

1 **Intestinal Colonization with Extended-Spectrum Cephalosporin- and**
2 **Colistin-Resistant *Enterobacteriaceae* in HIV-Positive Individuals in Switzerland:**
3 **Molecular Features and Risk Factors**

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16 **Short running title:** HIV-positive colonized with MDR *E. coli*

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26 Sir,
27 The increasing rates of gut colonization by extended-spectrum cephalosporin-resistant (ESC-R-)
28 and/or colistin-resistant *Enterobacteriaceae* (COL-R-Ent) carrying the plasmid-mediated *mcr-1*
29 gene in healthy humans raises serious concerns [1, 2]. Nevertheless, colonization prevalence and
30 risk factors associated with HIV+ individuals are unknown. This population undergoes a decrease
31 in CD4+ T cells in the gut-associated lymphoid tissue which is linked with a microbiota dysbiosis
32 possibly favoring colonization [3]. The aim of this pilot study was to better understand this
33 phenomenon to improve the management of these patients.

34 Between March 2015 and April 2016, 101 HIV+ individuals on suppressive anti-retroviral
35 therapy (ART) donated a stool sample and filled out a questionnaire (Table S1 and S2). To detect
36 ESC-R-, carbapenem-, and/or COL-R-Ent, stools were enriched in broth and plated on selective
37 agar plates [1, 2]. Species identification was achieved by using the MALDI-TOF MS (Bruker
38 Daltonics, Leipzig, Germany). MICs were obtained using the Sentitre™ GNX2F plate (Trek
39 Diagnostic Systems, Independence, OH, USA). β -lactamase genes were identified using both
40 CT103XL microarray (Check-Points, Wageningen, Netherlands) and PCR/sequencing [1, 2]. COL-
41 R-Ent were screened for *mcr-1/2* and PmrAB two-component system with PCR/sequencing [1].
42 Whole genome sequencing was carried out with MinION (Oxford Nanopore, Oxford, UK) [1].
43 Clonality was assessed with standard MLST (<http://mlst.ucc.ie/mlst/dbs/Ecoli>) and phylogenetic
44 grouping [2]. The PBRT kit (Diatheva, Cartoceto, Italy) was used to type plasmids. Conjugation
45 was performed using *E. coli* JF33 [1]. Univariate analysis was performed to compare colonized and
46 non-colonized subjects (GraphPad Prism software, version 7.0; La Jolla, CA, USA). Continuous
47 variables were analyzed using Mann–Whitney U test, whereas categorical variables with Fisher's
48 exact test.

49 Seven volunteers (6.9%) resulted colonized with ESC-R *E. coli* (other *Enterobacteriaceae* were
50 not detected). This data is consistent with that reported in the healthy population in Switzerland and
51 in other European countries [1, 2]. However, when looking into *Enterobacteriaceae* causing

52 infections in HIV+ subjects, the prevalence of ESC-R-Ent dramatically increased to 50%, though
53 these studies were either conducted in countries with high occurrence of these multidrug-resistant
54 organisms (MDROs) or have enrolled individuals not under ART [4 , 5].

55 Most ESC-R *E. coli* recovered in the present study were CTX-M-15 producers and associated with
56 F plasmids (Table 1) [2]. Each isolate yielded an individual sequence type (ST), including the high
57 risk clones (HiRC) ST131, ST73, ST405 and ST410 that might be associated with the frequent
58 contact of HIV+ people with the healthcare facilities [2].

59 Four volunteers (4.0%) were also colonized with COL-R *E. coli* (other *Enterobacteriaceae*
60 were not detected), of which one (31349) was of B1-ST5 and *mcr-1*-positive. The remaining three
61 strains failed to demonstrate a plasmid-mediated mechanism of colistin resistance after conjugation
62 experiments [1]. Analysis of the PmrAB system indicated that these strains had amino acid
63 substitutions possibly driving colistin resistance (Table 1) [1]. Demographic, clinical and
64 epidemiological data about the four volunteers colonized with COL-R *E. coli* are shown in Table
65 S1. Notably, volunteer 31349 was a 58 year-old male who in the last year was hospitalized in
66 Switzerland and received antibiotics, but he did not traveled anywhere.

67 Screening of raw genome data by PlasmidFinder-1.3 and ResFinder-2.1 indicated that *E. coli* strain
68 31349 possessed IncF, IncL, and IncX4 plasmids together with the *mcr-1* (no other resistance genes
69 were detected) [1]. *Mcr-1* was located in a 33.3 Kb plasmid that shared >99% identity (≤ 30
70 mismatches) with IncX4 plasmids reported in *E. coli* from pig faeces in China (GenBank:
71 KX254343) and from river water in Switzerland (GenBank: KZ129783). This is the first report of
72 *mcr-1*-carrying *Enterobacteriaceae* in the gut of HIV+ people and highlights the global
73 dissemination of this life-threatening resistance mechanism [1]. Despite the fact this COL-R strain
74 carrying *mcr-1* was pan-susceptible to all antibiotics, the detection of this gene in a population
75 carrying ESBL-producing HiRC is of great concern because these MDROs can further acquire the
76 *mcr-1* and disseminate across different settings.

77 Regarding the univariate analysis, only the total CD4 cells count at the time of stool sampling
78 was significantly lower among the colonized than those non-colonized subjects (median 301 vs. 706
79 cells/ μ l; P=0.02; Table S2 and Figure S1). This observation may have important implications for
80 empirical therapy when HIV+ individuals have serious bacterial infections. However, given the
81 small sample size, our study lacks statistical power. This hinders meaningful multivariable analysis
82 and potentially other associations in the univariate analysis. Additionally, it is possible that our
83 results are representative only for a country (Switzerland) with low-prevalence of ESC-R *E. coli*
84 colonization in the general population alongside HIV+ people under successful ART. Finally, as
85 already reported in the general population, travelling to South-East Asia (P=0.07) and
86 hospitalization abroad (P=0.13) could also be potentially correlated with ESC-R-Ent colonization
87 [1, 2]. It is important to note that we were unable to include a control group due to the lack of
88 information about HIV status and CD4 cell count of healthy people.

89 This is the first study analyzing the presence of MDR *Enterobacteriaceae* in the intestinal tract
90 of HIV+ people. When under ART, these subjects seem to acquire such pathogens as likely as the
91 general population. However, the identification of HiRC underlines the potential for developing
92 future difficult-to-treat extra-intestinal infections. Moreover, the identification of a strain carrying
93 *mcr-1* on a plasmid with high similarity with others present in food animals and environment
94 highlights potential sources of acquisition of these pathogens. Finally, a low total CD4 count might
95 be an additional risk factor favoring intestinal colonization. Elucidating this phenomenon with a
96 larger cohort will be crucial for a better management of HIV+ patients.

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108 **COMPETING INTERESTS**

109 None declared

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111 **ETHICAL APPROVAL**

112 The study was approved by Kantonale Ethikkommission Bern (KEK): *Schweizerische HIV*
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114 **REFERENCES**

115 [1] Bernasconi OJ, Kuenzli E, Pires J, Tinguely R, Carattoli A, Hatz C, et al. Travelers Can Import
116 Colistin-Resistant Enterobacteriaceae, Including Those Possessing the Plasmid-Mediated *mcr-1*
117 Gene. *Antimicrob Agents Chemother.* 2016;60:5080-4.

118 [2] Pires J, Kuenzli E, Kasraian S, Tinguely R, Furrer H, Hilty M, et al. Polyclonal Intestinal
119 Colonization with Extended-Spectrum Cephalosporin-Resistant *Enterobacteriaceae* upon Traveling
120 to India. *Frontiers in Microbiology.* 2016;7.

121 [3] Zilberman-Schapira G, Zmora N, Itav S, Bashiardes S, Elinav H, Elinav E. The gut microbiome
122 in human immunodeficiency virus infection. *BMC Medicine.* 2016;14:1-11.

123 [4] Padmavathy K, Padma K, Rajasekaran S. Extended-spectrum β -lactamase/AmpC-producing
124 uropathogenic *Escherichia coli* from HIV patients: do they have a low virulence score? *Journal of*
125 *Medical Microbiology.* 2013;62:345-51.

126 [5] Marwa KJ, Mushi MF, Konje E, Alele PE, Kidola J, Mirambo MM. Resistance to
127 Cotrimoxazole and Other Antimicrobials among Isolates from HIV/AIDS and Non-HIV/AIDS
128 Patients at Bugando Medical Centre, Mwanza, Tanzania. *AIDS Research and Treatment.*
129 2015;2015:8.

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