacturers.

Results

Rational design of dimeric Y_1 receptor affine peptide antagonists

In order to generate and identify competitive and Y_1 receptor affine antagonists suitable for breast cancer targeting, we took advantage of previously reported structure-activity relationship studies. Using a known dimeric Y_1 -selective antagonist (analog **1**, Table 1) as our template [23], we first looked at modifications supposedly

increasing the antagonistic property and/or selectivity of NPY analogs. As a matter of fact, the replacement of Tyr³² (NPY numbering) by a Trp and introduction of a hydrophobic and bulky residue such as norleucine (Nle) in position 34 were shown to increase Y_1 receptor-binding affinity [24]. The DOTA-conjugated counterparts of peptides **1** and **3** (Table 1) were then obtained through the addition of the chelator moiety at the N-termini of the peptide dimers (analog **2** and **4**, Table 1). Other sites for the introduction of the chelating derivative were also considered. Based on previously reported structure-activity relationship studies, only the residue at position 29, i.e., Asn, was tolerant to amino acid replacement [25,26]. The C-terminal part of NPY, i.e., regions 32 to 36, is known to be directly involved in Y_1 -receptor binding [26,27] and was therefore not considered in our study. Similarly, the proline residue at position 30 is essential for the antagonistic behavior of such analogs and was consequently not replaced [28]. Nevertheless, we investigated the possibility to replace the proline residue by a cis or trans amino-proline moiety to which DOTA could be coupled. However, this substitution resulted in a significant reduction of binding affinity (data not shown). Thus, we replaced Asn²⁹ with a diaminopropionic (Dpr) (**5**) or a lysine (Lys) residue (**9**), and attached the DOTA moiety to the β or ϵ amino group of these residues, respectively. Analog **6** and **10**, with two chelators, exhibited lower binding affinities than their parent peptides (Table 2). These two amino acids, varying only by the number of carbon in their side chain, will also help us investigate the impact of the spatial proximity between the DOTA moiety and the peptide backbone on the biological activity. Addition of DOTA derivative was often followed by a concomitant reduction of binding affinity, most probably due to the steric hindrance of such residue, as exemplified with somatostatin analogs [29]. With no understanding on how the conjugation of the chelator on each segment of the peptide dimer will impact the binding affinity, selectivity, and biological activity, we investigated the possibility to generate heterodimer bearing only one DOTA derivative (analog **8** and **11**, Table 1) thus keeping the other peptide segment intact to ensure high binding affinity. Each purified monomer was usually obtained in a 15% to 25% yield calculated on the base of the substitution of the starting resin. After completion of the DOTA-conjugation and the removal of the Fmoc protecting group (if necessary), the purified monomeric DOTA-conjugated derivatives were generally obtained in a 25% to 30% yield based on the starting material. Finally, following the dimerization and the subsequent purification steps, the final products were isolated in a 10% to 15% yield based on the equimolar concentration of the starting materials. In total, 11 peptides were synthesized, purified and analyzed by RP-HPLC, CZE and MALDI-TOF

Table 1 Physicochemical properties of free and DOTA-coupled dimeric peptide NPY analogs

Analog number	Amino acid residues					Purity (%)		MS ^c	
	Xa ²⁹	Xa ^{29'}	Xb	Xc	N-term DOTA	HPLC ^a	CZE ^b	calc	obs
1	Asn	Asn	Tyr	Leu	No	98	98	2,389.2	2,390.3
2	Asn	Asn	Tyr	Leu	Yes	97	96	3,161.6	3,162.6
3	Asn	Asn	Trp	Nle	No	94	90	2,435.2	2,436.5
4	Asn	Asn	Trp	Nle	Yes	95	92	3,207.6	3,208.8
5	Dpr	Dpr	Trp	Nle	No	99	96	2,379.3	2,380.1
6	Dpr(DOTA)	Dpr(DOTA)	Trp	Nle	No	94	93	3,151.6	3,152.4
7	Dpr	Asn	Trp	Nle	No	99	99	2,407.3	2,408.5
8	Dpr(DOTA)	Asn	Trp	Nle	No	98	97	2,793.5	2,794.9
9	Lys	Lys	Trp	Nle	No	88	84	2,463.4	2,464.5
10	Lys(DOTA)	Lys(DOTA)	Trp	Nle	No	92	96	3,236.3	3,236.8
11	Lys (DOTA)	Asn	Trp	Nle	No	83 ^d	83	2,834.5	2,835.3

^aPercentage purity determined by HPLC using buffer system: A = TEAP (pH 2.5) and B = 60% CH₃CN/40% A with a gradient slope of 1% B/min, at flow rate of 0.2 mL/min on a Vydac C₁₈ column (0.21 cm × 5 cm, 5-μm particle size, 300-Å pore size). Detection at 214 nm. ^bPercentage purity determined by capillary zone electrophoresis (CZE) using a Beckman P/ACE System 2050; field strength of 15 kV at 30°C. Buffer, 100 mM sodium phosphate (85:15, H₂O:CH₃CN), pH 2.50, on a Agilent μSil bare fused-silica capillary (75 μm i.d. × 40 cm length). Detection at 214 nm. ^cMALDI mass spectral analysis (m/z). The observed [M + H]⁺ of the monoisotopic mass (obs) compared with the calculated m/z of the monoisotope (calc). ^dThe detectable contaminant, produced during the dimerization process of this heterodimer, was identified as analog 3.

Table 2 Binding affinities at NPY Y₁- and Y₂-receptors and Y₁-related functional characteristics

Analog number	Binding affinity ^a		Functional assay cAMP for Y ₁
	Y ₁	Y ₂	
DOTA-free analogs			
1	11 ± 7	> 1,000	Antagonist
3	9.0 ± 3	> 1,000	Antagonist
5	143 ± 20	> 1,000	ND
7	19 ± 5	> 1,000	Antagonist
9	127 ± 50	> 1,000	Antagonist
DOTA-coupled analogs			
2	143 ± 37	> 1,000	Antagonist
4	294 ± 33	> 1,000	Antagonist
6	> 1000	> 1,000	ND
8	29 ± 7	> 1,000	Antagonist
10	283 ± 52	> 1,000	Antagonist
11	13 ± 3	> 1,000	Antagonist

Functional characteristics for free and DOTA-coupled dimeric peptide NPY analogs. ^aIC₅₀ values in nanomolars; mean ± SEM; n ≥ 3. ND, not determined.

spectrometry. Data are listed in Table 1. The structure of a DOTA-free homodimeric (**3**), a DOTA-conjugated homodimeric (**10**) and a DOTA-conjugated heterodimeric (**11**) Y₁ receptor affine antagonist are shown in Figure 2.

Binding affinity profile

The DOTA-free and DOTA-coupled analogs listed in Table 1 were analyzed in receptor autoradiography experiments for NPY Y₁ and Y₂ receptor-binding affinities on SK-N-MC cells endogenously expressing Y₁ and SH-SY5Y cells endogenously expressing Y₂, respectively (Table 2). Pharmacological displacement experiment using SK-N-MC cell membrane pellet sections for compounds **9**, **10**, and **11** are shown in Figure 3. The IC₅₀ values for all tested compounds are listed in Table 2. The addition of two DOTA moieties to the homodimeric analogs does decrease the Y₁ binding affinity up to 2- to 30-fold. However, the addition of only one DOTA to heterodimer **7** did markedly improve the binding affinity to an IC₅₀ of 29 nM (**8**), as compared to the symmetric, bi-DOTA-linked parent scaffold (**6**) showing an IC₅₀ over 1,000 nM. Finally, increasing the distance between the chelator and the peptide backbone, i.e., analog **11** (Figure 2), enhanced the binding affinity (IC₅₀ = 13 ± 3 nM; Table 2) of this heterodimer for the Y₁ receptor. None of the tested compounds showed Y₂ binding affinity.

In vitro forskolin-stimulated adenylate cyclase activity

Since the addition of a DOTA moiety can change the functional characteristics of a compound as recently shown in the somatostatin receptor field for sst₃ [19], where a sst₃ antagonist switched to an agonist upon the addition of a DOTA, compounds having a high or

moderate Y₁ affinity were analyzed in an adenylate cyclase activity assay for their agonistic or antagonistic properties. The results are shown in Figure 4. While the Y₁-selective agonist [Leu³¹, Pro³⁴]-hPYY, used as positive control, efficiently inhibited forskolin-stimulated cAMP accumulation when applied at concentrations of 20 μM and 100 nM, all tested compounds, DOTA-free and DOTA-coupled analogs behaved like full antagonists (Figure 4; Table 2). Given alone at a high concentration of 20 μM the analogs were not able to inhibit forskolin-stimulated cAMP accumulation but they efficiently antagonized the agonistic effect of 100 nM [Leu³¹, Pro³⁴]-hPYY. Moreover, Figure 5 shows that 20 μM of **11**, the best compound of this series, given together with an increasing concentration of [Leu³¹, Pro³⁴]-hPYY in the range from 10 nM up to 20 μM is able to shift by at least three orders of magnitude the dose response curve of [Leu³¹, Pro³⁴]-hPYY to the right, indicating that **11** efficiently antagonizes the agonist effect of [Leu³¹, Pro³⁴]-hPYY. Thus, these DOTA-conjugated heterodimers, i.e., analogs **8** and **11**, with their high binding affinity and antagonist property represent potential candidates for *in vivo* tumor targeting.

Discussion

The recent demonstration that radiolabeled antagonists considerably improved the sensitivity of *in vivo* diagnostic procedures and might improve the efficacy of receptor-mediated radiotherapy suggested the generalized use of peptide antagonists rather than agonists for *in vivo* tumor detection [5,30]. However, selective peptide antagonists suitable for radiolabeling are not available for each receptor candidate and therefore they need to be developed. The high density and incidence of Y₁ receptors in invasive and metastatic breast cancers, also expressing Y₂ but not Y₄ and Y₅ receptors, make these neoplasms important targets for diagnosis and therapy with NPY-related drugs [8]. Thus, the aim of the present study was to design Y₁ receptor affine antagonists suitable for molecular imaging.

As demonstrated, NPY exists in equilibrium between monomer and dimers in aqueous solution [31]. These dimeric structures were found to be not mandatory for the binding of NPY to Y₂ receptors but were responsible for the increased Y₁-affinity and selectivity. Proof of this concept was supported by the synthesis and the characterization of several dimeric NPY analogs retaining high Y₁-affinity and selectivity over Y₂ [23,25]. We thus derived a conceptual and experimental approach to the design of new dimeric Y₁ receptor antagonists using the previously described dimeric scaffold, i.e., Bis(31/31') [Pro³⁰, Cys³¹, Tyr³², Leu³⁴]NPY(28-36)-NH₂ (analog **1**), formed by the covalent linkage of two identical entities mimicking the C-terminal region of the full length NPY through a disulfide bridge between Cys moiety at

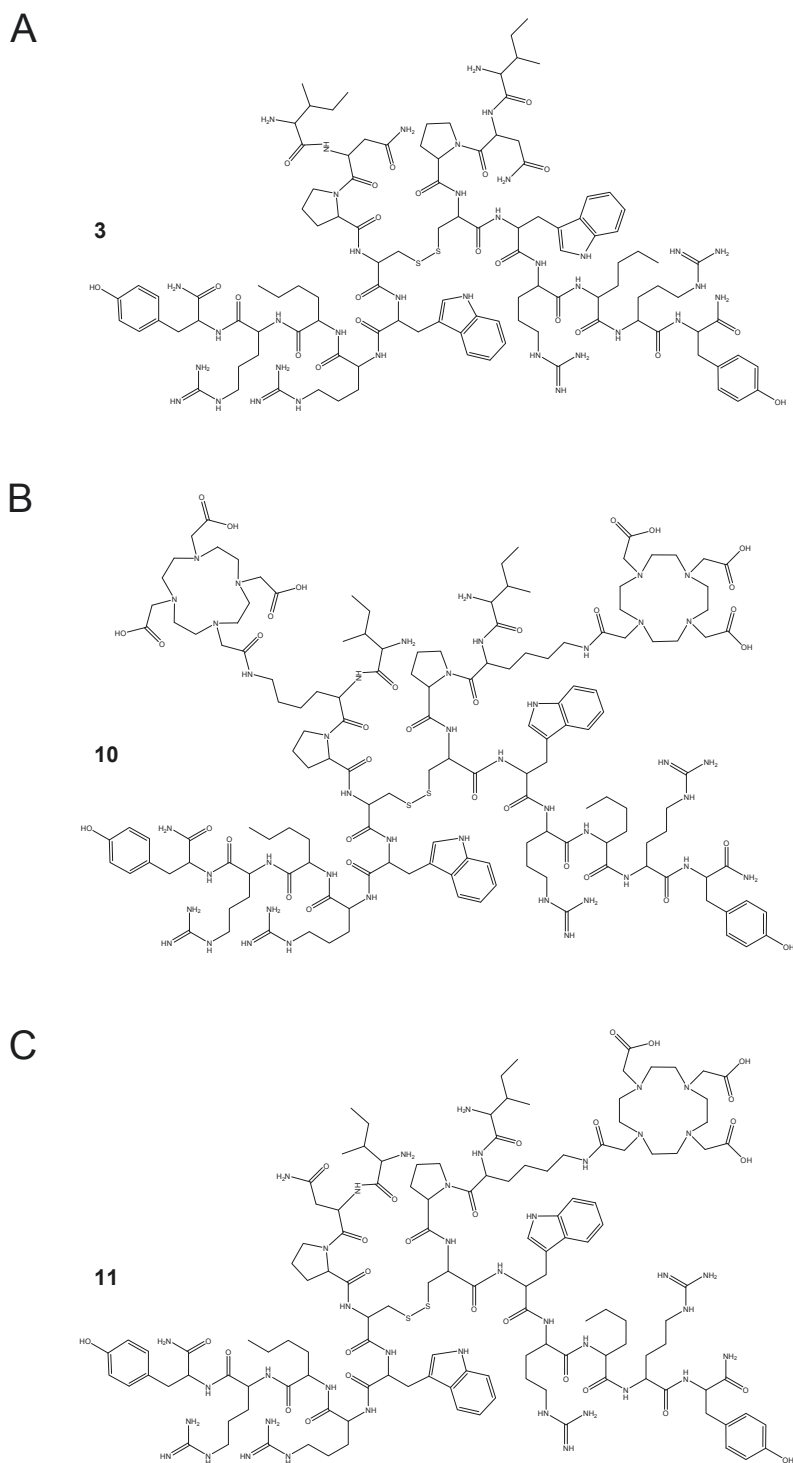
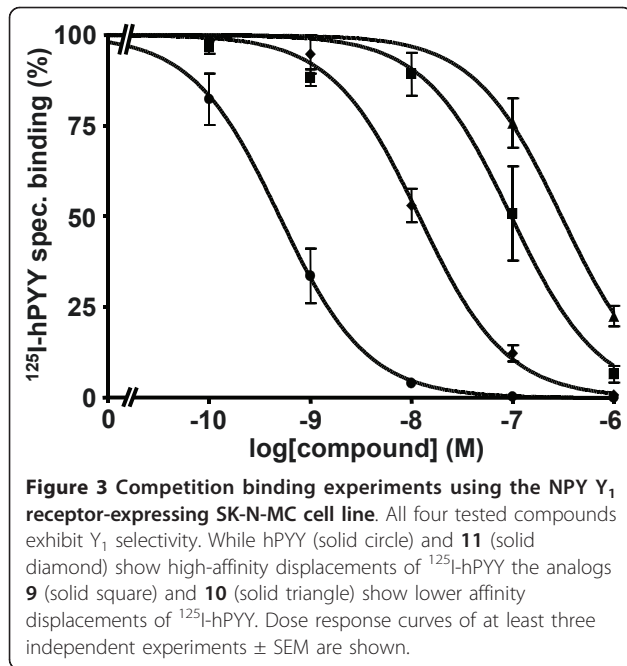


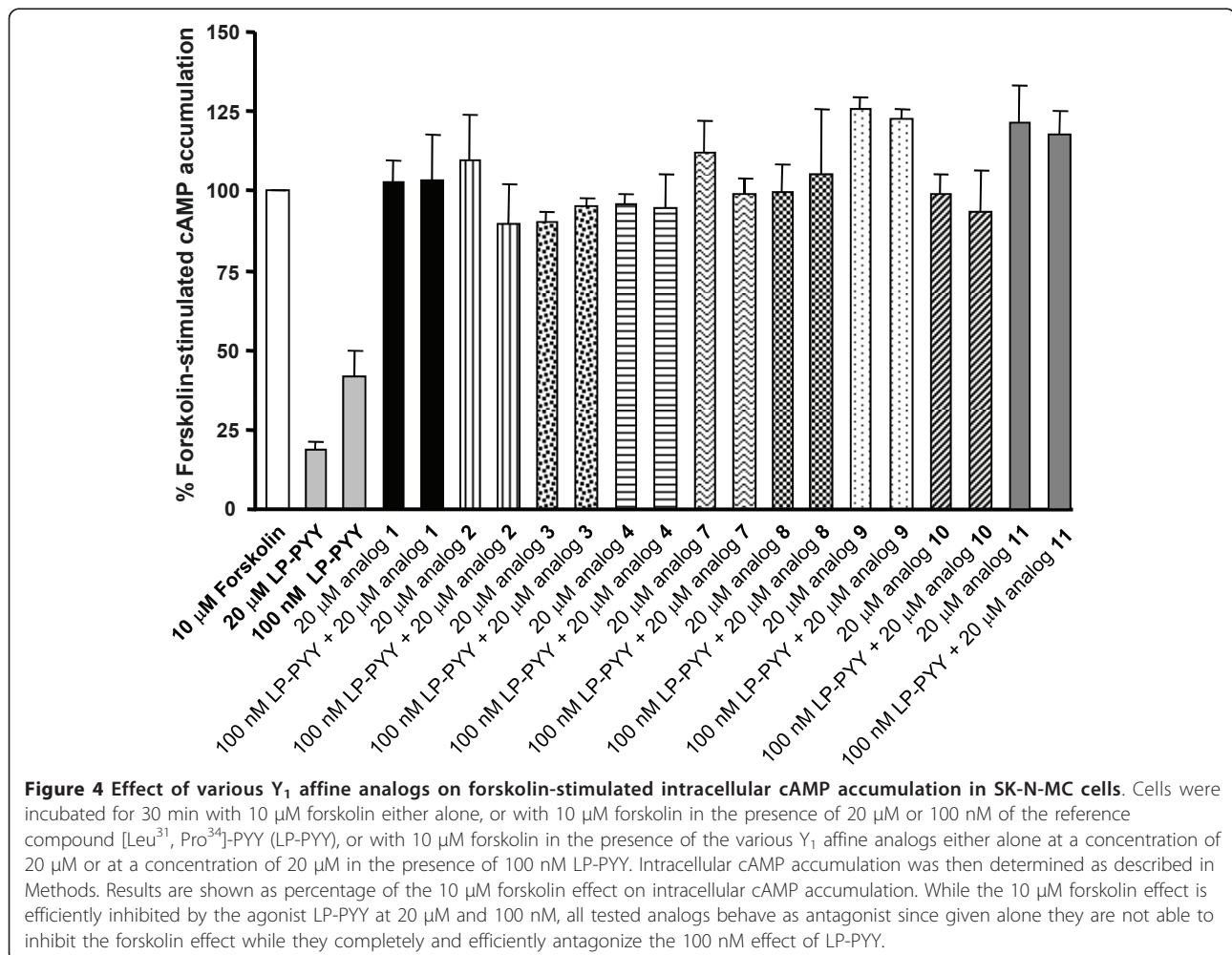
Figure 2 Structure of the DOTA-free dimeric Y_1 receptor affine antagonist (**3**) and its DOTA-conjugated dimeric counterparts. Structure of (A) the DOTA-free dimeric Y_1 receptor affine antagonist (**3**) and its DOTA-conjugated dimeric counterparts (B) **10** and (C) **11**.

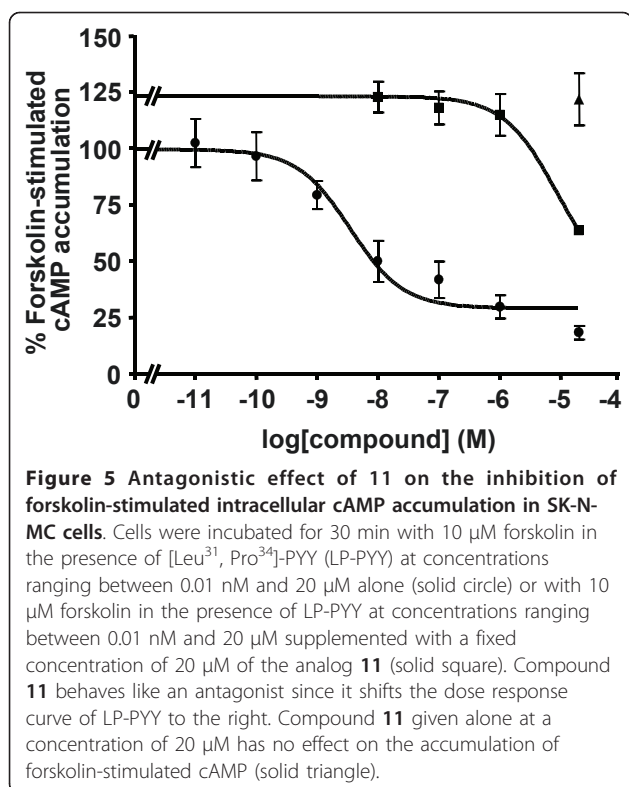
position 31 [32]. As supported by a previous report, replacement of Tyr³² by Trp and Leu³⁴ by Nle (**3**) did not produce a significant change in binding affinity or selectivity. However, adjunction of DOTA moieties to

the N-termini of these dimeric peptides resulted in a dramatic loss of binding affinity (**2** and **4**) suggesting the importance of the N-terminus for receptor recognition. In accordance, evidence demonstrating that N-terminal



acylation might compromise receptor affinity and selectivity were already reported in the literature [33,34]. Such observation prompted us to evaluate different sites for the introduction of the DOTA. Thus, introduction of the DOTA moiety was achieved through its selective addition to the β or ϵ side chain amino group of a Dpr or Lys residue, respectively, introduced at position 29 which was found to be tolerant to amino acid replacement [25,26]. However, such substitution in each segment of the peptide dimers (**6** and **10**) resulted in a loss of Y_1 receptor-binding affinity compared to their DOTA-free counterparts (**5** and **9**). Most likely, this reduction of affinity is probably due to the size and the negatively charged character of the DOTA moiety. As observed by NMR in BVD15, a monomer segment analogous to our peptide dimer, the Arg³³, which was found to be particularly involved in the Y_1 receptor recognition [27], and Asn²⁹ side chains seem to be spatially closed [35]. It is thus probable that coupling the DOTA moiety to the Dpr residue might disrupt the orientation or the character of the positively charged





Arg residue and thus alter the overall binding affinity. Supporting this hypothesis, increasing the distance between the DOTA moiety and the peptide backbone, i. e., analogs **6** and **10**, resulted in a less dramatic loss of binding affinity probably by reducing the electrostatic effect of the carboxyl groups of the DOTA moiety on the guanidinium derivative. The higher binding affinity of homodimeric NPY peptides was often related to the propensity of NPY receptors to form homodimers as recently demonstrated [36]. As such, by keeping intact one segment, original interaction between heterodimers (**8** and **11**) and their receptor was restored; these two compounds being almost equipotent compared to the lead precursor (**1**). Although preliminary in nature, our results, and more particularly those of analog **11**, represent the first step towards the development of dimeric DOTA-coupled Y₁ receptor antagonist for nuclear medicine application. Even if the stability of these analogs has not been evaluated in the present study, these compounds should prove to be sufficiently stable for tumor targeting purpose. Indeed, Bis(31/31')[Cys³¹, Nva³⁴]NPY (27-36)-NH₂, presenting a high sequence and scaffold homology with our compounds was able to stimulate *in vivo* the food intake in rats in a similar manner than that observed by other non-dimeric Y₅ selective agonists [37].

In analogy to somatostatin receptor targeting of tumors, it has been proposed to use NPY analogs to target NPY receptors for tumor therapy. NPY analogs

suitable for this purpose have indeed already been developed, such as a daunorubicin-coupled cytotoxic NPY analogs [16] and a Y₂-selective ^{99m}Tc-labeled radioactive NPY analog [17], all being agonists. Assuming that the observation of the superiority of somatostatin receptor antagonists, but also of bombesin receptor antagonists [30] for tumor targeting can be generalized, the present DOTA-coupled high-affinity Y₁ receptor antagonist may be a useful tool for the diagnostic and radiotherapeutic targeting of Y₁-expressing tumors. Breast tumors with their high Y₁-receptor density would represent first choice candidate tumors. Other tumor types, such as renal cell carcinomas, ovarian cancers, adrenal tumors and embryonic tumors, may also be targets of interest. The same general principles as for somatostatin receptor targeting could be applied. Advantages that should be put forward are a more favorable benefit-toxicity profile compared with conventional radio- or chemotherapy and the rarity of side effects. The radiotargeting of NPY receptor-expressing tumor blood vessels alone or together with NPY receptor-expressing tumor cells may also represent an attractive strategy for therapy. Finally, since many of the NPY receptor-expressing tumors can express multiple peptide receptors concomitantly, NPY receptors may be suitable for a multireceptor targeting with a cocktail containing NPY and other therapeutic peptide analogs directed against various peptide hormone receptors. For such a multireceptor approach, good candidate tumors seem to be breast tumors targeted with NPY and bombesin analogs.

Conclusions

In the course of designing and synthesizing DOTA-coupled dimeric NPY Y₁ receptor antagonists, we found that the addition of a DOTA moiety to both peptide segments negatively influenced the binding affinity of all dimeric compounds synthesized and that the asymmetric introduction of a DOTA to one of the segments of the peptide heterodimer yields compounds exhibiting high binding affinity and Y₁-selectivity over Y₂. Analog **8** and **11** are the first dimeric high-affinity DOTA-coupled Y₁ receptor antagonists. They may, when linked to an adequate radiometal, become useful tools for *in vivo* tumor targeting of Y₁-positive tumors, particularly breast tumors.

Abbreviations

The abbreviations for the common amino acids are in accordance with the recommendations of [38]. Additional abbreviations: AcM: acetamidomethyl; Boc: *tert*-butoxycarbonyl; BSA: bovine serum albumin; cAMP: 3',5'-cyclic adenosine monophosphate; ¹⁴C: ¹⁴Carbon; CZE: capillary zone electrophoresis; DCM: dichloromethane; DIC: N, N'-diisopropylcarbodiimide; DIPEA: N, N'-diisopropylethylamine; DMF: dimethylformamide; DMSO: dimethylsulfoxide; DOTA: 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid; Dpr: 2,3-diaminopropionic acid; FBS: fetal bovine serum; Fmoc: 9-

fluorenylmethoxycarbonyl; IBMX: 3-isobutyl-1-methylxanthine; ¹²⁵I: ¹²⁵Iodine; MALDI-TOF-MS: matrix assisted laser desorption/ionization-time off light mass spectrometry; MBHA: 4-methylbenzhydrylamine; MEM: minimum essential medium; NHS: N-hydroxysuccinimide; NMR: nuclear magnetic resonance; Nle: norleucine; NPY: neuropeptide Y; Nva: norvaline; PP: pancreatic polypeptide; PYY: peptide YY; RP-HPLC: reversed-phase high-performance liquid chromatography; TEA: triethylamine; TEAP: triethylammonium phosphate; TFA: trifluoroacetic acid.

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DC: carried out the synthesis of the peptides and participated in the manuscript draft.

RC: carried out the functional assays.

BW: carried out the binding assays.

JE: carried out the synthesis of the peptides and reviewed the chemistry data. JER: participated in the conception, design and coordination of the study, and reviewed the chemistry data.

JCR: participated in the conception, design and coordination of the study, and in the interpretation of the data. He reviewed the draft manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Reubi JC: Peptide receptors as molecular targets for cancer diagnosis and therapy. *Endocr Rev* 2003, **24**:389-427.
2. Maecke HR, Reubi JC: Somatostatin receptors as targets for nuclear medicine imaging and radionuclide treatment. *J Nucl Med* 2011, **52**:841-4.
3. Ginj M, Zhang H, Eisenwiener KP, Wild D, Schulz S, Rink H, Cescato R, Reubi JC, Maecke HR: New pansomatostatin ligands and their chelated versions: affinity profile, agonist activity, internalization, and tumor targeting. *Clin Cancer Res* 2008, **14**:2019-27.
4. Wild D, Schmitt JS, Ginj M, Macke HR, Bernard BF, Krenning E, De Jong M, Wenger S, Reubi JC: DOTA-NOC, a high-affinity ligand of somatostatin receptor subtypes 2, 3 and 5 for labelling with various radiometals. *Eur J Nucl Med Mol Imaging* 2003, **30**(10):1338-47.
5. Ginj M, Zhang H, Waser B, Cescato R, Wild D, Wang X, Erchegyi J, Rivier J, Mäcke HR, Reubi JC: Radiolabeled somatostatin receptor antagonists are preferable to agonists for in vivo peptide receptor targeting of tumors. *Proc Natl Acad Sci USA* 2006, **103**:16436-41.
6. Wild D, Fani M, Behe M, Brink I, Rivier J, Reubi JC, Maecke HR, Weber WA: First clinical evidence that imaging with somatostatin receptor antagonists is clinically feasible. *J Nucl Med* 2011.
7. Reubi JC, Gugger M, Waser B, Schaer JC: Y1-mediated effect of neuropeptide Y in cancer: breast carcinomas as targets. *Cancer Res* 2001, **61**:4636-41.
8. Korner M, Reubi JC: NPY receptors in human cancer: a review of current knowledge. *Peptides* 2007, **28**:419-25.
9. Michel MC, Beck-Sickinger A, Cox H, Doods HN, Herzog H, Larhammar D, Quirion R, Schwartz T, Westfall T: XVI. International union of pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. *Pharmacol Rev* 1998, **50**:143-50.
10. Balasubramaniam A: Clinical potentials of neuropeptide Y family of hormones. *Am J Surg* 2002, **183**:430-4.
11. Kamiji MM, Inui A: Neuropeptide Y receptor selective ligands in the treatment of obesity. *Endocr Rev* 2007, **28**:664-84.
12. Korner M, Waser B, Reubi JC: Neuropeptide Y receptors in renal cell carcinomas and nephroblastomas. *Int J Cancer* 2005, **115**:734-41.
13. Korner M, Waser B, Reubi JC: High expression of neuropeptide Y1 receptors in ewing sarcoma tumors. *Clin Canc Res* 2008, **14**:5043-49.
14. Korner M, Reubi JC: Neuropeptide Y receptors in primary human brain tumors: overexpression in high-grade tumors. *J Neuropathol Exp Neurol* 2008, **67**:741-49.
15. Khan IU, Beck-Sickinger AG: Targeted tumor diagnosis and therapy with peptide hormones as radiopharmaceuticals. *Anticancer Agents Med Chem* 2008, **8**:186-99.
16. Langer M, Kratz F, Rothen-Rutishauser B, Wunderli-Allenspach H, Beck-Sickinger AG: Novel peptide conjugates for tumor-specific chemotherapy. *J Med Chem* 2001, **44**:1341-48.
17. Langer M, La Bella R, Garcia-Garayoa E, Beck-Sickinger AG: 99 mTc-Labeled Neuropeptide Y Analogues as Potential Tumor Imaging Agents. *Bioconjug Chem* 2001, **12**:1028-34.
18. Khan IU, Zwanziger D, Bohme I, Javed M, Naseer H, Hyder SW, Beck-Sickinger AG: Breast-cancer diagnosis by neuropeptide Y analogues: from synthesis to clinical application. *Angew Chem Int Ed Engl* 2010, **49**:1155-8.
19. Reubi JC, Erchegyi J, Cescato R, Waser B, Rivier JE: Switch from antagonist to agonist after addition of a DOTA chelator to a somatostatin analog. *Eur J Nucl Med Mol Imaging* 2010, **37**:1551-8.
20. Korner M, Waser B, Reubi JC: Neuropeptide Y receptor expression in human primary ovarian neoplasms. *Lab Invest* 2004, **84**:71-80.
21. Reubi JC, Schaer JC, Waser B, Wenger S, Heppeler A, Schmitt J, Mäcke HR: Affinity profiles for human somatostatin receptor sst1-sst5 of somatostatin radiotracers selected for scintigraphic and radiotherapeutic use. *Eur J Nucl Med* 2000, **27**:273-82.
22. Erchegyi J, Cescato R, Grace CR, Waser B, Piccand V, Hoyer D, Riek R, Rivier JE, Reubi JC: Novel, potent, and radio-iodinatable somatostatin receptor 1 (sst1) selective analogues. *J Med Chem* 2009, **52**:2733-46.
23. Balasubramaniam A, Dhawan VC, Mullins DE, Chance WT, Sherif S, Guzzi M, Prabhakaran M, Parker EM: Highly selective and potent neuropeptide Y (NPY) Y1 receptor antagonists based on [Pro(30), Tyr(32), Leu(34)]NPY (28-36)-NH2 (BW1911U90). *J Med Chem* 2001, **44**:1479-82.
24. Balasubramaniam A, Zhai W, Sherif S, Tao Z, Chance WT, Fischer JE, Eden P, Taylor J: Bis(31/31') ([Cys(31), Trp(32), Nva(34)] NPY-(31-36)): a specific NPY Y-1 receptor antagonist. *J Med Chem* 1996, **39**:811-3.
25. Daniels AJ, Matthews JE, Slepatis RJ, Jansen M, Viveros OH, Tademalli A, Harrington W, Heyer D, Landavazo A, Leban JJ: High-affinity neuropeptide Y receptor antagonists. *Proc Natl Acad Sci USA* 1995, **92**: 9067-71.
26. Beck-Sickinger AG, Wieland HA, Wittneben H, Willim KD, Rudolf K, Jung G: Complete L-alanine scan of neuropeptide Y reveals ligands binding to Y1 and Y2 receptors with distinguished conformations. *Eur J Biochem* 1994, **225**:947-58.
27. Merten N, Lindner D, Rabe N, Rompler H, Morl K, Schoneberg T, Beck-Sickinger AG: Receptor subtype-specific docking of Asp6.59 with C-terminal arginine residues in Y receptor ligands. *J Biol Chem* 2007, **282**:7543-51.
28. Leban JJ, Heyer D, Landavazo A, Matthews J, Aulabaugh A, Daniels AJ: Novel modified carboxy terminal fragments of neuropeptide Y with high affinity for Y2-type receptors and potent functional antagonism at a Y1-type receptor. *J Med Chem* 1995, **38**:1150-7.
29. Reubi JC, Schaer JC, Wenger S, Hoeger C, Erchegyi J, Waser B, Rivier J: sst3-selective potent peptidic somatostatin receptor antagonists. *Proc Natl Acad Sci USA* 2000, **97**:13973-78.
30. Cescato R, Maina T, Nock B, Nikolopoulou A, Charalambidis D, Piccand V, Reubi JC: Bombesin receptor antagonists may be preferable to agonists for tumor targeting. *J Nucl Med* 2008, **49**:318-26.
31. Cowley DJ, Hoflack JM, Pelton JT, Saudek V: Structure of neuropeptide Y dimer in solution. *Eur J Biochem* 1992, **205**:1099-106.
32. Daniels AJ, Heyer D, Landavazo O, Leban JK, Spaltenstein A: Neuropeptide Y antagonists 1994, WO94/00486.
33. Murase S, Yumoto N, Petukhov MG, Yoshikawa S: Acylation of the alpha-amino group in neuropeptide Y(12-36) increases binding affinity for the Y2 receptor. *J Biochem* 1996, **119**:37-41.

34. Guerin B, Dumulon-Perreault V, Tremblay MC, Ait-Mohand S, Fournier P, Dubuc C, Authier S, Bénard F: [Lys(DOTA)4]BVD15, a novel and potent neuropeptide Y analog designed for Y1 receptor-targeted breast tumor imaging. *Bioorg Med Chem Lett* 2010, **20**:950-3.
35. Jois SD, Balasubramaniam A: Conformation of neuropeptide Y receptor antagonists: structural implications in receptor selectivity. *Peptides* 2003, **24**:1035-43.
36. Dinger MC, Bader JE, Kobor AD, Kretzschmar AK, Beck-Sickingler AG: Homodimerization of neuropeptide y receptors investigated by fluorescence resonance energy transfer in living cells. *J Biol Chem* 2003, **278**:10562-71.
37. Balasubramaniam A, Sheriff S, Zhai W, Chance WT: Bis(31/31')[[Cys(31), Nva(34)]NPY(27-36)-NH(2)]: a neuropeptide Y (NPY) Y(5) receptor selective agonist with a latent stimulatory effect on food intake in rats. *Peptides* 2002, **23**:1485-90.
38. IUPAC-IUB Joint Commission on Biochemical Nomenclature: Nomenclature and symbolism for amino acids and peptides. *Eur J Biochem* 1984, **138**:9-37.

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