autosomal dominant and recessive patterns, and the latter is typically associated with phenotypic variation. In the recessive form, mutational analysis has been performed in the respective genes. It has been suggested that a combination of genetic and environmental factors may contribute to the development of GA.


timepoint for atrophy and therefore can be used as a prognostic determinant. The fundus autofluorescence signal is mainly generated by RPE lipofuscin, a mixture of fluorophores originating from by-products of the visual cycle. As a result of RPE loss and the subsequent decrease of lipofuscin that occurs in advanced GA, there is a loss of signal in the FAE. However, in the junctional zone surrounding atrophy, FAE has been shown to be increased. The increased autofluorescence signal could be a result of increased phagocytosis with lipofuscin accumulation or formation of RPE multilayers. Alternatively, increased bisretinoid formation within impaired photoreceptors has been proposed to be causative for increased autofluorescence in the junctional zone. High-resolution optical coherence tomography (OCT) has advanced the study of GA with its unprecedented resolution, and thus it has provided further insights in the disease process and has been reported to be useful to identify areas of foveal sparing. OCT characteristics of GA include choroidal signal enhancement and alterations at the level of RPE, the outer nuclear layer (ONL) and outer plexiform layer (OPL), and the external limiting membrane. Time-resolved fundus autofluorescence imaging (FLIO) is a relatively novel imaging method that allows topographic mapping of endogenous fluorescence lifetimes within the retina. The fluorescence lifetime represents the time a molecule spends in its excited state after excitation with a laser.
impulse before returning to its ground state by emission of a photon with a longer wavelength. Autofluorescence lifetimes are specific for each fluorophore and sensitive to the metabolic environment.18

The aim of the present study was to characterize autofluorescence lifetimes in GA as a result of AMD and to identify GA phenotypes using FLIO and high-resolution spectral-domain OCT imaging.

**METHODS**

This study is registered at ClinicalTrials.gov (NCT01981148). It was carried out with the approval of the local ethics committee and is in accordance with the International Ethical Guidelines for Biomedical Research involving Human Subjects (Council for International Organizations of Medical Sciences) and the Declaration of Helsinki.

**Patients**

Patients with GA and AMD were recruited at the Outpatient Department of Ophthalmology at the Bern University Hospital, Bern, Switzerland, and signed an informed consent prior to study entry. None of the patients reported taking carotenoid supplementation.

Patient selection was relayed on the clinical diagnosis that was confirmed by OCT and FAF imaging. Eyes with a history or signs of neovascular AMD and other ophthalmic diseases affecting the macula as well as significant lens opacities were excluded.

**Geographic Atrophy**

Areas of GA were defined funduscopically with clearly visible marked boundaries of decreased retinal pigmentation and exposed underlying choroidal structures. In FAF, one or more clearly demarked areas of hypoautofluorescence were present, corresponding to areas in OCT with choroidal signal enhancement because of the absence of the retinal pigment epithelium and loss of the external limiting membrane within the fovea. Foveal sparing was defined as a central foveal island with an intact retinal layer structure that was preserved and surrounded by well-demarcated areas of GA for at least 270°.

**Patient Examinations**

For all of the patients included in this study, best corrected visual acuity (BCVA) was assessed (Early Treatment Diabetic Retinopathy Study [ETDRS] letters).19 Subsequently, pupils were maximally dilated with tropicamide 0.5% and phenylephrine hydrochloride 2.5%, and the fundus was examined biomicroscopically. Color fundus images (Zeiss FF 450plus, Zeiss, Oberkochen, Germany), FAF images, and OCT scans of the macula (Heidelberg Spectralis HRA+OCT; Heidelberg Engineering, Heidelberg, Germany) were obtained. OCT was performed using a high-speed mode and a scan pattern size of 20' x 20' centered on the fovea, resulting in 49 B scans per eye.

Fluorescence lifetime images were acquired using a fluorescence lifetime imaging ophthalmoscope, which is based on an HRA Spectralis system (Heidelberg Engineering).

**Fluorescence Lifetime Imaging Ophthalmoscope**

The fluorescence lifetime imaging ophthalmoscope uses a 473-nm pulsed blue laser light at an 80-MHz repetition rate for excitation of retinal autofluorescence. Emitted fluorescence light was registered by time-correlated single-photon counting modules using highly sensitive hybrid photon-counting detectors (Becker&Hickl, Berlin, Germany). Two detection channels with distinct wavelengths were used: a short spectral channel (SSC, 498–560 nm) and a long spectral channel (LSC, 560–720 nm). Simultaneously, a confocal high-contrast infrared image was recorded by the system to record each photon at the correct spatial location. A minimal photon count of 1000 photons per pixel was acquired for every location within the image, resulting in a total scan duration of approximately 90 seconds. The Becker&Hickl software (SPCImage 4.6) was used for the analysis of recorded lifetime data, and a biexponential decay model was applied using a binning factor of one. A triexponential fitting procedure, which requires a higher binning factor, did not substantially contribute to a lower χ² value, which indicates the goodness of the fit. Therefore, we used a binning of 3 x 3 pixels and two decay components to gain spatial resolution.

This procedure resulted in a short (T1) and a long (T2) lifetime component with corresponding amplitudes x1 and x2. For each pixel, an intensity-weighted mean fluorescence lifetime value $T_m$ was calculated as follows:

$$T_m = \frac{x_1 T_1 + x_2 T_2}{x_1 + x_2} \quad (1)$$

The basic characteristics of the FLIO device and the analysis method have been further described in previous reports.15

**Analysis of Fluorescence Lifetime Data**

Areas of GA within FAF images were classified according to Binnewald et al.15 The size of GA was measured manually within FAF images. Fluorescence lifetime data was further analyzed by using the FLIO reader software (ARTORG Center for Biomedical Engineering Research, University of Bern, Bern, Switzerland). A circle of interest with a diameter of 0.2 mm was applied for the quantitative analysis of detailed structures. This circle was placed within predefined areas of a standard early treatment of a diabetic retinopathy (ETDRS) grid with a central area (d = 1 mm), an inner ring (d = 3 mm), and an outer ring (d = 6 mm). Image detail sections (d = 0.2 mm) were analyzed within the foveal center and the outer ETDRS ring from the unaffected retina and areas within GA. The image detail section measurements represent a mean value of three individual measurement locations. Fluorescence lifetime data of both spectral channels were analyzed separately. If both eyes showed GA, the eye with the better image quality was chosen.

**Analysis of OCT Data**

Thickness measurements of retinal layers were performed using a spectral-domain OCT. The inbuilt automated retinal layer segmentation of the Heidelberg software was applied. Total retinal thickness within the foveal center was assessed after manual correction of the automatically segmented inner limiting membrane and the basal membrane. The values were extracted from the central subfield of the ETDRS grid, and a small marker was placed within the foveal center. Choroidal thickness was measured beneath the foveal center.

**Statistical Analysis**

For statistical analysis, Prism Graph Pad (Prism 6; GraphPad Software, Inc., La Jolla, CA, USA) was used. Within the outer ring of the ETDRS grid, retinal areas within GA were compared with surrounding retinal tissue and with lifetime values from the foveal center. Group comparison was done by one-way ANOVA and Tukey’s multiple-comparison posttest analysis.
et al., 6 10% of the eyes (4 patients) showed no increased
(63%, 17 patients). According to GA classification of Bindewald
lesions (37%, 10 patients) or multiple confluent atrophic areas
were either grouped in main atrophy with satellite
(27 patients) presented with multiple areas of GA. Multifocal
(14 patients) showed one isolated unifocal lesion, whereas 66%
(22) presented with multiple areas of GA. Multifocal
lesions were either grouped in main atrophy with satellite
(37%, 10 patients) or multiple confluent atrophic areas
(63%, 17 patients). According to GA classification of Bindewald
et al., 6 10% of the eyes (4 patients) showed no increased
 autofluorescence surrounding GA, 10% (4 patients) presented
 with focal, 49% (20 patients) with diffuse (reticular, branching,
 fine granular, trickling, granular with peripheral punctuate
 spots), 29% (12 patients) with banded, and 2% (1 patient) with
 patchy hyperfluorescence.

RESULTS
A total of 41 eyes with AMD and GA of 41 patients were
analyzed (22 right eyes and 19 left eyes). The mean age ±
standard deviation of the patients was 80 ± 7 years (range, 66–
94) and 54% (22) of the patients were female. Of the
investigated eyes, 66% (27 patients) were pseudophaclic.
Foveal sparing was seen in 24% (10 patients). The mean
area of GA (± standard error of the mean [SEM]) was 14.0 ±
1.8 mm² (range, 0.2–50 mm²). Of the investigated eyes, 34%
(14 patients) showed one isolated unifocal lesion, whereas 66%
(27 patients) presented with multiple areas of GA. Multifocal
lesions were either grouped in main atrophy with satellite
lesions (37%, 10 patients) or multiple confluent atrophic areas
(63%, 17 patients). According to GA classification of Bindewald
et al., 6 10% of the eyes (4 patients) showed no increased
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 patchy hyperfluorescence.

Fluorescence Lifetimes in GA
Representative images of a patient with GA are shown in Figure
1. Areas of decreased autofluorescence intensity in EAF correlated
well with the areas of abnormal mean fluorescence
lifetime values. Different patterns of GA were associated
with specific autofluorescence lifetime patterns (Fig. 2). The mean
autofluorescence lifetime measured in areas of GA was 1055 ± 52
ps (SEM) in the SSC and 924 ± 36 ps in the LSC (Fig. 3).
Therefore, Tm was significantly prolonged by 624 ± 276 ps
(+152%) in the SSC and 418 ± 186 ps (+83%) in the LSC when
compared with the surrounding retinal tissue with mean
lifetimes of 411 ± 11 ps and 506 ± 10 ps, respectively (P <
0.0001). The coefficient of variation in the SSC was 17% for the
retina and 31% within GA, and in the LSC it was 13% in the
retina and 24% within GA. Within GA, the mean autofluore-
cence lifetime of the foveal center was shorter when compared
with the surrounding atrophic area (Figs. 1–4), irrespective of
foveal sparing (Fig. 5). In seven eyes with progressed atrophy
of the outer retinal layers in OCT, the mean fluorescence
lifetime distribution did not show a peak of short lifetimes
within the macular center but, rather, a speckled pattern (Figs.
6A, 6B).

Fluorescence Lifetimes in Pseudophakic Versus
Phakic Patients
In the SSC, the mean fluorescence lifetime values were
significantly dependent on the lens status (P = 0.05 in the
foveal center, <0.001 in the normal retina, and 0.003 in GA
areas; multiple linear regression analysis). In the LSC, the
values were completely independent of the lens status (P =
0.41 in the foveal center, 0.4 in the normal retina, and 0.84 in
GA areas). However, because fluorescence lifetimes were
analyzed and compared within individual eyes, the lens status
as a potential confounder was eliminated.

Fluorescence Lifetimes in GA With Foveal Sparing
Patients with GA and foveal sparing (n = 10) showed uniformly
prolonged mean autofluorescence lifetimes inside the areas of
atrophy. Within the foveal center, typical short mean fluores-
cence lifetimes were observed (SSC: 295 ± 27 ps; LSC: 401 ±
28 ps; Fig. 5). Mean fluorescence lifetimes within the foveal
center of patients with central GA did not significantly differ
from values from patients with foveal sparing (P = 0.67 and
0.31; Fig. 5B).

FLIO and Visual Acuity
The mean visual acuity in the study eyes was 42 ± 3 ETDRS
letters (range, 0–84 letters). There was a significant correlation
between BCVA and the mean fluorescence lifetime of the
central ETDRS subfield in both spectral channels when
analyzing the data of all study eyes (r² = 0.19, P = 0.004; Fig.
7). In a subgroup analysis of eyes with foveal sparing (n = 10),
r² was 0.16 and 0.13, respectively (P = 0.03 and 0.04).

FLIO and OCT Analysis
Retinal thickness within atrophic areas was clearly reduced in
eyes with GA as a result of AMD (143 ± 12 μm; Figs. 1, 4–6).
Outer retinal layers, including the RPE, the photoreceptors,
and the ONL and OPL were significantly thinned or completely
missing. The RPE showed drusenoid deposits and irregularities
that appeared as highly hyper-reflective in the OCT scans.
Mean subfoveal choroidal thickness was 139 ± 11 μm.
The mean retinal thickness of the central subfield showed a weak correlation with the BCVA, whereby increased retinal thickness was associated with better ETDRS letter scores ($r^2 = 0.16$, $P = 0.01$). Mean fluorescence lifetimes were independent of the central retinal thickness. Longer lifetimes had a weak but significant association with thicker subfoveal choroidal thickness (SSC: $r^2 = 0.13$, $P = 0.02$; LSC: $r^2 = 0.1$, $P = 0.045$).

Next, we analyzed the seven cases without short autofluorescence lifetimes identifiable within the fovea. In these cases, the OPL was not identifiable in OCT (Figs. 6A, B).

**FLIO in GA Borders**

In FLIO, the borders of GA were identified to feature mean fluorescence lifetime values shorter than within the main atrophy but longer than the surrounding retina (SSC: 768 ± 43 ps, LSC: 756 ± 20 ps; all $P < 0.0001$). In the autofluorescence intensity images, these intermediate lifetime borders were located within the hypofluorescent area. In areas of hyperfluorescence surrounding the hypofluorescent atrophic areas, only marginally prolonged mean fluorescence lifetimes were measured when compared with the surrounding retina (SSC: 78 ± 12 ps, LSC: 61 ± 8 ps; both $P = $ not significant). Adjacent zones of GA were clearly identifiable in all GA FAF pattern using two-dimensional (2D) analysis (Fig. 8) and phasor analysis (Supplementary Fig. S1).²⁰

**Analysis of Individual Fluorescence Lifetime Components**

Because $Tm$ is a function of individual lifetimes ($T1$ and $T2$) and corresponding amplitudes ($a1$ and $a2$), we investigated the influence of these components toward the changes seen in $Tm$. Areas outside GA showed a characteristic distribution of the short lifetime component $T1$ versus the long component $T2$, which is depicted in a uniform distribution cloud within the 2D histogram in both spectral channels (Fig. 8). Areas with GA typically showed prolonged lifetime components, especial-
ly T2, resulting in an overall increase of the mean fluorescence lifetime \(Tm\). A separate lifetime entity was identified for image pixels located in the adjacent zone of atrophy.

Within GA, the shortest \(Tm\) values from the macular center resulted from a short \(T1\) component with a high amplitude \(a1\), whereby \(T2\) was equally distributed within the whole area of GA.

**DISCUSSION**

Time-resolved autofluorescence lifetime characteristics within areas of GA were investigated in a cohort of 41 patients with dry AMD. Areas of GA featured significantly longer retinal autofluorescence lifetimes when compared with the surrounding retinal tissue. There is a certain degree of variability of absolute FAF lifetimes between different patients because of various factors such as lens status. Because of this, data analysis within individual patients and comparisons of diseased tissue with surrounding unaffected retina was performed.

The adjacent zone bordering on areas of atrophy was clearly identifiable using 2D analysis of fluorescence lifetime values. These diagrams allow for differentiation and visualization of potential tissue at risk for GA known from FAF intensity studies as hyperautofluorescent areas.\(^{21,22}\) The main fluorescence within the retina originates from lipofuscin, a complex mixture of bisretinoid fluorophores from the visual cycle metabolism.\(^{7,25}\) According to conventional opinion, hyperfluorescent borders around GA are supposed to contain increased concentrations of lipofuscin.\(^{24}\) However, in FLIO measurements, corresponding mean fluorescence lifetime values still differ only marginally from values of the surrounding normal retina even though significantly prolonged lifetimes originating from lipofuscin (1262 ps) derivates might be expected.\(^{25,26}\) Rather, new insights indicate that increased autofluorescence might be caused by vertically superimposed cells and cellular fragments and not by increased lipofuscin concentrations.\(^{12,27}\) This hypothesis can be supported by our fluorescence lifetime measurements of these areas.

Typically areas of GA show loss of the RPE and the overlaying photoreceptors in OCT. In FLIO, this absence may allow increased contribution of autofluorescence decay times from retinal layers or metabolites with relatively longer lifetimes. They may either originate from the underlying choroid or from within the inner retinal layers. On the other hand, the presence of long lifetimes in patients with GA supports our hypothesis that short decay times mainly come from the RPE. However, in areas of generalized RPE atrophy in FAF and OCT, we found preservation of relatively short autofluorescence lifetimes. A recent study has provided evidence for a strong impact of macular pigment (MP) on macular FAF lifetimes by correlating MP optical density measurements with lifetimes in the fovea.\(^{28}\) MP consists of the two hydroxycarotenoids, lutein and zeaxanthin, with peak concentrations in the foveal center of the macula.\(^{29}\) MP is very effective at absorbing short-wavelength blue light with an absorbance peak at 460 nm.\(^{30,31}\) This can be seen in FAF, where MP is known to decrease an autofluorescence intensity signal, resulting in the typical hypofluorescent macula.\(^{29}\) MP is very effective at absorbing short-wavelength blue light with an absorbance peak at 460 nm.\(^{30,31}\) This can be seen in FAF, where MP is known to decrease an autofluorescence intensity signal, resulting in the typical hypofluorescent macula.\(^{29}\) Assuming that short lifetimes in the fovea are generated by...
MP, it is a striking finding that MP is preserved even in the presence of atrophy. Given that MP is mainly found in the plexiform layers and the photoreceptor axon layers, the persistence of MP may reflect the health of the plexiform layers. Interestingly, in some cases the characteristic short lifetimes were absent in the fovea. Correlation with OCT in these cases showed loss of the ONL and OPL within the area of GA. This strongly supports the evidence that short fluorescence lifetimes within the macular center originate from MP. However, given the persistence of short lifetimes even in the presence of central atrophy, short central retinal fluorescence lifetimes alone cannot be used for differentiation of foveal sparing.

In keeping with these findings, long fluorescence lifetimes in patients with central GA were associated with worse visual acuity and thinner subfoveal choroidal thickness. Both findings may point toward an advanced disease stage of GA. These results are in accordance with other studies in which the thinning of the choroid in patients with GA was found and even an association with the progression rate was identified. In this late stage, the layer structure of the other retinal layers is mostly destroyed including the Henle fiber layer and the OPL containing the macular pigment.

An advantage of time-resolved autofluorescence is the ability to investigate various lifetime components because \( Tm \) is a function of individual lifetimes and amplitudes. The individual clustering of pixel values in specific regions can be analyzed using 2D analysis. It is noteworthy that the lifetime cloud of the adjacent zone falls between the surrounding retina and the atrophic zone, which implies that the adjacent zone is influenced by fluorophores that predominate the atrophic area. In this context, a high spatial resolution of FLIO images is important, and therefore the curve-fitting parameters we applied seem to be appropriate.

Our findings of fluorescence lifetime measurement in GA may have an important impact on disease monitoring because they may be an indicator for degenerative processes in the outer retina.

This study shows a broad range of clinical presentations of GA. However, the number of included patients is limited and larger cohorts will have to be analyzed to investigate and evaluate pattern-specific characteristics in the different GA subgroups. Furthermore, additional ex vivo lifetime data will be necessary to dissect the influence of individual components within the border zone of atrophy and the surrounding retina. In addition, longitudinal follow-up examinations and further

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**Figure 4.** Fluorescence lifetimes within GA. (A) FLIO SSC and OCT scans of indicated lines 1 to 6. (B) Correlation of mean fluorescence lifetimes of the short (above) and long (below) spectral channel with central retinal thickness (dotted line, not significant) and central choroidal thickness (gray line, SSC: \( r^2 = 0.15, P = 0.02 \); LSC: \( r^2 = 0.1, P = 0.045 \)). AF, fundus autofluorescence intensity; CF, color fundus.
correlation with functional parameters are needed to identify markers for disease progression.

CONCLUSIONS

Fluorescence lifetime analysis of areas with GA provides specific lifetime patterns. Short fluorescence lifetimes within the macular center may provide information about the integrity of the OPL and ONL. The analysis of GA borders by FLIO might emerge as a useful tool for the visualization of local tissue remodeling and disease monitoring.

**FIGURE 5.** GA with foveal sparing. (A) OCT scan of the indicated line. (B) Comparison of fluorescence lifetimes within the foveal center in eyes with foveal sparing (F’spare) and central atrophy (GA) \(n = 10\) F’spare; \(n = 31\) GA). AF, fundus autofluorescence intensity LSC; CF, color fundus; FLIO SSC/LSC, mean fluorescence lifetime images of the short and the long spectral channels; ns, not significant.

**FIGURE 6.** FLIO (SSC) and OCT in GA without short central autofluorescence lifetimes. (A, B) The ONL and the OPL are completely missing within the area of GA. (C) Example with preserved ONL and OPL and short mean autofluorescence lifetimes within the foveal center.

**FIGURE 7.** Correlation of mean fluorescence lifetime values with best corrected visual acuity (BCVA, ETDRS letters). Mean values from the central subfield with error bars (95%, dotted lines) are shown for the short and the long spectral channels (both \(r^2 = 0.19, P = 0.004\)). SSC = black; LSC = gray.
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Figure 8. Distribution of fluorescence lifetime components in GA. Distribution histogram of the short decay component T1 versus the long decay component T2 (see also Equation 1). Specific lifetime distribution clusters were identified for the unaffected retina, GA, and the adjacent zone of GA.