Introduction

Based on biochemical characteristics, *Yersinia enterocolitica* is divided into several distinct biotypes (BTs) (BT 1A, 1B, 2, 3, 4 and 5), of which the pathogenic BTs (1B, 2, 3, 4 and 5) are responsible for the fourth most common foodborne zoonosis in Europe (EFSA, 2013). The most common human pathogenic *Y. enterocolitica* in the European Union and Switzerland are of bioserotype 4/O:3 (Fredriksson-Ahomaa et al., 2012; EFSA, 2013). Pigs have been characterized as the main reservoir for pathogenic *Y. enterocolitica* BT 4 and are supposed to be the most important infection hazard for humans (EFSA, 2013), but other mammals, e.g., sheep, cattle, cats and dogs, are also discussed as sources of human *Y. enterocolitica* BT 4 infection (Fearnley et al., 2005; Stamm et al., 2013). BT 4 strains are detected in carcasses and in pig feces at slaughter, as well as in tonsils, where they are present at a up to 6-fold higher detection rate than in feces (Fredriksson-Ahomaa et al., 2001; Nesbakken et al., 2003). However, the route of infection and the impact of different sources (tonsils versus feces) have not been clarified.

Although previous studies on molecular typing have demonstrated the genetic relatedness of porcine and human *Y. enterocolitica* (Kuehni-Boghenbor et al., 2006; Laukkanen et al., 2009), the characterization of virulence capability by screening for virulence-associated genes is required to clarify the role of porcine BT 4 in human disease. All pathogenic *Y. enterocolitica* isolates host the 70-kb virulence plasmid (pYV), which is indispensable for full virulence (Cornelis et al., 2006; Laukkanen et al., 2009).
Various virulence-associated genes located on the pYV were used in previous studies as a target to detect pathogenic Y. enterocolitica strains by PCR (Fredrikkson-Ahomaa and Korkeala, 2003). The yadA gene encodes the Yersinia adhesion A protein, relevant for host cell attachment and for complement antibiotic resistance (Galindo et al., 2011). The virF gene product is a key factor for transcription of yop genes (Comelis et al., 1998) and is, therefore, fundamental for the type III secretion system. Because pYV can be lost during the culturing process (Thoerner et al., 2003), chromosomal genes should be analyzed as well. The ail (attachment invasion locus) gene has been described as a target for virulence screening (Fredrikkson-Ahomaa and Korkeala, 2003; Thisted-Lambertz et al., 2008), as it is only found in strains associated with human infection (Miller et al., 1989). Several studies have reported that certain BT 1A isolates may host the ail gene as well. For this reason, the potential pathogenicity of BT 1A strains has previously been addressed (Bumens et al., 1996; Kraushaar et al., 2011; Paixão et al., 2013a,b; Sihvonen et al., 2011a). The inv gene encodes invasin, a protein functioning in the invasion of and adhesion to the host cell. This gene is present in all Y. enterocolitica isolates but non-functional in BT 1A (Pierson and Falkow, 1990). The expression of inv is positively regulated by rovA (regulator of virulence) and negatively regulated by ymoA (Yersinia modulating protein) (Ellison et al., 2003; Revell and Miller, 2000). The ystA (Yersinia stable toxin A) gene is uniquely found in pathogenic BTs, whereas BT 1A frequently bears the ystB (Yersinia stable toxin B) gene (Bhagat and Virdi, 2007; Stephan et al., 2013). Another virulence-associated gene is the mytA (mucoid Yersinia factor) gene, which encodes fimbriae (Revell and Miller, 2001).

In previous studies, the characterization of Y. enterocolitica biosootypes by analyzing the virulence gene pattern has produced strong evidence of swine as the main reservoir for human pathogenic strains. However, several of these studies focused solely on porcine isolates (Bonardi et al., 2013; Paixão et al., 2013a,b) or human strains (Stephan et al., 2013; Fredrikkson-Ahomaa et al., 2012). Or only a small number of virulence-associated genes were analyzed (Thoerner et al., 2003). To our knowledge, no previous study has used this same approach to analyze BT 4 strains from porcine tonsils, porcine feces and human feces for a complete spectrum of relevant virulence-associated genes.

Y. enterocolitica is known to be highly susceptible to most antibiotics with the exception of penicillin, ampicillin and first-generation cephalosporins. This intrinsic antibiotic resistance is based on 2 well-known chromosomal beta-lactamase genes, blaA and blaB (Fàbrega and Vila, 2012). The occurrence of multiresistant human pathogenic Y. enterocolitica strains and a high proportion of antibiotic resistance to chloramphenicol, streptomycin and sulfonamides in porcine strains in Italy have recently been described (Sihvonen et al., 2011b; Bonardi et al., 2014). These findings raise questions about the current situation in Swiss isolates. Previous studies have used the disc diffusion method to conduct antimicrobial susceptibility testing for Swiss Y. enterocolitica isolates (Baumgartner et al., 2007; Fredrikkson-Ahomaa et al., 2012). However, this method is known to produce high rates of incorrect results (Meyer et al., 2011). Accordingly, broth microdilution was used in this study.

Thus, the aim of this study was to i) characterize Y. enterocolitica BT 4 from human clinical cases in comparison with porcine BT 4 strains for a broad spectrum of relevant virulence-associated genes, ii) to include porcine Y. enterocolitica isolates from both tonsils and feces to detect possible source-dependent gene patterns and iii) to characterize Y. enterocolitica BT 4 isolates from Switzerland for their antibiotic susceptibility pattern based on the broth microdilution method.

### 2 Materials and Methods

#### 2.1 Y. enterocolitica strains

Eighty-seven BT 4 Y. enterocolitica were analyzed. Thirty-eight strains were isolated from Swiss slaughter pigs from March 2012 to February 2013 and another thirteen strains were isolated with fecal swabs from Swiss slaughter pigs between July and December 2013 (Büttrner et al., 2013). Isolation and biotyping was based on the International Standards Organization (ISO 10273:2003) isolation method for Y. enterocolitica in food (unpublished data). The strains were serotyped for O:3, O:5, O:8, O:9 and O:27 by slide agglutination with commercially available antisera (SIFIN, Berlin, Germany).

Thirty-six human fecal BT 4 isolates had been collected between 2001 until 2003 at the Federal Office of Public Health (FOPH) (Kuehni-Boghenbor et al., 2006).

All strains were preserved in Trypticase Soy Bouillon (TSA) (Becton Dickinson, Franklin Lakes, New Jersey, USA) with 30% glycerol and stored at −80 °C until further analysis.

#### 2.2 Detection of virulence-associated genes by PCR

Virulence-associated genes of Y. enterocolitica isolates were detected by conventional PCR. DNA was prepared from overnight-growth colonies on TSA at 30 ± 1 °C. A few colonies were transferred to lysis buffer (0.1 M Tris–HCl, pH 8.5, 0.05% Tween 20, 0.24 mg/ml protease K) and incubated for 1 h at 60 °C following 15 min at 97 °C. DNA extracts were stored at −20 °C for further analyses. Nine virulence-associated genes were tested chromosomal genes (ail, inv, rovA, ymoA, ystA, ystB and mytA) and 2 plasmid-borne genes (yadA, virF). For virF, ail, inv, ymoA, ystA, ystB and mytA, the primers and cycling parameters described by Bhagat and Virdi (2007) were used. For rovA the protocols defined by Divya and Varadaraj (2011) and for yadA the protocols by Thoerner et al. (2003) were used (Table 1). Two microliters of template was added to 28 µl of reaction mixture consisting of 11 µl PCR buffer (Solis BioDyne, Tartu, Estonia), 2.5 mM MgCl₂ (Solis BioDyne, Tartu, Estonia), 200 µM dNTPs (Roche, Rotkreuz, Switzerland), 0.25 µl FIREPol DNA polymerase (Solis BioDyne, Tartu, Estonia) and primers (Microsynth, Balgach, Switzerland) at a final concentration of 0.25 µM.

| Table 1. Y. enterocolitica biotype 4 serotypes and virulence<sup>–</sup>gene patterns from strains isolated out of different sources. |
|-----------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| **Material**    | **ST** | **No.** | **ail** | **inv** | **ystA** | **ystB** | **mytA** | **rovA** | **ymoA** | **yadA** | **virF** |
| **Soy Bouillon**|       |        |       |       |       |       |       |       |       |       |       |
2.3 Antimicrobial testing

The minimal inhibitory concentration (MIC) of the antibiotics was determined by broth microdilution in cation-adjusted Mueller-Hinton broth using the Sensititre susceptibility plate EUMVS2 (Sensititre, TREK Diagnostic System, Cleveland, Ohio, USA; TREK Diagnostic System, East Grinstead, West Sussex, England). The antibiotics and the concentration ranges are presented in Tables 2 and 3. The plates were incubated for 18–24 h at 30 ± 1 °C. Isolates were classified as susceptible or resistant according to clinical breakpoints for Enterobacteriaceae issued by the European Committee on Antimicrobial Susceptibility Testing (EUCAST) guidelines version 4.0 (www.eucast.org). If EUCAST breakpoints were unavailable, human clinical breakpoints for Enterobacteriaceae from Clinical and Laboratory Standards Institute (CLSI) documents M100-S22 (Clinical and Laboratory Standards Institute, 2013) were used. For florfenicol and streptomycin, no breakpoints from EUCAST or CLSI were available. Therefore, MIC50 and MIC90 were calculated. Escherichia coli ATCC 25922 was used as the control strain with 37 ± 1 °C as the incubation temperature. Measured MICs were in the appropriate range for this strain.

Table 2 Minimal inhibitor concentration (MIC) from of porcine tonsil Y. enterocolitica biotype 4 isolates.

<table>
<thead>
<tr>
<th>Antimicrobial agent</th>
<th>No of strains with MIC (reg L)</th>
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<tbody>
<tr>
<td>Ampicillin</td>
<td>54/2</td>
</tr>
<tr>
<td>Amoxicillin</td>
<td>32/2</td>
</tr>
<tr>
<td>Chloramphenicol</td>
<td>3/2</td>
</tr>
<tr>
<td>Ciproflaxacin</td>
<td>58/30</td>
</tr>
<tr>
<td>Cotrimoxazole</td>
<td>58/2</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>55/25</td>
</tr>
<tr>
<td>Kanamycin</td>
<td>55/25</td>
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<tr>
<td>Netilmicin</td>
<td>55/25</td>
</tr>
<tr>
<td>Tetracycline</td>
<td>55/25</td>
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<tr>
<td>Trimethoprim</td>
<td>55/25</td>
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</tbody>
</table>

Numbers indicate the number of isolates with corresponding MIC value. White areas indicate the range of dilutions tested for each antimicrobial agent; values above or below this range denote MIC values greater than the highest concentration tested and MIC values smaller than or equal to the lowest concentration tested, respectively. Vertical lines indicate clinical breakpoints, when two vertical lines exist, the lower breakpoint indicate susceptibility and the higher breakpoint...
This finding is in contrast to the results of other studies that determined almost 100% bioserotype 4/O:3 for both porcine and human samples. This discrepancy can be explained by methodological differences in serotyping. In other studies, only antiserum O:3 was used, whereas sera against O:8 or O:9 were used in this study. The results show that the detection rate in the current study, with 92.1% of yadA-positive porcine tonsil strains, was still higher than that in recent studies on porcine or human BT 4 strains (54–83%) (Zheng et al., 2008; Stephan et al., 2013; Bonardi et al., 2013). As we tested 2 plasmid-borne genes and the strains still carry the virF gene, it is unlikely that the lack of the yadA gene is a consequence of a plasmid loss during the cultivation process. Two tested BT 4 strains from porcine tonsils lacked the inv gene. It has been reported that inv-negative strains still have the ability to invade the host cells (Pepe and Miller, 1993). Therefore, it is possible that the 2 porcine isolates lacking inv are still pathogenic. In addition to the abovementioned virulence-associated gene patterns, the ymoA and rovA genes were tested. Both of these genes are involved in the regulation of inv expression. Moreover, ymoA is also relevant for the modulation of the expression of the virF and yst genes (Revell and Miller, 2000) (Ellison et al., 2003). Our results showed that, independent of the source, all tested isolates possessed both effector and regulator genes. To the authors' knowledge these findings are not demonstrated up to now elsewhere.

### 3.2 Serotype distribution

The tested isolates comprised a broad spectrum of known STs (O:3; O:6; O:27; O:3,8,27; O:8,27 and O:9,27). The most frequent STs were O:3 and O:3,27, followed by O:8,27, except for human isolates, which did not show ST O:8,27 (Table 1). This finding is in contrast to the results of other studies that determined almost 100% bioserotype 4/O:3 for both porcine and human strains. This discrepancy can be explained by methodological differences in serotyping. In other studies, only antiserum O:3 (Laukkonen et al., 2009) or a different panel of antisera was used (Bonardi et al., 2013). In the present study, the virulence-associated gene pattern was independently detected from STs for both porcine and human isolates. Thus, determination of STs is of limited value for the characterization of pathogenic Y. enterocolitica isolates.
3.3 Antimicrobial testing

In general, the majority of all strains of this study were susceptible to almost all the tested antibiotics. In contrast, 100% antibiotic resistance to ampicillin was found (Tables 2 and 3). The high rate of resistance to β-lactam antibiotics has been well described (Fábrega and Vila, 2012), and these findings also agree with previous studies of the antimicrobial antibiotic resistance of porcine BT 4 strains from Switzerland (Fredriksson-Ahomaa et al., 2007; Baumgartner et al., 2007). Although human BT 4 isolates shared the overall antibiotic susceptibility phenotype with porcine isolates, 2 human BT 4 isolates possessed antibiotic resistance to chloramphenicol and nalidixic acid. One strain was also resistant to sulfamethoxazole. Y. enterocolitica antibiotic resistance to chloramphenicol is rarely described in the literature, but Bonardi et al. (2014) recently found BT 4 strains resistant to chloramphenicol. Additionally, Sihvonen et al. (2011b) detected antibiotic resistance to chloramphenicol in human Y. enterocolitica strains. For streptomycin, the human and the porcine tonsil strains showed an MIC50 of 8 mg/L and an MIC90 of 16 mg/L. In contrast, the 2 chloramphenicol-resistant strains showed an elevated MIC of >128 mg/L, which indicated that they are resistant to streptomycin as well. The origin of this antibiotic resistance pattern (chloramphenicol, streptomycin and sulfonamide) is most likely a conjugative plasmid, as chloramphenicol is rarely described in the literature, but these findings also agree with previous studies of the antimicrobial antibiotic resistance of porcine BT 4 strains from Switzerland (Fredriksson-Ahomaa et al., 2012), and a substantial increase in nalidixic acid resistance in human clinical isolates has been detected in Spain between 1995 and 2002 (Capilla et al., 2004). MIC50 and the MIC90 of florfenicol were 4 mg/L each for isolates from porcine tonsils and from porcine feces. Only 1 porcine isolate exhibited an elevated MIC to florfenicol (MIC = 8 mg/L). For human isolates MIC50 of florfenicol was 2 mg/L and the MIC90 = 4 mg/L, respectively.

4 Conclusions

The virulence-associated gene pattern detected in the human strains from clinical cases is supposed to be sufficient for human infection. The same virulence-associated gene pattern was found in the strains isolated from porcine tonsils as well as porcine feces, demonstrating the potential to cause human infection for all of the Swiss porcine Y. enterocolitica BT 4 strains tested. Therefore, measures must be taken at slaughter to avoid Y. enterocolitica cross-contamination from tonsils as well as fecal contamination of the carcasses.

Recent studies in Europe have reported increasing antibiotic resistance of both human and porcine Y. enterocolitica isolates (Sihvonen et al., 2011b; Bonardi et al., 2014). Low level of antibiotic resistance found in Swiss human strains from 2001 to 2003 underline the increasing tendency in human strains, whereas the rate of acquired antibiotic resistance in recently isolated predominant Swiss porcine Y. enterocolitica BT 4 strains is currently very low. However, the use of antibiotics in Swiss livestock is still considerable. Therefore, the prudent use of antimicrobials in veterinary medicine is important for the control of antibiotic resistance phenomenon in Swiss Y. enterocolitica isolates.

References


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**Highlights**

- The same virulence-associated gene pattern were found in *Y. enterocolitica* BT 4 strains isolated from porcine tonsils and as well as from porcine feces are equal.
- Porcine *Y. enterocolitica* BT 4 strains are capable to infect humans despite the source of isolation and serotype.
- The rate of acquired resistance in predominant Swiss porcine *Y. enterocolitica* BT 4 strains is currently low.

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**Queries and Answers**

**Query:**

The data “O3”, “O8”, “O9” and “O27” are also presented in the text as “O:3”, “O:8”, “O:9” and “O:27”. Please check if this should be made consistent, and amend if necessary.

**Answer:** Corrections were made in the table, so that data are consistent.

**Query:**

Your article is registered as a regular item and is being processed for inclusion in a regular issue of the journal. If this is NOT correct and your article belongs to a Special Issue/Collection please contact p.sivakumar@elsevier.com immediately prior to returning your corrections.

**Answer:** Our article do not belongs to special issue.

**Query:**

Please confirm that given names and surnames have been identified correctly.

**Answer:** The given names and surnames are correct.

**Query:**

Highlights should only consist of 125 characters per bullet point, including spaces. The highlights provided are too long; please edit them to meet the requirement.

**Answer:** The first highlight was shortened. The second and third highlight were not shortened as they consist of 112 and 108 characters, respectively.
The citation “Paixão et al., 2013” has been changed to match the author name/date in the reference list. Please check here and in subsequent occurrences, and correct if necessary.

Answer: The citations are correct.

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The citation “Paixão et al. 2013” has been changed to match the author name/date in the reference list. Please check here and in subsequent occurrences, and correct if necessary.

Answer: The citations are correct.

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Please provide the volume number and page range for the bibliography in 'Stephan et al., 2013'.

Answer: The volume number and page range is provided.