Antibiotic-Resistant *Neisseria gonorrhoeae* Spread Faster with More Treatment, Not More Sexual Partners

Stephanie M. Fingerhuth¹,²*, Sebastian Bonhoeffer¹, Nicola Low², Christian L. Althaus²

¹ Institute of Integrative Biology, ETH Zurich, Zurich, Switzerland, ² Institute of Social and Preventive Medicine (ISPM), University of Bern, Bern, Switzerland

*stephanie.fingerhuth@env.ethz.ch

Abstract

The sexually transmitted bacterium *Neisseria gonorrhoeae* has developed resistance to all antibiotic classes that have been used for treatment and strains resistant to multiple antibiotic classes have evolved. In many countries, there is only one antibiotic remaining for empirical *N. gonorrhoeae* treatment, and antibiotic management to counteract resistance spread is urgently needed. Understanding dynamics and drivers of resistance spread can provide an improved rationale for antibiotic management. In our study, we first used antibiotic resistance surveillance data to estimate the rates at which antibiotic-resistant *N. gonorrhoeae* spread in two host populations, heterosexual men (HetM) and men who have sex with men (MSM). We found higher rates of spread for MSM (0.86 to 2.38 y⁻¹, mean doubling time: 6 months) compared to HetM (0.24 to 0.86 y⁻¹, mean doubling time: 16 months). We then developed a dynamic transmission model to reproduce the observed dynamics of *N. gonorrhoeae* transmission in populations of heterosexual men and women (HMW) and MSM. We parameterized the model using sexual behavior data and calibrated it to *N. gonorrhoeae* prevalence and incidence data. In the model, antibiotic-resistant *N. gonorrhoeae* spread with a median rate of 0.88 y⁻¹ in HMW and 3.12 y⁻¹ in MSM. These rates correspond to median doubling times of 9 (HMW) and 3 (MSM) months. Assuming no fitness costs, the model shows the difference in the host population’s treatment rate rather than the difference in the number of sexual partners explains the differential spread of resistance. As higher treatment rates result in faster spread of antibiotic resistance, treatment recommendations for *N. gonorrhoeae* should carefully balance prevention of infection and avoidance of resistance spread.

Author Summary

More and more infectious disease treatments fail because the causative pathogens are resistant to the drugs used for treatment. For the treatment of *Neisseria gonorrhoeae*, a sexually transmitted bacterium, drug resistance is a particularly big problem: there is only a single antibiotic left that is recommended for treatment. We aimed to understand how antibiotic-resistant *N. gonorrhoeae* spread in a sexually active host population and how the spread of
resistance can be slowed. From antibiotic resistance surveillance data, we first estimated the rate at which antibiotic-resistant \textit{N. gonorrhoeae} spread. Second, we reproduced the observed dynamics in a mathematical model describing the transmission between hosts. We found that antibiotic-resistant \textit{N. gonorrhoeae} spread faster in host populations of men who have sex with men than in host populations of heterosexuals. We could attribute the faster spread of resistant pathogens to higher treatment rates. This finding implies that promoting screening to control antibiotic-resistant \textit{N. gonorrhoeae} could in fact accelerate their spread.

\section*{Introduction}

Antibiotic-resistant \textit{Neisseria gonorrhoeae} can evolve and spread rapidly \citep{1}. Resistance is commonly observed against the antibiotic classes penicillin, tetracycline and fluoroquinolones \citep{2–4}. Resistance also emerged against cefixime, an oral third generation cephalosporin, in recent years \citep{2, 3}. Since 2010, cefixime is no longer recommended as first-line treatment \citep{5} following guidelines from the World Health Organization (WHO) that an antibiotic should not be used when more than 5\% of \textit{N. gonorrhoeae} isolates are resistant \citep{6}. Injectable ceftriaxone, in combination with oral azithromycin, is now the last antibiotic remaining as recommended first-line treatment \citep{7}. Although other antibiotics are being tested for their safety and efficacy for \textit{N. gonorrhoeae} treatment \citep{8}, no new classes of antibiotics are currently available \citep{4} and management of antibiotics is urgently needed to preserve their efficacy. The current management strategy tries to reduce the overall burden of \textit{N. gonorrhoeae} infection by expanded screening and treatment of hosts \citep{9, 10}, but the outcome of this strategy for resistance is uncertain. Understanding the drivers of resistance spread and anticipating future resistance trends will provide rationales for antibiotic management and help to improve antibiotic treatment strategies.

Men who have sex with men (MSM) are host populations that have higher levels of antibiotic-resistant \textit{N. gonorrhoeae} than heterosexual host populations \citep{3}. In a study \citep{5} based on the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP) in England and Wales, cefixime-resistant \textit{N. gonorrhoeae} were mainly found in MSM until 2011. The authors suggested that cefixime resistance was circulating in a distinct sexual network of highly active MSM and that bridging between MSM and heterosexuals was necessary for subsequent spread among heterosexual hosts. However, cefixime-resistant \textit{N. gonorrhoeae} might have already been spreading undetected in the heterosexual host population.

Mathematical models can help explain the differential observations of antibiotic-resistant \textit{N. gonorrhoeae} in different host populations. In 1978, Yorke et al. \citep{11} introduced the concept of core groups to model the transmission of \textit{N. gonorrhoeae}. The concept of core groups posits that an infection can only be maintained in a host population if a highly sexually active group of hosts is responsible for a disproportionate amount of transmissions. More recent modeling studies have examined the transmission of antibiotic-resistant \textit{N. gonorrhoeae}. Chan et al. \citep{12} found that prevalence rebounds more quickly to a pre-treatment baseline when treatment is focused on the core group. Xiridou et al. \citep{13} developed a \textit{N. gonorrhoeae} transmission model to determine the impact of different treatment strategies on the prevalence of \textit{N. gonorrhoeae} in Dutch MSM. They found that increased treatment rates could increase the spread of resistance, whereas re-treatment could slow it down. Hui et al. \citep{14} used an individual-based \textit{N. gonorrhoeae} transmission model in a heterosexual host population to investigate the effect of a molecular resistance test on the time until 5\% resistance are reported. None of these studies has investigated or explained the differences in the spread of antibiotic-resistant \textit{N. gonorrhoeae} in MSM and heterosexual host populations.
In this study, we investigated the dynamics and determinants of antibiotic-resistant *N. gonorrhoeae* spread using surveillance data and mathematical modeling. We estimated the rates at which resistance spreads in heterosexual men (HetM) and MSM using surveillance data from the USA and from England and Wales. We then developed a mathematical model of *N. gonorrhoeae* transmission to reconstruct the observed dynamics of resistance spread. This allowed us to determine the major driver of resistance spread, and to explore the expected rates at which resistance spreads in MSM and heterosexual host populations.

**Methods**

**Data**

**Data sources.** We used data from the GRASP [15, 16] and the Gonococcal Isolate Surveillance Project (GISP) [17]. GRASP is a program of Public Health England (PHE) that monitors antibiotic-resistant *N. gonorrhoeae* in England and Wales. GISP is an equivalent program from the Centers for Disease Control and Prevention (CDC) in the USA. We used Plot Digitizer 2.6.6 [18] to digitize data on the proportion of cefixime- and ciprofloxacin-resistant *N. gonorrhoeae* from figures that were published online (see S1 and S2 Tables).

**Rate of spread.** We determined the rate of resistance spread by assuming that the proportion of antibiotic-resistant *N. gonorrhoeae* follows logistic growth. We used the least squares function *nls* from the R software environment for statistical computing [19] to fit the following function to the data:

\[
f(t) = \frac{c}{1 + a \times \exp(-bt)}.
\]

*f(t)* represents the proportion of antibiotic-resistant *N. gonorrhoeae* at time *t*, *c* is the maximal proportion of antibiotic-resistant *N. gonorrhoeae* (carrying capacity), *a* is the ratio between antibiotic-sensitive and -resistant *N. gonorrhoeae* at time 0, and *b* is the rate at which the proportion of antibiotic-resistant *N. gonorrhoeae* increases in the initial exponential growth phase. We only used data from the years before the first decline in the proportion of resistant *N. gonorrhoeae* because we were interested in the rate of resistance spread during the initial exponential growth phase and while the antibiotic was still used.

**Model**

**Transmission model.** We developed a mathematical model to describe the spread of antibiotic-resistant *N. gonorrhoeae* in a given host population [12]:

\[
\dot{S}_i = -S_i \pi \sum_{j \in G} \rho_j \beta_j \frac{I_{senj} + I_{resj}}{N_j} + v(I_{senj} + I_{resj}) + \tau(1 - \mu)I_{seni} - \alpha S_i + \gamma S_i + \gamma N \sum_{j \in G} S_j,
\]

\[
\dot{I}_{seni} = S_i \pi \sum_{j \in G} \rho_j \beta_j \frac{I_{senj} + I_{resj}}{N_j} - vI_{seni} - \tau I_{seni} - \alpha I_{seni} + \gamma I_{seni} + \gamma N \sum_{j \in G} I_{senj},
\]

\[
\dot{I}_{resi} = S_i \pi \sum_{j \in G} \rho_j \beta_j \frac{I_{resj} + I_{senj}}{N_j} - vI_{resi} + \tau I_{resi} - \alpha I_{resi} + \gamma I_{resi} + \gamma N \sum_{j \in G} I_{resj},
\]
Sen and Res indicate the antibiotic-sensitive and -resistant *N. gonorrhoeae* strains, \( G = \{ L, H \} \) is the set of low and high sexual activity groups and \( i \in G \) (Fig 1). Each sexual activity group \( N_i \) consists of susceptible hosts, \( S_i \), hosts infected with an antibiotic-sensitive strain, \( I_{\text{Sen},i} \), and hosts infected with an antibiotic-resistant strain, \( I_{\text{Res},i} \). To account for individual heterogeneity in sexual behavior [20], hosts are redistributed to either the same or the other sexual activity group at rate \( \gamma \). Redistribution is proportional to the size of the sexual activity group, i.e. hosts from the larger sexual activity group are less likely to change their sexual behavior than hosts from the smaller sexual activity group. Hosts can also enter or leave the population at rate \( \alpha \). Susceptible hosts become infected depending on the partner change rate, \( \pi_i \), the transmission probability per partnership, \( \beta_{ij} \), and the sexual mixing matrix, \( \rho_{ij} \), which describes how many partnerships are formed within and outside the host’s activity group:

\[
\rho_{ij} = \epsilon \delta_{ij} + (1 - \epsilon) \frac{\pi_i N_i}{\sum_{k \in G} \pi_k N_k},
\]

where \( \delta_{ij} = 1 \) if \( i = j \) and \( \delta_{ij} = 0 \) if \( i \neq j \). \( \epsilon \) is the sexual mixing coefficient [21]. It ranges from 0 (random or proportionate mixing) to 1 (assortative mixing, i.e. all partnerships are formed with hosts from same group). Hosts infected with an antibiotic-sensitive strain can recover spontaneously at rate \( \nu \) or receive treatment at rate \( \tau \). Treatment occurs both when the host seeks treatment for a symptomatic infection or is screened and diagnosed with an asymptomatic infection. Hosts receiving treatment recover at rate \( \tau (1 - \mu) \) and develop resistance during treatment with probability \( \mu \). Hosts infected with an antibiotic-resistant strain can only recover spontaneously at rate \( \nu \). We assumed equal fitness of antibiotic-sensitive and -resistant strains in absence of treatment, i.e. no fitness costs for the antibiotic-resistant strain. We evaluated the impact of fitness costs on the model outcomes in a sensitivity analysis (see S1 Appendix).

**Parameters.** Model parameters were estimated from sexual behavior data, calibrated through model simulation or informed by literature. The partner change rate and the proportions of the host population in each sexual activity group were estimated from the second
British National Survey of Sexual Attitudes and Lifestyles (Natsal-2) [22], a population-based cross-sectional survey. For the heterosexual men and women (HMW) model population, we used the number of new heterosexual partners in the last year of all male and female participants between 16–44 years who reported never having had a homosexual partner. For the MSM model population, we used the number of new homosexual partners in the last year of all male participants between 16–44 years who reported ever having had a homosexual partner. For each host population, the number of partners per year was weighted with weights provided in Natsal-2 to adjust for unequal selection probabilities in the survey. We estimated the partner change rate by assuming that the reported numbers of new sexual partners can be described by two Poisson distributions with means \( \pi_L \) and \( \pi_H \), weighted by the proportion of individuals in each sexual activity group [23]. For HMW, the sexual partner change rates are \( \pi_L = 0.25 \text{ y}^{-1} \) and \( \pi_H = 4.57 \text{ y}^{-1} \) with \( N_H = 6.3\% \) of the population being in the high sexual activity group and \( N_L = 1 - N_H \). The obtained partner change rates for MSM are \( \pi_L = 0.41 \text{ y}^{-1} \) and \( \pi_H = 30.49 \text{ y}^{-1} \) with \( N_H = 5.3\% \) of the population belonging to the high sexual activity group and \( N_L = 1 - N_H \).

We calibrated the sexual mixing coefficient, \( \epsilon \), the fraction of diagnosed and treated infections, \( \phi \), the average duration of infection, \( D \), and the per partnership transmission probabilities within the low, \( \beta_{LL} \), and the high sexual activity group, \( \beta_{HH} \), to \( N. \text{gonorrhoeae} \) prevalence and incidence using the following algorithm:

1. Define prior parameter distributions (Table 1).
2. Define the ranges for the expected prevalence and incidence of diagnosed infections (Table 2) of urethral and cervical \( N. \text{gonorrhoeae} \) infections for HMW, and urethral, rectal and pharyngeal infections for MSM.
3. Randomly draw 10^7 parameters sets from prior distributions.
4. Simulate the transmission model until it approaches a resistance-free (\( \mu = 0 \)) endemic equilibrium using the ordinary differential equation solver runsteady from the R [19] package rootSolve [24].
5. Select the parameters sets (posterior distributions) that result in prevalences and incidences within the defined range.

Information about parameter estimates for \( N. \text{gonorrhoeae} \) is scarce, so we chose to use non-informative priors for all parameters except the duration of infection which was informed by Garnett et al. [25]. The ranges for the expected prevalence and incidence of diagnosed infections are described in Table 2.

Table 1. Prior distributions and posterior estimates of model parameters.

<table>
<thead>
<tr>
<th>parameter</th>
<th>description</th>
<th>priors</th>
<th>M_{MSM}</th>
<th>IQR_{MSM}</th>
<th>M_{HMW}</th>
<th>IQR_{HMW}</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \epsilon )</td>
<td>sexual mixing coefficient</td>
<td>( U(0, 1) )</td>
<td>0.57</td>
<td>0.30–0.80</td>
<td>0.73</td>
<td>0.53–0.89</td>
</tr>
<tr>
<td>( \phi )</td>
<td>fraction of diagnosed and treated infections</td>
<td>( U(0, 1) )</td>
<td>0.64</td>
<td>0.48–0.81</td>
<td>0.50</td>
<td>0.36–0.66</td>
</tr>
<tr>
<td>( D )</td>
<td>average duration of infection (years)</td>
<td>( \Gamma(2, 0.125) )</td>
<td>0.19</td>
<td>0.14–0.25</td>
<td>0.55</td>
<td>0.46–0.66</td>
</tr>
<tr>
<td>( \beta_{LL} )</td>
<td>transmission probability within low activity group</td>
<td>( U(0, 1) )</td>
<td>0.59</td>
<td>0.42–0.77</td>
<td>0.87</td>
<td>0.79–0.94</td>
</tr>
<tr>
<td>( \beta_{HH} )</td>
<td>transmission probability within high activity group</td>
<td>( U(0, \beta_{LL}) )</td>
<td>0.30</td>
<td>0.25–0.40</td>
<td>0.72</td>
<td>0.63–0.81</td>
</tr>
</tbody>
</table>

We assumed that the duration of infection is described by a gamma distribution \( \Gamma(k, \theta) \) with shape parameter \( k = 2 \) and scale parameter \( \theta = 0.125 \text{ y} \) resulting in an average infectious duration of 0.25 y. Because highly sexually active hosts have fewer sex acts per partnership, we assumed that the transmission probability within the high activity group cannot be higher than the transmission probability within the low activity group. M and IQR represent the median and interquartile range of the posterior distributions.

doi:10.1371/journal.ppat.1005611.t001
Table 2. Prevalence and incidence ranges used for model calibration.

<table>
<thead>
<tr>
<th>parameter</th>
<th>infection site</th>
<th>host population</th>
<th>sexual activity group</th>
<th>range</th>
</tr>
</thead>
<tbody>
<tr>
<td>prevalence</td>
<td>urethral, cervical</td>
<td>HMW</td>
<td>low</td>
<td>0–0.38%</td>
</tr>
<tr>
<td>prevalence</td>
<td>urethral, cervical</td>
<td>HMW</td>
<td>high</td>
<td>0.16–100%</td>
</tr>
<tr>
<td>prevalence</td>
<td>urethral, cervical</td>
<td>HMW</td>
<td>either</td>
<td>0.16–0.38%</td>
</tr>
<tr>
<td>prevalence</td>
<td>pharyngeal, anal, urethral</td>
<td>MSM</td>
<td>low</td>
<td>0–2.79%</td>
</tr>
<tr>
<td>prevalence</td>
<td>pharyngeal, anal, urethral</td>
<td>MSM</td>
<td>high</td>
<td>1.19–100%</td>
</tr>
<tr>
<td>prevalence</td>
<td>pharyngeal, anal, urethral</td>
<td>MSM</td>
<td>either</td>
<td>1.19–2.79%</td>
</tr>
<tr>
<td>incidence</td>
<td>urethral, cervical</td>
<td>HMW</td>
<td>either</td>
<td>0.12–0.36% person(^{-1}) y(^{-1})</td>
</tr>
<tr>
<td>incidence</td>
<td>pharyngeal, anal, urethral</td>
<td>MSM</td>
<td>either</td>
<td>5.88–7.19% person(^{-1}) y(^{-1})</td>
</tr>
</tbody>
</table>

Prevalence and incidence ranges for HMW were based on the National Health and Nutrition Examination Survey [26] and surveillance data [27], both from CDC. For MSM, prevalence and incidence ranges were based on the Health in Men Study in Australia [28, 29]. The upper and lower bound of the ranges for the low and high sexual activity groups are given by the lower and upper bound of the overall population.

doi:10.1371/journal.ppat.1005611.t002

Table 3. Composite model parameters.

<table>
<thead>
<tr>
<th>parameter</th>
<th>description</th>
<th>formula</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \tau )</td>
<td>treatment rate per year</td>
<td>( \frac{\phi D}{\phi + D} )</td>
</tr>
<tr>
<td>( \nu )</td>
<td>spontaneous recovery rate per year</td>
<td>( \frac{1}{\phi} )</td>
</tr>
<tr>
<td>( \beta_{LH} )</td>
<td>transmission probability per partnership between low and high sexual activity host</td>
<td>( \sqrt{\beta_{LL} \beta_{HH}} )</td>
</tr>
<tr>
<td>( \beta_{HL} )</td>
<td>transmission probability per partnership between high and low sexual activity host</td>
<td>( \sqrt{\beta_{LL} \beta_{HH}} )</td>
</tr>
</tbody>
</table>

The composite model parameters \( \tau \) and \( \nu \) relate to other model parameters with \( D = \frac{1}{\phi} \) and \( \phi = \frac{1}{\psi} \). We assumed that the transmission probabilities between hosts of different sexual activity groups are given by the geometric mean of the transmission probabilities for hosts within each group.

doi:10.1371/journal.ppat.1005611.t003
Results

We fitted a logistic growth model to the proportion of antibiotic-resistant *N. gonorrhoeae* as observed in the two gonococcal surveillance programs (Fig 2). The proportion of cefixime-resistant *N. gonorrhoeae* in GRASP appears to increase for both HetM and MSM after 2006. Ciprofloxacin-resistant *N. gonorrhoeae* in HetM and MSM were spreading in all observed host populations after the year 2000. For a given antibiotic and surveillance program, the rates of resistance spread were consistently higher for MSM than for HetM (Table 4). The average rate of resistance spread was 0.53 y\(^{-1}\) for HetM and 1.46 y\(^{-1}\) for MSM, corresponding to doubling times of 1.3 y (HetM) and 0.5 y (MSM) during the initial exponential growth phase.

Next, we studied the transmission of *N. gonorrhoeae* and the spread of resistance in the dynamic transmission model. We calibrated five model parameters to expected prevalence and incidence in MSM and HMW host populations. The posterior distributions of the parameters were based on 2,779 parameter sets for HMW and 65,699 parameter sets for MSM (Fig 3, Table 1). Distributions of the modeled prevalence and incidence of diagnosed infections after

![Graph showing increase in antibiotic-resistant *N. gonorrhoeae*. Points show data from antibiotic resistance surveillance programs (GRASP and GISP). Dashed lines indicate the fit of the logistic growth model to the data. For a given antibiotic and surveillance program, the rates of spread in MSM (green) are consistently higher than those in HetM (blue).](image)

**Table 4. Rates of resistance spread in *N. gonorrhoeae* surveillance programs.**

<table>
<thead>
<tr>
<th>program</th>
<th>antibiotic</th>
<th>years</th>
<th>host population</th>
<th>rate (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GRASP</td>
<td>Cefixime</td>
<td>2004–2010</td>
<td>HetM</td>
<td>0.86 (0.73–1.00) y(^{-1})</td>
</tr>
<tr>
<td>GRASP</td>
<td>Cefixime</td>
<td>2004–2010</td>
<td>MSM</td>
<td>2.38 (1.72–3.03) y(^{-1})</td>
</tr>
<tr>
<td>GRASP</td>
<td>Ciprofloxacin</td>
<td>2000–2009</td>
<td>HetM</td>
<td>0.24 (0.03–0.45) y(^{-1})</td>
</tr>
<tr>
<td>GRASP</td>
<td>Ciprofloxacin</td>
<td>2000–2009</td>
<td>MSM</td>
<td>1.15 (0.76–1.54) y(^{-1})</td>
</tr>
<tr>
<td>GISP</td>
<td>Ciprofloxacin</td>
<td>1995–2006</td>
<td>HetM</td>
<td>0.50 (0.45–0.55) y(^{-1})</td>
</tr>
<tr>
<td>GISP</td>
<td>Ciprofloxacin</td>
<td>1995–2006</td>
<td>MSM</td>
<td>0.86 (0.66–1.06) y(^{-1})</td>
</tr>
</tbody>
</table>

Estimated rates of resistance spread from the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP, England and Wales) and from the Gonococcal Isolate Surveillance Project (GISP, USA). CI: confidence interval.
calibration are provided as Supporting Information (S1 and S2 Figs, S3 Table). The sexual mixing coefficient showed a tendency towards assortative mixing in both MSM and HMW (Fig 3a). The fraction of diagnosed and treated infections tended to be higher in MSM compared to HMW (Fig 3b), whereas the infectious duration was considerably shorter in MSM (median: 2.3 months, IQR: 1.7–3.0 months) than in HMW (median: 6.6 months, IQR: 5.5–7.9 months) (Fig 3c). The transmission probabilities per partnership were generally higher in HMW than in MSM (Fig 3d and 3e).

After calibration, we used the dynamic transmission model to study the spread of antibiotic-resistant \( N. \) gonorrhoeae. The proportion of antibiotic-resistant \( N. \) gonorrhoeae increased faster in MSM than in HMW (Fig 4). The fraction of diagnosed and treated infections tended to be higher in MSM compared to HMW (Fig 3b), whereas the infectious duration was considerably shorter in MSM (median: 2.3 months, IQR: 1.7–3.0 months) than in HMW (median: 6.6 months, IQR: 5.5–7.9 months) (Fig 3c). The transmission probabilities per partnership were generally higher in HMW than in MSM (Fig 3d and 3e).

Antibiotic-sensitive and -resistant \( N. \) gonorrhoeae share the same resource for growth, i.e. the susceptible hosts. The rate at which one strain replaces the other strain in the host population is given by the difference in their net growth rates. We assume that the transmission probabilities and the infectious duration of the two strains are the same. Since the probability of resistance during treatment is very small (\( \mu \ll 1 \)), the difference in net growth rates between the strains is approximated by the treatment rate \( \tau \) and corresponds to the rate of spread of
antibiotic-resistant \textit{N. gonorrhoeae}. The observed distributions of treatment rates from the
transmission model hardly overlap between HMW and MSM (Fig 5). The median treatment
rates, i.e. the approximated median rates of resistance spread in the transmission model are
3.12 y\(^{-1}\) (MSM) and 0.88 y\(^{-1}\) (HMW).

We tested whether changes in the probability of resistance during treatment, \(\mu\), and fitness
costs in the antibiotic-resistant strain alter the model outcomes. Higher probabilities of resis-
tance during treatment accelerate the establishment of antibiotic-resistant \textit{N. gonorrhoeae}
in the population and hence reduce the time until 5% resistance is reached (S3 Fig). Higher
probabilities of resistance during treatment, however, do not affect rates of spread, unless the

\[ \text{Fig 4. Spread of antibiotic resistance in the transmission model.} \quad \text{Ranges indicating 50\% of all simulations are shown in dark color, and ranges indicating}
95\% of all simulations are shown in light color. The continuous lines describe the median proportion of antibiotic-resistant \textit{N. gonorrhoeae} for all simulations.}
The black dotted line indicates the 5\% threshold.\]

doi:10.1371/journal.ppat.1005611.g004

\[ \text{Fig 5. Distribution of treatment rates in HMW and MSM.} \quad \text{Treatment rates closely approximate the rates of resistance spread. The median treatment}
rate was 0.88 y\(^{-1}\) in HMW and 3.12 y\(^{-1}\) in MSM.} \]

doi:10.1371/journal.ppat.1005611.g005
probability of resistance during treatment is unrealistically high (10%) (S4 Fig). Fitness costs in the antibiotic-resistant strain result in rates of resistance spread that are lower than the treatment rate $\tau$ (Fig. B in S1 Appendix). Fitness costs that reduce the transmission probability per partnership, $\beta_{ij}$, have a stronger effect than fitness costs that reduce the duration of infection. The effects of fitness costs are independent of the sexual partner change rate, $\pi$, and $\beta_{ij}$ if they affect the duration of infection, but can vary with $\pi$ and $\beta_{ij}$ if they affect the transmission probability per partnership (Fig. C in S1 Appendix). While high fitness costs can prevent the spread of antibiotic-resistant strains (Fig. A in S1 Appendix), fitness costs between 0%–10% have only small effects on the rates of resistance spread (Fig. B in S1 Appendix).

**Discussion**

In this study, we quantified the rate at which antibiotic-resistant *N. gonorrhoeae* spread in heterosexual and MSM populations. We used data from two different surveillance programs and estimated that the proportion of ciprofloxacin- and cefixime-resistant *N. gonorrhoeae* doubles on average every 1.3 y in HetM and 0.5 y in MSM. The faster spread of antibiotic-resistant *N. gonorrhoeae* in MSM than in heterosexual hosts was corroborated using a dynamic transmission model, which was calibrated to observed prevalence and incidence rates. The model allowed us to identify the higher treatment rates in MSM, compared with heterosexual hosts, as the major driver for the faster spread of antibiotic-resistant *N. gonorrhoeae*.

To our knowledge, this is the first study to have analyzed and interpreted *N. gonorrhoeae* antibiotic resistance surveillance data in a dynamic and quantitative manner. The transmission model was parameterized using sexual behavior data for HMW and MSM from Natsal-2 [22], a large probability sample survey of sexual behavior. Calibrating the model to observed prevalence and incidence rates allowed us to use largely uninformative priors for the model parameters. The calibration makes our model more robust to changes in parameters than using fixed parameter values, especially since for *N. gonorrhoeae* available parameter values are very uncertain [31]. It also allowed us to rely on few assumptions about the natural history of *N. gonorrhoeae* infection.

The limitations to our study need to be taken into consideration when interpreting the findings. First, we used data from different sources, although all were collected in high income countries. The antibiotic resistance surveillance data are from programs in England and Wales and the USA. The mathematical transmission model was parameterized using British sexual behavior data [22] and calibrated to prevalence and incidence rates from the USA (HMW) [26, 27] and Australia (MSM) [28, 29]. For simplicity, we modeled the heterosexual and MSM host populations separately although there is some mixing between them. We assumed the sexual behavior of heterosexual men and women to be the same and pooled their behavioral data. Second, we assumed complete resistance against the antibiotic, i.e. 100% treatment failure. We further assumed that treatment of the sensitive strain is 100% efficacious. Both assumptions might explain why antibiotic-resistant *N. gonorrhoeae* spread at somewhat higher rates in the dynamic transmission model than estimated from data. Third, we restricted our model to resistance to one antibiotic with no alternative treatment or interventions. This is why we observe complete replacement of the antibiotic-sensitive strain in the model, a phenomenon that has not been observed in surveillance data. Fourth, resistance in our model is treated as a generic trait, but it likely depends on the underlying molecular mechanisms and possibly the genetic background of the *N. gonorrhoeae* strain. Different resistance mechanisms might explain some of the differences in the rates of resistance spread between the model and the different antibiotics from the surveillance data. Fifth, we did not include co- and superinfection with antibiotic-sensitive and -resistant *N. gonorrhoeae* strains. Since genetic typing provides evidence for
mixed infections [32], it is worth speculating how they would affect the rate of spread from the transmission model. If antibiotic-sensitive and -resistant strains co-existed in a host and acted independently, we would not expect significant effects on the rate of spread. In contrast, if there was competition between the two strains within a host, the rate of spread would increase if the antibiotic-resistant strain outcompetes the -sensitive strain, and decrease otherwise.

Sixth, we do not consider importation of resistance from another population. For example, importation of resistance from other countries might play a particularly important role during the early phase of resistance spread, when stochastic events can lead to extinction of the antibiotic-resistant strain. We expect that a high rate of importation of antibiotic resistance shortens the time to reach 5% resistance drastically, but that once the resistant strain is established in the population, importation hardly affects the rate of resistance spread. Finally, we assumed that the transmission probabilities per partnership and the durations of infection in the model represent average values for *N. gonorrhoeae* infections at different infection sites (urethral, pharyngeal, anal, cervical).

The estimated posterior distributions of the parameters fit within the range of previously used values, and provide some insights into sexual mixing and the natural history of *N. gonorrhoeae*. The sexual mixing coefficient tends to be assortative for both HMW and MSM host populations in our model. Quantifying the degree of sexual mixing is difficult and largely depends on the study population, but our finding is consistent with other studies indicating assortative sexual mixing in the general population [30, 33]. The posterior estimates of the fraction of diagnosed and treated infections are consistent with the notion that a large proportion of *N. gonorrhoeae* infections are symptomatic, and that this proportion is expected to be higher in men than in women [34–36]. The average duration of infection was the only parameter with an informative prior, but we found marked differences between the duration of infection in HMW (6.6 months) and MSM (2.3 months). Per sex act transmission probabilities are generally considered to be lower from women to men than vice versa [37–39]. In our model, the median of the transmission probability per partnership was lower in MSM hosts than in HMW for both sexual activity groups. This could be explained by different numbers of sex acts per partnership in the two populations. The low transmission probability within the highly active MSM group (median: 30%) could reflect a single or a small number of sex acts per partnership. In contrast, the high transmission probability for HMW within the low sexual activity group (median: 87%) could be a result of a larger number of sex acts per partnership in those individuals. Furthermore, condom use is more frequent in MSM than in HMW [22], which could explain part of the observed differences in transmission probabilities.

Our study found that the treatment rate is the driving force of resistance spread. Xiridou et al. [13] found that resistance could spread faster when the treatment rate was higher, but they did not identify the treatment as the major driver of resistance spread. Chan et al. [12] found that focusing treatment on the core group leads to a faster rebound to pre-treatment prevalence than equal treatment of the entire host population. Unfortunately, our findings cannot be compared with Chan et al. because they do not report the proportion of antibiotic-resistant *N. gonorrhoeae*.

It was shown previously that treatment is the main selective force acting on antibiotic evolution due to the selective advantage to the resistant pathogen [40, 41]. We now expand this concept by showing that, assuming no fitness costs, treatment rates determine the rates of resistance spread even when the host populations has a heterogeneous contact structure. The intuitive argument that a faster spread of an infection, due to a higher number of sexual partners, will result in a faster spread of resistance does not hold. Instead, the proportion of resistant infections spreads equally in host populations with different number of partners as long as they receive treatment at the same rate and there are no fitness costs associated with the
transmission probability per partnership. For \textit{N. gonorrhoeae}, this insight challenges the current management strategy that aims to lower the overall burden of infection by expanding screening and treatment of hosts \cite{9,10}. As soon as antibiotic-resistant pathogens are frequent enough to evade stochastic extinction, expanded treatment will foster their spread and increase the burden of \textit{N. gonorrhoeae}. Additionally, we show that fitness costs can decelerate or even prevent the spread of antibiotic-resistant \textit{N. gonorrhoeae} strains. Fitness costs therefore might explain why highly resistant strains, such as the ceftriaxone-resistant \textit{N. gonorrhoeae} strain H041, do not spread in the host population after their first detection \cite{42}. Our findings also show that bridging between the HetM and the MSM host populations might not have been necessary for cefixime-resistance to spread in the HetM population after 2010 \cite{5}. It is likely that cefixime-resistant \textit{N. gonorrhoeae} had already been present in the HetM population but were spreading at a lower rate than in the MSM population.

The results of our study will be useful for future \textit{N. gonorrhoeae} research and for guiding treatment recommendations. The \textit{N. gonorrhoeae} transmission model describes observed prevalence and incidence rates well and can reconstruct the spread of antibiotic-resistant \textit{N. gonorrhoeae}. Estimating rates of resistance spread is useful for projecting future resistance levels and the expected time it will take until a certain threshold in the proportion of antibiotic-resistant \textit{N. gonorrhoeae} is reached. Until now, treatment recommendations for \textit{N. gonorrhoeae} are subject to change when 5\% of \textit{N. gonorrhoeae} isolates show resistance against a given antibiotic \cite{6}. Our study shows the importance of the rate of spread: a level of 5\% resistance results in a marginal increase to 8\% in the following year if resistance spreads logistically at rate 0.53 y$^{-1}$ (HetM mean estimate from Table 4), but reaches 18\% in the next year if resistance spreads at rate 1.46 y$^{-1}$ (MSM mean estimate from Table 4). Public health authorities could use surveillance data and adapt thresholds for treatment recommendation change to specific host populations using the method we describe. Our study challenges the currently prevailing notion that more screening and treatment will limit the spread of \textit{N. gonorrhoeae}, as higher treatment rates will ultimately result in faster spread of antibiotic resistance. Future treatment recommendations for \textit{N. gonorrhoeae} should carefully balance prevention of \textit{N. gonorrhoeae} infection and avoidance of the spread of resistance.

**Supporting Information**

\textbf{S1 Table. Digitized data from the Gonococcal Resistance to Antimicrobials Surveillance Programme (GRASP).} Data for heterosexual men (HetM) and men who have sex with men (MSM).

(PDF)

\textbf{S2 Table. Digitized data from the Gonococcal Isolate Surveillance Project (GISP).} Data for men who have sex with women (MSW) and men who have sex with men (MSM). We used the MSW data as heterosexual men (HetM) data.

(PDF)

\textbf{S3 Table. Prevalence and incidence of diagnosed and treated infections after model calibration.}

(PDF)

\textbf{S1 Fig. Posterior distributions of (a) prevalence and (b) incidence of diagnosed and treated infections for MSM.}

(TIFF)
S2 Fig. Posterior distributions of (a) prevalence and (b) incidence of diagnosed and treated infections for HMW.
(TIFF)

S3 Fig. Sensitivity of time to 5% resistance towards changes in the probability of resistance during treatment, $\mu$. The time to 5% resistance of both MSM (green) and HMW (blue) are sensitive towards $\mu$. Lower and upper bound of the box indicate the first and third quartiles, bar in the box indicates median, whiskers span 1.5 times IQR. Outliers are shown in orange and are outside 1.5 times IQR.
(TIFF)

S4 Fig. Sensitivity of rate of spread towards changes in the probability of resistance during treatment, $\mu$. The rates of spread of both MSM (green) and HMW (blue) are only sensitive towards $\mu$ when $\mu$ is unrealistically high. Lower and upper bound of the box indicate the first and third quartiles, bar in the box indicates median, whiskers span 1.5 times IQR. Outliers are shown in orange and are outside 1.5 times IQR.
(TIFF)

S1 Appendix. Fitness costs and spread of resistance.
(PDF)

Acknowledgments
We would like to thank Sandro Gsteiger and Denise Kühnert for advice on parameter calibration, and Fengyi Jin and Andrew Grulich for providing the MSM prevalence and incidence data from the Health in Men Study.

Author Contributions
Conceived and designed the experiments: SMF SB NL CLA. Performed the experiments: SMF. Analyzed the data: SMF SB NL CLA. Wrote the paper: SMF SB NL CLA.

References


