RETICULAR PSEUDODRUSEN ON INFRARED IMAGING ARE TOPOGRAPHICALLY DISTINCT FROM SUBRETINAL DRUSENOID DEPOSITS ON EN FACE OPTICAL COHERENCE TOMOGRAPHY

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Purpose: To evaluate the quantitative and topographic relationship between reticular pseudodrusen (RPD) on infrared reflectance (IR) and subretinal drusenoid deposits (SDD) on en face volumetric spectral domain optical coherence tomography.

Methods: Reticular pseudodrusen were marked on IR images by a masked observer. Subretinal drusenoid deposits were visualized on en face sections of spectral domain optical coherence tomography below the external limiting membrane and identified by a semiautomated technique. Control RPD lesions were generated in a random distribution for each IR image. Binary maps of control and experimental RPD and SDD were merged and analyzed in terms of topographic localization and quantitative drusen load comparison.

Results: A total of 54 eyes of 41 patients diagnosed with RPD were included in this study. The average number of RPD lesions on IR images was 320 ± 44.62 compared with 127 \pm 26.02 SDD lesions on en face (P < 0.001). The majority of RPD lesions did not overlap with SDD lesions and were located $>30 \ \mu$ m away (92%). The percentage of total SDD lesions overlapping RPD was 2.91 \pm 0.87% compared with 1.73 \pm 0.68% overlapping control RPD lesions (P < 0.05). The percentage of total SDD lesions between 1 and 3 pixels of the nearest RPD lesion was 5.08 \pm 1.40% compared with 3.33 \pm 1.07% between 1 and 3 pixels of the nearest control RPD lesion (P < 0.05).

Conclusion: This study identified significantly more RPD lesions on IR compared with SDD lesions on en face spectral domain optical coherence tomography and found that a large majority of SDD (>90% of lesions) were >30 μ m away from the nearest RPD. Together, our findings indicate that RPD and SDD are two entities that are only occasionally topographically associated, suggesting that at some stage in their development, they may be pathologically related.

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Age-related macular degeneration (AMD) is the leading cause of blindness among the elderly in developed countries, with 1.75 million people affected in the United States alone.¹ Significant visual impairment results from late-stage AMD compared with early-stage AMD, which has little impact on visionrelated function.² Given the high prevalence and potential morbidity of AMD, the identification and characterization of risk factors for progression to late-stage AMD is warranted. One risk factor that has garnered recent attention is reticular pseudodrusen (RPD), a distinctive phenotypic characteristic observed in patients with AMD. Reticular pseudodrusen was first described in 1990 as having a peculiar yellowish pattern more visible under blue light.³ Although multiple imaging modalities have subsequently been used to detect RPD, infrared reflectance (IR) has proven to be the most sensitive for its visualization.⁴ The increased recognition of RPD has lead to better understanding of its clinical

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implications, most notably the association with an increased incidence of progression to late-stage AMD.⁵⁻¹²

Another finding associated with AMD are subretinal drusenoid deposits (SDD), aggregations of membranous material in the subretinal space. In 1988, Sarks et al¹³ first described membranous debris in multiple locations including within the subretinal space. The composition of this subretinal debris has been shown to include highly enriched unesterified cholesterol particles.¹⁴ These particles are morphologically similar to drusen on histopathology but differ in that they contain little esterified cholesterol, thereby designating these subretinal deposits as drusenoid.^{14,15} Although initial studies identified SDD using histopathology and low-magnification transmission electron microscopy, subsequent research has correlated these lesions with findings on spectral domain optical coherence tomography (SD-OCT).

The association between SDD and RPD has been a topic of recent debate. A study reevaluating three eyes initially used to assess the composition of SDD found evidence to suggest SDD is the histologic manifestation of RPD, although these eyes were not verified with clinical RPD at autopsy.¹⁶ Other studies have found evidence that SDD are adjacent to rather than overlying RPD, suggesting an association without being the same entity.^{17,18} Adding to the controversy, a recent study showed histologic evidence of SDD in eyes without RPD clinically.¹⁹ In this study, our goal was to analyze the quantitative and topographic relationship between RPD and SDD to examine the association between these two findings.

Methods

Study Design

The study was designed to evaluate the quantitative and topographic relationship between RPD and SDD. Reticular pseudodrusen were marked on IR images by a masked observer (J.K.F.), with a second observer

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(A.A.F.) marking a subset of images to assess accuracy. Subretinal drusenoid deposits were marked on optical coherence tomography (OCT) C-scans by a semiautomated process described below.

Study Sample

We retrospectively identified patients diagnosed with RPD between March 2012 and March 2014 at the Department of Ophthalmology of the Northwestern University Feinberg School of Medicine. IRB protocols were approved by the Scientific Review Committee of Northwestern University. This research followed the tenets of the Declaration of Helsinki, and informed consent was waived for this retrospective study. Inclusion criteria were patients with a diagnosis of RPD and a complete ophthalmologic examination, including IR, fundus autofluorescence imaging, and SD-OCT (Cirrus high-definition OCT [HD-OCT]; Carl Zeiss Meditec Inc, Dublin, CA) and the Spectralis HRA + OCT (Heidelberg Engineering Inc, Dossenheim, Germany). Exclusion criteria were <50 RPD, the RPE being too irregular on OCT for the Cirrus software to trace, or the IR images or OCT scans being of poor quality. Poor quality was defined qualitatively by the inability to accurately identify SDD and RPD on IR and OCT images and quantitatively by the Cirrus OCT image quality score <6.

The diagnosis of RPD was based on previously defined criteria.⁴ Reticular pseudodrusen were identified on IR reflectance images in concordance with previous reports demonstrating superior sensitivity of IR compared with fundus autofluorescence and red-free images in detecting RPD.¹⁷ We used the following criteria to define RPD on IR reflectance imaging: distinct hyporeflective lesions on a background of mild hyperreflectance and/or regular reticular patterns within well-defined domains.

Image Acquisition

Images were acquired on the Cirrus HD-OCT 5000 device (Carl Zeiss Meditec Inc) using a superluminescent diode at 840 nm, which achieves a 5 μ m axial and a 15 μ m transverse tissue resolution. The device captures 27,000 A-scans per second at 2 mm of depth. The images were viewed with the Cirrus HD-OCT software (version 6.5.0.772; Carl Zeiss Meditec Inc). Imaging was performed with a 512 × 128 macular cube volume scan consisting of 128 equally spaced horizontal B-scans (each composed of 512 A-scans) over a 6-mm square grid. In the 512 × 128 scan protocol, the pixel separation in the horizontal direction is 6,000/512 or 11.7 μ m, whereas the pixel separation in the vertical direction is 6,000/128 or 46.9 μ m. The

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pixel spacing in the axial direction for both cubes is 2,000/1,024 or approximately 2 μ m, whereas the axial resolution is approximately 5 μ m. The line scanning laser ophthalmoscope feature obtained a registered IR fundus image for each data cube. No image manipulation was performed to ensure alignment of IR and en face images (Figure 1). All scans were evaluated for alignment error and verified by retinal vessel comparison (Figure 2). Only volume scans free of motion artifact were selected for this study, as determined by assessment of the OCT volume projection to ensure complete lack of vessel alignment artifacts and/or horizontal lines across the volume, which also indicate motion.

Identification of Reticular Pseudodrusen on Infrared Reflectance Images

A masked investigator (J.K.F.) used the paintbrush tool on ImageJ software (NIH, Bethesda, MD) to individually mark each RPD lesion visualized on the IR image, using the criteria described above (Figure 3, A and B). This investigator was masked to the en face OCT images. Accuracy of lesion identification was studied using five randomly selected patients, where another masked reader (A.A.F.) independently identified the RPD lesions, and the intergrader correlation was analyzed.

Another investigator (M.J.H.) performed the subsequent steps of the analysis. The marked IR image was thresholded until only the RPD lesion markings were visible. The "find maxima" function (output type: single point) was used to identify all of the lesions. The resulting map contained single pixel dots with a thresholded value of 255, each representing the center of an individual RPD lesion. The pixel values were divided by 255, so that each RPD was represented by a single pixel with a value of 1.

A set of control (software generated) RPD lesions was generated to compare the study RPD lesions with a randomly distributed sample. The "add specified noise" function of ImageJ was used to generate a control image with equal number of RPD lesions for each corresponding study IR image. The "find maxima" and "divide" by 255 functions were applied to these control images, so that they could be used for the subsequent analysis and compared with the study IR images.

Identification of Subretinal Drusenoid Deposits on En Face Optical Coherence Tomography Sections

Optical coherence tomography volume scans were reviewed on Cirrus version 6.0 software using the advanced visualization feature. As previously described,^{17,19} the RPE feature was used to obtain en face slices, or C-scans, which were contoured based on each patient's RPE curvature. En face slabs, 4 pixels in thickness, were generated above the level of the RPE and just below the external limiting membrane (Figure 3, E and F). Subretinal drusenoid deposits were visualized as white circular spots, which were reviewed for each patient to ensure correspondence with the B-scan. The level of each en face slab was chosen to maximize the number of SDD visualized, while avoiding a confluence of the lesions, to allow for precise localization of individual lesions (Figure 4). Using the RPE slice overlay feature, these slabs were registered to the Cirrus IR fundus image with guidance from the SD-OCT scan.

A masked investigator (M.J.H.) performed the subsequent steps of the analysis on NIS-Elements AR (Nikon Instruments, Inc, Tokyo, Japan), a biologic image analysis software. The IR and C-scan images were overlaid and compared (Figure 5). Areas corresponding to retinal vessels on the C-scan image were removed. All marked RPD lesions that corresponded to these removed areas were excluded from the analysis. A topographic binary map of the SDD was

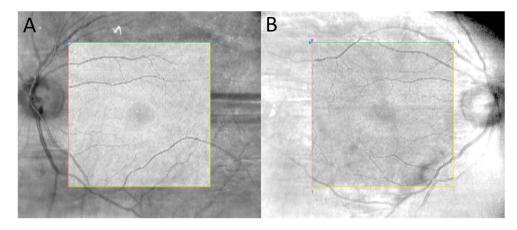


Fig. 1. Examples of OCT volume projection overlaid onto IR scanning laser ophthalmoscope image to demonstrate alignment accuracy. A and B. Two different examples of OCT volume projection within the square demonstrating excellent alignment with IR image outside square. Images obtained using overlay feature of Cirrus HD-OCT software (Carl Zeiss Meditec) with transparency setting of 100%.

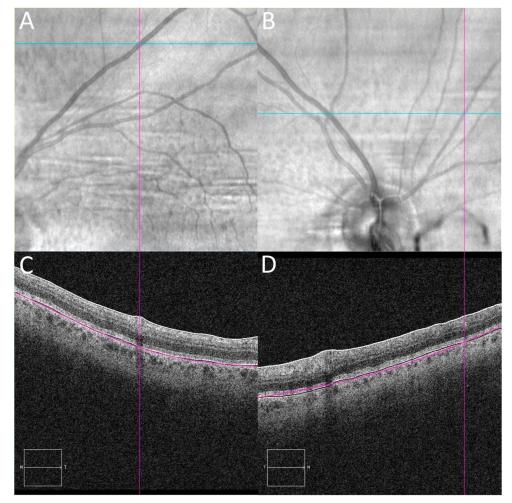


Fig. 2. Infrared reflectance and OCT B-scan alignment validation using retinal vessels. A and B. Infrared reflectance image with the cyan line marking the section of OCT B-scan image and the purple line positioned over the center of the retinal vessel along that section. C and D. Optical coherence tomography B-scan image with vertical purple line marking the retinal vessel

created by applying Gaussian noise reduction (median filter set to 5) and thresholding of the en face C-scan, based on minimum size (5 pixel diameter = 150 μ m) and pixel value (0–255) (Figure 3, C and D). The resulting SDD map had a binary set of pixels, which carried a value of either 0 (SDD, white) or 255 (background, black).

Image Analysis

The number of control (software generated) and study RPD overlapping SDD was determined using the "image calculator" function of ImageJ, by multiplying the RPD map (each lesion represented by 1 pixel with a value of 1) and the thresholded SDD binary map (each lesion represented by a group of pixels with values of 255). A histogram of the resulting multiplied image was used to determine the total number of pixels with a value of 255, representing the number of RPD lesions that overlap the SDD lesions.

Further analysis was performed to evaluate the proximity of the nonoverlapping RPD to the nearest

SDD. The thresholded SDD binary map was inverted. The "distance map" function was applied to the inverted image. The "image calculator" function was used to multiply the resulting distance map with the RPD map. A histogram of the resulting multiplied image was used to determine linear distance from each nonoverlapping RPD to the nearest SDD. The proximity of nonoverlapping RPD was categorized into 2 groups: 1 to 3 pixels and >3 pixels away from the nearest SDD.

Statistical Analysis

Statistical analysis was performed using Excel (Microsoft Excel for Mac 2011; Microsoft Corporation, Redmond, WA). One-way independent analysis of variance test was used for all experiments, and a *P* value <0.05 was considered significant. All results were expressed as mean \pm standard error of the mean. Interobserver calculation was performed by J. K. Fernandes and A. A. Fawzi for RPD identification (intraclass correlation coefficient).

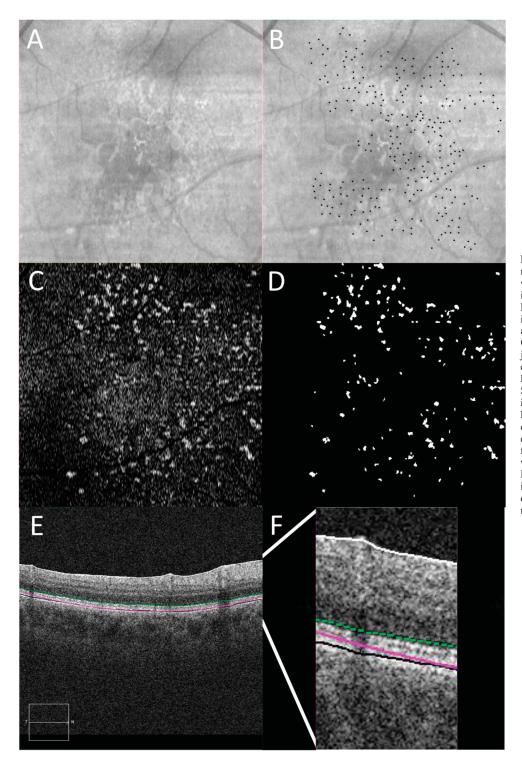
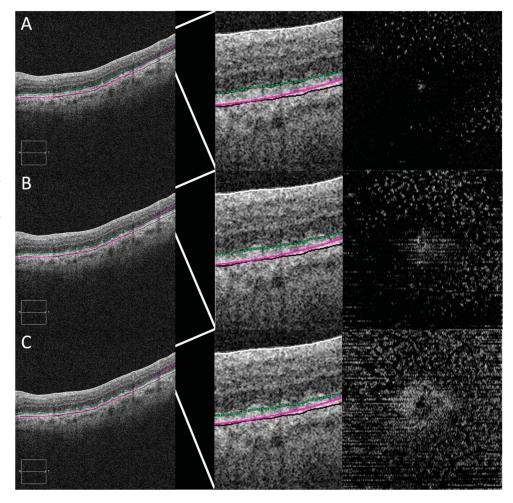


Fig. 3. Reticular pseudodrusen marked on IR and SDD on OCT with representative en face OCT image. A. Infrared reflectance. B. Infrared reflectance with individual RPD lesions manually marked with black dots. C. En face OCT C-scan taken just below the level of the external limiting membrane. D. En face OCT with individual SDD lesions semiautomatically identified, thresholded, and labeled in white. E. Optical coherence tomography B-scan demonstrating the location of en face OCT C-scan in (C) marked with a green dotted line. F. Magnified view of OCT B-scan in (E) to illustrate the location of the en face OCT C-scan marked with a green dotted line.

Results

Retrospective review identified 64 patients as having RPD on IR imaging between March 2012 and March 2014. Of the 128 eyes, 22 had incomplete ophthalmologic examinations and/or were missing either IR or OCT at the time of analysis. Twentyseven IR images were determined to be of insufficient quality to reliably identify RPD. Fourteen eyes had <50RPD lesions and were excluded, as in a previous study.¹⁸ Eleven OCT images were excluded because of failure of the Cirrus software RPE feature, leading Fig. 4. Subretinal drusenoid deposits on OCT B-scan and en face images at varying levels above RPE. Far left column: OCT B-scans with location of en face OCT (far right) marked with a green dotted line. Middle column: Magnified view of OCT B-scan (far left) with slab of en face OCT marked with a green dotted line. A. En face level taken too high above RPE resulting in too few SDD. B. En face level taken at lower location above RPE resulting in maximum number of welldefined SDD. C. En face level taken too close to RPE resulting in confluent SDD that cannot be precisely localized.



to artifacts in SDD slab. The remaining 54 eyes of 41 patients had good quality imaging and were included in this study.

Quantification of Reticular Pseudodrusen and Subretinal Drusenoid Deposits

For identification of RPD lesions on IR, there was excellent interrater reliability with an intraclass correlation coefficient of 0.96. The average number of RPD lesions in each eye as identified on IR images was 320 ± 44.62 . The average number of SDD lesions in each eye identified on C-scan images was 127 ± 26.02 . The difference was statistically significant using a oneway independent analysis of variance (P < 0.001; Figure 6).

Topographic Correlation Between Reticular Pseudodrusen and Subretinal Drusenoid Deposits

The percentage of total SDD overlapping study RPD in each eye was $2.91 \pm 0.87\%$ compared with $1.73 \pm 0.68\%$ SDD overlapping control (software

generated) RPD (P < 0.05). The SDD not overlapping RPD were categorized based on distance from the nearest RPD into 2 groups: either 1 to 3 pixels or greater than 3 pixels (30 μ m) away from RPD. There were 5.08 ± 1.40% of total SDD that were between 1 and 3 pixels from the nearest study RPD compared with 3.33 ± 1.07% control RPD (P < 0.05). There were 92.01 ± 13.14% of total SDD that were greater than 3 pixels from the nearest study RPD, which was not a statistically significant difference from control (software generated) RPD (94.94 ± 14.03%; Figure 7).

Discussion

In this study, quantification of IR and en face OCT images revealed significantly fewer RPD lesions compared with SDD lesions. Topographic analysis demonstrated that the majority of SDD were >30 μ m (3 pixels) away from the nearest RPD, not different from random sample (control RPD). The percentage of RPD overlapping SDD (2.9%) was significantly

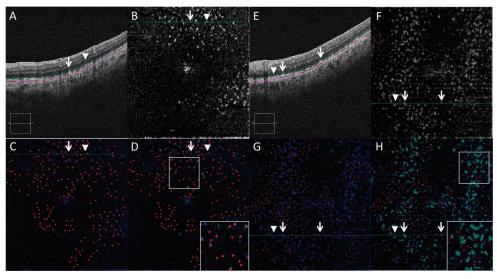


Fig. 5. Subretinal drusenoid deposits thresholding on en face SD-OCT image correlates with SDD on OCT B-scan image. White arrow: SDD lesion; white arrowhead: no SDD present. A and E. Optical coherence tomography B-scan with level of en face OCT C-scan marked with a green dotted line. B and F. En face OCT C-scan taken just below the level of the external limiting membrane. C and G. Composite image; blue channel: en face OCT C-scan taken just below the level of the external limiting membrane; red channel; individual RPD lesions marked with dots; channels merged using ImageJ. D and H. Composite image; blue channel: en face OCT C-scan taken just below the level of the external

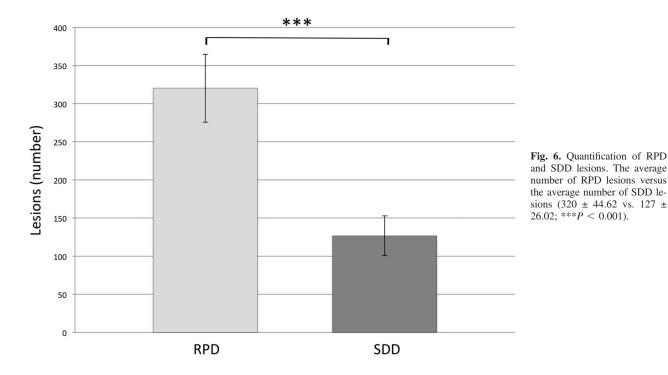
limiting membrane; red channel: individual RPD lesions marked with dots; green channel: thresholded SDD lesions on en face OCT C-scan; channels merged using ImageJ.

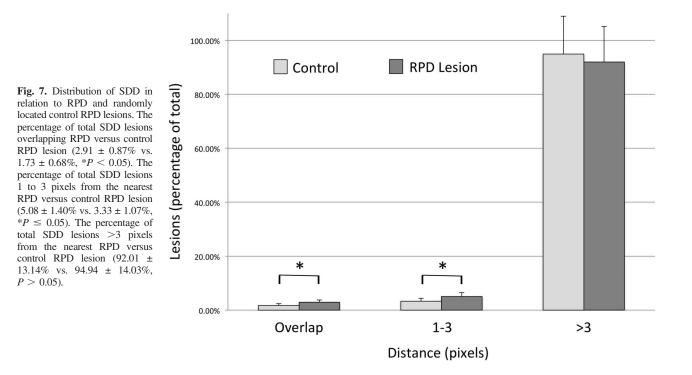
greater compared with control RPD lesions. Overall, these results provide evidence that these two lesions are topographically distinct, but it is possible that, at some stage of their evolution, they could be pathologically interrelated.

The relationship between RPD and SDD was first suggested by Sarks et al¹³ in 1988 who described small round dots between 25 μ m and 50 μ m in size seen in red-free photographs, appearing as hyperfluorescent lesions on fluorescein angiography. They speculated that these dots were caused by the rounded

mounds of membranous debris that had been characterized in numerous locations in histologic samples. However, this study failed to find histologic correlation of these dots, although the membranous debris had been well characterized.

Zweifel et al¹⁶ presented qualitative data to suggest the correlation between the lesions, based on the histologic features of SDD, suggesting interconnected subretinal material by sinuous bands. This interconnectedness was suspected to be a manifestation of the reticular pattern seen on evaluation under blue





light. However, the eye used for the histologic sections was not viewed under blue light before sectioning; hence, the direct relationship with RPD was not confirmed. In a single clinically confirmed RPD-containing specimen examined by Sarks et al, collections of membranous material were found in the subretinal space. More interestingly, similar collections of subretinal debris of the same composition have been demonstrated in specimens without any associated RPD lesions.¹³ The same group suggested that SDD could be viewed as localized hyporeflectivity from the RPE and account for the dark appearance of RPD seen on autofluorescence.²⁰

Sohrab et al¹⁷ characterized RPD on IR, autofluorescence, red-free images, and choroidal SD-OCT scans. This study suggested that RPD could not be seen in all areas of subretinal deposits or inner segment/outer segment disruptions on en face OCT sections but tended to be located adjacent to the SDD. Furthermore, these authors showed that RPD lesions bordered the edges of the underlying large choroidal vessels. The first study to suggest choroidal association of RPD was reported by Arnold et al⁵ when they examined a single donor eye with RPD, demonstrating a reticular arrangement of fibrotic changes to the choroidal stroma and a decrease in the number of large choroidal veins. The same specimen was found to have choroidal thinning and a reduction in choroidal melanocytes, with those remaining melanocytes being located in the outer choroid near the large arteries.

Numerous other studies have provided further evidence of choroidal changes associated with RPD.^{4,18,21–24} Querques et al used indocyanine green angiography along with SD-OCT to show RPD closely abutting, but not overlapping, the large choroidal vessels, with similar choroidal thinning as previously described. One observation made in multiple eyes with RPD has been impaired choroidal filling, which is consistent with a choroidal association and may provide a potential mechanism for RPD pathogenesis.^{3,5,25}

Although a choroidal association with RPD is an area of research focus, there has been little investigation into the relationship between the choroid and SDD. Recently, Vongkulsiri et al²⁶ provided the first evidence that SDD are not correlated with large choroidal vessels using a novel random stereological analysis. However, no other groups have corroborated the lack of large choroidal vessel involvement with either SDD or RPD, and the evidence supporting a relationship with RPD remains substantial.

The proposed mechanisms of both SDD and RPD pathogenesis offer a potential link between the two lesions. Therefore, the initial recognition of a choroidal association with RPD, choroidal ischemia, and potential fibrosis has been discussed as a potential mechanism for RPD formation and the appearance of RPD. Loss of the choriocapillaris²² and replacement of stroma between fewer large choroidal veins would lead to ischemia in the overlying RPE. Derangement

of the RPE could then cause aberrant accumulation of debris (SDD) above the RPE, perhaps due to metabolic derangements. The theories of pathogenesis of SDD have been more diverse than RPD, given their unusual localization to the subretinal space. The same derangement of the RPE leading to RPD could cause a loss of RPE cell anchors to the basement membrane and adjacent cells resulting in shedding debris into the subretinal space.²⁷ However, the membranous debris may also be a reaction to RPE cell stress, precipitated by ischemic, oxidative, or traumatic injury. The blebbing of cell membrane under these circumstances may represent exocytosis of damaging or damaged cell contents.²⁸ Conversely, the debris may represent material that failed to be phagocytized such as from the photoreceptor outer segments. Another suggested pathogenesis of SDD includes the disruption of lipoprotein transport between the photoreceptor outer segment and the RPE.²⁹ Further characterization of these lesions along with identification of associated clinical, optical, and histologic findings will help elucidate their pathogenesis.

In a recent study, Spaide³⁰ reported colocalization of RPD and SDD in 10 eyes using IR and en face SD-OCT, respectively. The study by Spaide differs from this study in its manual unmasked selection of a subsection of each experimental image. In addition, an image manipulation photoshop technique called "puppet warping" was used to align the IR and OCT images, with unknown potential effects on the relationship of SDD and RPD in the respective images. Puppet warping is a user-dependent technique with the potential for intrauser and interuser variability. Figure 8 demonstrates image distortion, resulting in a change in surface area of SDD lesions rather than image realignment alone, which was the initial goal of this technique. Therefore, in our study, we chose to exclude any OCT images with displacement artifacts that warrant manual OCT line realignment because we believe that any software manipulations could potentially confound our attempts at studying colocalization. These corrections would require the ability to realign each OCT B-scan individually, otherwise we may be concerned that manual realignment software approaches are bound to induce potential artifactual image distortion (Figure 8). Although image quality is a necessary limitation of all studies, choosing a portion of each image introduces another degree of selection bias to this study. Therefore, we excluded in our study any images with poor quality, even with sections of poor quality. Regarding the potential for mismatches between OCT and corresponding IR images, point-to-point colocalization errors of large magnitude have been reported, especially in Spectralis, and were shown to be directly proportional to the length of time taken to scan.³¹ Hence, in the study by Spaide³⁰ where dense rasters and averaging were performed to obtain these scans, these errors are more likely. In contrast, in this study, as well as previously published studies,¹⁷ manual registration and overlay was performed between IR and OCT cube from the same machine without the need for averaging or software registration,

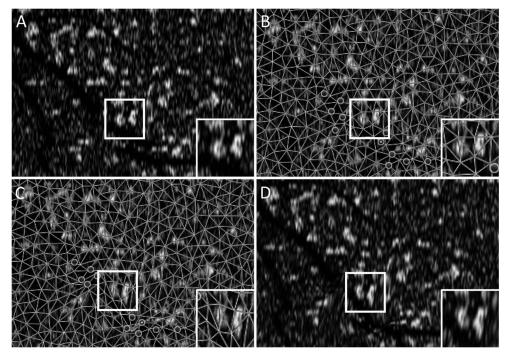


Fig. 8. Blood vessel with displacement artifact and adjacent SDD before and after puppet warping. A. Original en face image showing an artificially misaligned blood vessel broken by a displacement artifact. Inset image demonstrating unmodified SDD morphology. B. Same en face image with puppet warping anchors set before image modification. C. Modified en face image with "puppet warping" anchors adjusted to align a blood vessel with displacement artifact. Inset image demonstrating SDD morphology after puppet warping application. D. Modified en face image showing a realigned blood vessel using puppet warping.

which minimized potential mismatch error (Figures 1 and 2). 30

Limitations

This study has important limitations. We were not able to use a single threshold for all the en face C-scans because of variations in noise and signal strength during the time of image capture. However, a masked observer confirmed the accuracy of SDD lesion identification in five randomly selected images minimizing the likelihood of bias. Another potential source of bias was the manual marking of RPD on IR images, but again, a second masked observer confirmed these markings, with high interrater correlation. The lack of universally accepted criteria for RPD could lead to decreased generalizability of the results of this study. However, this is a commonly encountered limitation among early research subjects that was minimized using the most widely accepted definition of RPD. We have therefore chosen to use the IR reflectivity definition for RPD diagnosis in this study and all our previous studies. Finally, the exclusion of sample images based on set quality criteria introduces potential selection bias. However, the improved certainty of lesion identification with improved image quality far outweighs this potential for bias or artifacts induced by attempts at manual realignment and is therefore acceptable in the absence of more definitive or automated modalities for RPD lesion identification.

Conclusion

In a previous pilot study, we showed qualitative evidence that RPD and SDD may not be topographically identical.¹⁷ In this study, we provide further quantitative evidence in a larger number of eyes, demonstrating a significant quantitative and topographic difference between RPD and SDD. Based on this evidence, further investigation of the relationship between these two entities is warranted.

Key words: age-related macular degeneration, en face optical coherence tomography, infrared reflectance, reticular pseudodrusen, subretinal drusenoid deposits.

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