

# Ecuzumab hepatotoxicity in pediatric aHUS

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Received: 26 June 2013 / Revised: 25 August 2014 / Accepted: 9 October 2014  
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## Abstract

**Background** Ecuzumab is a humanized anti-C5 antibody approved for the treatment of atypical hemolytic uremic syndrome (aHUS). Its use is increasing in children following reports of its safety and efficacy.

**Methods** We reviewed biochemical and clinical data related to possible drug-induced liver injury in 11 children treated with ecuzumab for aHUS in a single center.

**Results** Elevated aminotransferases were observed in 7 children aged 6 to 11 years following ecuzumab treatment for aHUS. Internationally accepted liver enzyme thresholds for drug-induced liver injury were exceeded in 5 cases. In all cases, liver injury was classified as mixed hepatocellular and cholestatic. Infectious and other causes were excluded in each case. One patient with no pre-existing liver disease developed tender hepatomegaly and liver enzyme derangement exceeding 20 times the upper limit of normal following initiation of ecuzumab. Recurrent liver injury following re-challenge

with ecuzumab necessitated its discontinuation and transition to plasma therapy.

**Conclusions** Hepatotoxicity in association with ecuzumab is a potentially important yet previously unreported adverse event. We recommend monitoring liver enzymes in all patients receiving ecuzumab. Further research is required to clarify the impact of this adverse event, to characterize the mechanism of potential hepatotoxicity, and to identify which patients are most at risk.

**Keywords** Atypical hemolytic uremic syndrome · Thrombotic microangiopathy · Complement · Ecuzumab · Transaminitis · Hepatotoxicity

## Introduction

Ecuzumab is a first-in-class humanized anti-C5 antibody approved for the treatment of paroxysmal nocturnal hemoglobinuria (PNH) and atypical hemolytic uremic syndrome (aHUS) in adults and children. It was first established as a treatment for PNH, for which it is reported to be safe and effective [1–3]. Its use in treating aHUS is increasing in children following case series and recent treatment trials reporting safety and efficacy in inducing thrombotic microangiopathy (TMA) response [4–8].

In treatment trials of aHUS, various nonspecific side effects have been reported in association with ecuzumab [8]. Other adverse events reported to date, but likely unrelated to ecuzumab treatment, include septic shock [9], photodermatitis [10], melanoma [11], and a case of desquamating rash and hyperammonemia [12]. No hepatic side effects have been reported to our knowledge.

We report a series of 5 children with aHUS who experienced hepatotoxicity following ecuzumab treatment. Two further children experienced transient liver enzyme

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derangement, which did not reach the threshold for liver injury as defined by Aithal et al. [13]. Recurrent symptoms necessitated discontinuation of eculizumab and transition to plasma therapy for aHUS in one patient. In each case, workup for infective and autoimmune causes of hepatitis was non-explanatory, including hepatitis A, B, C, Epstein-Barr virus (EBV), cytomegalovirus (CMV), parvovirus, adenovirus, HHV6, HHV7, alpha-1-antitrypsin, ceruloplasmin, copper, total immunoglobulin G (IgG), antinuclear antibodies, smooth muscle cell antibodies, and liver–kidney microsomal antibodies.

## Materials and methods

Eculizumab was dosed according to a standardized weight-based regimen. The induction schedule comprised four weekly doses of 900 mg for patients weighing 40 kg or more, and two weekly doses of 600 mg for patients weighing 30–39.9 kg. The maintenance regimen comprised 1,200 mg every 2 weeks for patients weighing 40 kg or more, and 900 mg for patients weighing 30–39.9 kg (Table 1).

Investigations for genetic mutations in complement proteins CFI, CFH, MCP/CD46, THBD/CD141, C3, and CFB and anti-CFH antibodies were performed in all patients. No known pathogenic mutations were identified. Variants of unknown significance were found in C3 and CFH in cases 2 and 4 respectively. One patient tested positive for anti-CFH antibodies (case 5).

Drug-induced liver injury was defined by internationally accepted criteria of a greater than 5-fold increase in alanine transaminase (ALT) or aspartate aminotransferase (AST) above the upper limit of normal (ULN) or an increase in hepatic alkaline phosphatase (ALK) above two times the ULN [13]. Liver injury was further classified as mixed hepatocellular and cholestatic in each case [13].

## Case reports

We observed derangement in liver enzymes in association with eculizumab therapy in 5 of the 11 patients receiving eculizumab for aHUS in our center. Transient mild liver enzyme disturbance was observed in 2 further cases. The time course of liver enzyme derangement was similar in each case

with fluctuation in ALT, AST, ALK and gamma-glutamyl transpeptidase (GGT) 10–29 days following the first eculizumab dose with resolution within 14–21 days (Fig. 1).

In 4 of the 5 affected patients, the biochemical derangement was asymptomatic and maintenance eculizumab was continued. In 1 patient (case 1), the magnitude of elevation in liver enzymes was more marked; concern about progressive elevation in transaminases and clinical symptoms of tender hepatomegaly and jaundice necessitated discontinuation of eculizumab.

The pattern of liver injury was classified as mixed hepatocellular/cholestatic in each case with R values ranging from 2.8 to 4.9 [13].

The time course of transaminitis relative to eculizumab dosing for cases 1 and 3 is illustrated in Fig. 1. Patient characteristics and patterns of liver enzyme dysfunction are summarized in Table 2.

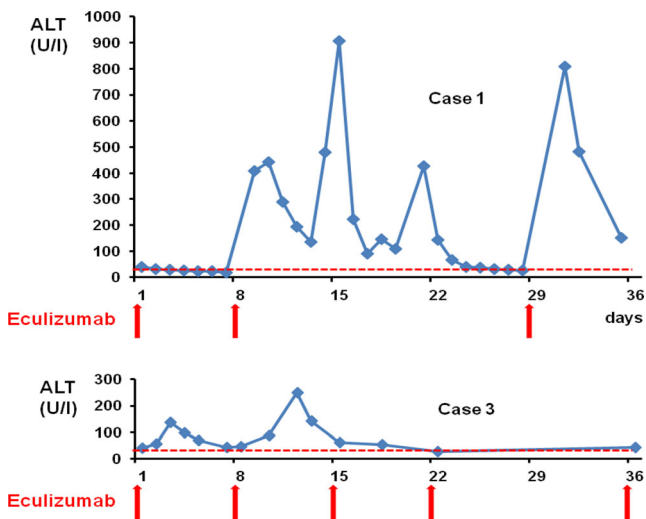
### Case 1

A boy aged 8 years and 11 months with hereditary spherocytosis and previous splenectomy developed aHUS for which eculizumab was commenced. Three days following the second dose, markedly elevated liver enzymes were noted (Fig. 1). ALT, AST, and GGT rose to over 20 times the ULN; the international normalized ratio (INR) and albumin remained within normal limits. Clinical symptoms of right upper quadrant pain and signs of tender hepatomegaly were evident. Right upper quadrant abdominal pain in the context of spherocytosis can be reflective of gall bladder disease; however, ultrasound imaging showed a normal gall bladder, mild hepatomegaly with a coarse parenchymal appearance, and normal hepatic and portal vasculature. Investigations for underlying causes of liver disease were normal, including viral hepatitis, autoimmune workup, alpha-1-antitrypsin, copper, ceruloplasmin, and creatinine phosphokinase.

Liver enzyme abnormalities fluctuated following the second dose of eculizumab with two further significant peaks in ALT (Fig. 1). A third dose was suspended pending normalization of liver enzymes. A liver biopsy taken after the second dose when liver enzymes had normalized showed mild minimal pathological change, with mild hepatocellular cytoplasmic swelling and clearing, mild Kupffer cell hemosiderosis, mild pericentral sinusoidal fibrosis, and mild sinusoidal

**Table 1** Induction and maintenance regimens according to patient weight

Patient weight (kg)	Induction regimen	Maintenance regimen
≥ 40	900 mg weekly×4	1,200 mg week 5; 1,200 mg every 2 weeks
30–39.9	600 mg weekly×2	900 mg week 3; 900 mg every 2 weeks
20–29.9	600 mg weekly×2	600 mg week 3; 600 mg every 2 weeks
10–19.9	600 mg weekly×1	300 mg week 2; 300 mg every 2 weeks
5–9.9	300 mg weekly×1	300 mg week 2; 300 mg every 3 weeks



**Fig. 1** Time course of alanine transaminase (ALT) in cases 1 and 3 in relation to eculizumab dosing. *Dashed lines* represent the upper limit of normal

dilatation. There was no evidence of autoimmune or infectious hepatitis. The presence of sinusoidal eosinophils and hepatocellular cytoplasmic changes raised concerns with regard to a drug effect.

The third eculizumab dose was then given, after which marked liver enzyme elevation recurred (Fig. 1). Eculizumab was therefore discontinued, with transition to plasma therapy. Follow-up to 4 weeks revealed no further abnormalities in liver biochemistry.

#### Case 2

A boy aged 10 years and 3 months with a background of Evans syndrome and recurrent thrombocytopenia developed marked hemolysis and thrombotic microangiopathy consistent with aHUS. Plasma exchange was initiated with partial remission, following which eculizumab was commenced. Asymptomatic elevation of ALT (maximum 175 IU/l) and AST (maximum 409 IU/l) were noted 7 days after the third eculizumab dose with spontaneous resolution over the subsequent 14 days. Whilst elevated liver enzymes can be observed in patients with Evans syndrome, such findings were not previously observed in this patient.

#### Case 3

An 11.5 year-old boy had a history of obesity, uniform hyperechogenicity of the liver parenchyma on ultrasound imaging, and normal liver biochemistry, thought to be consistent with non-alcoholic fatty liver disease. He developed aHUS and achieved remission with plasma therapy, following which he was transitioned to eculizumab. Three days following the first eculizumab dose, elevated liver enzymes were noted (Table 2). ALT rose again after a further dose but

subsequently normalized (Fig. 1). Again, elevated liver enzymes can be observed in the context of the underlying disease, but were not previously observed in this patient.

#### Case 4

A boy aged 6.5 years had severe systemic sequelae from his initial presentation with thrombotic microangiopathy including end-stage renal disease (ESRD), limb ischemia with resulting loss, ischemic colitis requiring colectomy, testicular infarct, and skin necrosis requiring grafting. Following a period of partial remission on plasma therapy, he was transitioned to eculizumab.

Fluctuating asymptomatic elevations in ALT, AST, and ALK with spontaneous resolution were noted in association with eculizumab doses. One year following initial presentation the patient developed abdominal pain with elevated liver enzymes. ERCP showed a choledochal cyst. Liver biopsy showed mild chronic eosinophilic portal inflammation with mild fibrosis and moderate iron accumulation. Transaminitis was felt to be unlikely to be related to these findings.

#### Case 5

An 8-year-old boy with no pre-existing liver disease experienced derangement in ALT and AST exceeding internationally accepted thresholds for drug-induced liver injury following initiation of eculizumab for aHUS. Liver enzyme abnormalities were not associated with specific symptoms and resolved after 56 days.

## Discussion

Hepatotoxicity in association with various biological therapies is rare but well recognized [14–25]. However, to our knowledge, this is the first report of potential hepatotoxicity in association with eculizumab therapy. Eculizumab is a humanized monoclonal anti-C5 antibody [26] that has been used for over a decade in patients with paroxysmal nocturnal hemoglobinuria (PNH) [1, 3] and aHUS [8] in patients of all age groups. The serious adverse event (SAE) profile of eculizumab is nonspecific; hepatotoxicity (including elevation in liver enzymes) has not been reported, yet was observed in two adult PNH patients, although it was clearly linked to an underlying liver disease preceding eculizumab treatment in these cases (Camille Bedrosian, personal communication, 8 October 2014). On the other hand, eculizumab was found to reverse progressive thrombosis and hepatic failure in a patient with PNH and Budd–Chiari syndrome unresponsive to anticoagulation therapy [27].

**Table 2** Patient characteristics with peak values (baseline) for alanine aminotransferase (ALT, upper limit 40 U/l), aspartate aminotransferase (AST, upper limit 45 U/l), gamma-glutamyl transpeptidase (GGT, upper limit 45 U/l), alkaline phosphatase (ALP, upper limit 180 U/l) and conjugated bilirubin (upper limit 2 μmol/l) with platelet count, LDH and haptoglobin on the day of the first eculizumab dose

Case	1	2	3	4	5
Age at disease onset	8 years 11 months	10 years	11 years	4 years 1 month	8 years 4 months
Gender	Male	Male	Male	Male	Male
Diagnosis	aHUS	aHUS	aHUS	aHUS	aHUS
Complement mutation	None identified	Homozygous variant of unknown significance in C3 gene (c.304C>G(p.Arg102Gly))	None identified	Variant of unclear significance in CFH gene (c.245-7_245-8delinsTTCA)	None identified
Anti-CFH antibodies	Pending	Negative	Pending	Pending	Positive
Concomitant disease	Congenital spherocytosis	Evans syndrome	Increased cranial pressure	Ischemic complications of TMA (skin, intestinal, and limb)	None
Treatment prior to eculizumab	Plasma exchange	Plasma exchange	Plasma exchange	Plasma exchange	Plasma exchange
Duration of disease at first dose of eculizumab	1 ½ weeks	25 days	2 months	2 years 3 months	14 days
Eculizumab doses preceding changes in liver enzymes	2	1	2	4	1
Creatinine (μmol/l) at first dose of eculizumab	175	63	89	382	67
Dialysis at first dose of Eculizumab	No	No	No	Hemodialysis	No
ALT max (baseline)	908 (19)	175 (21)	249 (24)	360 (16)	129 (19)
AST max (baseline)	1107 (40)	491 (89)	244 (23)	373 (44)	262 (26)
GGT max (baseline)	1039 (26)	40 (24)	445 (29)	33 (14)	29 (19)
ALK max (baseline)	1023 (114)	240 (56)	229 (90)	571 (304)	173 (75)
Conjugated bilirubin max (baseline)	168 (0)	35 (0)	125 (0)	7 (0)	6 (0)
Platelets (×10 <sup>9</sup> /l) at first dose of eculizumab	561	202	176	282	243
LDH at first dose of eculizumab	3250	1034	650	1062	922
Haptoglobin at first dose of eculizumab	0.59	0.87	1.28	1.07	<0.10
Liver biopsy	Hepatocellular cytoplasmic swelling, Kupffer cell hemosiderosis, sinusoidal fibrosis	N/A	N/A	Mild chronic eosinophilic portal inflammation, mild fibrosis, moderate iron accumulation	N/A
Follow up on eculizumab without liver injury	0 months	11 months	8 months	2 months	17 months

A possible explanation for the observed liver injury in this series is the underlying disease process itself. Hepatitis was reported in 50 % of 27 patients with anti-CFH autoantibody-mediated aHUS, although the degree of liver enzyme derangement was not characterized [28]. In our case series, however, liver injury was temporally associated with eculizumab dosing rather than with aHUS disease presentation. We observed liver enzyme derangement 10 to 29 days following initiation of eculizumab, but 10 days to 2 years 3 months following presentation with aHUS.

In 4 of the 5 cases in this series, liver enzyme derangement was transient, asymptomatic, and did not compromise treatment. In one case, however, clinical symptoms of tender hepatomegaly coupled with recurrent liver enzyme derangement and jaundice led us to discontinue eculizumab and change to plasma therapy. Discontinuation of biological therapy for hepatotoxicity is relatively common in cancer treatment (e.g., discontinuation of erlotinib or imatinib for cytolytic hepatitis or gemtuzumab for cytolytic hepatitis); however, this is a novel complication in the treatment of aHUS [29].

The underlying mechanisms of hepatotoxicity associated with biological therapies remain incompletely understood [29]. Certain therapies are associated with specific patterns of hepatotoxicity, for example, sinusoidal obstruction syndrome secondary to gemtuzumab ozogamicin [30]. Specific targeting of hepatic Kupffer cells was initially thought to be the major mechanism responsible [30], but accumulation of antibody–toxin conjugates in hepatocytes causing calicheamicin-induced damage has subsequently been demonstrated [31].

The mechanism of hepatotoxicity potentially related to eculizumab in this case series has not yet been elucidated. A potential mechanism for our observations relates to the role of C5 in normal liver regeneration and its role in hepatocyte defense mechanisms during inflammation. C5 activity was found to be important in liver regeneration in a murine model of liver toxicity [32]. Further animal studies have demonstrated a potential role for C5 in defense reactions of Kupffer cells and hepatocytes [33]. Eculizumab may therefore disrupt hepatic regeneration and defense reactions.

Other possible mechanisms include a hypersensitivity reaction, off-target effects of the monoclonal antibody [34], and local hepatic side effects of C5 terminal complement blockade. Recent data from renal biopsies following long-term eculizumab therapy suggest direct antibody binding to tissue C5 [35]. If a similar process occurs in the liver, this may play a role in hepatotoxicity.

Some studies would lead one to hypothesize a hepatoprotective effect of eculizumab. Hillebrandt et al. found the Hc gene (which encodes complement C5) to be a significant modifier of liver fibrogenesis in animal studies. In addition, an association between genetic haplotypes associated with higher serum C5 levels and aggravated liver fibrogenesis in

human patients with hepatitis C has been observed [36]. Given these findings, one could hypothesize that C5 blockade might slow the progression of liver fibrosis, as has been suggested in case reports [27].

The long-term consequence of recurrent subclinical hepatocellular injury, which may be related to maintenance eculizumab therapy, is not known. Reports of cirrhosis and chronic hepatitis following acute idiosyncratic drug-induced liver injury are concerning in this regard [37].

To our knowledge, hepatotoxicity in association with eculizumab has not previously been reported. This may be an effect that has escaped the attention of clinicians, as it was subclinical in all but one case in this series. The true prevalence of this side effect remains to be established as hepatic enzyme monitoring has not been part of routine clinical monitoring for patients receiving eculizumab to date.

In light of the consistent finding of hepatotoxicity in this series we recommend regular monitoring of hepatic enzymes in all patients receiving eculizumab and caution in its use in patients with pre-existing liver disease. In patients with excessive hepatic enzyme disturbance or clinical symptoms of hepatitis in association with eculizumab, plasma therapy may have to be considered as an alternative treatment. Further research is required to clarify the impact of this adverse effect, to characterize the mechanism of potential hepatotoxicity, and to identify which patients are most at risk.

**Acknowledgements** The authors wish to thank Camille Bedrosian, MD, CMO, Alexion Pharmaceuticals, for helpful discussion of this manuscript.

**Disclosures** CL is advisory board member of Alexion Pharmaceuticals and Achillon Pharmaceuticals. He has received travel and speaker stipends as well as unrestricted research funds from Alexion Pharmaceuticals.

**Conflict of interest** The other authors declare no conflict of interest.

## References

- Hillmen P, Hall C, Marsh JC, Elebute M, Bombara MP, Petro BE, Cullen MJ, Richards SJ, Rollins SA, Mojcik CF, Rother RP (2004) Effect of eculizumab on hemolysis and transfusion requirements in patients with paroxysmal nocturnal hemoglobinuria. *N Engl J Med* 350:552–559
- Brodsky RA, Young NS, Antonioli E, Risitano AM, Schrezenmeier H, Schubert J, Gaya A, Coyle L, de Castro C, Fu CL, Maciejewski JP, Bessler M, Kroon HA, Rother RP, Hillmen P (2008) Multicenter phase 3 study of the complement inhibitor eculizumab for the treatment of patients with paroxysmal nocturnal hemoglobinuria. *Blood* 111:1840–1847
- Hillmen P, Young NS, Schubert J, Brodsky RA, Socie G, Muus P, Roth A, Szer J, Elebute MO, Nakamura R, Browne P, Risitano AM, Hill A, Schrezenmeier H, Fu CL, Maciejewski J, Rollins SA, Mojcik CF, Rother RP, Luzzatto L (2006) The complement inhibitor

- eculizumab in paroxysmal nocturnal hemoglobinuria. *N Engl J Med* 355:1233–1243
4. Schmidtko J, Peine S, El-Housseini Y, Pascual M, Meier P (2013) Treatment of atypical hemolytic uremic syndrome and thrombotic microangiopathies: a focus on eculizumab. *Am J Kidney Dis* 61:289–299
  5. Hodgkins KS, Bobrowski AE, Lane JC, Langman CB (2012) Clinical grand rounds: atypical hemolytic uremic syndrome. *Am J Nephrol* 35:394–400
  6. Gruppo RA, Rother RP (2009) Eculizumab for congenital atypical hemolytic-uremic syndrome. *N Engl J Med* 360:544–546
  7. Numberger J, Philipp T, Witzke O, Opazo Saez A, Vester U, Baba HA, Kribben A, Zimmerhackl LB, Janecke AR, Nagel M, Kirschfink M (2009) Eculizumab for atypical hemolytic-uremic syndrome. *N Engl J Med* 360:542–544
  8. Legendre CM, Licht C, Muus P, Greenbaum LA, Babu S, Bedrosian C, Bingham C, Cohen DJ, Delmas Y, Douglas K, Eitner F, Feldkamp T, Fouque D, Furman RR, Gaber O, Herthelius M, Hourmant M, Karpman D, Lebranchu Y, Mariat C, Menne J, Moulin B, Numberger J, Ogawa M, Remuzzi G, Richard T, Sberro-Soussan R, Severino B, Sheerin NS, Trivelli A, Zimmerhackl LB, Goodship T, Loirat C (2013) Terminal complement inhibitor eculizumab in atypical hemolytic-uremic syndrome. *N Engl J Med* 368:2169–2181
  9. Gleesing J, Chiwane S, Rongkavilit C (2012) Gonococcal septic shock associated with eculizumab treatment. *Pediatr Infect Dis J* 31:543
  10. Balagula Y, Newman SB, Lacouture ME (2010) Photodermatosis associated with eculizumab (Soliris): a novel monoclonal antibody directed against the complement protein C5. *Am J Hematol* 85:392–393
  11. Manganoni AM, Pavoni L, Facchetti F, Farisoglio C, Sereni E, Calzavara-Pinton P (2012) Melanoma in a patient in treatment with eculizumab. *Ann Hematol* 91:135–136
  12. Knoll BM, Letendre L, Steensma DP (2008) Life-threatening desquamating rash and hyperammonemia following administration of eculizumab for paroxysmal nocturnal hemoglobinuria. *Am J Hematol* 83:881–883
  13. Aithal GP, Watkins PB, Andrade RJ, Larrey D, Molokhia M, Takikawa H, Hunt CM, Wilke RA, Avigan M, Kaplowitz N, Bjornsson E, Daly AK (2011) Case definition and phenotype standardization in drug-induced liver injury. *Clin Pharmacol Ther* 89:806–815
  14. Unizony S, Arias-Urdaneta L, Miloslavsky E, Arvikar S, Khosroshahi A, Keroack B, Stone JR, Stone JH (2012) Tocilizumab for the treatment of large-vessel vasculitis (giant cell arteritis, Takayasu arteritis) and polymyalgia rheumatica. *Arthritis Care Res* 64:1720–1729
  15. Navarro-Millan I, Singh JA, Curtis JR (2012) Systematic review of tocilizumab for rheumatoid arthritis: a new biologic agent targeting the interleukin-6 receptor. *Clin Ther* 34:788–802
  16. Hassan R, Cohen SJ, Phillips M, Pastan I, Sharon E, Kelly RJ, Schweizer C, Weil S, Laheru D (2010) Phase I clinical trial of the chimeric anti-mesothelin monoclonal antibody MORAb-009 in patients with mesothelin-expressing cancers. *Clin Cancer Res* 16:6132–6138
  17. Langenberg MH, Witteveen PO, Lankheet NA, Roodhart JM, Rosing H, van den Heuvel JJ, Beijnen JH, Voest EE (2010) Phase I study of combination treatment with PTK 787/ZK 222584 and cetuximab for patients with advanced solid tumors: safety, pharmacokinetics, pharmacodynamics analysis. *Neoplasia* 12:206–213
  18. Trarbach T, Moehler M, Heinemann V, Kohne CH, Przyborek M, Schulz C, Sneller V, Gallant G, Kanzler S (2010) Phase II trial of mapatumumab, a fully human agonistic monoclonal antibody that targets and activates the tumour necrosis factor apoptosis-inducing ligand receptor-1 (TRAIL-R1), in patients with refractory colorectal cancer. *Br J Cancer* 102:506–512
  19. Ierardi E, Valle ND, Nacchiero MC, De Francesco V, Stoppino G, Panella C (2006) Onset of liver damage after a single administration of infliximab in a patient with refractory ulcerative colitis. *Clin Drug Invest* 26:673–676
  20. Menghini VV, Arora AS (2001) Infliximab-associated reversible cholestatic liver disease. *Mayo Clin Proc* 76:84–86
  21. Saleem G, Li SC, MacPherson BR, Cooper SM (2001) Hepatitis with interface inflammation and IgG, IgM, and IgA anti-double-stranded DNA antibodies following infliximab therapy: comment on the article by Charles et al. *Arthritis Rheum* 44:1966–1968
  22. Germano V, Picchianti Diamanti A, Baccano G, Natale E, Onetti Muda A, Priori R, Valesini G (2005) Autoimmune hepatitis associated with infliximab in a patient with psoriatic arthritis. *Ann Rheum Dis* 64:1519–1520
  23. Tobon GJ, Canas C, Jaller JJ, Restrepo JC, Anaya JM (2007) Serious liver disease induced by infliximab. *Clin Rheumatol* 26:578–581
  24. Soto-Fernandez S, Gonzalez-Carro P, De Pedro-Esteban A, Legaz-Huidobro ML, Perez-Roldan F, Roncero Garcia-Escribano O, Valbuena-Gonzalez M, Ruiz-Carrillo F (2006) Infliximab-induced hepatitis in a patient with Crohn's disease. *Gastroenterol Hepatol* 29:321–322
  25. Ozorio G, McGarity B, Bak H, Jordan AS, Lau H, Marshall C (2007) Autoimmune hepatitis following infliximab therapy for ankylosing spondylitis. *Med J Aust* 187:524–526
  26. Rother RP, Rollins SA, Mojcik CF, Brodsky RA, Bell L (2007) Discovery and development of the complement inhibitor eculizumab for the treatment of paroxysmal nocturnal hemoglobinuria. *Nat Biotechnol* 25:1256–1264
  27. Brodsky A, Mazzocchi O, Sanchez F, Khursigara G, Malhotra S, Volpacchio M (2012) Eculizumab in paroxysmal nocturnal hemoglobinuria with Budd-Chiari syndrome progressing despite anticoagulation. *Exp Hematol Oncol* 1:26
  28. Dragon-Durey MA, Sethi SK, Bagga A, Blanc C, Blouin J, Ranchin B, Andre JL, Takagi N, Cheong HI, Hari P, Le Quintrec M, Niaudet P, Loirat C, Fridman WH, Fremeaux-Bacchi V (2010) Clinical features of anti-factor H autoantibody-associated hemolytic uremic syndrome. *J Am Soc Nephrol* 21:2180–2187
  29. Loriot Y, Perlemuter G, Malka D, Penault-Llorca F, Boige V, Deutsch E, Massard C, Armand JP, Soria JC (2008) Drug insight: gastrointestinal and hepatic adverse effects of molecular-targeted agents in cancer therapy. *Nat Clin Pract Oncol* 5:268–278
  30. McKoy JM, Angelotta C, Bennett CL, Tallman MS, Wadleigh M, Evens AM, Kuzel TM, Trifilio SM, Raisch DW, Kell J, DeAngelo DJ, Giles FJ (2007) Gemtuzumab ozogamicin-associated sinusoidal obstructive syndrome (SOS): an overview from the research on adverse drug events and reports (RADAR) project. *Leuk Res* 31:599–604
  31. Maniecki MB, Hasle H, Friis-Hansen L, Lausen B, Nielsen OJ, Bendix K, Moestrup SK, Moller HJ (2008) Impaired CD163-mediated hemoglobin-scavenging and severe toxic symptoms in patients treated with gemtuzumab ozogamicin. *Blood* 112:1510–1514
  32. Mastellos D, Papadimitriou JC, Franchini S, Tsonis PA, Lambris JD (2001) A novel role of complement: mice deficient in the fifth component of complement (C5) exhibit impaired liver regeneration. *J Immunol* 166:2479–2486
  33. Schieferdecker HL, Schlaf G, Jungermann K, Gotze O (2001) Functions of anaphylatoxin C5a in rat liver: direct and indirect actions on nonparenchymal and parenchymal cells. *Int Immunopharmacol* 1:469–481
  34. Cao Y, Marks JD, Huang Q, Rudnick SI, Xiong C, Hittelman WN, Wen X, Marks JW, Cheung LH, Boland K, Li C, Adams GP, Rosenblum MG (2012) Single-chain antibody-based immunotoxins targeting Her2/neu: design optimization and impact of affinity on antitumor efficacy and off-target toxicity. *Mol Cancer Ther* 11:143–153

35. Herlitz LC, Bomback AS, Markowitz GS, Stokes MB, Smith RN, Colvin RB, Appel GB, D'Agati VD (2012) Pathology after eculizumab in dense deposit disease and C3 GN. *J Am Soc Nephrol* 23:1229–1237
36. Hillebrandt S, Wasmuth HE, Weiskirchen R, Hellerbrand C, Keppeler H, Werth A, Schirin-Sokhan R, Wilkens G, Geier A, Lorenzen J, Kohl J, Gressner AM, Matern S, Lammert F (2005) Complement factor 5 is a quantitative trait gene that modifies liver fibrogenesis in mice and humans. *Nat Genet* 37:835–843
37. Andrade RJ, Lucena MI, Kaplowitz N, Garcia-Munoz B, Borraz Y, Pachkoria K, Garcia-Cortes M, Fernandez MC, Pelaez G, Rodrigo L, Duran JA, Costa J, Planas R, Barriocanal A, Guarner C, Romero-Gomez M, Munoz-Yague T, Salmeron J, Hidalgo R (2006) Outcome of acute idiosyncratic drug-induced liver injury: Long-term follow-up in a hepatotoxicity registry. *Hepatology* 44:1581–1588