The Total Number of Acini Remains Constant throughout Postnatal Rat Lung Development

Sébastien F. Barré¹, David Haberthür², Tiziana P. Cremona¹,
Marco Stampanoni²,³, and Johannes C. Schittny¹.

¹ Institute of Anatomy Bern, University of Bern, Bern, Switzerland
² Swiss Light Source, Paul Scherrer Institute, Villigen, Switzerland
³ Institute for Biomedical Engineering, Swiss Federal Institute of Technology and University of Zürich, Zürich, Switzerland.

Running Head. Constant Number of Acini during Rat Lung Development

Correspondence. Johannes C. Schittny
University of Bern, Institute of Anatomy,
Baltzerstrasse 2, CH-3012 Bern, Switzerland
Phone +41 31 631 4635, FAX +41 31 631 3807,
E-mail: johannes.schittny@ana.unibe.ch
Abstract

The pulmonary airways are subdivided into conducting and gas-exchanging airways. The small tree of gas-exchanging airways which is fed by the most distal conducting airway represents an acinus. Very little is known about the development of the number of acini. The goal of this study was to estimate their number throughout rat postnatal development. Right middle rat lung lobes were obtained at postnatal day 4-60, stained with heavy metals, paraffin embedded, and scanned by synchrotron radiation based X-ray tomographic microscopy or imaged using micro computed tomography after critical point drying. The acini were counted by detection of the transitional bronchioles (bronchioalveolar duct junction; BADJ) using morphological criteria (thickness of the walls of airways and appearance of alveoli) during examination of the resulting 3D image stacks. Between postnatal days 4-60, the number of acini per lung remained constant (5840 ± 547 acini), but their volume increased significantly. We conclude that the acini are formed before the end of the saccular stage (before postnatal day 4) and that the developmental increase of the lung volume is achieved by an increase of the acinar volume and not by an increase of their number. Furthermore, our results propose that the bronchioalveolar stem cells, which are residing in the BADJ, are as constant in their location at the BADJ itself.

Keywords. Pulmonary acinus, Lung development, bronchioalveolar duct junction, BADJ bronchioalveolar stem cells, BASCs
Introduction

In 1974, P. H. Burri explained (8): “Another type of alveolar formation consists in the transformation of originally purely conducting into respiratory airways” and in 1984 (7): “The branching pattern and structure of conducting airways are mature at birth, so that except for the terminal bronchiole (where transformation into respiratory passage may occur) the adult bronchial tree is the replica of the newborn one”. His statements were based on observations made by Boyden and Tompsett on bronchial trees of human infants and dog puppies (5, 6). To our best knowledge — due to the complexity of lung architecture and to technical limitations — nobody has quantitatively followed the development of the pulmonary acini until now.

The term acinus (plural: acini) is generally used to describe the few generations of the bronchial tree which are located distally of a terminal bronchiole. The latter is defined as the most distal purely conducting generation of airways. One acinus represents the functional unit of the lung. Because rat belong to the animals which do not possess respiratory bronchioles (41) a transitional bronchiole opens directly into the most proximal alveolar duct(s) of the acinus. The transitional bronchioles contain the bronchioalveolar duct junction where the bronchioalveolar stem cells are located. In human lungs, which possess respiratory bronchioles (41) the entrance of the acini is defined at the junction of the terminal and respiratory bronchioles. The bronchioalveolar junction is defined as the entrance of a ventilator unit (38). Again at this junction the bronchioalveolar stem cells are located. Therefore, the best human correlate of the rat acinus is the human ventilator unit.
Due to their complex three-dimensional structure, acini cannot be detected by two-dimensional investigations. Therefore, several approaches were proposed to overcome this limitation. Casting methods were used by Yeh et al. (46) and by Rodriguez et al. (24). Serial-sectioning was used by Mercer and Crapo (21), while Wulfsohn et al. (45) developed an estimation technique based on a disector of five consecutive sections. However, as explained in Barré et al. (3) these methods require tedious work, and thus are not suited for studies with larger number of samples. Several publications proposed approaches based on X-ray tomography (10, 17, 18, 20, 34-36, 42). Even if the latter approach would be suitable for the analysis of larger number of sample, to our best knowledge, it has not been done until now, especially not for the analysis of the development of the acini.

Lung developmental is divided in the embryonic, fetal, and postnatal periods (7, 30, 33, 44). In the embryonic period, the lung anlage appears as two independent outpouchings of the foregut forming the two lung buds. The formation of the airways starts shortly afterwards by branching morphogenesis which is defined as a repetitive branching and outgrowth of the future airways. Typically, in human lungs, the branching of the airways follows a dichotomous pattern. This means that the airways symmetrically branch into two smaller airways of the same diameter. With the appearance of the pleura the embryonic period blends over into the fetal period. Branching morphogenesis continues and at the end of the pseudoglandular stage approximately 20 airway generations are formed in humans (16). The airway formation continues during the next step, the canalicular stage. At the end of this stage the transition between conducting and gas-exchanging airways is detectable, resulting in the “birth of the acinus” (4) even if the proximal part of the acinar airways is already formed during the pseudoglandular stage. Branching morphogenesis
comes to its end at latest during the saccular stage, by the arising of saccules and of the type 1 and 2 alveolar epithelial cells at the distal ends of the bronchial tree. The next and final stage, alveolarization, is composed of two phases (classic and continued) (32, 39) which ends during young adulthood (12, 14, 31). This description of the stages of lung development is in general valid for all mammalians independently of their airways branching pattern (28). The main differences are found in the duration of the stages and at the time point of birth.

While human lungs possess a dichotomous branching pattern, lungs of rats and other rodents possess a monopodial one (23, 46). At most of the branching points the airway divides asymmetrically into one larger and one smaller airway. In rats, the trachea divides into two main bronchi from which main lobar bronchi, one per lobe, emerge to ventilate the five lung lobes. A system of secondary and tertiary lobar bronchi arises from the next larger category to ventilate the peripheral parts of the lobes. Acini are directly connected by transitional bronchioles to purely conducting airways of various generations, even including secondary lobar bronchi (3, 24).

The goal of this study was to estimate the number of acini for rats at different ages (postnatal days 4, 10, 21, 36, and 60). Taking P.H. Burri’s statements (7, 8) as starting point, we studied the development of the pulmonary acini in rats. We used a time-efficient estimation protocol developed in our laboratory (3) in order to investigate large amounts of samples. Surprisingly, the number of acini showed no statistical differences between the days studied indicating that the number of acini remains constant during lung development. Furthermore we conclude that the positions of the entrance of the acini, as well as the sites where the bronchioalveolar
stem cells are residing, are defined in the bronchial tree before the end of the saccular stage.

**Materials and Methods**

**Animals.** Lungs of twenty-seven male rats (postnatal days 4, 10, 21, 36, and 60; Wistar Bern) were obtained after fixation with 4% paraformaldehyde in phosphate buffered saline via tracheal instillation at a constant pressure of 20 cm water column. The lung was removed from the chest cavity and the pressure was maintained during fixation for a minimum of two hours at 4°C in order to prevent a recoiling of the lung (19, 22, 40). After fixation, all lobes were separated and their volume was measured by water displacement (26). A second volume measurement was performed using the Cavalieri principle (9) on the tomographic dataset to determine the shrinkage factor of the samples.

All animal studies were approved by and conducted in accordance with the Veterinary Service of the Canton of Bern, Switzerland and the Swiss Federal Agency for Environment, Forest and Landscape.

**X-ray Tomography.** Two tomographic methods were used during this study: synchrotron radiation based x-ray tomographic microscopy (SRXTM) and micro computed tomography (μCT). The right middle lung lobes were prepared, as already described (3, 25), either by critical point drying for μCT (15) or by heavy metal
staining with osmium tetroxide and uranyl acetate (33), and paraffin embedding for SRXTM.

**Synchrotron radiation based x-ray tomographic microscopy.** 20 samples were scanned at the TOMCAT beamline (37) of the Swiss Light Source synchrotron facility at the Paul Scherrer Institute (Villigen, Switzerland). X-rays at energy of 20 keV were converted to visible light by a scintillator (20 µm thick LuAG:Ce or 18 µm thick YAG:Ce, Crytur Ltd., Turnov, Czech Republic) after passing the samples. An optic microscope magnified the visible light in order to obtain effective voxel size lengths of 1.75 to 3.5 µm. In order to visualize the entire lung lobe, the field of view of the microscope was increased perpendicular to the rotational axis using ‘wide-field SRXTM’ (three field of views) (11) or 360° scans (two field of views). In addition, five to seven wide-field scans were stacked parallel to the rotational axis, resulting in 3D-stacks of 8 bit grayscale images of up to 7500x7500 pixels per slice.

**Micro computed tomography.** Seven samples were imaged by a µCT device (SkyScan 1172, Bruker, Billerica, MA, USA) at 33 kV and 204 µA without filtering. Two to three oversize scans in the vertical direction were needed to visualize the entire sample at 2.5-3.5 µm voxel side length. The GPU reconstruction software (NReconServer64bit, Bruker, Billerica, MA, USA) was used on a GeForce GTX 680 graphic card (Nvidia Corp., Santa Clara, Ca, USA) to create 8 bit grayscale image stacks of approximately 4000x4000x2500 voxels per scan. Additional samples (right upper, right lower, cardiac lobe and left lung) at days 4 and 60 were also scanned in the µCT.
**Detection and Counting of Acini.** The manual acini counting followed the protocol described by Barré et al. (3). The acini entrances (i.e. transition from conducting to gas-exchanging airways) were detected based on morphological criteria (thickness of airway walls and appearance of alveoli) by scrolling through tomographic data sets representing right middle lung lobes (n = 27). In addition, three entire lungs were counted at days 4 and 60 to validate that the right middle lobe is a valid estimator of the entire lung. The software Fiji (27) was used to crop and display sub-stacks of 250 to 500 images, in order to reduce the computing power requirement, and to manually label the detected acini entrances. The labels were manually counted using a laboratory counter (Clay Adams, New York, USA). After one sub-stack was counted, labels of overlapping acini were reported to the next sub-stack to exclude double counting. All data were analyzed on a Dell Precision T5500 work station (Intel Xeon X5650 (six Core, 2.67 GHz), 24 GB RAM, Windows 7 Professional 64).

**Three-dimensional Visualization.** The conducting airways of five right middle lung lobes (one per developmental time point) were visualized three-dimensionally adapting a protocol described by Barré et al. and Haberthür et al. (3, 10). Briefly, several images stacks of 250-500 eight bit greyscale images were analyzed using MeVisLab (version 2.1, 2010-07-26 Release, MeVis Medical Solutions and Frauenhofer MEVIS-Institute for Medical Image Computing, Bremen, Germany). The sub-stacks were loaded as TIFF files and segmentation stoppers were manually set at the acinar entrances to separate them from the conducting airways. The conducting airways were segmented using a gray-level threshold-based region growing algorithm (47) after down-sampling with a factor of 2. The segmentation seed points were manually set within the conducting airways. All segmentations were reconstructed as one 3D model using a custom-made MeVisLab pipeline. This basic
pipeline stacked all segmentations and displayed spheres at the position of the segmentation stoppers.

Statistics. The statistical analysis was done using Microsoft Excel (version 14.0.7106.5003, 32-bit) and Prism (version 5.04, GraphPad Software Inc.). The values are expressed as means (± standard deviation). The linear regression of the observed values and the $R^2$ (coefficient of determination) were calculated. This coefficient indicates how well the observed values fit the ideal values of a linear regression. In this case, $R^2$ is the square of the Pearson correlation coefficient ($r$). No correlation between parameters was assumed when $R^2 \leq 0.5$ ($|r| \leq 0.7$), weak correlation when $0.5 < R^2 \leq 0.7$ ($0.7 < |r| \leq 0.84$), and strong correlation when $0.7 < R^2$ ($0.84 < |r|$). In addition, multiple regression and ANOVA F-tests were used (both using the data analysis toolbox on Microsoft Excel)(1). A multiple regression test was used to determine if observations groups were correlating. To do so, one group was expressed as the dependent variable and the others as independent variables. The predicted values (i.e. regression results) were compared to the values of the group set as dependent variable using the coefficient of determination. The standard score (difference of observation and group mean divided by group standard deviation) was used to normalize the data prior to regression analysis. The significance of difference between the means of observation groups was achieved using a T-test or using one way ANOVA F-test. A significance level of $\alpha = 95\%$ was used for both tests, detecting significant difference if $p \leq 0.05$ or non-significant difference if $p > 0.05$. In addition to ANOVA, a Bonferroni’s multiple comparison tests was performed to define which group was significantly different from the other groups (1). T-test and ANOVA require normal distributed data. Therefore QQ-plots were used to test for normal-distributed values.
Calculations. The total number of acini present in a rat lung was calculated by dividing the counted number of acini in the right middle lobe by the parenchymal volume of the corresponding lobe and by multiplying this result with the total parenchymal volume. The mean acinar volume (tissue plus airspace of an acinus) was calculated by dividing the lobe volume by the counted number of acini and multiplied by the volume density of parenchyma (Tab. 1). The parenchymal volume density was estimated according to the ATS guidelines (13).

Results

Detection of the acinar entrance throughout lung development. In tomographic datasets morphological criteria (thickness of the airways walls and appearance of alveoli) were used to detect the entrances of the acini (Fig. 1) as described in Barre et al. (2). Because the acini are not fully developed and the alveoli not yet formed at day 4, we used the appearance of the gas-exchanging capillaries in the walls of the airspaces as additional, newly introduced criterion for the detection of the acinus entrance. Due to the iron contained within the erythrocytes, the contrast inside the capillaries was large enough to be safely detected. This criterion was also used at day 10, while it was not necessary to be used for lungs at older age. Although acinar airways are still immature, the alveolar duct already presents its typical shape (airway walls covered with uprising new alveolar septa) at the beginning of the stage of alveolarization. The latter contrasted with the tubular wall of the conducting airways.
Therefore, we used it as the second additional criterion for the detection of the transition between conducting and gas-exchanging airways.

**Growth of lobe volume.** We compared the growth of the five lung lobes throughout postnatal lung development (Tab. 2). In particular, we asked if its fraction of the total lung volume changes during development. No significant changes were observed for the right upper, right middle, and right lower lobe (Tab. 2). However, the fraction of the left lung decreased between postnatal days 4-10 and subsequently stays constant. The volume fraction of the cardiac lobe increases inversely proportional to the left lung between postnatal days 4-10. Afterward only a small additional increase was observed until day 60.

**Validation of the sampling.** According to Barré et al. (3), the right middle lung lobe is a valid estimator of the number of acini for the entire lung for adult rats. The estimation was based on the parenchymal volume of the entire lung and of the right middle lobe. In order to test if the right middle lobe is also a valid estimator for immature lungs, all acini of three entire lungs at postnatal day 4 were counted manually. This counting showed that the mean acinar volume of the right middle lobe (0.066 $\mu$l) was not statistically different from the mean acinar volume of right upper (0.065 $\mu$l) and right lower lobes, as well as of the entire lung (0.073 $\mu$l) and of the sum of the left lung and the cardiac lobe (0.081 $\mu$l / Tab. 3). We chose this sum, because during postnatal development the volume of the cardiac lobe growth unproportional to the three other right lung lobe (see above). However, the sum of the volume of the left lung and the cardiac lobe growth proportional to the three other lobes (Tab. 2). Based on the mean acinar volume the mean number of acini (6469 ± 720 acini) was estimated for these three samples following the method presented
previously (3). No statistical difference was observed between the counted (5865 ± 465 acini) and the estimated (6469 ± 720 acini) number of acini at postnatal day 4. The counted number of acini per lobe (N=3 for right upper, right middle, right lower, cardiac, and left lung) between days 4 and 60 (3) showed no statistical differences. In addition, the mean of the observed difference between estimated and counted total number of acini was approximately 10%. Collectively, these findings support the view that the right middle lung lobe is a valid estimator for the total number of acini at any stage of postnatal lung development.

Number of Acini. The number of acini in the right middle lung lobes were counted at postnatal days 4, 10, 21, and 36 as previously described and compared to data of the adult lung (day 60) (3). No significant differences were observed between the five age groups (Tab. 4). Therefore, in opposite to body weight and lung volume, the number of acini is constant throughout postnatal lung development. We detected no direct correlation between these three lung parameters. However, the variation to the mean can be calculated for all individuals over the five time points. To do so, in a multiple regression analysis the standard scores (difference of mean and individual value divided by the standard deviation) of body weight and lung volume of all 27 animals were set as the independent variables, and the number of acini of the right middle lobe was set as the dependent variable. The analysis provided a value (Fig. 2, predicted curve) for every sample. Assuming a linear relationship between all three tested parameters, the predicted values should match with the observed number of acini (Fig. 2). This was not the case ($R^2 = 0.08$), and thus the number of acini, body weight, and lung volume showed no linear correlations.
**Mean acinar volumes.** The mean acinar volume (Tab. 5), defined as the parenchymal lobe volume divided by the counted number of acini, was calculated for the right middle lobe at day 4, 10, 21, 36, and 60. Based on the specific acinar volume (Tab. 5) a bi-phasic growth of the acinar volume was observed (Fig. 3). Hence, the acinar volume increases proportionally to body weight until day 21, while after day 21 body weight grows faster than acinar volume. This kind of bi-phasic growth was already reported for the lung volume, the anlage of new alveolar septa, and for the number of alveoli by Burri (8), Schittny (32), and Tschanz (40), respectively.

**Shrinkage of samples.** Large shrinkage factors have been reported for paraffin embedding (28.70 ± 0.62 %) and for critical point drying (62.0 ± 1.5 %) both measured at day 60 (3). In the present study, we analyzed the shrinkage factor of the right middle lobe for all five remaining time points. The volume of paraffin embedded samples showed a reduction of 24.6 % (± 4.4) and the ones of the critical point dried samples decreased by 62 % (± 1.5).

**Reconstruction of conducting airways.** In order to visualize our results, we reconstructed the conducting airways of one right middle lobe per time point (Fig. 4). The visualization highlighted the high similarity of the conducting airways pathway at any time point and between the individuals. Most of the conducting airways ended with one acinar entrance being located on a transitional bronchiole. However, we also detected acinar entrances very close to each other. In this case, a branching point was located inside the transitional bronchiole (Fig. 4) as already reported in Barré (3).
Discussion

Previous studies by Burri et al. (7, 8) and Boyden & Tompsett (5, 6) proposed that originally purely conducting bronchioles may be transformed into respiratory airways during early alveolarization. To our best knowledge the presented study represents the first quantitative investigation of the development of the number of acini throughout lung development in rats. We were able to show that the number of acini stays constant from day 4 to young adulthood (Tab. 4). From the structural point of view we did not observe any shift of the acinus entrance or the bronchoalveolar duct junction within the bronchial tree. Initially, we expected a proximal shift of the acinus entrances and a dramatic, approximately factor of 2 decrease of the number of acini. Our investigations throughout postnatal lung development did not show any statistical difference of the number of acini. At an early stage of alveolarization (day 4 postnatal) 6326 ± 497 acini were estimated for the entire lung. A similar number (5612 ± 547) was observed in young adult rats at postnatal day 60. We conclude that the acini are formed before the end of the saccular stage (before postnatal day 4) and that the developmental increase of the lung volume is achieved by an increase of the acinar volume and not by an increase of the number of acini. According to Kitaoka et al. (16) the formation of the airways is completed up to the acinar airways at the end of the pseudoglandular stage. During the canalicular stage, epithelial differentiation takes place and the bronchioalveolar duct junction, the border between the conducting airways and the gas-exchange region, is formed (29). Because no shift of the acinus entrance occurs and no additional conducting airways are created, we conclude that the number of acini will not change during alveolarization.
The bronchioalveolar duct junction contains the so-called bronchioalveolar stem cells which are very important for homeostasis and repair. Because the location of the acinar entrance or better the bronchioalveolar duct junction does not move during alveolarization, the site where these stem cells are residing, stays also constant.

Our conclusions, based on rat lungs, are at a first view in contradiction with the observations of Boyden and Tompsett (5, 6). They observed a reduction of the non-respiratory generation between newborn and adult in humans and dogs. The difference may be explained by the different architecture of rat, dog, and human airways. Rats possess one generation of transitional bronchioles instead of few generations of respiratory bronchioles (24) which are found in dogs and humans. Thus, for rats, the acinus entrance is located at the bronchioalveolar duct junction where the epithelium of the bronchioles (club cells and ciliated cells) blends over into the alveolar epithelium. The correlating structural in humans is the ventilatory unit and not the acinus (see introduction). Therefore, Boyden and Tompsett (5, 6) described the alveolarization of the respiratory bronchioles and not a movement of the bronchioalveolar duct junction. If the correlating structure of humans and rat are correctly compared, no contradiction between the presented data and Boyden and Tompsett observations (5, 6) are present.

Our detection method did not only focus on the appearance of alveoli for the detection of the acini entrances, but also on the airway wall thickness, on the appearance of alveolar capillaries, and on the shape of the airways. Because of that, we were able to detect the transition between the conducting and the gas-exchanging parts of the transitional bronchiole even before alveoli were present. As a consequence, the number of acini could be counted prior to the beginning of the
alveolarization. We conclude that, for rat lungs, the final number of acini is reached at
the latest by the end of the saccular stage.

The localization of the acini entrances represents an important additional observation
(Fig. 4, colored spheres). They are inhomogenously distributed over the entire lobe,
in close proximity to the larger airways. As a result a cortical region exists near the
pleura which is free of any acinus entrances. Due to the monopodial branching
pattern (46) the most proximal acinus is located at intra-lobar generation four,
whereas distal acini are located at much higher generations (Fig. 4). Lee et al. (17, 18)
used X-ray tomography of silicon casts of the airways to analyze and count the
generations and segments of the conducting airways. We propose that an estimation
of the number of acini, based on their method may be biased due to the
inhomogeneous distribution of the entrances of the acini in species possessing a
monopodial branching pattern. To eliminate any dependency of the results on the
kind of branching pattern, we counted all acinar entrances in one lobe.

We compared the bronchial tree down to the acinar entrances between days 4 and
60 (Fig. 4). The shown bronchial trees were obtained of five different individuals.
Therefore, we are comparing different developmental stages and individual animals
at the same moment. We observed a high similarity and no change in complexity
between the five analyzed bronchial trees. We conclude that once the bronchial tree
is formed it stays very constant during lung development and that a proportional
growth takes place during postnatal rat lung development. In addition, the individual
alterations appear to be small and at a similar level as variations observed in the
branching of larger blood vessels. The number and the localization of the secondary
and tertiary lobar airways (43) demonstrated this similarity.
Previously, we demonstrated that the right middle lung lobe is a valid estimator for the number of acini for the entire lung (3). However, our method assumes a direct correspondence between the number of acini per lobe and the fraction of total lung volume of this particular lobe. A variation of the fraction of total lung volume was observed for the left lung and the cardiac lobe between days 4 and 10 (Tab. 2). To investigate if these variations influenced the estimation method, the number of acini of three entire lungs were manually counted at day 4 (5865 ± 465 acini) and compared with the results obtained at day 60 (5943 ± 521 acini) (3). No statistical differences were observed between the counted numbers of acini at these two time points. This demonstrated that at day 4 the acinar development of cardiac lobe is at a similar state as the other four lobes. The observed variations of the fraction of total lung volume had no influence on the estimation of the total number of acini based on the right middle lobe. However, due to these variations the number of acini cannot be estimated for the single lobes at postnatal day 4, as proposed in Barré et al. (3). To overcome this problem cardiac lobe and left lung have to be considered as one entity (Tab. 3) for the calculations. When the cardiac lobe and the left lung are combined as one entity, the mean acinar volume of the right middle lobe does not statistically differ with the mean acinar volume of any other lobe (p = 0.198, Tab. 3). Thus, we conclude that the right middle lobe is also a valid estimator for the entire lung development.

In summary, we conclude that the total number of acini is constant throughout lung development but can differ between individuals. No relationships were detected with other parameters like total lung volume, surface area, etc. Combining our method with others (10, 17) it will be possible to further characterize conducting airways and
gas-exchanging regions. This will hopefully lead to a better understanding of clinical relevant topics, for instance air-flow within the lung, pulmonary particle depositions, or lung regeneration.

Acknowledgement

We thank Rajmund Mokso, Goran Lovric, Bernd Pinzer, and Federica Marone for their continuous support at the TOMCAT beamline and Eveline Yao for expert technical assistance. We are thankful for the support of the Swiss National Science Foundation (grants 310030-125397, 310030-153468 and CR23I2-135550).

Disclosures. No conflict of interest, financial or otherwise

Author Contributions. S.F. Barré, obtained and scanned samples, developed the procedure and performed the counting of the acini, analyzed data and drafted the manuscript. D. Haberthür obtained and scanned samples, contributed to the development of the procedure of the counting of the acini and to the writing of the manuscript. T.P. Cremona contributed to writing. M. Stampanoni designed and built the beamline. J.C. Schittny conceived and designed the study, obtained and scanned samples, analyzed data and contributed to writing.
References


Figure 1. Acini, as they appear throughout lung development. At day 4 (a) only sacculi and no alveoli are present. The walls of the conducting airways are smooth and not much, but significantly thicker than the inter airspace wall of the parenchyma. At day 10 (b) alveolarization evidently started; resulting in smaller parenchymal airspace and a larger difference between the thickness of the walls of the conducting and gas-exchanging airways. At the later days (21 – 60, c-e) the difference of the wall thicknesses are becoming even more pronounced. Dotted lines mark acinar entrances. Scale bar 250 µm.
No linear correlation was observed between number of acini, body weight, and lung volume. Thus, the number of acini cannot be estimated by these parameters.

**Figure 2. Relationship between number of acini, lung volume, and body weight.**
Figure 3. Bi-phasic growth of the mean acinar airspace volume. The mean acinar volume (tissue and airspace) grows in two distinct phases. Days 4-21: proportional growth of acinar volume and body weight ($R^2 = 0.954$, dotted line). Days 36-60: no proportional growth ($R^2 = 0.887$, dashed line).
Figure 4. Trees of conducting airways throughout postnatal lung development.

The walls of the conducting airways are shown in grey and the spheres represent the acini entrances. These three-dimensional visualizations show the large similarity of the conducting airways structure at days 4, 10, 21, 36, and 60 and between different individuals.
### Volume density of parenchyma

<table>
<thead>
<tr>
<th>Postnatal day</th>
<th>RUL</th>
<th>RML</th>
<th>RLL</th>
<th>LC</th>
<th>LL</th>
</tr>
</thead>
<tbody>
<tr>
<td>4</td>
<td>0.865</td>
<td>0.818</td>
<td>0.856</td>
<td>0.827</td>
<td>0.865</td>
</tr>
<tr>
<td>10</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.846</td>
</tr>
<tr>
<td>21</td>
<td></td>
<td></td>
<td></td>
<td>0.869</td>
<td></td>
</tr>
<tr>
<td>36</td>
<td></td>
<td></td>
<td>0.870</td>
<td></td>
<td></td>
</tr>
<tr>
<td>60</td>
<td>0.888</td>
<td>0.839</td>
<td>0.843</td>
<td>0.865</td>
<td>0.862</td>
</tr>
</tbody>
</table>

**Table 1. Volume density of the parenchyma.** No significant differences were observed. RUL: right upper lobe; RML: right middle lobe; RLL: right lower lobe; LC: cardiac lobe; LL: left lung.
<table>
<thead>
<tr>
<th>Postnatal Day</th>
<th>Fraction of Total Lung Volume [%]</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RUL</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
</tr>
<tr>
<td>4</td>
<td>11.15</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>11.75</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>21</td>
<td>11.04</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>36</td>
<td>10.74</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>60</td>
<td>10.89</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean of all days</td>
<td>11.09</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2. Fraction of the lung lobes of the total lung volume throughout lung development. Except of the cardiac lobe and the left lung the lobe volume increases in parallel to the growth of the total lung volume. At the cost of the left lung, the cardiac lobe increases disproportionately. RUL: right upper lobe; RML: right middle lobe; RLL: right lower lobe; LC: cardiac lobe; LL: left lung; SD: standard deviation; *: differs significantly (ANOVA) from the other time points for the same lobe.
Table 3. Comparison of the mean acinar volume and the number of acini per lobe at postnatal day 4. To validate the right middle lobe as an estimator for the entire lung, three entire lungs were manually counted. We observed that the right upper, middle, and low lobe, but not the cardiac lobe and the left lung represent a valid sample for the entire lung. RUL: right upper lobe; RML: right middle lobe; RLL: right lower lobe; LC: cardiac lobe, LL: left lung, SD: standard deviation, *: significantly differs (ANOVA) from the other time points for the same lobe.
<table>
<thead>
<tr>
<th>Postnatal Day</th>
<th>Number of Samples</th>
<th>Body Weight [g]</th>
<th>Lung Volume [ml]</th>
<th>Number of Acini (RML) [# Acini]</th>
</tr>
</thead>
<tbody>
<tr>
<td>4</td>
<td>4</td>
<td>10.15</td>
<td>0.54</td>
<td>0.506</td>
</tr>
<tr>
<td>10</td>
<td>5</td>
<td>29.42</td>
<td>3.22</td>
<td>1.32</td>
</tr>
<tr>
<td>21</td>
<td>5</td>
<td>59.65</td>
<td>2.05</td>
<td>2.32</td>
</tr>
<tr>
<td>36</td>
<td>5</td>
<td>84.75</td>
<td>9.65</td>
<td>2.80</td>
</tr>
<tr>
<td>60</td>
<td>8</td>
<td>295.82</td>
<td>16.77</td>
<td>7.51</td>
</tr>
<tr>
<td>Mean of all days</td>
<td>27</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 4. Comparison of the body weight, lung volume and number of acini.

RML: right middle lobe, SD: standard deviation, *: no significant difference.
<table>
<thead>
<tr>
<th>Postnatal Day</th>
<th>Mean Acinar Volume [µl]</th>
<th>Specific Mean Acinar Volume [µl / 100 g]</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
</tr>
<tr>
<td>4</td>
<td>0.069*</td>
<td>0.007</td>
</tr>
<tr>
<td>10</td>
<td>0.196*</td>
<td>0.017</td>
</tr>
<tr>
<td>21</td>
<td>0.355*</td>
<td>0.027</td>
</tr>
<tr>
<td>36</td>
<td>0.410*</td>
<td>0.050</td>
</tr>
<tr>
<td>60</td>
<td>1.157*</td>
<td>0.138</td>
</tr>
</tbody>
</table>

Table 5 Mean acinar volume. Parenchymal lobe volume divided by the counted number of acini, Specific mean acinar volume: mean acinar volume per 100 g body weight, *: differs significantly (ANOVA) from the other time points, +: the mean acinar volume of days 21 and 36 are not significantly different from each other, however they are both significantly different from the three other time points.