



Does dienogest influence the inflammatory response of endometriotic cells? A systematic review

Giovanni Grandi^{1,3} · Michael Mueller^{2,3} · Nick A. Bersinger^{2,3} · Angelo Cagnacci¹ · Annibale Volpe¹ · Brett McKinnon^{2,3}

Received: 7 October 2015 / Revised: 18 November 2015 / Accepted: 19 November 2015 / Published online: 9 December 2015
© Springer International Publishing 2015

Abstract

Objective and design A systematic review of all literature was done to assess the ability of the progestin dienogest (DNG) to influence the inflammatory response of endometriotic cells.

Main outcome measures In vitro and in vivo studies report an influence of DNG on the inflammatory response in eutopic or ectopic endometrial tissue (animal or human).

Results After strict inclusion criteria were satisfied, 15 studies were identified that reported a DNG influence on the inflammatory response in endometrial tissue. These studies identified a modulation of prostaglandin (PG) production and metabolism (PGE₂, PGE₂ synthase, cyclooxygenase-2 and microsomal PGE synthase-1), pro-inflammatory cytokine and chemokine production [interleukin (IL)-1 β , IL-6, IL-8, tumor necrosis factor- α , monocyte chemoattractant protein-1 and stromal cell-derived factor-1], growth factor biosynthesis (vascular endothelial growth factor and nerve growth factor) and signaling kinases, responsible for the control of inflam-

mation. Evidence supports a progesterone receptor-mediated inhibition of the inflammatory response in PR-expressing epithelial cells. It also indicated that DNG inhibited the inflammatory response in stromal cells, however, whether this was via a PR-mediated mechanism is not clear.

Conclusions DNG has a significant effect on the inflammatory microenvironment of endometriotic lesions that may contribute to its clinical efficacy. A better understanding of the specific anti-inflammatory activity of DNG and whether this contributes to its clinical efficacy can help develop treatments that focus on the inhibition of inflammation while minimizing hormonal modulation.

Keywords Dienogest · Inflammation · Endometriosis · Cytokines · Chemokines · Prostaglandin · Growth factors · Progestin · Progestogen

Introduction

Endometriosis is characterized by the growth of endometrial epithelial and stromal cells outside the uterine cavity and affects up to 15 % of reproductive age women [1]. It is associated with chronic pelvic pain, infertility [2] and a reduced quality of life [3]. The growth of endometriotic lesions is estrogen dependent and produces an increase in pro-inflammatory cytokine [4, 5], chemokine [6, 7] and growth factor [8] concentrations in the local environment. Current treatments for endometriosis target the estrogen dependence of the disease predominantly with progestogens and gonadotropin releasing hormone analogs (GnRHa) that create hypo-estrogenic and hyper-progestogenic environment via the inhibition of ovarian follicle development and the subsequent reduction in estrogen

Responsible Editor: Bernhard Gibbs.

✉ Brett McKinnon
brett.mckinnon@dkf.unibe.ch

¹ Azienda Ospedaliero-Universitaria Policlinico, University of Modena and Reggio Emilia, Via del Pozzo 71, 41124 Modena, Italy

² Department of Obstetrics and Gynaecology, Inselspital, Berne University Hospital, Effingerstrasse 102, 3010 Berne, Switzerland

³ Department of Clinical Research, University of Berne, Murtenstrasse 35, 3010 Berne, Switzerland

production and serum concentrations [9–11]. The long-term induction of a hypo-estrogenic state, however, will create significant side effects, as well as negatively impacting fertility options and are thus less than optimal for the treatment of reproductive age women. Improved treatment options are required.

Dienogest (DNG) is an orally active progestogen, highly selective for the progesterone receptor (PR) that exists in two major isoforms, PR-A and PR-B [12]. DNG is indicated for the long-term treatment of endometriotic pain [13, 14] and is associated with an increase in the patients' quality of life [15, 16] that is maintained even after the discontinuation of therapy [17]. DNG treatment with 2 mg day⁻¹, however, reduces estrogen concentrations only to levels that remain within the normal range of women in the early follicular phase of the menstrual cycle and are still higher than those mediated by GnRH_a buserelin and leuproreline, but with a comparable improvement in clinical symptoms [10]. This suggests that it may not be required to eliminate the estrogen production completely for effective treatment, or that another, non-hormonal activity may contribute to DNG effectiveness.

In addition to estrogen dependence, the inflammatory microenvironment of endometriotic lesions can significantly contribute to both disease progression [18] and symptomology [19], and therefore, may also represent a viable target for the treatment of endometriosis that has not yet been fully exploited [20]. Progesterone receptor (PR) activation, the primary target for DNG also regulates inflammation in endometrium. Progesterone inhibits inflammation in endometrial myometrial cells through regulation of the nuclear factor (NF) κ B, the transcription factor responsible for the pro-inflammatory cytokine production in endometrial tissue [21]. Furthermore, a reciprocal relationship between PR-B expression and NF κ B exists in endometrial tissue [22]. Therefore, given the minimal decrease of systemic estrogen and the importance of inflammation to endometriotic disease progression, it is possible that an anti-inflammatory mechanism may also contribute to the effectiveness of DNG treatment in endometriotic women. If present, such a mechanism of action may be a valid therapeutic target for endometriosis treatment that could be optimized to limit hormonal modulation.

Objectives

As a preliminary investigation, we therefore examined the literature to determine whether there was sufficient evidence to support a DNG influence on the inflammatory response of endometriotic cells, and secondly, to determine whether the mechanism is mediated by PRs activation.

Materials and methods

A systematic review of the *in vitro* and *in vivo* effects of DNG on endometrial and endometriotic cells was performed in accordance with Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines. All aspects of the study were decided before the literature search and no post hoc changes were made. The relevant articles were identified through a search of the Medline, PubMed and EMBASE databases with the keywords 'dienogest' in combination with 'endometriosis' and 'inflammation' by one author (G.G.) and identified references were reviewed by a second author (B.D.M.). All searches were made in June 2015, and the databases were searched from January 1, 1987 through May 31, 2015. We assessed all potentially relevant articles and examined the reference lists for additional publications. Only English language articles were considered. Both published articles and accepted manuscripts were included. We did not consider abstracts and case reports.

The inclusion criteria were the examination of the inflammatory response of endometrial/endometriotic tissue after exposure to DNG. Primary outcomes were a change in the production, or concentration of pro-inflammatory molecules, including prostaglandins (PG), cytokines, chemokines and growth factors and intracellular signaling kinases that mediate inflammation. Institutional Review Board approval was not obtained for this systematic review, as it was determined as unnecessary since all information were extracted from previously published studies.

Results

Study selection

The predefined search terms generated 90 references in PubMed. Of these 90 references, 68 were excluded because they did not meet the eligibility criteria and were concerned mainly with clinical efficacy, safety and tolerability of DNG in endometriosis patients, leaving 22 remaining publications [23–44]. Six studies were excluded, as the outcomes were not considered linked to inflammation, but rather progesterone metabolism/activity and cell cycle interference [25, 27, 28, 30–32]. One additional study was excluded as it was performed only on cells from breast and endometrial cancers [24]. The final sample consisted of 15 references [23, 26, 29, 33–44] (Fig. 1). A total of 11 studies [26, 29, 33–41] were identified that examined inflammatory effects on endometrial or endometriotic cells after DNG treatment *in vitro* and three studies [23, 42, 43] that

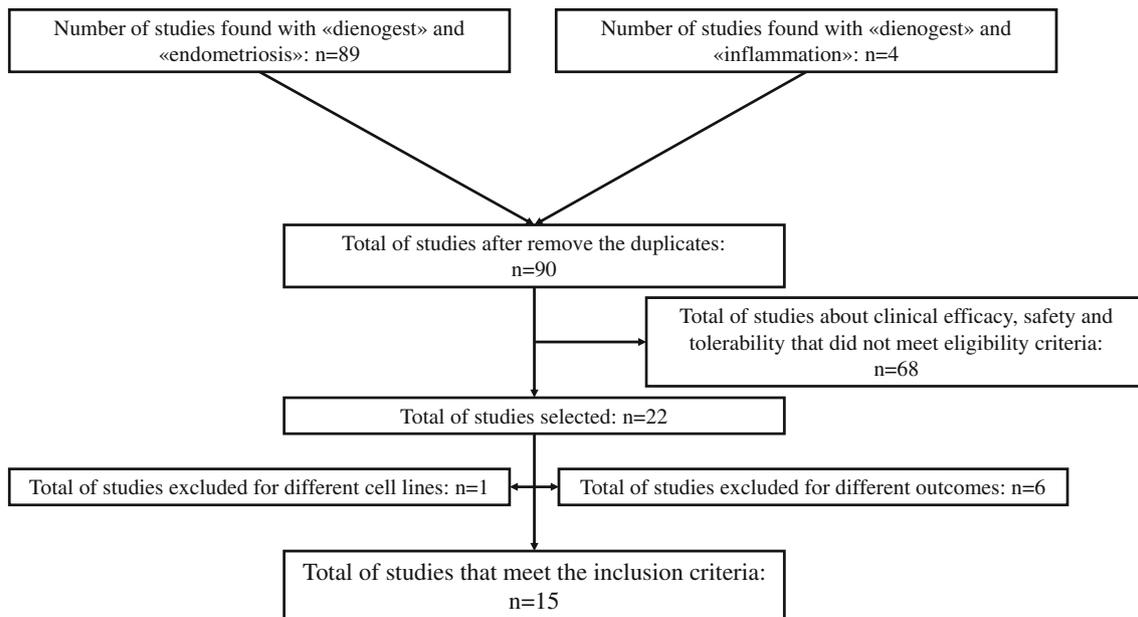


Fig. 1 Flowchart of the systematic review process used during this study

examined DNG effect on inflammation in animals (mice). The final study is an in vivo study [44] that evaluated the impact of DNG on endometrial tissue of human subjects.

Study characteristics and risk of bias

When assessing in vitro studies the use of different cell culture models and conditions is a potential source of bias. In this review, all epithelial cell studies were performed on immortalized cell lines, except one [35]. Stromal cell cultures were isolated from either the eutopic or the ectopic tissue of endometriotic women. Furthermore, both monolayer cell cultures [37, 38] and spheroid cell cultures [33, 35] have been included, which may also introduce a degree of bias. Both cell type and the conditions for each study have been clearly stated throughout the manuscript. DNG concentrations are also a source of potential bias and are clearly reported where appropriate, and range between 10^{-5} and 10^{-9} M. One final source of variation was the expression of PRs, known to vary in endometriosis patients [45], cell types and primary cell preparations (Tables 1, 2).

DNG influence on the inflammatory response

Prostaglandins and converting enzymes

In an in vitro model of immortalized epithelial cells transfected with either PRA or PRB, the treatment with DNG significantly inhibited CYP19A1, prostaglandin E2 Synthase (PGES) and microsomal (m)PGES-1 mRNA and prostaglandin E2 (PGE2) protein concentrations in PRA+/

B+ epithelial cells, an effect that could be attenuated by the specific anti-progesterone RU486 [33]. DNG also inhibited CYP19A1 in immortalized epithelial cells from a peritoneal lesion (12Z) [29]. In a series of immortalized epithelial cell lines derived from ovarian endometriomas that utilized either the endogenous (PRA−/PRB−; EMosis-CC/TERT1) or the overexpression PR isoforms (PRA; EMosis-CC/TERT1/PRA, PRB; EMosis-CC/TERT1/PRA−/PRB+) there was an inhibition of the cyclooxygenase (COX)-2, mPGES and CYP19A1 mRNA in both the PRA+ and PRB+ cells, but not in the endogenous PRA−/B− cells when treated with DNG (10^{-7} M) [33]. Epithelial cells of endometrioma tissues collected in women after treatment with DNG also expressed a lower proportion of aromatase positive cells compared with controls [44].

Other studies indicate a similar influence of DNG on stromal cells. In primary stromal cells isolated from endometriomas and grown in a spheroid cell culture, DNG treatment (10^{-7} M) significantly reduced the expression of both COX2 and CYP19A1 mRNA and PGE2 protein expression [34]. Aromatase inhibition was also confirmed in stromal cell spheroid cultures from ovarian endometriomas [35]. Neither the PR subtype ratio nor the influence of specific PR antagonists was investigated.

Inflammatory cytokines and chemokines

Using the immortalized epithelial ectopic and eutopic cell culture models transfected with either PRA or PRB described previously [33], DNG treatment (10^{-7} M) mediated an inhibition of interleukin (IL)-6, IL-8 and

Table 1 The influence of DNG on the inflammatory reaction of endometrial epithelial cells in vitro and the potential mechanism of action

Effect of DNG	Expression levels evaluated			PRs dependent activity evaluated		References
	mRNA	Proteins	Others	Yes/ no	Way	
Inhibition of PGE2		X		Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
	X	X		Yes	Reversed by RU-486 Not present in cells with low levels of PRs	Shimizu et al. [26]
Reduction of COX-2	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
	X			No		Shimizu et al. [26]
Reduction of PGE2 synthase	X			No		Shimizu et al. [26]
Reduction of mPGES-1	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
	X			No		Shimizu et al. [26]
IL-6 downregulation	X			Yes	Limited to the cells that express PRA or B	Mita et al. [36]
IL-8 downregulation		X (LPS/HMGB1 activated)		Yes	Limited to the cells that express PRB	Mita et al. [36]
	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
Reduction of MCP-1		X (LPS/HMGB1 activated)		Yes	Limited to the cells that express PRB	Mita et al. [36]
	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
Reduction of aromatase	X			Yes	Not present in cells with low levels of PRs	Shimizu et al. [26]
	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
	X			No		Beranič and Rižner [29]
NF-κB inactivation			Binding activity to DNA (EMSA)	No		Shimizu et al. [26]
			NF-κB gene activity (reporter gene assays)	Yes	Limited to the cells that express PRB	Mita et al. [36]
VEGF downregulation	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]
NGF downregulation	X	X	Bioactivity (assaying neurite outgrowth of rat pheochromocytoma cells-12 cells)	Yes	PRA or B both mediate the effect that is reduced in PR negative cells	Mita et al. [40]
	X			Yes	Limited to the cells that express PRA or B	Ichioka et al. [33]

LPS/HMGB1 lipopolysaccharide and high-mobility group box 1, *PR* progesterone receptor, *NF-κB* nuclear factor-κB, *PG* prostaglandin, *COX* cyclo-oxygenase, *mPGES* microsomal PGE synthase, *IL* interleukin, *MCP-1* monocyte chemoattractant protein-1, *VEGF* vascular endothelial growth factor, *NGF* nerve growth factor, *EMSA* electrophoretic mobility shift assay

monocyte chemotactic protein (MCP)-1 in both the PRA+ and PRB+ cells, but not in the PRA/B− cells [33]. DNG (10^{-7} to 10^{-9} M) mediated these anti-inflammatory effects

through the inhibition of the toll-like receptor 4 (TLR4) mRNA expression and a subsequent inhibition of TLR4 attenuated IL-8 production [36]. In primary endometriotic

Table 2 The influence of DNG on the inflammatory reaction of endometrial stromal cells and the potential mechanism of action

Effect of DNG	Expression levels evaluated			PRs dependent activity evaluated		References
	mRNA	Proteins	Others	Yes/ no	Mechanism of action	
Inhibition of PGE2		X		No		Yamanaka et al. [34]
Reduction of COX-2	X	X		No		Yamanaka et al. [34]
IL-8 downregulation		X (TNF- α and E2 activated)		Yes	The effect was not attenuated by the coadministration of RU486	Horie et al. [37]
Reduction of SDF-1	X (E2 activated)	X (E2 activated)		No		Okada et al. [38]
Reduction of aromatase	X	X		No		Yamanaka et al. [34]
NF- κ B inactivation			NF- κ B activation (EMSA) (TNF α and E2 activated)	No		Horie et al. [37]
		X	NF- κ B DNA-binding activity (ELISA)	No		Yamanaka et al. [34]
VEGF downregulation	X (E2 activated)	X (E2 activated)		No		Okada et al. [38]

E2 estradiol, PR progesterone receptor, NF- κ B nuclear factor- κ B, PG prostaglandin, COX cyclo-oxygenase, IL interleukin, SDF-1 stromal cell-derived factor-1, VEGF vascular endothelial growth factor, EMSA electrophoretic mobility shift assay, TNF α tumor necrosis factor α , ELISA enzyme-linked immunosorbent assay

stromal cells, DNG (10^{-7} M) reduced tumor necrosis factor (TNF) α -induced (0.1 ng/mL) IL-8 expression, which could not be reversed by coadministration of RU486 [37]. DNG also attenuated estradiol (E2)-induced stromal cell-derived factor (SDF)-1 mRNA and protein production in endometrial stromal cells from women with fibroids but without endometriosis [38]. DNG (10^{-7} M) also reduced TNF α production and secretion by peritoneal macrophages [39], and in an autotransplanted model of Sprague–Dawley rats DNG reduced IL-1 β expression in isolated peritoneal macrophages [23].

Growth factors

DNG treatment reduced the mRNA expression of vascular endothelial growth factor (VEGF) and nerve growth factor (NGF) in PRA+ and PRB+, but not PRA–/B– immortalized endometrial epithelial cells [33]. DNG (10^{-7} to 10^{-9} M) also inhibited NGF mRNA and protein expression and reduced neurite outgrowth of rat pheochromocytoma cells, which are positive for PR expression [40]. In endometrial stromal cells from non-endometriotic women, VEGF mRNA and protein production was also inhibited by DNG treatment (10^{-7} to 10^{-9} M) [38]. In S-180 mouse tumor cells, topical DNG inhibited embryonic angiogenesis in a dose-dependent manner and its oral administration ($1 \text{ mg kg}^{-1} \text{ day}^{-1}$) significantly suppressed angiogenesis [42]. In a rat endometrial autograft model, oral DNG

(1 mg/kg body weight/day) reduced the size of the microvascular network, decreased microvessel density and influenced the structure of the newly formed microvessels compared to controls [43].

Cellular-based modulators of the inflammatory response

DNG (10^{-7} M) inhibited NF κ B-binding activity in immortalized endometriotic epithelial cells [26] and in lipopolysaccharide and high-mobility group box 1 (LPS/HMGB1) stimulated endometrial epithelial cells overexpressing PR–B [36]. In endometriotic stromal cells, treatment with DNG (10^{-7} M) attenuated TNF α stimulated NF κ B activation in both monolayer [37] and spheroid cultures [34]. Lastly, it has also been shown that there was a lower expression of HLA-DR expression in peritoneal macrophages from women with endometriosis and that incubation with DNG (10^{-7} M) could restore this to similar levels as controls [39], suggesting DNG may influence immune cells.

Discussion

DNG is used for the long-term treatment of endometriosis by creating a hypo-estrogenic environment that inhibits endometriotic lesion growth. However, it is possible that DNG has additional mechanisms of actions that contribute

to lesion reduction. Given the ability of progesterone to influence inflammation, we postulated that DNG might also influence the inflammatory reaction that supports endometriotic lesion progression. Through this systematic review, we identified a total of 15 studies that reported an influence of DNG on the production of PGs (PGE₂, COX-2 and mPGES-1) [26, 33–35], pro-inflammatory cytokines (IL-1 β , IL-6 and TNF- α) [33, 36, 37], chemokines (IL-8, MCP-1 and SDF-1) [33, 36, 38] and growth factors (VEGF and NGF) [33, 38, 40] in endometriotic related cells. Although the number of studies that met the strict inclusion criteria for this manuscript is small, we believe that they provide sufficient preliminary evidence to indicate a local, anti-inflammatory role for DNG on endometriotic cells that deserves further attention.

Inflammation is a significant factor in the progression of endometriosis and the inflammatory mediators found to be regulated by DNG all contribute to endometriotic lesion progression. TNF- α stimulates the production of downstream cytokines in both epithelial and stromal cells [46, 47], leading to an increase in peritoneal fluid levels of numerous pro-inflammatory mediators including IL-6, MCP-1 [48, 49] and IL-8 [6] in endometriotic women. The response to inflammation and, therefore, the ability of DNG to influence it may also be lesion dependent as SDF-1 is significantly increased in the peritoneal fluid of women with deep infiltrating endometriosis (DIE) [50] and MCP-1, IL-6 and TNF α production is enhanced in recto-vaginal septum lesions compared to lesions from other locations [51]. Inflammation also influences the surrounding microenvironment as VEGF and NGF also stimulate angiogenesis and nerve growth. The ability of DNG to attenuate the production of all these cytokines and growth factors, therefore, will not only influence the viability of the endometriotic cells themselves, but also the supporting endothelial and stromal cells and extend its influence beyond hormonal modulation.

The mechanism by which DNG mediates this anti-inflammatory effect in endometriotic cells is difficult to infer from the data available. The majority of studies that we identified reported a DNG influence on inflammation via *in vitro* experiments. These *in vitro* models provide the advantage of being isolated from the influence of reduced follicular estrogens, thereby allowing the direct effects of DNG to be unmasked, confirming a local effect. How this local effect is mediated, however, is not clear. Endometriotic lesions produce their own estrogenic supply and are considered progesterone resistant and the modified PR activity is one of the primary mechanisms that allow endometriotic lesions to remain at ectopic locations through a dysregulation of decidualization [20, 52]. In epithelial cells, a DNG-mediated effect on inflammation occurred only when PR protein was present through an

exogenous transfection, supporting a PR-based mechanism. In the stromal cells, such a mechanism could not be confirmed from the data available as the presence of PR was rarely examined, although in all studies a DNG effect on inflammation was observed without the need for exogenous PR expression. A progesterone resistance would suggest that local DNG effects occur through non-PR-mediated mechanisms, although PR expression and progesterone resistance in endometriotic lesions remain controversial [45, 53]. The exact mechanism of local DNG effects and whether these effects can be replicated *in vivo*, therefore, remains an open question.

Another consideration is whether anti-inflammatory DNG effects can also be mediated by its systematic influence on circulating estrogen concentrations. Estrogen can both stimulate and suppress the immune response, dependent on the cell type and estrogen concentrations [54]. In dendritic cells, estrogen stimulates inflammation [55, 56] whereas it suppresses the inflammatory response to influenza [57]. In endometriosis, both ER α and β have been observed in endometriotic cells [58] and estrogen treatment creates an inflammatory response in human endometrial stromal cells [59]. Therefore, the reduction of circulating estrogen by DNG could have a significant anti-inflammatory activity on both the immune and endometriotic cells in the peritoneal cavity.

Furthermore, there is also an interdependent relationship between estrogen and progesterone. For example, in breast cancer the PR is capable of influencing the cellular response to estrogen [60], whereas the physiological effects of progesterone may be amplified in the presence of estrogens due to the increased expression of PR [61]. Therefore, the imbalance of PR A and B expression in endometriotic lesions [62] could have a significant influence to how these cells respond to DNG through altered estrogen concentrations.

The potential of an estrogen-regulated inflammatory response by DNG treatment, whether through local or systemic effects may also have significant implications for other estrogen-driven diseases that have an inflammatory component. Autoimmune diseases such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), autoimmune thyroid diseases (ATD) and multiple sclerosis (MS) all have a higher incidence in the female population [63]. Obese postmenopausal women are also at a higher risk of breast cancer due to the production of estrogen [64]. There may be, therefore, the possibility of using an anti-estrogen, anti-inflammatory drug, such as DNG in other estrogen-driven immune conditions. The therapeutic potential of targeting both inflammation and estrogen has already been proposed for MS [65].

The strongest evidence of a physiological effect of DNG on the inflammatory endometriotic environment comes

from the stromal cell studies; first, because these studies were performed in primary cell cultures and second, because transfection of PR was not required to detect an influence of DNG on the inflammatory response of these cells (Fig. 2). In normal functioning, endometrium stromal cells are the major target for progesterone [66] and are expected to have a significantly stronger PR expression; however, as the PR expression was not assessed in the majority of these studies it was difficult to determine if the local effects were mediated directly through the nuclear receptors or through another mechanism. Some of the results presented support a possibility for non-PR-mediated mechanisms; DNG-induced anti-proliferative effects in a rat model of endometriosis were observed even in the presence of a PR antagonist [23] and a DNG-mediated increase in IL-8 mRNA expression [37] and cell proliferation [41] was not attenuated by RU486 (Table 2). These preliminary data suggest that a non-PR mechanism should be investigated.

In contrast to the stromal cells, the epithelial cell studies were performed predominantly after the presence of PR proteins was determined (Fig. 2). Importantly, only a limited effect of DNG was observed in PR negative cells under basal conditions. If cells were transfected with PR to increase expression, however, a significant effect of DNG treatment on the mRNA production of inflammatory mediators was observed. These data suggest that in these immortalized epithelial cells the local DNG effects are mediated directly through PR activation and therefore that the PR status of epithelial cells *in vivo* will be important as

to whether DNG has a physiological influence. When evaluated, the contribution of PR-A and PR-B isoforms has so far revealed similar biological activity [26, 33, 36, 40] (Tables 1, 2). Previous studies suggest that the inflammatory response of stromal cells is significantly stronger than the response of epithelial cells [47] and in fact, recent studies suggest that the stromal cell PR expression is sufficient for a progesterone-mediated inhibition of epithelial cell proliferation [12], as PR activation of stromal cells blocks the production of mitogenic mediators that influence epithelial cell viability [67].

Limitations

The major limitation of this systematic review is the small number of studies that met the strict inclusion criteria. However, we believe that this small number is reflective of the novelty and timeliness of this question, but was still sufficient to draw robust conclusions on the influence of DNG on inflammation. It was also not clear in some studies whether PR was present in the cell models examined, making it difficult to make conclusions on the mechanism of action.

Conclusions

This systematic review identifies sufficient evidence to support a local effect on endometriotic inflammation by DNG and identifies two important questions that remain

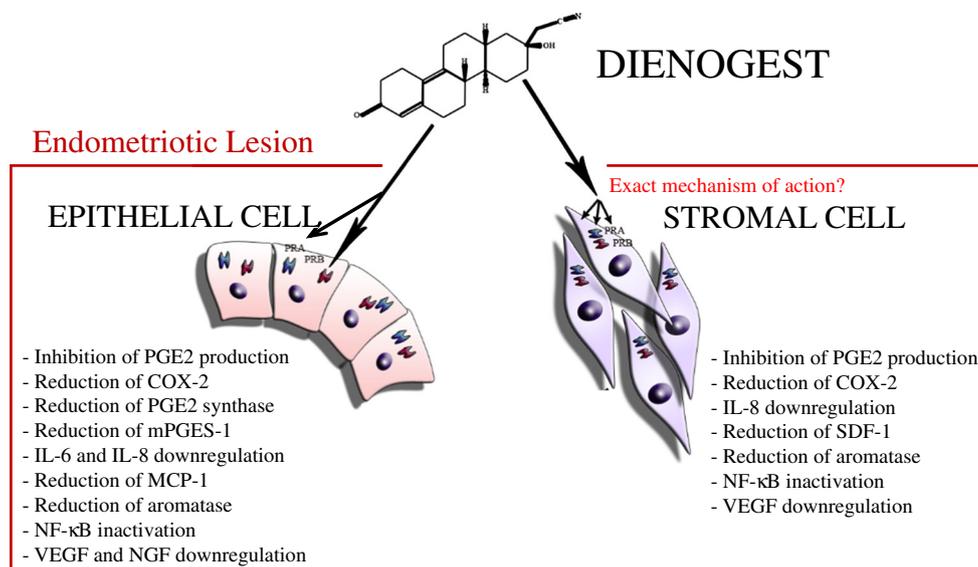


Fig. 2 In epithelial cells, DNG treatment was associated with a decrease in many different pro-inflammatory markers. This modulation, however, was only observed in cells positive for PR either through endogenous expression or PR transfection. DNG also

modulates the inflammatory response of endometriotic stromal cells, however, the evidence was not clear on the mechanism of action and it is possible that this occurred through a non-PR-mediated mechanism

unanswered and should be addressed in future studies. In epithelial cells, it is likely the DNG effect is mediated through PR; however, the use of immortalized cell lines transfected with PR means that PR expression in endometriotic epithelial cells in vivo needs to be clarified. In primary stromal cells, a direct effect of DNG on inflammation is also present, although whether this occurs directly through PR modulation also needs confirmation.

In summary, the symptomatology and the progression of endometriosis are intricately linked to the hyper-inflammatory microenvironment stimulated by the growth of the ectopic endometrium. A better understanding of the local effects of DNG and other progestogens on local endometriotic tissue could facilitate the development of novel therapeutics that emphasize non-hormonal functions of this class of drugs and greatly improve their clinical profile.

Acknowledgments The authors of this review have received no payment in preparation of their manuscript.

Compliance with ethical standards

Conflict of interest The authors of this review declare that they have no conflict of interest.

References

- Vercellini P, Viganò P, Somigliana E, Fedele L. Endometriosis: pathogenesis and treatment. *Nat Rev Endocrinol*. 2014;10(5):261–75.
- Bulun SE. Endometriosis. *N Engl J Med*. 2009;360(3):268–79.
- Simoens S, Dunselman G, Dirksen C, Hummelshoj L, Bokor A, Brandes I, et al. The burden of endometriosis: costs and quality of life of women with endometriosis and treated in referral centres. *Hum Reprod Oxf Engl*. 2012;27(5):1292–9.
- Bedaiwy MA, Falcone T, Sharma RK, Goldberg JM, Attaran M, Nelson DR, et al. Prediction of endometriosis with serum and peritoneal fluid markers: a prospective controlled trial. *Hum Reprod Oxf Engl*. 2002;17(2):426–31.
- Buyalos RP, Funari VA, Azziz R, Watson JM, Martinez-Maza O. Elevated interleukin-6 levels in peritoneal fluid of patients with pelvic pathology. *Fertil Steril*. 1992;58(2):302–6.
- Bersinger NA, von Roten S, Wunder DM, Raio L, Dreher E, Mueller MD. PAPP-A and osteoprotegerin, together with interleukin-8 and RANTES, are elevated in the peritoneal fluid of women with endometriosis. *Am J Obstet Gynecol*. 2006;195(1):103–8.
- Akoum A, Lemay A, McColl S, Turcot-Lemay L, Maheux R. Elevated concentration and biologic activity of monocyte chemotactic protein-1 in the peritoneal fluid of patients with endometriosis. *Fertil Steril*. 1996;66(1):17–23.
- Hirota Y, Osuga Y, Koga K, Yoshino O, Hirata T, Harada M, et al. Possible implication of midkine in the development of endometriosis. *Hum Reprod*. 2005;20(4):1084–9.
- McCormack PL. Dienogest: a review of its use in the treatment of endometriosis. *Drugs*. 2010;70(16):2073–88.
- Strowitzki T, Marr J, Gerlinger C, Faustmann T, Seitz C. Dienogest is as effective as leuprolide acetate in treating the painful symptoms of endometriosis: a 24-week, randomized, multicentre, open-label trial. *Hum Reprod Oxf Engl*. 2010;25(3):633–41.
- de Andres M. P, Lopes LA, Baracat EC, Podgaec S. Dienogest in the treatment of endometriosis: systematic review. *Arch Gynecol Obstet*. 2015;292(3):523–9.
- Kim JJ, Kurita T, Bulun SE. Progesterone action in endometrial cancer, endometriosis, uterine fibroids, and breast cancer. *Endocr Rev*. 2013;34(1):130–62.
- Köhler G, Faustmann TA, Gerlinger C, Seitz C, Mueck AO. A dose-ranging study to determine the efficacy and safety of 1, 2, and 4 mg of dienogest daily for endometriosis. *Int J Gynaecol Obstet Off Organ Int Fed Gynaecol Obstet*. 2010;108(1):21–5.
- Strowitzki T, Faustmann T, Gerlinger C, Seitz C. Dienogest in the treatment of endometriosis-associated pelvic pain: a 12-week, randomized, double-blind, placebo-controlled study. *Eur J Obstet Gynecol Reprod Biol*. 2010;151(2):193–8.
- Strowitzki T, Marr J, Gerlinger C, Faustmann T, Seitz C. Detailed analysis of a randomized, multicenter, comparative trial of dienogest versus leuprolide acetate in endometriosis. *Int J Gynaecol Obstet Off Organ Int Fed Gynaecol Obstet*. 2012;117(3):228–33.
- Harada T, Momoeda M, Taketani Y, Aso T, Fukunaga M, Hagino H, et al. Dienogest is as effective as intranasal buserelin acetate for the relief of pain symptoms associated with endometriosis—a randomized, double-blind, multicenter, controlled trial. *Fertil Steril*. 2009;91(3):675–81.
- Petraglia F, Hornung D, Seitz C, Faustmann T, Gerlinger C, Luisi S, et al. Reduced pelvic pain in women with endometriosis: efficacy of long-term dienogest treatment. *Arch Gynecol Obstet*. 2011;285(1):167–73.
- Giudice LC, Kao LC. Endometriosis. *Lancet*. 2004;364(9447):1789–99.
- McKinnon BD, Bertschi D, Bersinger NA, Mueller MD. Inflammation and nerve fiber interaction in endometriotic pain. *Trends Endocrinol Metab TEM*. 2015;26(1):1–10.
- Han SJ, O'Malley BW. The dynamics of nuclear receptors and nuclear receptor coregulators in the pathogenesis of endometriosis. *Hum Reprod Update*. 2014;20(4):467–84.
- Hardy DB, Janowski BA, Corey DR, Mendelson CR. Progesterone receptor plays a major antiinflammatory role in human myometrial cells by antagonism of nuclear factor-kappaB activation of cyclooxygenase 2 expression. *Mol Endocrinol*. 2006;20(11):2724–33.
- Guo S-W. Nuclear factor-kappaB (NF-kappaB): an unsuspected major culprit in the pathogenesis of endometriosis that is still at large? *Gynecol Obstet Invest*. 2007;63(2):71–97.
- Katsuki Y, Takano Y, Futamura Y, Shibutani Y, Aoki D, Udagawa Y, et al. Effects of dienogest, a synthetic steroid, on experimental endometriosis in rats. *Eur J Endocrinol*. 1998;138(2):216–26.
- Katsuki Y, Shibutani Y, Aoki D, Nozawa S. Dienogest, a novel synthetic steroid, overcomes hormone-dependent cancer in a different manner than progestins. *Cancer*. 1997;79(1):169–76.
- Ono YJ, Terai Y, Tanabe A, Hayashi A, Hayashi M, Yamashita Y, et al. Decorin induced by progesterone plays a crucial role in suppressing endometriosis. *J Endocrinol*. 2014;223(2):203–16.
- Shimizu Y, Mita S, Takeuchi T, Notsu T, Mizuguchi K, Kyo S. Dienogest, a synthetic progestin, inhibits prostaglandin E2 production and aromatase expression by human endometrial epithelial cells in a spheroid culture system. *Steroids*. 2011;76(1–2):60–7.
- Fu L, Osuga Y, Morimoto C, Hirata T, Hirota Y, Yano T, et al. Dienogest inhibits BrdU uptake with G0/G1 arrest in cultured endometriotic stromal cells. *Fertil Steril*. 2008;89(5 Suppl):1344–7.
- Prechapanich J, Kajihara T, Fujita K, Sato K, Uchino S, Tanaka K, et al. Effect of a dienogest for an experimental three-

- dimensional endometrial culture model for endometriosis. *Med Mol Morphol.* 2014;47(4):189–95.
29. Beranič N, Rižner TL. Effects of progestins on local estradiol biosynthesis and action in the Z-12 endometriotic epithelial cell line. *J Steroid Biochem Mol Biol.* 2012;132(3–5):303–10.
 30. Hayashi A, Tanabe A, Kawabe S, Hayashi M, Yuguchi H, Yamashita Y, et al. Dienogest increases the progesterone receptor isoform B/A ratio in patients with ovarian endometriosis. *J Ovarian Res.* 2012;5(1):31.
 31. Shimizu Y, Takeuchi T, Mita S, Mizuguchi K, Kiyono T, Inoue M, et al. Dienogest, a synthetic progestin, inhibits the proliferation of immortalized human endometrial epithelial cells with suppression of cyclin D1 gene expression. *Mol Hum Reprod.* 2009;15(10):693–701.
 32. Beranič N, Lanišnik Rižner T. Progestin effects on expression of AKR1C1-AKR1C3, SRD5A1 and PGR in the Z-12 endometriotic epithelial cell line. *Chem Biol Interact.* 2013;202(1–3):218–25.
 33. Ichioka M, Mita S, Shimizu Y, Imada K, Kiyono T, Bono Y, et al. Dienogest, a synthetic progestin, down-regulates expression of CYP19A1 and inflammatory and neuroangiogenesis factors through progesterone receptor isoforms A and B in endometriotic cells. *J Steroid Biochem Mol Biol.* 2015;147:103–10.
 34. Yamanaka K, Xu B, Suganuma I, Kusuki I, Mita S, Shimizu Y, et al. Dienogest inhibits aromatase and cyclooxygenase-2 expression and prostaglandin E₂ production in human endometriotic stromal cells in spheroid culture. *Fertil Steril.* 2012;97(2):477–82.
 35. Mori T, Ito F, Matsushima H, Takaoka O, Koshiba A, Tanaka Y, et al. Dienogest reduces HSD17β1 expression and activity in endometriosis. *J Endocrinol.* 2015;225(2):69–76.
 36. Mita S, Shimizu Y, Notsu T, Imada K, Kyo S. Dienogest inhibits Toll-like receptor 4 expression induced by costimulation of lipopolysaccharide and high-mobility group box 1 in endometrial epithelial cells. *Fertil Steril.* 2011;96(6):1485.e4–1489.e4.
 37. Horie S, Harada T, Mitsunari M, Taniguchi F, Iwabe T, Terakawa N. Progesterone and progestational compounds attenuate tumor necrosis factor alpha-induced interleukin-8 production via nuclear factor kappa B inactivation in endometriotic stromal cells. *Fertil Steril.* 2005;83(5):1530–5.
 38. Okada H, Okamoto R, Tsuzuki T, Tsuji S, Yasuda K, Kanzaki H. Progestins inhibit estradiol-induced vascular endothelial growth factor and stromal cell-derived factor 1 in human endometrial stromal cells. *Fertil Steril.* 2011;96(3):786–91.
 39. Maeda N, Izumiya C, Taniguchi K, Matsushima S, Mita S, Shimizu Y, et al. Dienogest improves human leucocyte antigen-DR under expression and reduces tumor necrosis factor-α production in peritoneal fluid cells from women with endometriosis. *Eur J Obstet Gynecol Reprod Biol.* 2014;177:48–51.
 40. Mita S, Shimizu Y, Sato A, Notsu T, Imada K, Kyo S. Dienogest inhibits nerve growth factor expression induced by tumor necrosis factor-α or interleukin-1β. *Fertil Steril.* 2014;101(2):595–601.
 41. Okada H, Nakajima T, Yoshimura T, Yasuda K, Kanzaki H. The inhibitory effect of dienogest, a synthetic steroid, on the growth of human endometrial stromal cells in vitro. *Mol Hum Reprod.* 2001;7(4):341–7.
 42. Nakamura M, Katsuki Y, Shibutani Y, Oikawa T. Dienogest, a synthetic steroid, suppresses both embryonic and tumor-cell-induced angiogenesis. *Eur J Pharmacol.* 1999;386(1):33–40.
 43. Katayama H, Katayama T, Uematsu K, Hiratsuka M, Kiyomura M, Shimizu Y, et al. Effect of dienogest administration on angiogenesis and hemodynamics in a rat endometrial autograft model. *Hum Reprod.* 2010;25(11):2851–8.
 44. Miyashita M, Koga K, Takamura M, Izumi G, Nagai M, Harada M, et al. Dienogest reduces proliferation, aromatase expression and angiogenesis, and increases apoptosis in human endometriosis. *Gynecol Endocrinol.* 2014;30(9):644–8.
 45. Shao R, Cao S, Wang X, Feng Y, Billig H. The elusive and controversial roles of estrogen and progesterone receptors in human endometriosis. *Am J Transl Res.* 2014;6(2):104–13.
 46. Bersinger NA, Günthert AR, McKinnon B, Johann S, Mueller MD. Dose-response effect of interleukin (IL)-1β, tumor necrosis factor (TNF)-α, and interferon-γ on the in vitro production of epithelial neutrophil activating peptide-78 (ENA-78), IL-8, and IL-6 by human endometrial stromal cells. *Arch Gynecol Obstet.* 2011;283(6):1291–6.
 47. Bersinger NA, Frischknecht F, Taylor RN, Mueller MD. Basal and cytokine-stimulated production of epithelial neutrophil activating peptide-78 (ENA-78) and interleukin-8 (IL-8) by cultured human endometrial epithelial and stromal cells. *Fertil Steril.* 2008;89(5 Suppl):1530–6.
 48. Bersinger NA, Dechaud H, McKinnon B, Mueller MD. Analysis of cytokines in the peritoneal fluid of endometriosis patients as a function of the menstrual cycle stage using the Bio-Plex[®] platform. *Arch Physiol Biochem.* 2012;118(4):210–8.
 49. Kalu E, Sumar N, Giannopoulos T, Patel P, Croucher C, Sherriff E, et al. Cytokine profiles in serum and peritoneal fluid from infertile women with and without endometriosis. *J Obstet Gynaecol Res.* 2007;33(4):490–5.
 50. Leconte M, Chouzenoux S, Nicco C, Chéreau C, Arkwright S, Santulli P, et al. Role of the CXCL12-CXCR4 axis in the development of deep rectal endometriosis. *J Reprod Immunol.* 2014;103:45–52.
 51. Bertschi D, McKinnon BD, Evers J, Bersinger NA, Mueller MD. Enhanced inflammatory activity of endometriotic lesions from the rectovaginal septum. *Mediators Inflamm.* 2013;2013:450950.
 52. Patel B, Elguero S, Thakore S, Dahoud W, Bedaiwy M, Mesiano S. Role of nuclear progesterone receptor isoforms in uterine pathophysiology. *Hum Reprod Update.* 2015;21(2):155–73.
 53. Bedaiwy MA, Dahoud W, Skomorovska-Prokvolit Y, Yi L, Liu JH, Falcone T, et al. Abundance and localization of progesterone receptor isoforms in endometrium in women with and without endometriosis and in peritoneal and ovarian endometriotic implants. *Reprod Sci.* 2015;22(9):1153–61.
 54. Beagley KW, Gockel CM. Regulation of innate and adaptive immunity by the female sex hormones oestradiol and progesterone. *FEMS Immunol Med Microbiol.* 2003;38:13–22.
 55. Seillet C, Rouquié N, Foulon E, Douin-Echinard V, Krust A, Chambon P, et al. Estradiol promotes functional responses in inflammatory and steady-state dendritic cells through differential requirement for activation function-1 of estrogen receptor α. *J Immunol.* 1950;2013(190):5459–70.
 56. Siracusa MC, Overstreet MG, Housseau F, Scott AL, Klein SL. 17β-estradiol alters the activity of conventional and IFN-producing killer dendritic cells. *J Immunol.* 1950;2008(180):1423–31.
 57. Robinson DP, Lorenzo ME, Jian W, Klein SL. Elevated 17β-estradiol protects females from influenza A virus pathogenesis by suppressing inflammatory responses. *PLoS Pathog.* 2011;7:e1002149.
 58. Lessey BA, Metzger DA, Haney AF, McCarty KS. Immunohistochemical analysis of estrogen and progesterone receptors in endometriosis: comparison with normal endometrium during the menstrual cycle and the effect of medical therapy. *Fertil Steril.* 1989;51:409–15.
 59. Chang K-K, Liu L-B, Li H, Mei J, Shao J, Xie F, et al. TSLP induced by estrogen stimulates secretion of MCP-1 and IL-8 and growth of human endometrial stromal cells through JNK and NF-κB signal pathways. *Int J Clin Exp Pathol.* 2014;7:1889–99.
 60. Mohammed H, Russell IA, Stark R, Rueda OM, Hickey TE, Tarulli GA, Serandour AA, Birrell SN, Bruna A, Saadi A, Menon S, Hadfield J, Pugh M, Raj GV, Brown GD, D'Santos C, Robinson JL, Silva G, Launchbury R, Perou CM, Stingl J, Caldas C, Tilley WD, Carroll JS. Progesterone receptor modulates ERα action in breast cancer. *Nature.* 2015;523(7560):313.

61. Tan IJ, Peeva E, Zandman-Goddard G. Hormonal modulation of the immune system—a spotlight on the role of progestogens. *Autoimmun Rev.* 2015;14(6):536–42.
62. Attia GR, Zeitoun K, Edwards D, Johns A, Carr BR, Bulun SE. Progesterone receptor isoform A but not B is expressed in endometriosis. *J Clin Endocrinol Metab.* 2000;85:2897–902.
63. Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol.* 2015;294:63–9.
64. Rose DP, Gracheck PJ, Vona-Davis L. The interactions of obesity, inflammation and insulin resistance in breast cancer. *Cancers.* 2015;7:2147–68.
65. Gold SM, Voskuhl RR. Estrogen treatment in multiple sclerosis. *J Neurol Sci.* 2009;286:99–103.
66. Deligdisch L. Hormonal pathology of the endometrium. *Mod Pathol.* 2000;13(3):285–94.
67. Li Q, Kannan A, DeMayo FJ, Lydon JP, Cooke PS, Yamagishi H, et al. The antiproliferative action of progesterone in uterine epithelium is mediated by Hand2. *Science.* 2011;331(6019):912–6.