A mathematical approach to human pterygium shape

Bojan Pajic1–4, Iraklis Vastardis1, Predrag Rajkovic5, Brigitte Pajic-Eggspuehler1, Daniel M. Aebersold6, Zeljka Cvejic2

1Eye Clinic ORASIS, Swiss Eye Research Foundation, Reinach AG, Switzerland; 2Department of Physics, Faculty of Sciences, University of Novi Sad, Novi Sad, Serbia; 3Faculty of Medicine of the Military Medical Academy, University of Defence, Belgrade, Serbia; 4Division of Ophthalmology, Department of Clinical Neurosciences, Geneva University Hospitals, Geneva, Switzerland; 5Department of Mathematics, Faculty of Mechanical Engineering, University of Niš, Niš, Serbia; 6Department of Radiation Oncology, Inselspital, Bern University Hospital, Bern, Switzerland

Correspondence: Bojan Pajic
Swiss Eye Research Foundation, Eye Clinic ORASIS, Titlisstrasse 44, 5734 Reinach, Switzerland
Tel +41 627 656 080
Email bpajic@datacomm.ch

Purpose: Pterygium is a common lesion affecting the population in countries with high levels of ultraviolet exposure. The final shape of a pterygium is the result of a growth pattern, which remains poorly understood. This manuscript provides a mathematical analysis as a tool to determine the shape of human pterygium.

Materials and methods: Eighteen patients, all affected by nasal unilateral pterygia, were randomly selected from our patient database independently of sex, origin, or race. We included all primary or recurrent pterygia with signs of proliferation, dry eye, and induction of astigmatism. Pseudopterygia were excluded from this study. Pterygia were outlined and analyzed mathematically using a Cartesian coordinate system with two axes (X, Y) and five accurate landmarks of the pterygium.

Results: In 13 patients (72%), the shape of the pterygia was hyperbolic and in five patients (28%), the shape was rather elliptical.

Conclusion: This analysis gives a highly accurate mathematical description of the shape of human pterygia. This might help to better assess the clinical results and outcome of the great variety of therapeutic approaches concerning these lesions.

Keywords: etiology, pterygium, limbal stem cells, stem cells dysfunction, mathematical shape analysis

Introduction

The medical term “pterygium” is derived from the Greek word “pteron”, meaning wing. Pterygia are common and frequently recurring ocular surface lesions, affecting predominately the population in warm climate countries near the equator, the Mediterranean, and the Asia Minor, where exposure to ultraviolet (UV) light is more prominent.1-2 Clinically, a pterygium is a bulking, vascular tissue, which expands toward the center of the cornea. Pterygia may originate from the corneal limbus, where the limbal epithelial stem cells are proliferating, using the Bowman’s membrane as a leading structure, destroying the latter and the epithelium during their advancement.3,4

In a normal ocular surface, the corneal epithelium has a thickness of ~50 μm and consists of up to five to seven layers of regularly arranged, nonkeratinized, squamous epithelial cells.5-8 The limbus represents a zone of transition of ten to twelve layers of epithelial cells containing melanocytes, Langerhans cells, and a network of blood vessels, the latter serving as repositories of corneal epithelial cells, which then would respond in replacing defective corneal epithelium in cases of corneal melting, recurrent erosion, or chemical burns. The limbal stroma is arranged in radial fibrovascular elevations, termed the palisades of Vogt. These are predominantly found in the upper and lower corneoscleral limbus, where the vessels seem to originate.9,10 No goblet cells are present in the limbal stroma. The bulbar conjunctiva consists of six to nine layers of epithelial cells. These cells are not as regularly and compactly arranged as in the cornea. They are also smaller...
and show wide intercellular spaces. The presence of secreting goblet cells is characteristic for the conjunctival epithelium; goblet cells comprise 7% of the basal cell population.11

In normal conditions, the corneal epithelium renews every 7 to 10 days,12 mimicking a state of constant healing; squamous epithelial cells are continuously shed into the tear pool and replaced by fresh migrating cells moving centrally from the limbal area. This theory was first assumed by Thoft and Friend in his X, Y, Z hypothesis,13 where he declared that if the corneal epithelium is to be maintained, cell loss must be balanced by cell replacement (X + Y = Z, where X stands for the proliferation of basal epithelial cells, Y stands for the contribution to the cell mass by centripetal movement of peripheral cells, and Z stands for the epithelial cell loss from the surface). This hypothesis gave birth to a multitude of studies on the location of stem cells, their centripetal migration to the cornea, and the healing pattern of the cornea under normal and pathological conditions. Cumulative evidence has clearly established the limbal basal epithelium as one of the repositories of stem cells for corneal epithelial cells.14,15 The centripetal migration and stem cells existence were established using staining materials16–18 in rabbit and mouse corneas or by identification of stem cell markers.19 In the course of corneal stem cell research, the concept of conjunctival transdifferentiation was born,11,20–22 a process in which the conjunctiva responds to corneal injury by cellular proliferation.20,22,23

Pterygia are pathological proliferative lesions of vascular inflamed tissue which affect the cornea, regarded by some as a deregulated wound healing4,24,25 and by others as a degenerative disease.25,26 A number of studies regarding pathogenesis, histology, and surgical treatment of these lesions have been performed in the past years. However, no study focusing on the shape of pterygia has been published.

We believe that the shape of these lesions may help support the hypothesis that the pterygium is a consequence of limbal stem cell deficiency or malfunction and the centripetal movement of the corneal cells.

Materials and methods

Eighteen patients (12 females) with a median age of 59±9.6 years and unilateral pterygium were recruited in a single center (ORASIS Eye Clinic, Reinach AG, Switzerland). Sixteen pterygia were primary and two recurrent, with sizes varying from 1.2 to 4.0 mm (mean 2.51±0.83 mm) and from 2.2 to 2.4 mm (mean 2.3±0.14 mm), respectively. Thirteen patients (72%) were of Swiss origin and had lived their entire life in Switzerland, and five patients (28%) were of Mediterranean origin and had grown up in their home countries. The research adhered to the tenets of the Declaration of Helsinki. According to the ethics committee of Ethikkommission Nordwest-und Zentralschweiz (EKNZ) as this is a mathematical model with anonymous pterygium data, ethics approval and patient consent is not required.

Pictures of each pterygium were taken at the initial ophthalmological examination using a camera (FF 450/FF 450 IR; Carl Zeiss Meditec AG, Jena, Germany) and analyzed. Slit lamp measurements of each pterygium’s length, size, and distance of the head to the corneal limbus were made in order to control the digital measurements. Corneal topography was performed using a video keratoscope (Orbscan II version 3.0, Bausch & Lomb Zyoptix; Bausch & Lomb Incorporated, Bridgewater, NJ, USA) which served to determine corneal astigmatism and white-to-white distance. All digitized pictures were magnified and the pterygium was outlined using AutoCAD® V 04 (Autodesk Inc., San Rafael, CA, USA). Later on, the same program was used to assess the various topographical values (diameter and digital outline of the pterygium). The goal was to generate a high-precision digital photo of every pterygium in a Cartesian coordinate system, using two axes (X, Y) and five accurate geographical points on the pterygium, in the same scale (mm) as in corneal topography. Point (F) is always passing through the intersection of the two axes F (0, 0) in order to have a landmark and a standard procedure in calculating the measurements and to reassure the integrity of results (Figure 1). The eye is a sphere, but if we consider that corneal topography represents the eye rather as a vertical dissection of the sphere, then the pterygium is simply a protrusion in the eye’s cutting plane.

Results

We examined the pictures of each pterygium taken at the initial ophthalmological examination using a camera. On the contour plane curves which were seen, we measured the coordinates of

Figure 1 Point (F) is always passing through the intersection of the two axes F (0, 0) in order to have a landmark.
Table 1 Coordinates of the five accurate points for all accounted pterygia

<table>
<thead>
<tr>
<th></th>
<th>X-axis (mm)</th>
<th>Y-axis (mm)</th>
<th></th>
<th>X-axis (mm)</th>
<th>Y-axis (mm)</th>
<th></th>
<th>X-axis (mm)</th>
<th>Y-axis (mm)</th>
<th></th>
<th>X-axis (mm)</th>
<th>Y-axis (mm)</th>
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<th>X-axis (mm)</th>
<th>Y-axis (mm)</th>
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<tbody>
<tr>
<td>Pat 1</td>
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<td>0.00</td>
<td>Pat 2</td>
<td>0.00</td>
<td>0.00</td>
<td>Pat 3</td>
<td>0.00</td>
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<td>Pat 4</td>
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<tr>
<td>Pat 5</td>
<td>-0.75</td>
<td>2.40</td>
<td>Pat 6</td>
<td>-0.05</td>
<td>1.58</td>
<td>Pat 7</td>
<td>0.00</td>
<td>0.00</td>
<td>Pat 8</td>
<td>0.00</td>
<td>0.00</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Pat 9</td>
<td>0.31</td>
<td>0.86</td>
<td>Pat 10</td>
<td>-1.05</td>
<td>1.58</td>
<td>Pat 11</td>
<td>0.00</td>
<td>0.00</td>
<td>Pat 12</td>
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<td>0.00</td>
<td></td>
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<td></td>
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<tr>
<td>Pat 13</td>
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<td>0.98</td>
<td>Pat 14</td>
<td>0.22</td>
<td>0.66</td>
<td>Pat 15</td>
<td>-0.4</td>
<td>0.99</td>
<td>Pat 16</td>
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<td>Pat 17</td>
<td>-0.74</td>
<td>-1.21</td>
<td>Pat 18</td>
<td>0.23</td>
<td>-0.66</td>
<td>Pat 19</td>
<td>0.00</td>
<td>0.00</td>
<td>Pat 20</td>
<td>0.00</td>
<td>0.00</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pat 21</td>
<td>0.32</td>
<td>0.66</td>
<td>Pat 22</td>
<td>0.21</td>
<td>0.65</td>
<td>Pat 23</td>
<td>0.15</td>
<td>-0.44</td>
<td>Pat 24</td>
<td>0.41</td>
<td>-0.79</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pat 25</td>
<td>0.15</td>
<td>0.59</td>
<td>Pat 26</td>
<td>0.55</td>
<td>-1.18</td>
<td>Pat 27</td>
<td>0.00</td>
<td>0.00</td>
<td>Pat 28</td>
<td>0.00</td>
<td>0.00</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Five points (Table 1). For some patients, a few points were too close to enable us to estimate curve’s type. Hence, we used the software to determine four more points on the contour curve.

We applied the spline approximations through those nine points which successfully reconstructed the curve. Reconstructed graphs of contour curve by spline approximation are shown in Figure 2. Since the original curves were very similar to conics, we took five points: the first is the point which is the nearest to the origin of supposed conic; a further two points distinguished from one side and two from the other side. The most simple class of curves which pass through five plane points is the class of conic sections which satisfies the equation of the form (Figure 3):

\[
\begin{align*}
\det(M) &= \begin{vmatrix} x^2 & xy & y^2 & x & y & 1 \\
x_1^2 & x_1y_1 & y_1^2 & x_1 & y_1 & 1 \\
x_2^2 & x_2y_2 & y_2^2 & x_2 & y_2 & 1 \\
x_3^2 & x_3y_3 & y_3^2 & x_3 & y_3 & 1 \\
x_4^2 & x_4y_4 & y_4^2 & x_4 & y_4 & 1 \\
x_5^2 & x_5y_5 & y_5^2 & x_5 & y_5 & 1 \\
\end{vmatrix} = 0.
\end{align*}
\]

The previous equation given by the determinant can be written in the form \(Ax^2 + 2Bxy + Cy^2 + 2Dx + 2Ey + F=0\).
Denote

\[ s = A + C, \quad \delta = \begin{vmatrix} A & B \\ B & C \end{vmatrix} = -(B^2 - AC), \quad \Delta = \begin{vmatrix} A & B & D \\ B & C & E \\ D & E & F \end{vmatrix} \]

(2)

According to the analytical geometry, this equation determines:

1. Ellipse if \( \delta > 0 \land s\Delta < 0 \)
2. Hyperbola if \( \delta < 0 \land \Delta \neq 0 \)
3. Parabola if \( \delta = 0 \land \Delta \neq 0 \)

By plotting original curve, spline, and conics, we obtained very close graphs for every patient (Table 2). So, we can conclude that the shape of pterygia is of conic form.

Equations of conics for five patients were calculated (Table 3). These equations show that the conics are slightly rotated around X-axis; therefore, the graphs are not symmetrically located by X-axis (Figure 4).

**Discussion**

A multitude of clinical studies have been published on pterygia over the years. The description of the pterygium shape has always been vague, referring to it as a triangular, bulky, or windy-shaped lesion. We decided to determine the shape of these lesions and found a conic shape in all the pterygia examined.

The pathogenesis of pterygium is complex. Its incidence and prevalence vary according to the geographical region, favoring regions with high exposure to UV radiation (290–400 nm).\(^1,2\) Dushku and Reid\(^27\) first showed that mutations in the TP53 gene expressed in the parental limbal basal cells following UV light exposure may be responsible for the production of abnormal elastotic material and for the invasion of the cornea with various MMPs, resulting in the excessive production of transforming growth factor-beta (TGF-\(\beta\)) via the p53-Rb-TGF-\(\beta\) pathway.\(^4\) Fibroblast growth factors, vascular endothelial growth factors responsible for angiogenesis, and stem cell factors are also upregulated.\(^5,7,28\) Girolamo et al\(^29\) reported the abundant expression of MMP-1 in pterygium tissue and only recently reinforced his study when he postulated that MMP-1 is a likely candidate enzyme for pterygium formation, because UV light regulates this enzyme at the level of transcription and translation, establishing a direct link between UV exposure and the induction of matrix-denaturing enzymes in pterygia.\(^30\)

In normal conditions, the corneal epithelium is constantly renewed every 7 to 10 days.\(^12\) This condition was
supported by Thoft and Friend in his X, Y, Z hypothesis, when he declared that if the corneal epithelium is to be maintained, cell loss must be balanced by cell replacement. These cells were initially assumed to migrate with centripetal movements from the periphery to the center of the cornea in cases of corneal damage tissue. Later, it was proved by Pellegrini et al that the stem cell population appears to be localized to the palisades of Vogt in the limbal region.

Thoft and Friend initially stated that “while the movement of the cells from the periphery of the cornea seems well established, the source of these cells were uncertain”. Later, they restated that the original, X, Y, Z hypothesis did not ascribe an origin of these cells, assuming that conjunctiva could simply drift across the limbus to provide either acute or chronic replacement of peripheral cells through a process called conjunctival transdifferentiation. Various studies in mice and rabbits have shown that the conjunctival cells can also migrate to the cornea after denudation of the corneal epithelium, resulting in vascularization of the cornea when the limbus is removed. Under normal circumstances, the limbal epithelium acts as a barrier and is able to exert an inhibitory growth pressure preventing migration of conjunctival epithelial cells onto the cornea.

Shapiro et al divided this process into five stages, depending on the presence and density of goblet cells and the degree of stratification of the conjunctival epithelium. The overall process has been extensively studied and the current hypothesis concludes that goblet cells do not migrate onto the cornea but rather develop de novo from non-goblet epithelial cells. Vascularization of the regenerated epithelium is associated with poor transdifferentiation and persistence of goblet cells. In a recent study, it was concluded that the cytology of surface cells in pterygium exhibits squamous metaplasia with increased goblet cell density and that a graded series of ocular surface changes exists throughout the bulbar conjunctiva (even the unaffected one) in eyes with pterygium, with the most advanced changes occurring directly over the pterygium surface.

Table 2 Equations of all 18 pterygia

<table>
<thead>
<tr>
<th>Patient no</th>
<th>Function</th>
<th>Equation</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Hyperbole</td>
<td>0.04076x² − 0.000828y² + 0.00236xy + 0.02772x + 0.000009y = 0</td>
</tr>
<tr>
<td>2</td>
<td>Ellipse</td>
<td>0.00599x² + 0.00757y² − 0.00056xy − 0.01841x + 0.00022y = 0</td>
</tr>
<tr>
<td>3</td>
<td>Hyperbole</td>
<td>0.03320x² − 0.02767y² − 0.00076xy + 0.04364x + 0.00109y = 0</td>
</tr>
<tr>
<td>4</td>
<td>Ellipse</td>
<td>0.00351x² + 0.00498y² + 0.00117xy − 0.01126x − 0.00006y = 0</td>
</tr>
<tr>
<td>5</td>
<td>Hyperbole</td>
<td>2.76910x² − 0.91154y² + 0.37047xy + 4.18353x − 0.04651y = 0</td>
</tr>
<tr>
<td>6</td>
<td>Hyperbole</td>
<td>0.00923x² − 0.00894y² − 0.00443xy + 0.01370x − 0.00035y = 0</td>
</tr>
<tr>
<td>7</td>
<td>Hyperbole</td>
<td>0.00029x² − 0.000011y² + 0.00009y − 0.00009y − 0.00002y = 0</td>
</tr>
<tr>
<td>8</td>
<td>Hyperbole</td>
<td>0.02800x² − 0.06723y² − 0.01546xy + 0.34070x + 0.00266y = 0</td>
</tr>
<tr>
<td>9</td>
<td>Ellipse</td>
<td>0.00054x² + 0.00035y² − 0.00007xy − 0.00077x − 0.00002y = 0</td>
</tr>
<tr>
<td>10</td>
<td>Hyperbole</td>
<td>0.09636x² − 0.22137y² + 0.01323xy + 0.40036x + 0.00350y = 0</td>
</tr>
<tr>
<td>11</td>
<td>Ellipse</td>
<td>0.00104x² + 0.00061y² − 0.00015xy − 0.00232x + 0.00004y = 0</td>
</tr>
<tr>
<td>12</td>
<td>Hyperbole</td>
<td>0.00187x² − 0.00574y² − 0.00025xy + 0.00536x + 0.00003y = 0</td>
</tr>
<tr>
<td>13</td>
<td>Hyperbole</td>
<td>0.02159x² − 0.04652y² − 0.00245xy + 0.07845x + 0.00327y = 0</td>
</tr>
<tr>
<td>14</td>
<td>Ellipse</td>
<td>0.00044x² + 0.00040y² − 1.93446x×10⁻⁶xy − 0.00087x − 4.66188x×10⁻⁴y = 0</td>
</tr>
<tr>
<td>15</td>
<td>Hyperbole</td>
<td>0.00057x² − 0.00148y² + 0.00007xy + 0.00331x + 0.00001y = 0</td>
</tr>
<tr>
<td>16</td>
<td>Hyperbole</td>
<td>0.00095x² − 0.00247y² − 0.0013xy + 0.00788x − 0.00003x = 0</td>
</tr>
<tr>
<td>17</td>
<td>Hyperbole</td>
<td>0.01106x² − 0.01755y² − 0.00246x + 0.02122x + 0.00043y = 0</td>
</tr>
<tr>
<td>18</td>
<td>Hyperbole</td>
<td>0.03332x² − 0.03374y² − 0.00432xy + 0.06269x + 0.00032y = 0</td>
</tr>
</tbody>
</table>

Table 3 Equations of conics for five patients

<table>
<thead>
<tr>
<th>Patient</th>
<th>Equations of hyperbolae</th>
</tr>
</thead>
<tbody>
<tr>
<td>16</td>
<td>-4.8924x² + 0.5976xy + 3.0387y² − 8.8581x + 0.0280y = 0</td>
</tr>
<tr>
<td>17</td>
<td>-5.9591x² − 0.0285xy + 5.2167y² − 10.1380x − 0.5713y = 0</td>
</tr>
<tr>
<td>18</td>
<td>-8.8767x² + 1.6141xy + 6.9860y² − 11.5030x − 0.4806y = 0</td>
</tr>
<tr>
<td>12</td>
<td>-1.8938x² + 0.7338xy + 2.8284y² − 3.5674x − 0.2207y = 0</td>
</tr>
<tr>
<td>15</td>
<td>-4.0144x² + 0.6306xy + 3.5808y² − 6.9281x − 0.1073y = 0</td>
</tr>
</tbody>
</table>

Figure 4 Overlapping of spline approximation and conics in the case of patient 18.
analysis of pterygium showed that conjunctival epithelial cells were also recognizable on the head of the lesion.37

Despite the multitude of recent studies, the exact shape of pterygia was not investigated. One of the rare attempts to characterize the growth pattern and shape observed in pterygia was given by Chui et al.38 They assumed that the direction of epithelial cell movements follows the radial curve of nerves. Furthermore, they detected a neuropeptide K (NK,) receptor and a sensory neuropeptide substance P in pterygia. The latter is a potent chemoattractant for pterygium fibroblasts and vascular endothelial cells, implying that substance P, given the radial pattern of the corneal innervation, may contribute to determine the shape of pterygia.

According to our mathematical calculations, all pterygia examined were of conic shape. Regarding other theories that reinforced the centripetal movements of stem cells and according to the data of the various publications regarding the pterygium pathogenesis mentioned earlier, we could conclude that the conjunctival epithelium is allowed to intrude the cornea, forming the pterygium with the known cytology.36,37 This centripetal movement of cells has been confirmed in animal models.39,40

If the centripetal movement is indeed the result of reactive forces and a result of centripetal force acting on these cells, with the principle that any matter would obey the laws of physics, then when this matter is inserted in that field, which is itself a convention designed around particular forces, then that body must obey the laws associated with that field. This is supported by studies where limbal transplant surgery results in healing with corneal epithelium.31,42 Conversely, in animal models, surgical removal of the limbus leads to healing with noncorneal epithelium.21

Nevertheless, we feel that this mathematical analysis of pterygium shape may represent a significant piece in the puzzle representing the pathogenesis of pterygia, proving that pterygia repeatedly adapt to same shape and form, in this case a conic shape.

Acknowledgment
Parts of our original work were presented in the poster session during the XXXIII Congress of the ESCRIS, 5–9 September, 2015, in Barcelona, Spain.

Disclosure
The authors report no conflicts of interest in this work.

References


